

# Supplementary Materials

## Novel Stemphol Derivatives from a Marine Fungus *Pleospora* sp.

Jinzhong Xu<sup>1</sup>, Peng Liu<sup>1</sup>, Xinyang Li<sup>1</sup>, Li-she Gan<sup>2</sup>, Pinmei Wang<sup>1\*</sup>

<sup>1</sup>Institute of Marine Biology, Ocean College, Zhejiang University, Zhoushan, 310026, China

<sup>2</sup>College of Pharmaceutical Science, Zhejiang University, Hangzhou, 310058, China

Four novel stemphol derivatives, pleosporols A-D (**1**, **2** and mixture of **3** and **4**) together with known compounds stemfolones (mixture of **5** and **6**), stemphol (**7**) were isolated from a marine fungus *Pleospora* sp. (PO4) derived from the gut of marine isopod *Ligia oceanica*. The planar structures of novel compounds were elucidated on the basis of mass and NMR spectral analysis and their stereochemistry was determined by CD spectra, NOESY data, coupling constants analysis and modified Mosher's method. Novel compounds contained  $\alpha,\beta$ -unsaturated cyclohexanone ring and possibly derived from the oxidation of stemphol. All novel ones showed strong antimicrobial activity against *S. epidermidis* CMCC26069 with MIC values less than 10  $\mu\text{g/mL}$ .

**Keywords:** Pleosporols; *Pleospora* sp.; Dialkylresorcinols; Antibacterial

---

\*Corresponding authors.  
E-mail address: [wangpinmei@zju.edu.cn](mailto:wangpinmei@zju.edu.cn)

## **1.General experimental procedures**

Optical rotation was measured on a Jasco P-1010 polarimeter. ECD spectra were obtained on an Applied Photophysics Chirascan spectrometer. The ultraviolet (UV) absorption spectra were measured in MeOH on a METASH UV-8000 spectrophotometer. IR spectra were recorded on a Bruker VECTOR 22 FT-IR spectrometer. NMR spectra were recorded in CDCl<sub>3</sub> (ALDRICH, St. Louis, MO, USA) or MeOH-d<sub>4</sub> (ALDRICH, St. Louis, MO, USA) with tetramethylsilane (TMS) as an internal standard, using a Bruker AVANCE-III 600 MHz NMR spectrometer. HR-ESIMS data were obtained on an Agilent 6224 TOF LC/MS, MS/MS data was collected by a TripleTOF® 5600+ LC/MS/MS system (AB SCIEX). Column chromatography (CC) was performed with silica (100-200 and 200-300 mesh, Qingdao Haiyang Chemical Co., Ltd., Qingdao, China).

## **2.Fungal material and fermentation**

The fungal colony (PO4) was isolated from the gut of marine isopod *Ligia oceanica* which was collected at Dongsha of Dinghai in Zhoushan, Zhejiang Province of China in May 2013. The fungal PO4 was determined as *Pleospora* sp. by 26S rDNA analysis.

26s rDNAD1/D2 sequence of PO4

CTCAAATTGAAATCTGGCTTTAGGGTCCGAGTTGTAATTGCAGA  
GGCGCTTGGCTTGGCAGCGGTCCAAGTCCTGGAACAGGACGTCAC  
AGAGGGTGAGAATCCCGTACGTGGTCGCTAGCTATTGCCGTAAAGCCC  
CTTCGACGAGTCGAGTTGGATGCAGCTCTAAATGGGAGGTAAAT  
TTCTTCTAAAGCTAAATATTGCCAGAGACCGATAGCGCACAGTAGAGT  
GATCGAAAGATGAAAAGCACTTGGAAAGAGAGTCAAACAGCACGTGAA  
ATTGTTGAAAGGAAGCGCTTGCAGCCAGACTTGCTGCAGTTGCTCATC  
CGGGCTTGGCCGGTGCACCTCTGTAGGCAGGCCAGCATCAGTTGGG  
CGGTGGATAAAGGTCTGTCACGTACCTCTTCGGGGAGGCCTTATA  
GGGGAGACGACATACCACCAAGCCTAGACTGAGGTCCGCGCATCTGCTAGG

ATGCTGGCGTAATGGCTGTAAGCGGC

The strain (PO4) was static cultured in rice cultural medium (rice 40g, artificial sea water 40 mL) at 28°C for 30 days in 500-mL Erlenmeyer-flasks (100 flasks).

### 3.Extraction and isolation

The cultured medium was extracted with EtOAc and evaporated under vacuum to recycle EtOAc. The extract was dissolved in MeOH and partitioned with petroleum ether to remove the lipidic substance. Finally 12.95g of secondary metabolites (SM) extract was obtained from EtOAc layer. All of SM extract was fractionated by silica gel column chromatography (CC) eluted in gradient from petroleum ether - EtOAc (10:1-2:1) to CH<sub>2</sub>Cl<sub>2</sub>-MeOH (50:1-1:1) to yield 19 fractions (Frs. 1-19) based on TLC analysis. Frs. 10 and 11 were subjected to ODS MPLC (Medium Pressure Liquid Chromatography) and eluted by MeOH/H<sub>2</sub>O (50%~100%) and the subfractions further purified by semi-preparative HPLC (COSMOSIL PACKED COLUMN, 5C18-MS-II column, 10ID×250mm, 3 mL/min). Compounds **1** (50% MeOH/H<sub>2</sub>O, t<sub>R</sub> = 14 min, 19.7 mg) and **2** (80% MeOH/H<sub>2</sub>O, t<sub>R</sub>=8 min, 11.0 mg) were isolated from different subfractions of Fr.10 while Compounds **3/4** (t<sub>R</sub>=10 min, 10.8 mg) and **5/6** (t<sub>R</sub>=11 min, 8.9 mg) were successively obtained from Fr.11-4 eluted by 55% MeOH/H<sub>2</sub>O. Compound **7** (t<sub>R</sub> = 10 min 13.0 mg) was isolated from Fr.18 through semi-preparative ODS-HPLC eluted by 20% MeOH/H<sub>2</sub>O.

### Physicochemical and spectral data

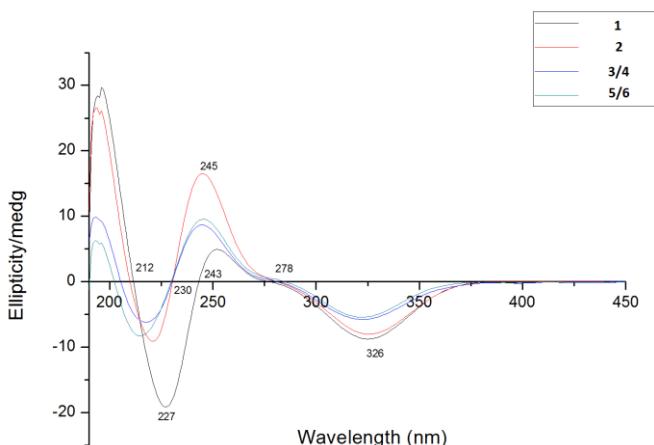
*Pleosporol A (1)*: colorless oil;  $[\alpha]^{25}_D +11.15$  (c=2.75, MeOH); UV (MeOH)  $\lambda_{max}(\log\epsilon)$  205 (3.28), 234 (3.75) nm; CD (c 1.03×10<sup>-4</sup>mol/L, MeOH),  $\lambda_{max}$  ( $\Delta\epsilon$ ) 325 (-8.76), 252 (4.93), 227 (-19.17); IR (KBr)  $\nu_{max}$  (cm<sup>-1</sup>): 3421, 2956, 2931, 2867, 1668, 1378, 1211, 1138, 1073, 1030, 985; HRESIMS *m/z* 269.1779 [M-H]<sup>-</sup> (Calcd. for C<sub>15</sub>H<sub>25</sub>O<sub>4</sub>, 269.1753); NMR data in the Tables 1 and 2.

*Pleosporol B (2)*: colorless oil;  $[\alpha]^{25}_D +3.27$  (c=3.40, MeOH); UV (MeOH)  $\lambda_{max}(\log\epsilon)$  204 (3.43), 233 (3.98) nm; CD (c 1.46×10<sup>-4</sup>mol/L, MeOH),  $\lambda_{max}$  ( $\Delta\epsilon$ ) 325 (-8.00), 245 (16.49), 221 (-9.10); IR (KBr)  $\nu_{max}$  (cm<sup>-1</sup>): 3421, 2956, 2932, 2867, 1670, 1378, 1210, 1138, 1072, 1032; HRESIMS *m/z* 269.1782 [M-H]<sup>-</sup> (Calcd. for C<sub>15</sub>H<sub>25</sub>O<sub>4</sub>,

269.1753); NMR data in the Tables 1 and 2.

*Pleosporols C/D (3/4)*: colorless oil;  $[\alpha]^{25}_D +2.96$  ( $c=2.96$ , MeOH); UV (MeOH)  $\lambda_{\max}(\log\epsilon)$  205 (3.16), 234 (3.79) nm; CD ( $c\ 2.68\times10^{-4}$  mol/L, MeOH),  $\lambda_{\max}$  ( $\Delta\epsilon$ ) 323 (-5.78), 245 (8.80), 218 (-6.10); IR (KBr)  $\nu_{\max}$  ( $\text{cm}^{-1}$ ): 3419, 2957, 2931, 2869, 1666, 1374, 1210, 1116, 1072, 1032; HRESIMS  $m/z$  269.1781 [M-H]<sup>-</sup> (Calcd. for C<sub>15</sub>H<sub>25</sub>O<sub>4</sub>, 269.1753); NMR data in the Tables 1 and 2.

