

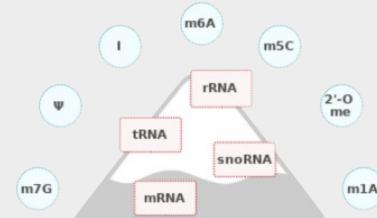
Signal level RNA modifications detection in eukaryotic ncRNAs

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The zoo of RNA post-transcriptional modifications

- Highly conserved feature found in archaea, bacteria and eukarya
- Impact on RNA structure and interaction properties
- More than 150 known RNA modifications



Meet Nanocompore



- Identifies signal-level differences between conditions (e.g. WT vs IVT/KD/KO)
- Based on Nanopolish resquigling
- Robust and flexible statistical framework
- Takes into account biological variability
- Allows for complex statistical designs (e.g. multi-factor designs, batch effects, etc.)

The Nanopore analysis workflow

nextflow

Automated data pre-processing workflow for multiple samples, including quality control.

Input dRNA-Seq data (fast5)

Control condition (low or no modifications)

- ↳ Replicate 1
- ↳ Replicate 2
- ↳ Replicate ...

Test condition (wild type)

- ↳ Replicate 1
- ↳ Replicate 2
- ↳ Replicate ...

Guppy
RNA basecalling



Minimap2

Transcriptome alignment
+
Samtools
reads filtering

Nanopolish
Events alignment
+
NanopolishComp
kmer level collapsing
and indexing

Basecalling
quality control



Alignment
quality control
(under development)

The Nanocompore analysis workflow

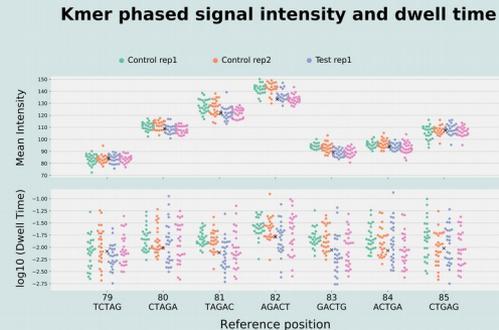
Compares 2 ONT dRNA sequencing datasets from different experimental conditions expected to have a significant impact on the epitranscriptome. Replicates aware framework with continuous integration testing.

Whitelist

Select reads mapped on transcripts with sufficient coverage

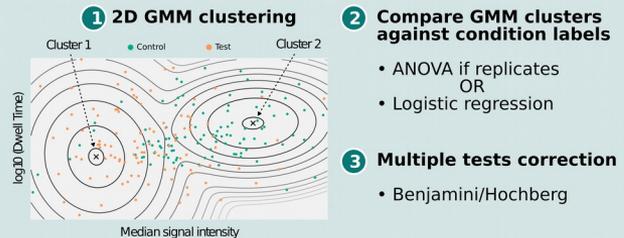
Position-wise data aggregation

Intensity and dwell time values are collected at transcript position level. The replicate structure is conserved



Statistical pairwise comparison

- Mann-Whitney, Kolmogorov-Smirnov or T test
- Gaussian mixture model clustering + Anova/logit
- Complex statistical design (multi-factor, batch effects ...)



4 Results saving and analysis

- Save data in lightweight GDBM database
- Extensive plotting API to explore data

Benchmarks on *in silico* generated data

A kmers model

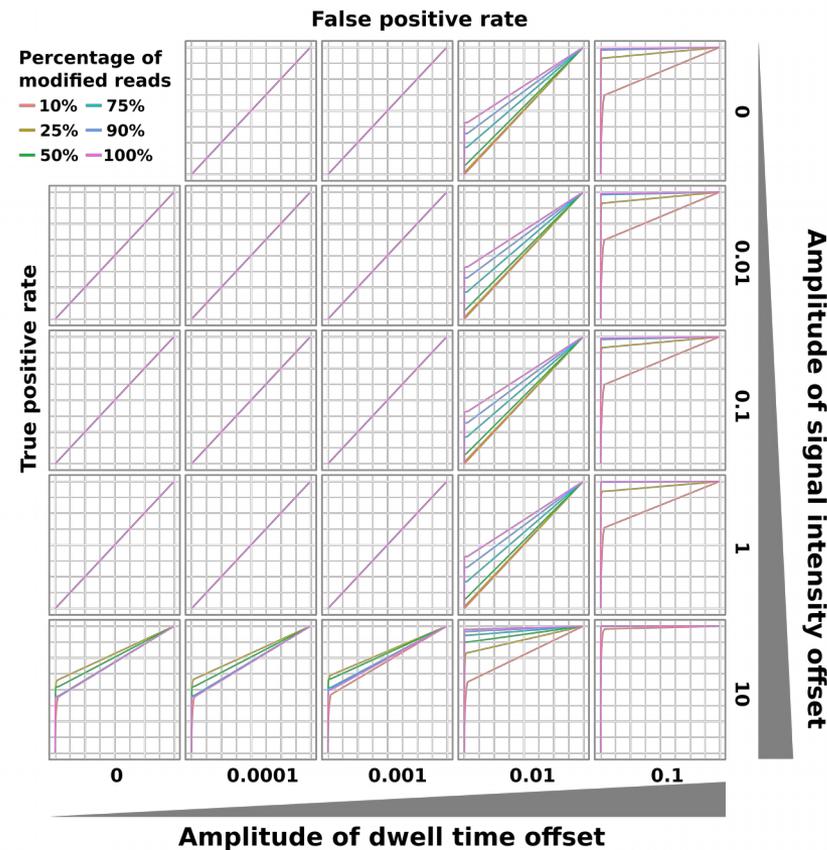
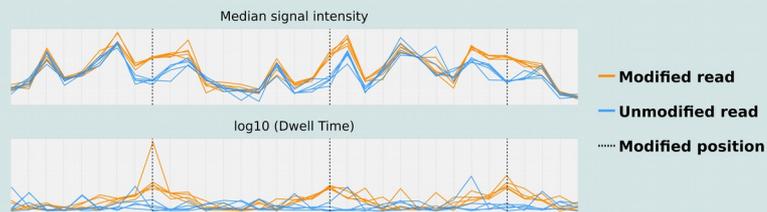
From an *in vitro* generated non-modified RNA dataset, find the best distribution to fit each kmers for both median intensities (*Logistic*) and dwell time (*Wald*)