Stemfolone B and 14-*epi*-stemfolone B(**5/6**): colorless oil;  $[\alpha]^{25}_D +4.32$  ( $c=1.20$ , MeOH); UV (MeOH)  $\lambda_{\max}(\log\epsilon)$  205 (3.29), 234 (3.79) nm; CD ( $c\ 5.44\times10^{-4}$  mol/L, MeOH),  $\lambda_{\max}$  ( $\Delta\epsilon$ ) 322 (-5.44), 245 (9.56), 215 (-8.30); IR (KBr)  $\nu_{\max}$  ( $\text{cm}^{-1}$ ): 3421, 2957, 2930, 2871, 1666, 1374, 1210, 1116, 1072, 1032; HRESIMS  $m/z$  269.1781 [M-H]<sup>-</sup> (Calcd. for C<sub>15</sub>H<sub>25</sub>O<sub>4</sub>, 269.1753); NMR data in the Tables 1 and 2.

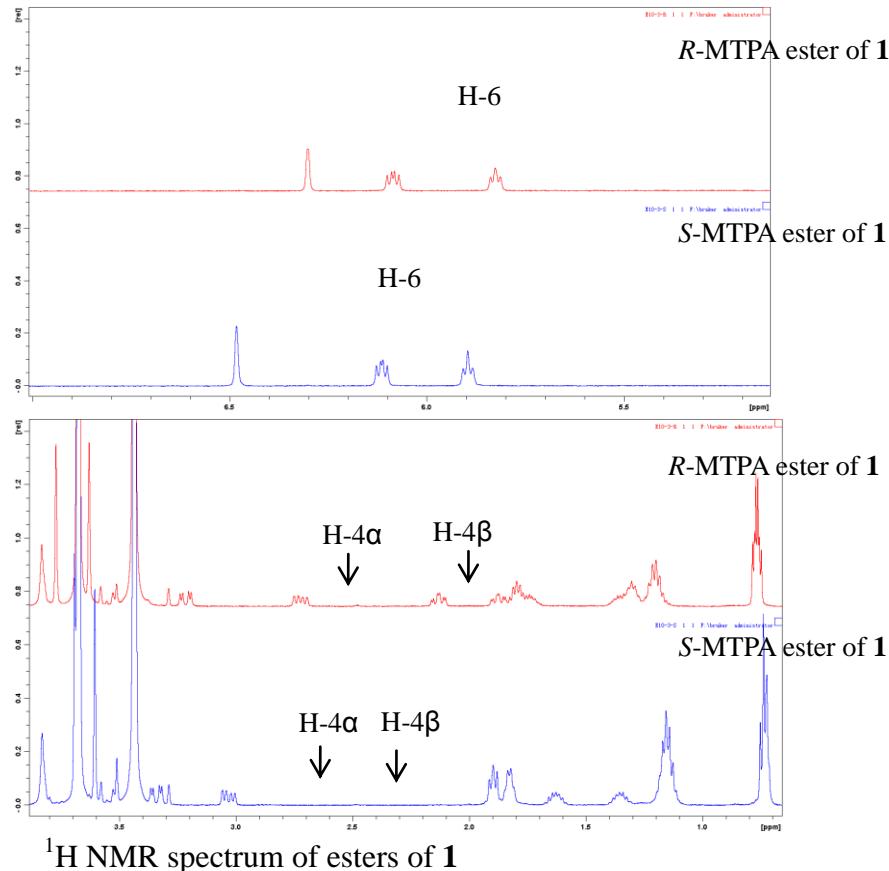


Measured CD spectra for **1-6**

#### 4. MTPA Esterification of **1**

Compound **1** was transferred into two clean NMR tubes (0.5mg in each tube) and dried completely under vacuum. Deuterated pyridine (400 $\mu$ L) was added to dissolve the sample and (*R*)-MTPA-Cl/(*S*)-MTPA-Cl (8 $\mu$ L) were quickly added into the tubes, respectively. All contents were mixed thoroughly by shaking the tubes carefully. The

reaction was performed at room temperature for 2.5h to obtained the *S*-MTPA and the *R*-MTPA ester (**1a** and **1b**), respectively<sup>1</sup>. The chemical shift differences ( $\Delta\delta = \delta_S - \delta_R$ ) calculated from the <sup>1</sup>H NMR spectra of the two diastereomeric esters **1a** and **1b** enabled us to determine the absolute configuration of C-3 as *R*.



## 5. Biological assay

Antibacterial activities of novel compounds were tested against Methicillin-resistant *S. aureus* ATCC43300, *P. Aeruginosa* ATCC27853, *E. coli* ATCC35218 and *S. epidermidis* CMCC26069 using broth dilution methods to determine the minimal inhibitory concentration. These bacteria were inoculated into nutrient broth medium in 96-well microtiter plate in the presence of serial dilution concentrations of compounds. After incubation for 24h, the MIC value was read. Vancomycin hydrochloride (0.5 $\mu$ g/ml) and aerosporin (0.5 $\mu$ g/ml) were used as positive controls against gram-positive and gram-negative bacteria, respectively.

## 6. The spectra for compounds 1-7

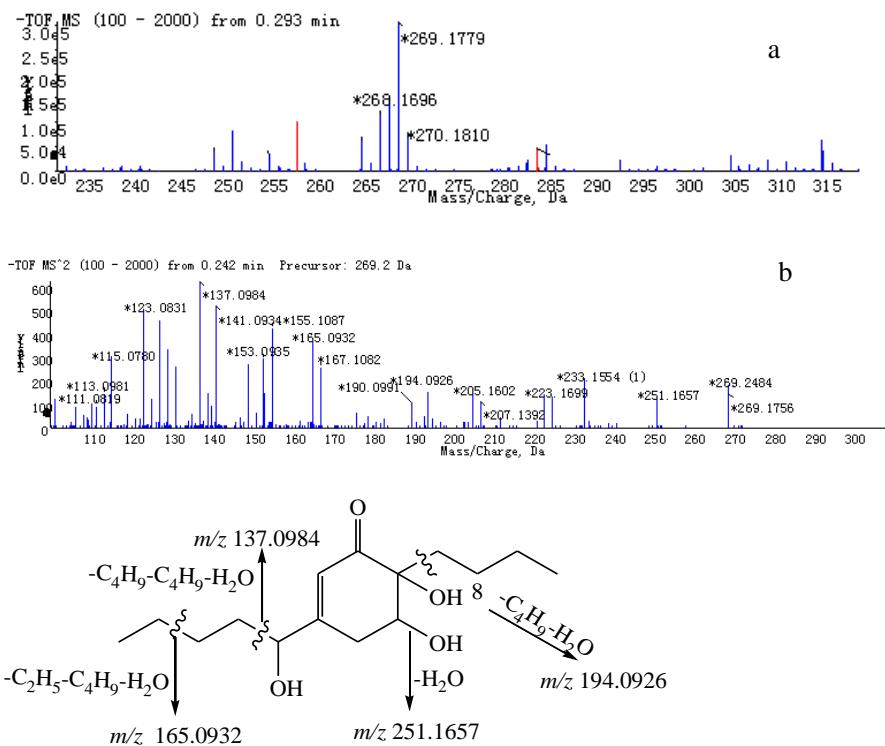
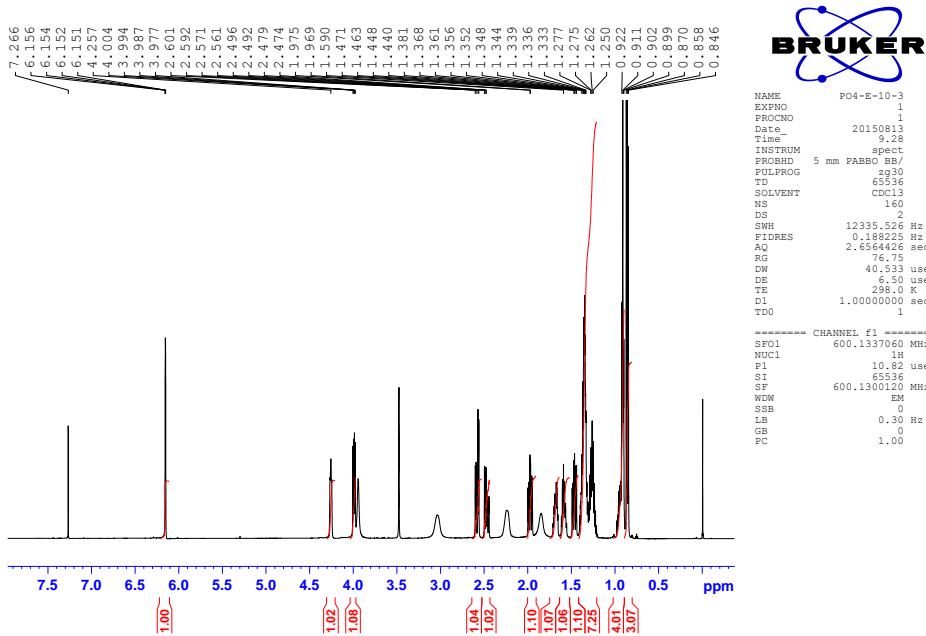
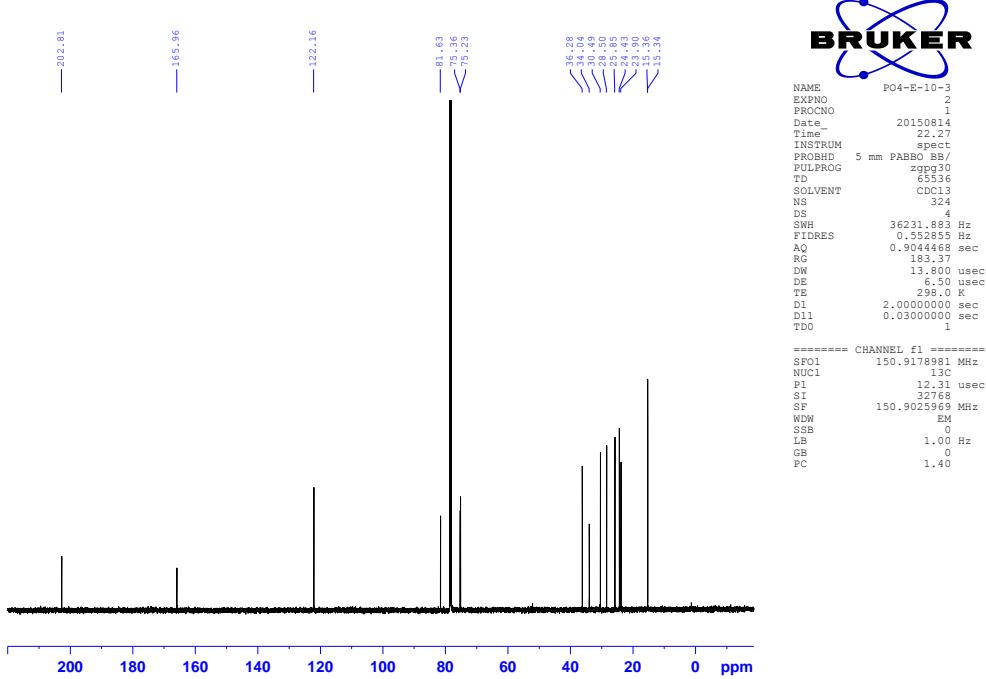
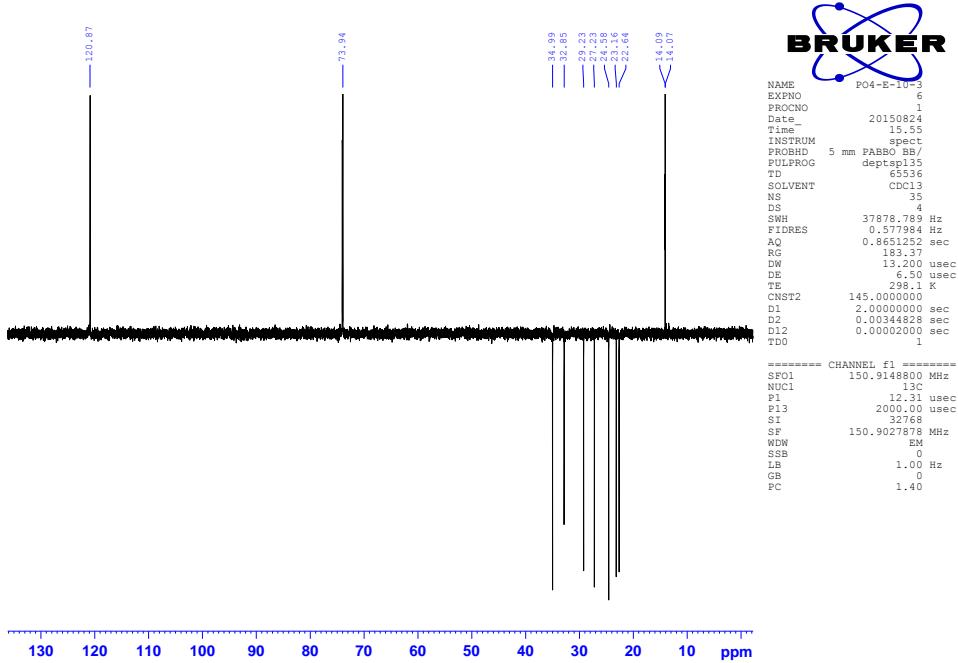


Figure S1 MS/MS fragment ion peaks of **1** (a: MS<sup>1</sup>, b: MS<sup>2</sup>)

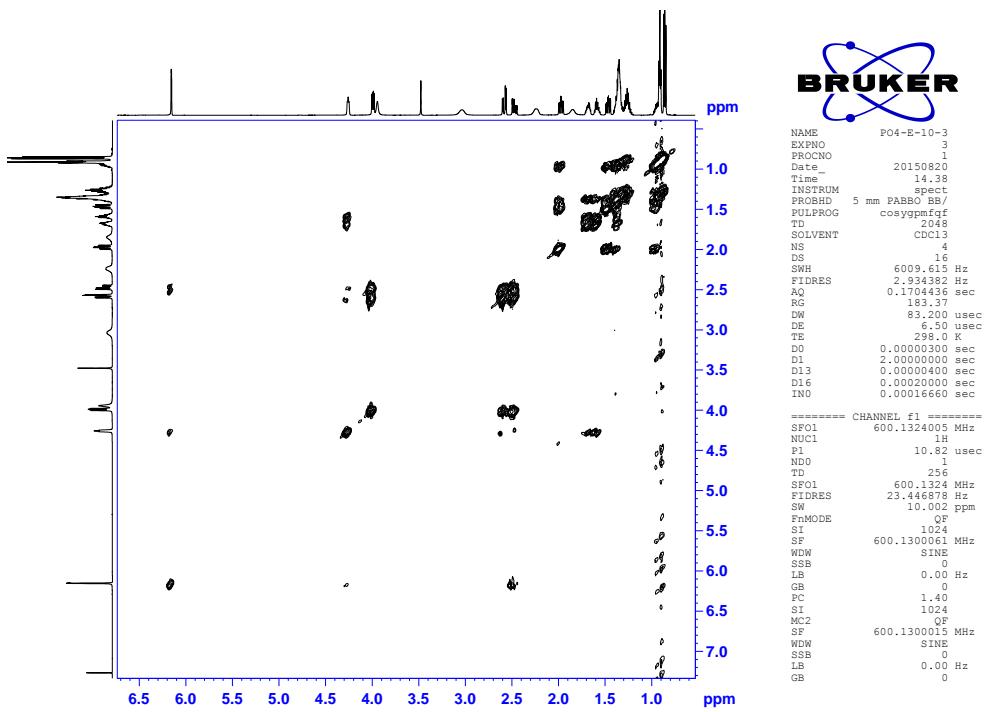




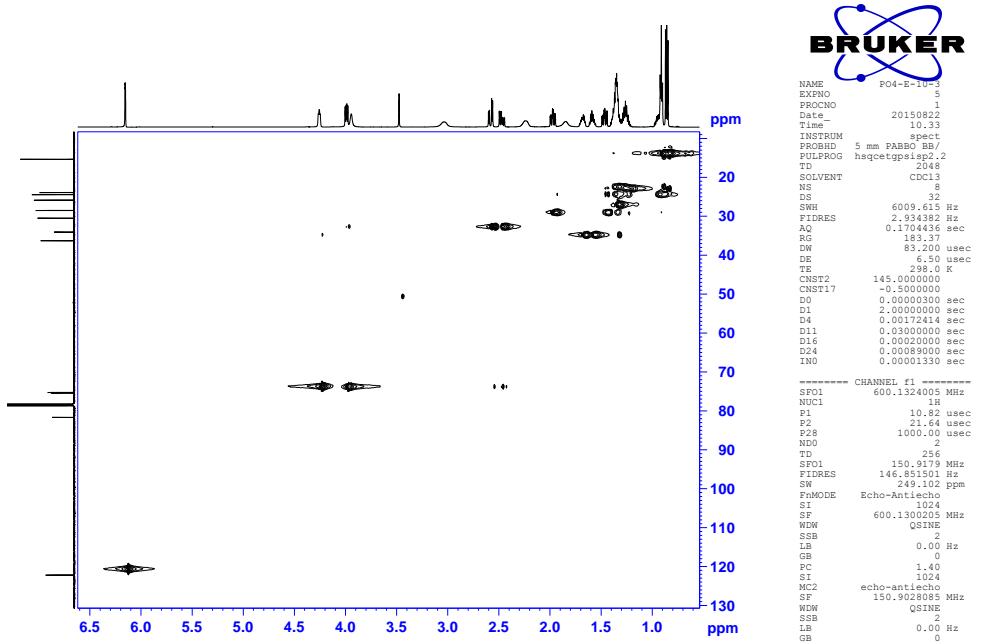
**Figure S3**  $^{13}\text{C}$  NMR spectrum of **1** in  $\text{CDCl}_3$