B Artificial reference

Set of 2000 semi-random references 500 bases long. Covers all 5-mers (median: 970 times per kmer) and 99.67% of the possible 9-mers (median: 4 times per kmer)

C *In silico* modifications simulation

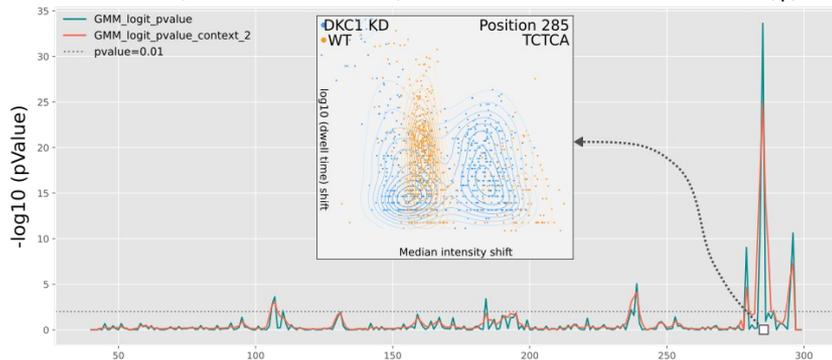
100 reads per reference using the probability density functions of the kmer models. Simulation of RNA modification effect on signal by offsetting the model density function.



Use-case 1: RNA modifications in targeted human ncRNAs

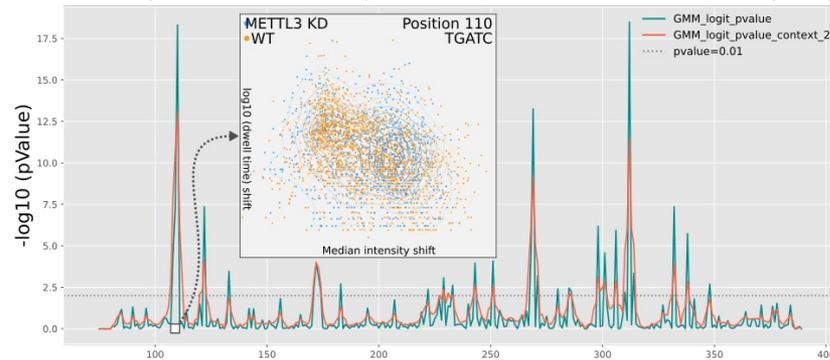
Pseudouridine

RMRP (ENST00000602361) / MOLM-13 cells WT vs DK1 KD (ψ)