**Figure S4** DEPT spectrum of **1** in  $\text{CDCl}_3$



**Figure S5**  $^1\text{H}$ - $^1\text{H}$  COSY spectrum of **1** in  $\text{CDCl}_3$



**Figure S6** HMQC spectrum of **1** in  $\text{CDCl}_3$

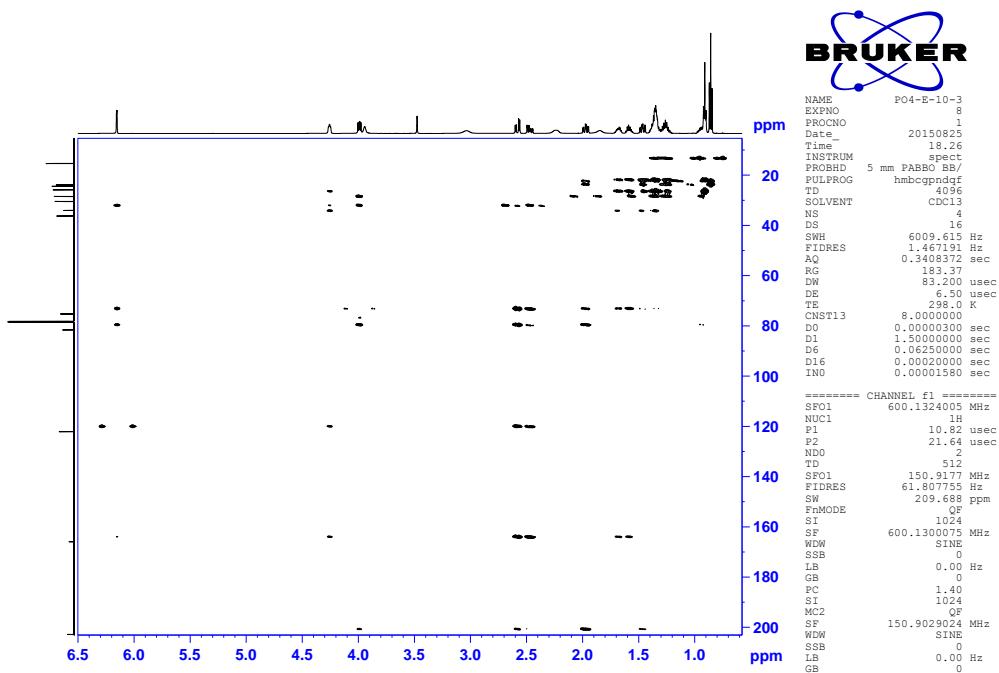


Figure S7 HMBC spectrum of **1** in  $\text{CDCl}_3$

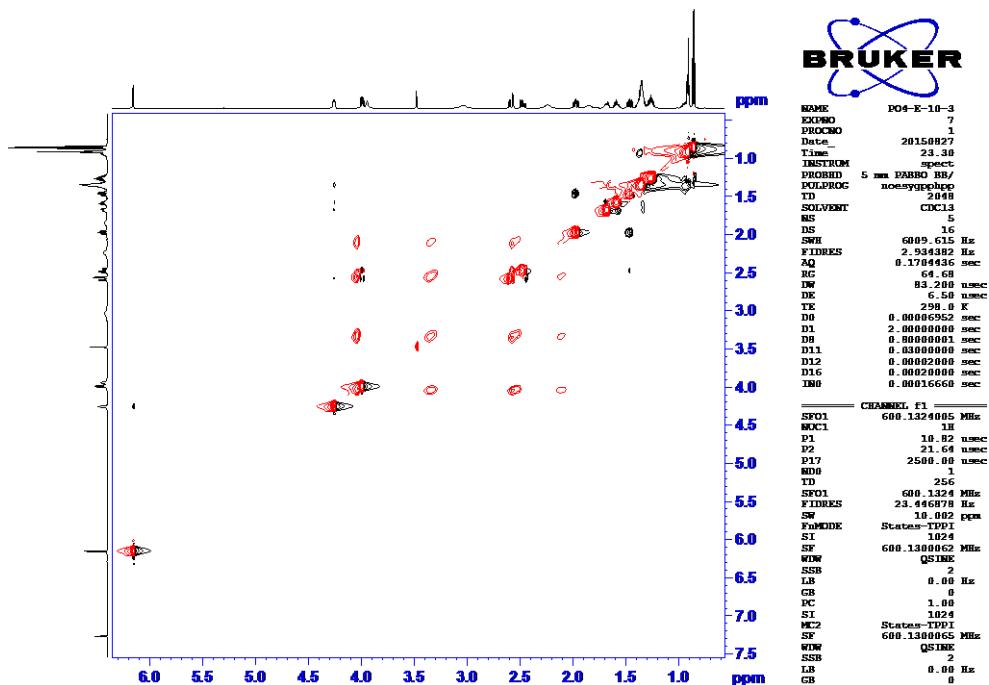
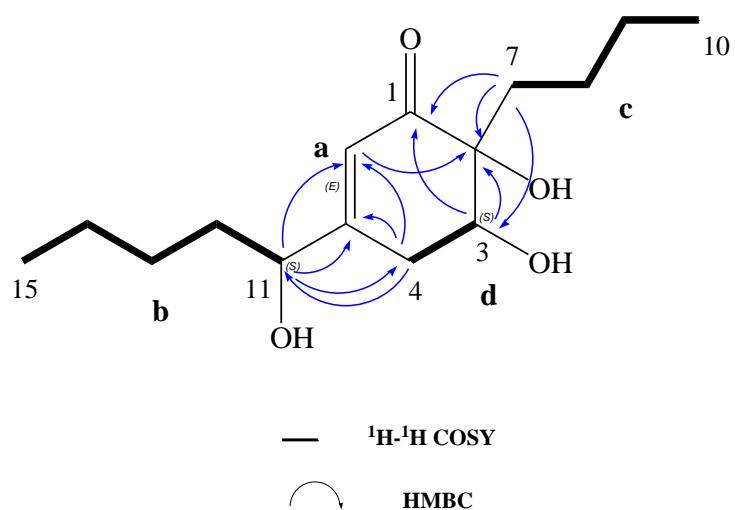
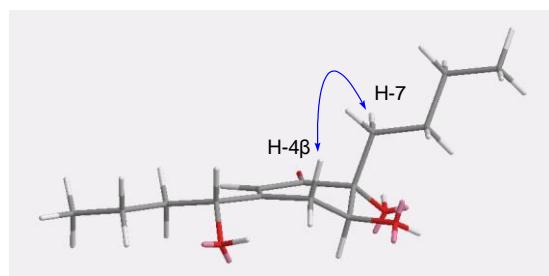


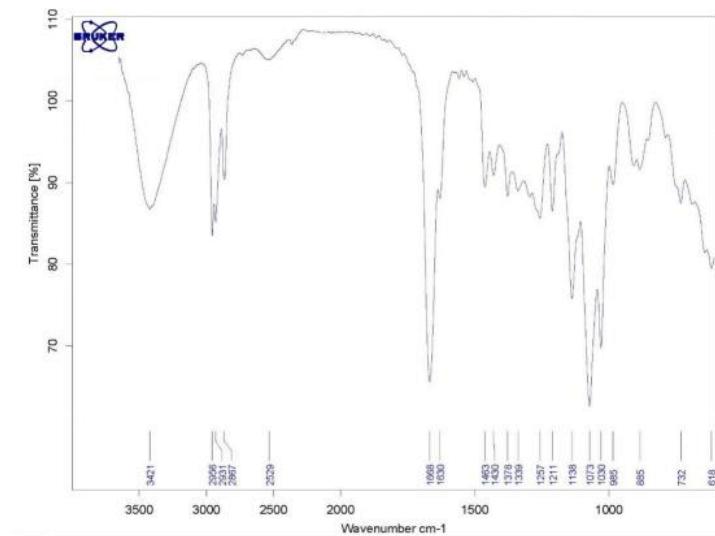
Figure S8 NOESY spectrum of **1** in  $\text{CDCl}_3$



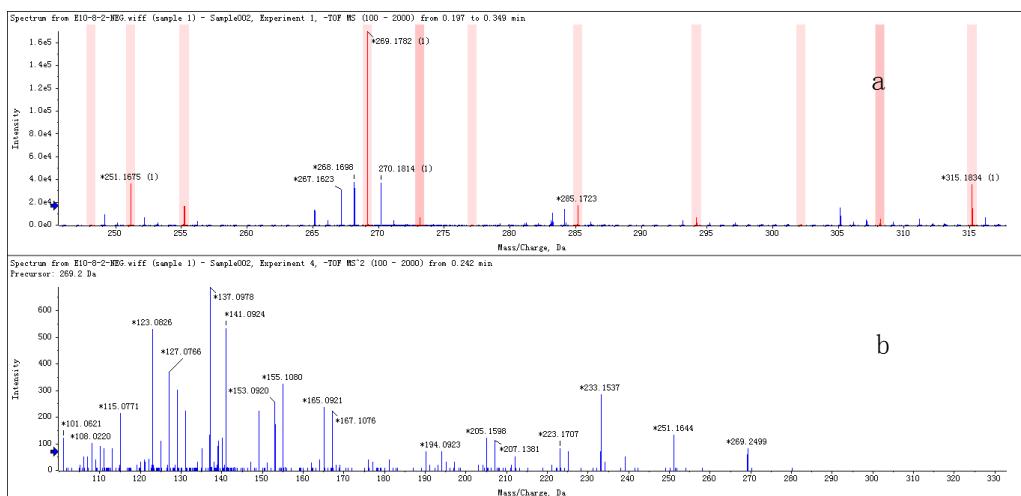
**Figure S9**  $^1\text{H}$ - $^1\text{H}$  COSY and key HMBC correlations for **1**



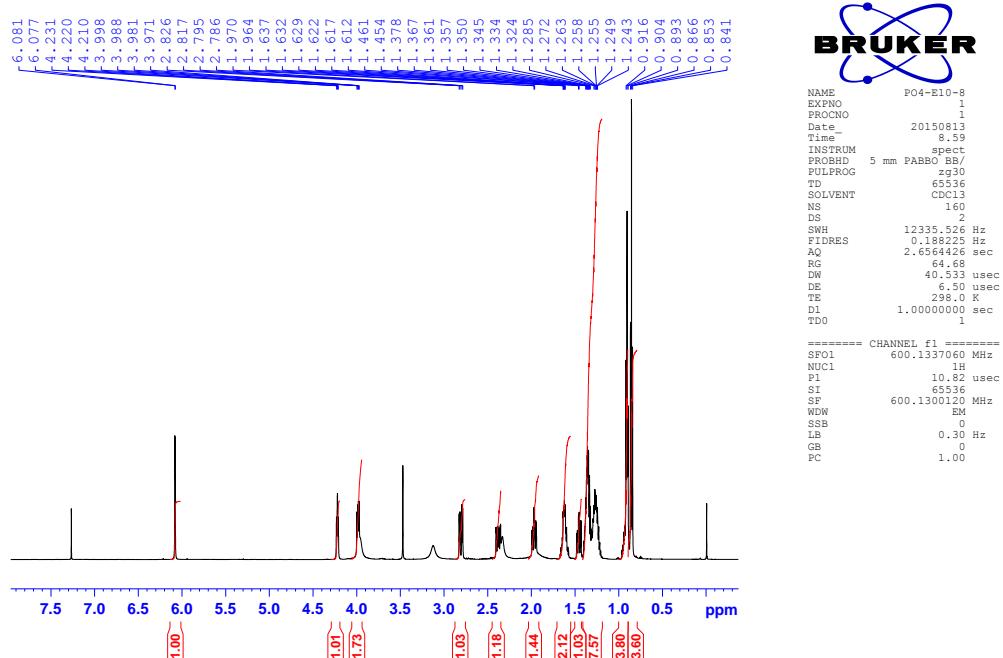
**Figure S10** key NOESY correlations for **1-6**



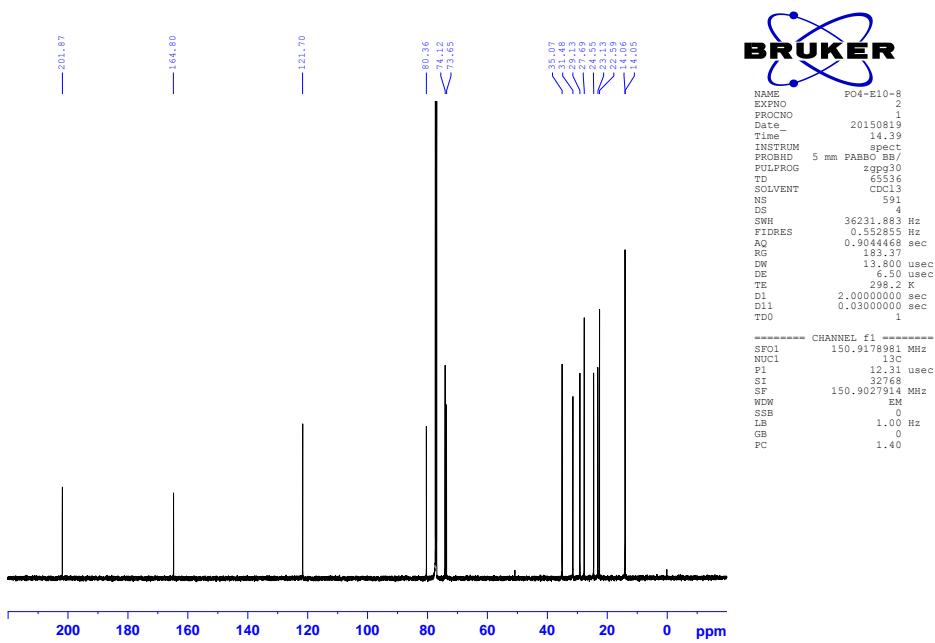
**Figure S11** IR spectrum of **1**



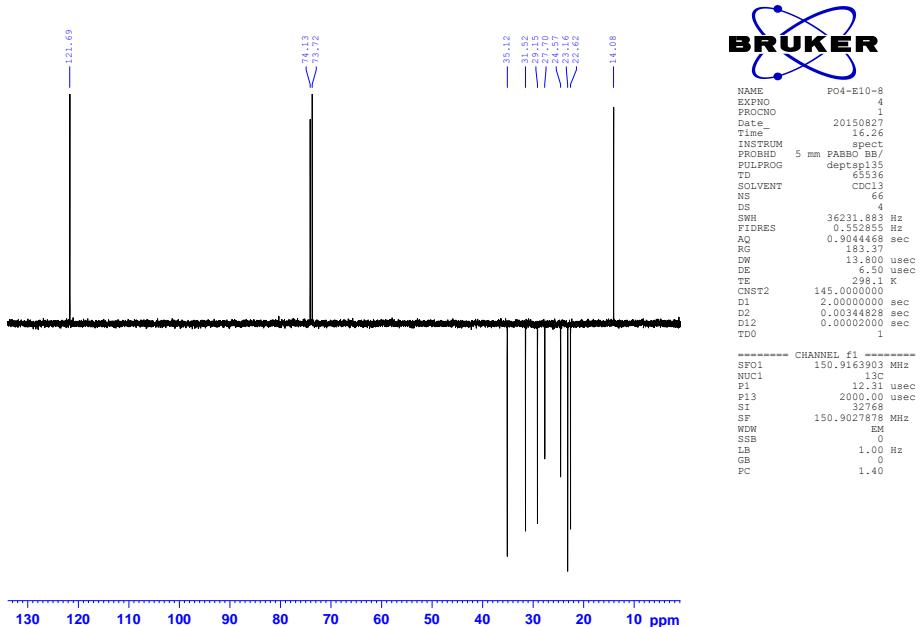
**Figure S12** MS/MS of **2** (a:  $\text{MS}^1$ , b:  $\text{MS}^2$ )



**Figure S13**  ${}^1\text{H}$  NMR spectrum of **2** in  $\text{CDCl}_3$



**Figure S14**  $^{13}\text{C}$  NMR spectrum of **2** in  $\text{CDCl}_3$



**Figure S15** DEPT spectrum of **2** in  $\text{CDCl}_3$

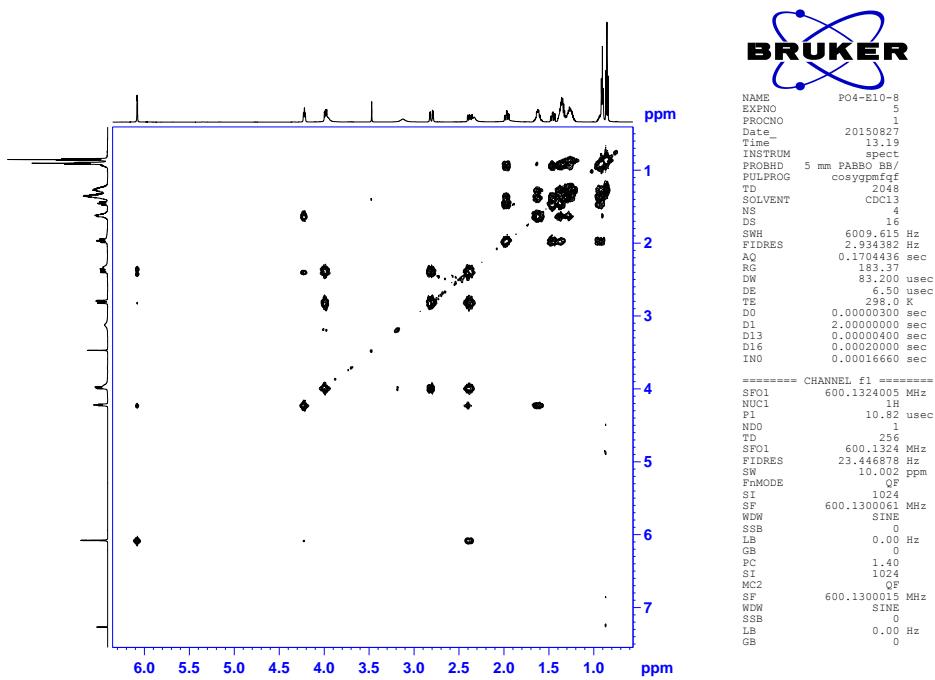


Figure S16  $^1\text{H}$ - $^1\text{H}$  COSY spectrum of **2** in  $\text{CDCl}_3$