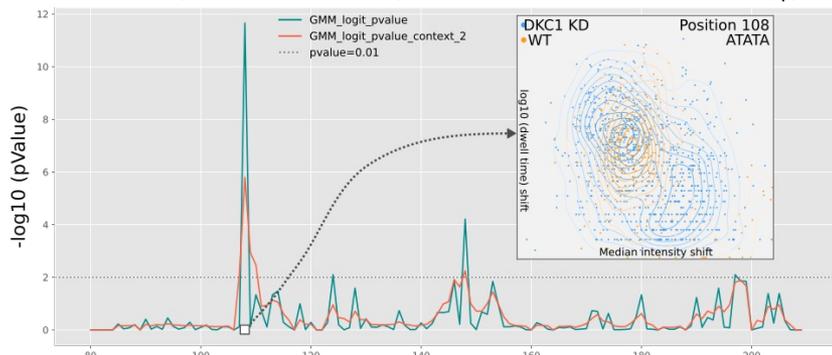


N6-methyl-adenosine

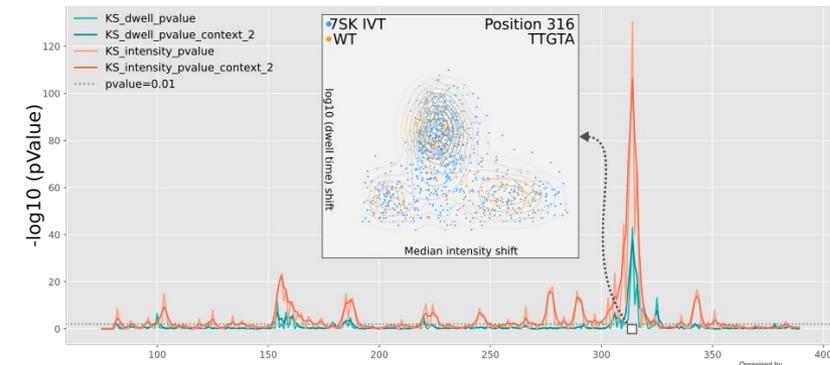
RN7SK (ENST00000636484) / MOLM-13 cells WT vs METTL3 KD (m6A)



RNU2-1 (ENST00000618664) / MOLM-13 cells WT vs DK1 KD (ψ)

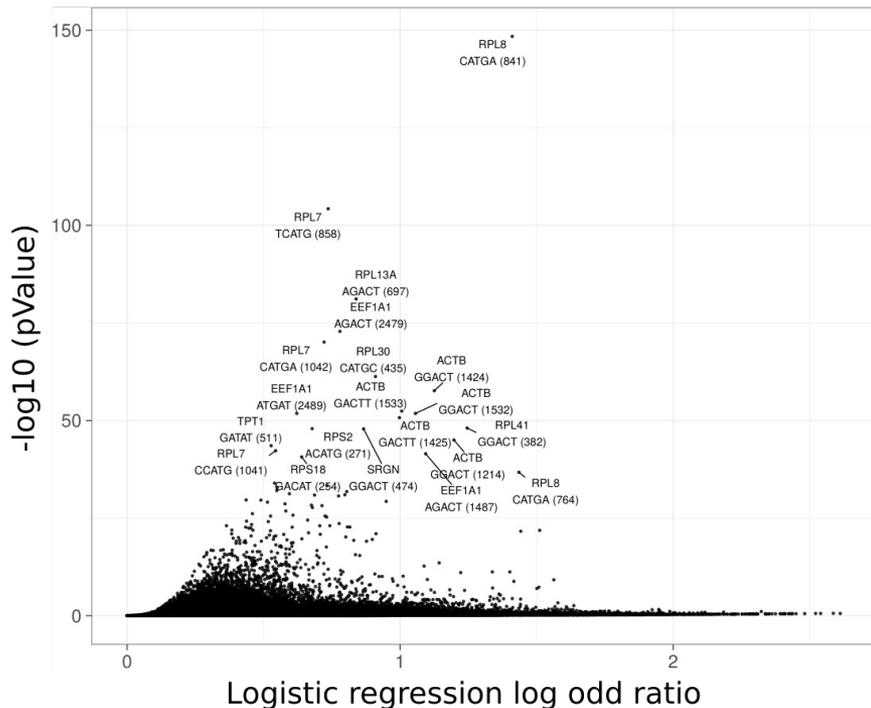


RN7SK (ENST00000636484) / MOLM-13 cells WT vs IVT 7SK (all mods)



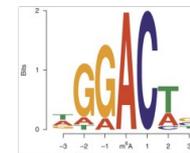
Use-case 2: m6A in a human polyA transcriptome

Top candidate kmer sites found by Nanocompare



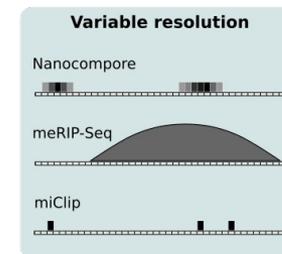
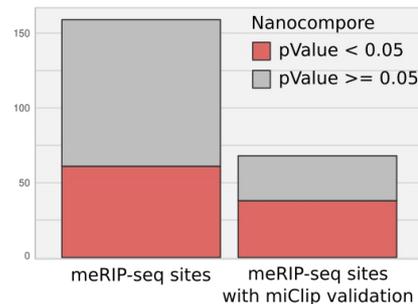
De novo motif enrichment analysis in top 100 hits

Rank	Motif	P-value	log P-pvalue	% of Targets
1		1e-2	-5.971e+00	22.45%
2		1e0	-1.172e-01	6.12%



Dominissini et al.
Nat Prot vol 8, (2013)

Orthogonal METTL3-dependent m6A mapping methods



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