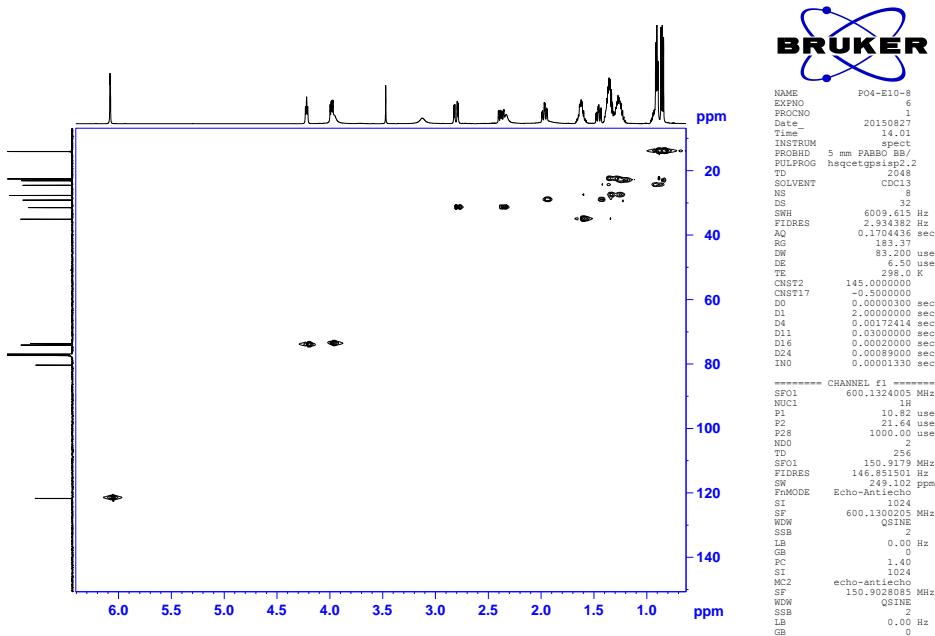
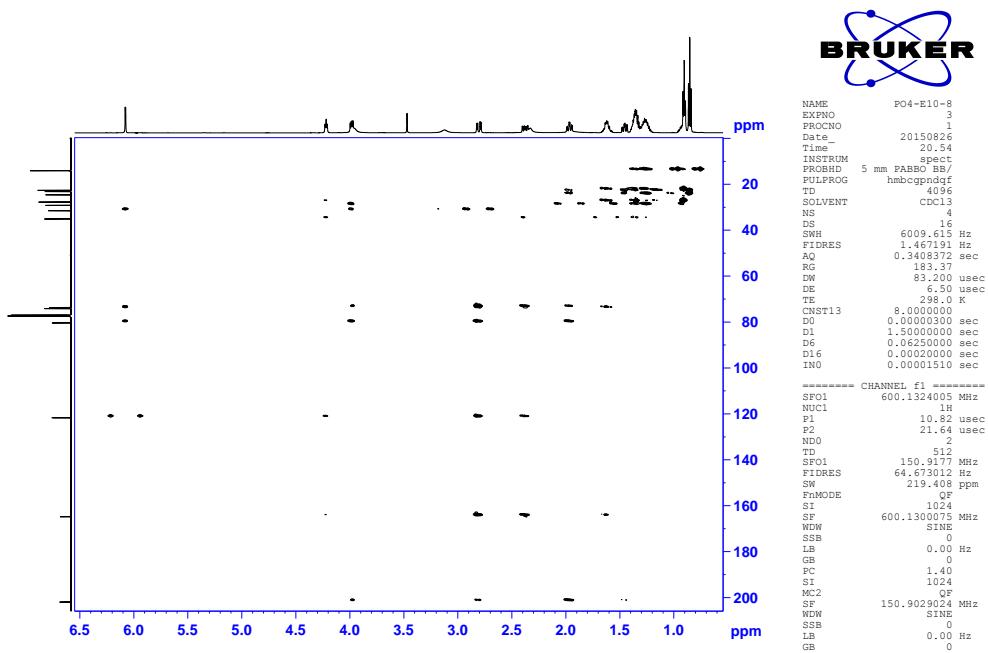
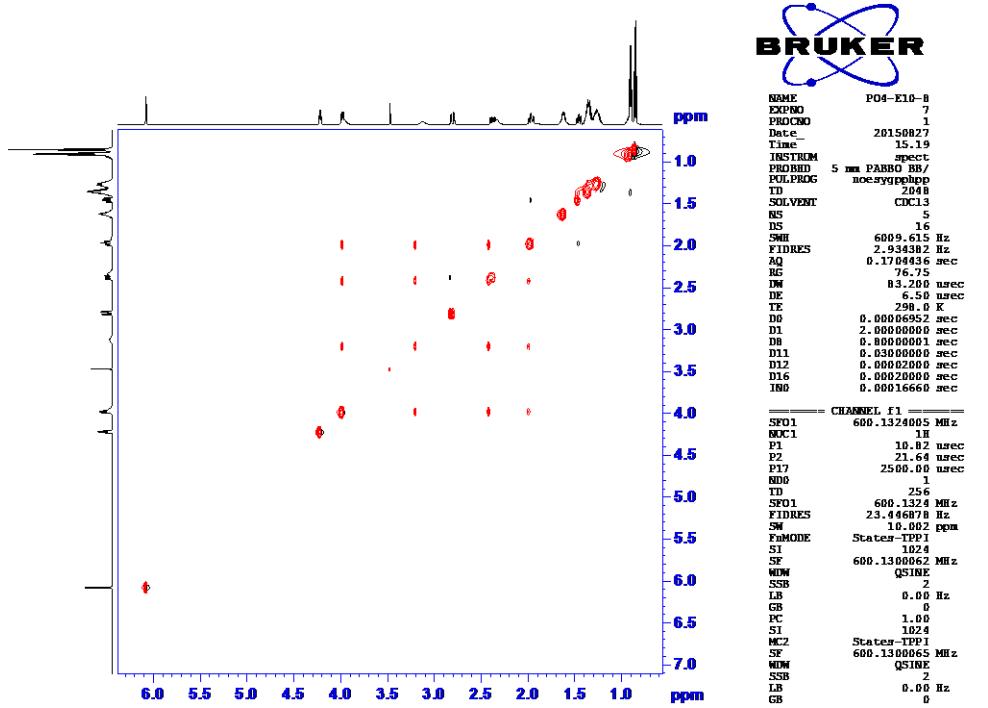


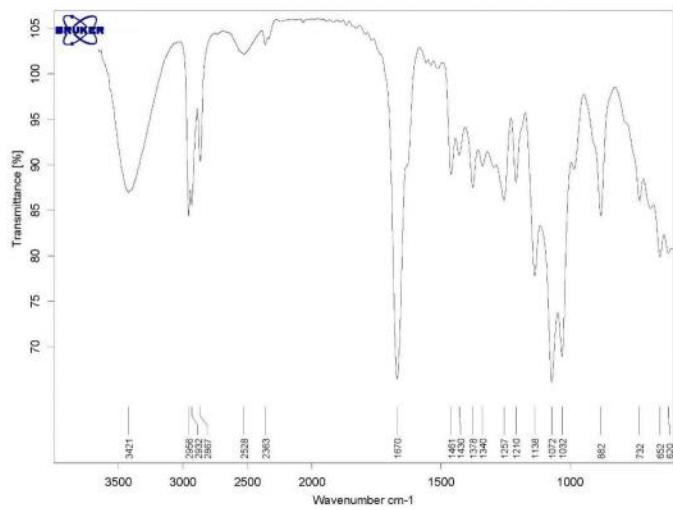
Figure S17 HMQC spectrum of **2** in  $\text{CDCl}_3$



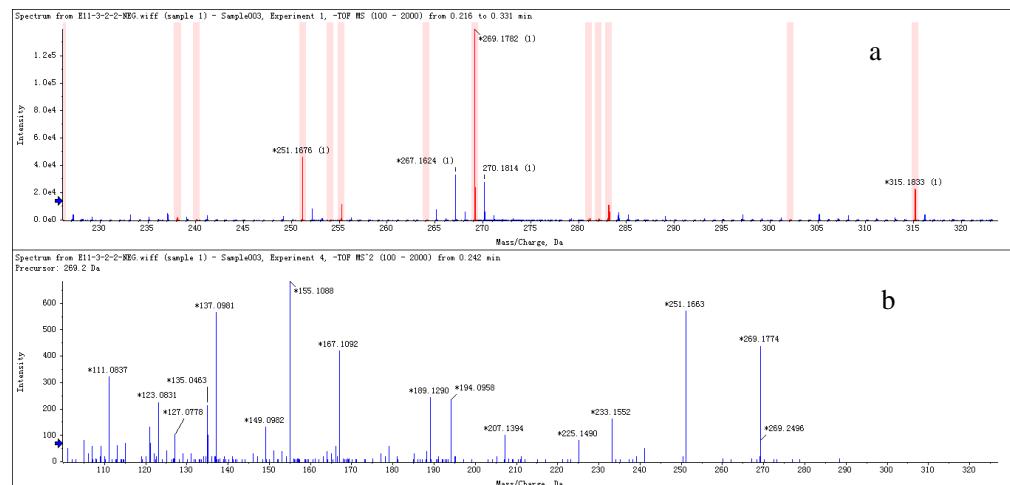
**Figure S18** HMBC spectrum of **2** in  $\text{CDCl}_3$



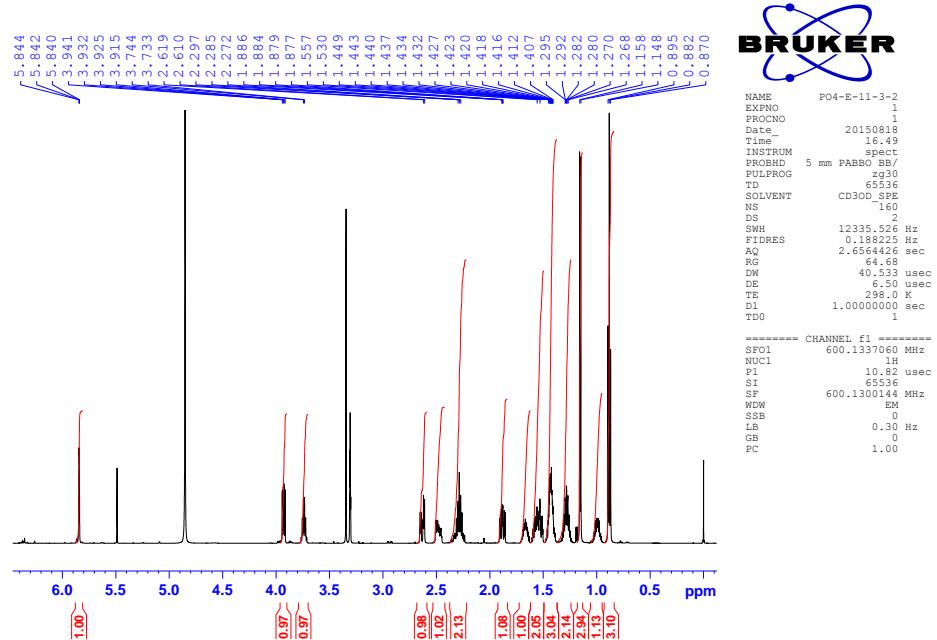
**Figure S19** NOESY spectrum of **2** in  $\text{CDCl}_3$



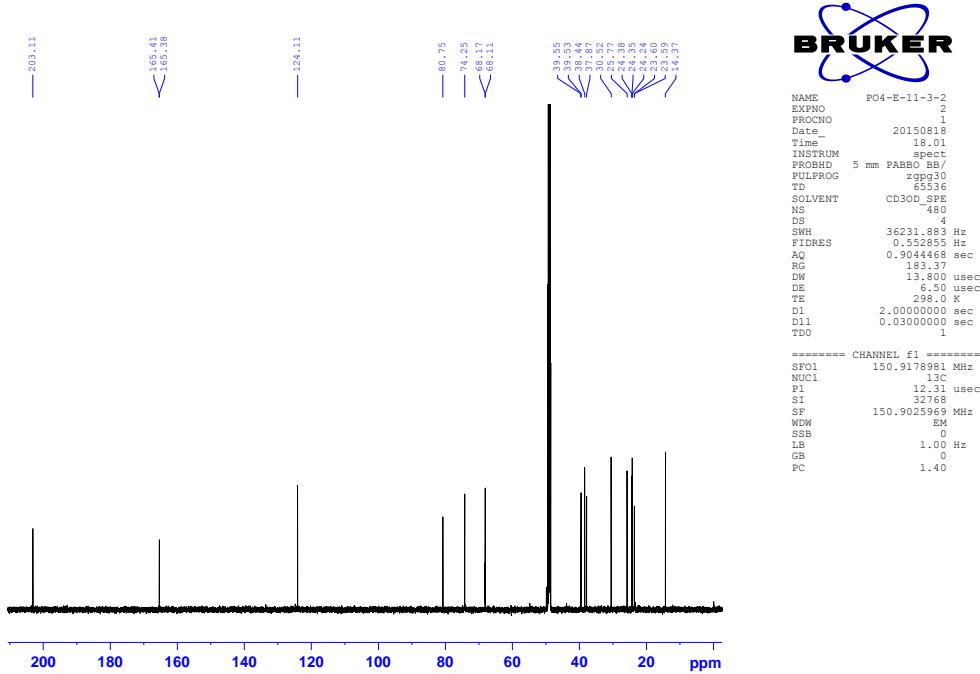
**Figure S20** IR spectrum of **2**



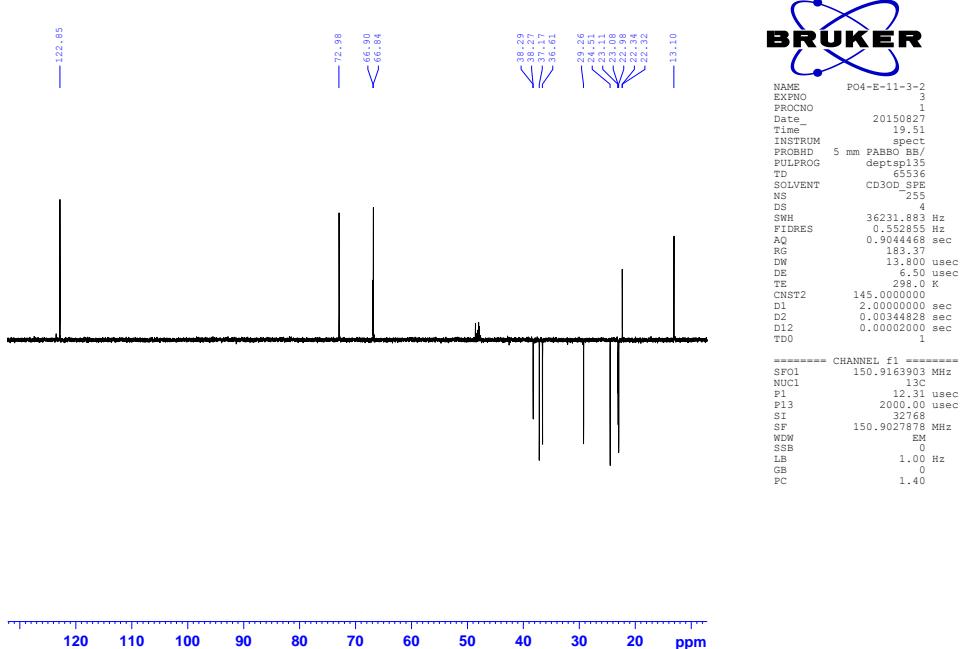
**Figure S21** MS/MS of 3/4 (a: MS<sup>1</sup>, b: MS<sup>2</sup>)



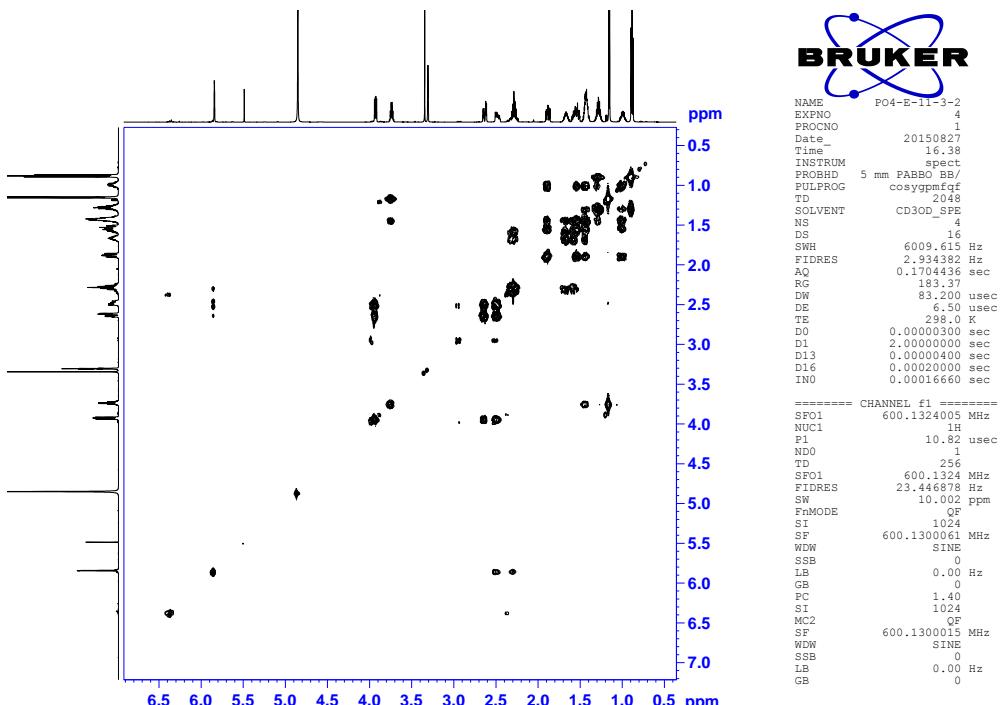
**Figure S22**  $^1\text{H}$  NMR spectrum of **3/4** in  $\text{CD}_3\text{OD}$



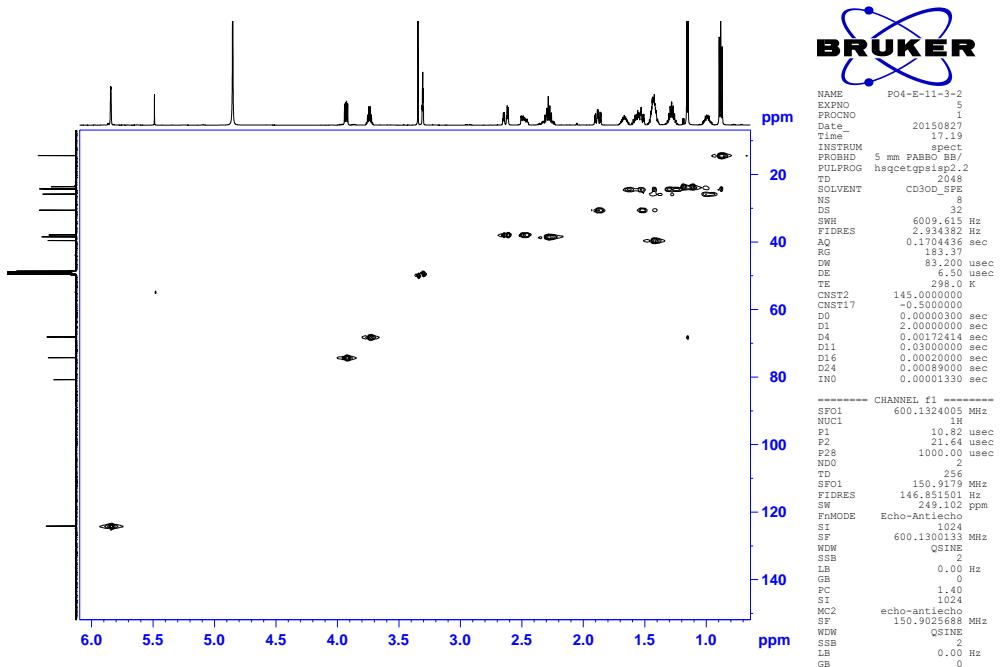
**Figure S23**  $^{13}\text{C}$  NMR spectrum of 3/4 in  $\text{CD}_3\text{OD}$



**Figure S24** DEPT spectrum of 3/4 in  $\text{CD}_3\text{OD}$



**Figure S25**  $^1\text{H}$ - $^1\text{H}$  COSY spectrum of **3/4** in  $\text{CD}_3\text{OD}$



**Figure S26** HMQC spectrum of **3/4** in  $\text{CD}_3\text{OD}$

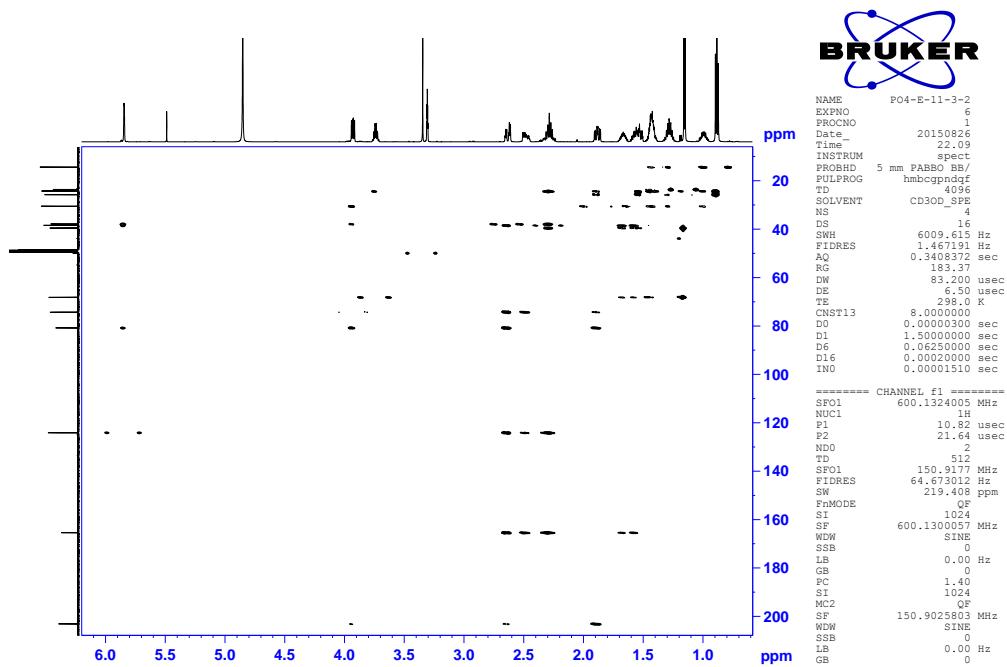


Figure S27 HMBC spectrum of 3/4 in  $\text{CD}_3\text{OD}$

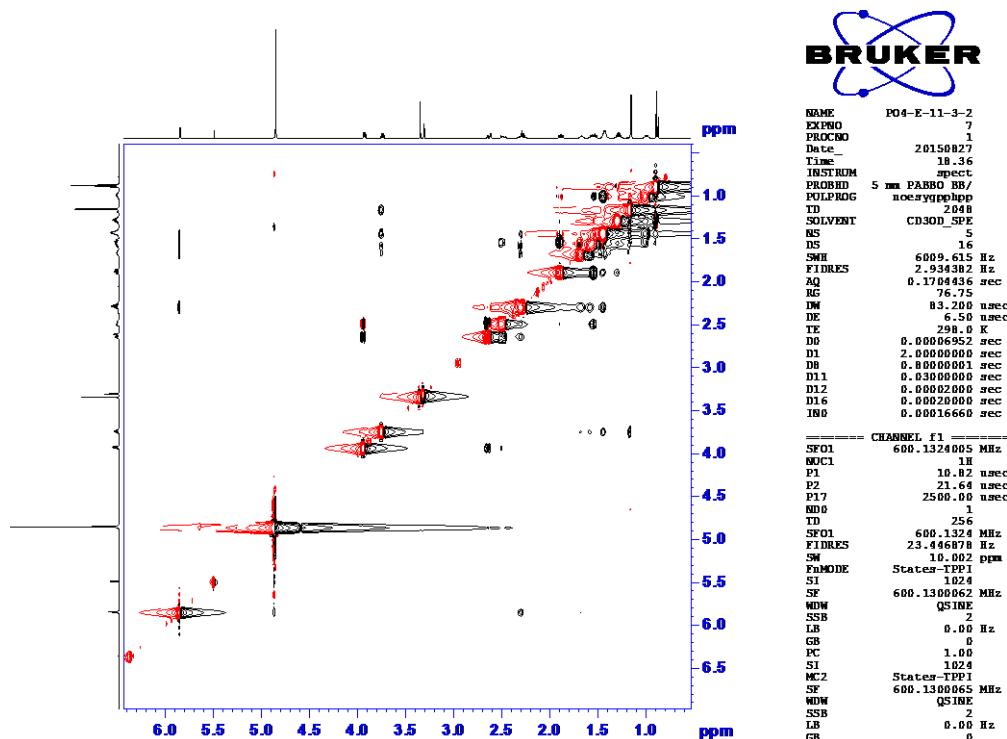
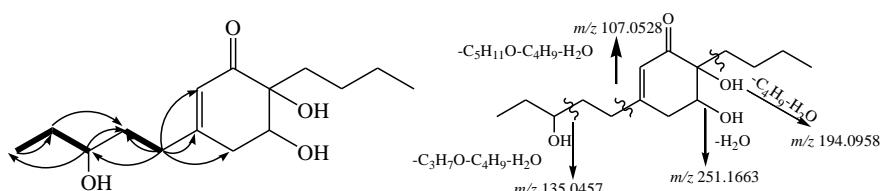
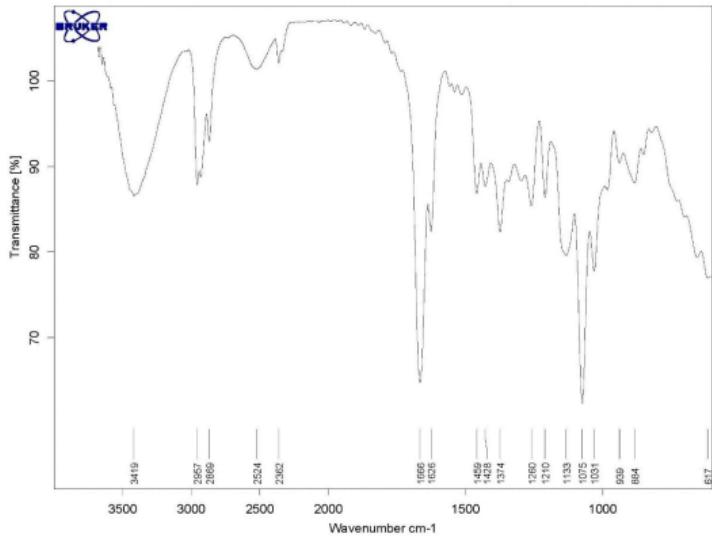


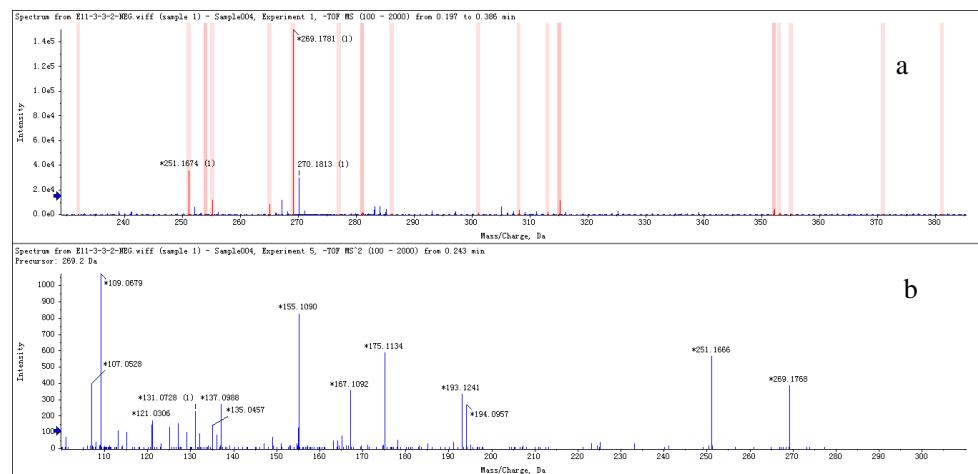
Figure S28 NOESY spectrum of 3/4 in  $\text{CD}_3\text{OD}$



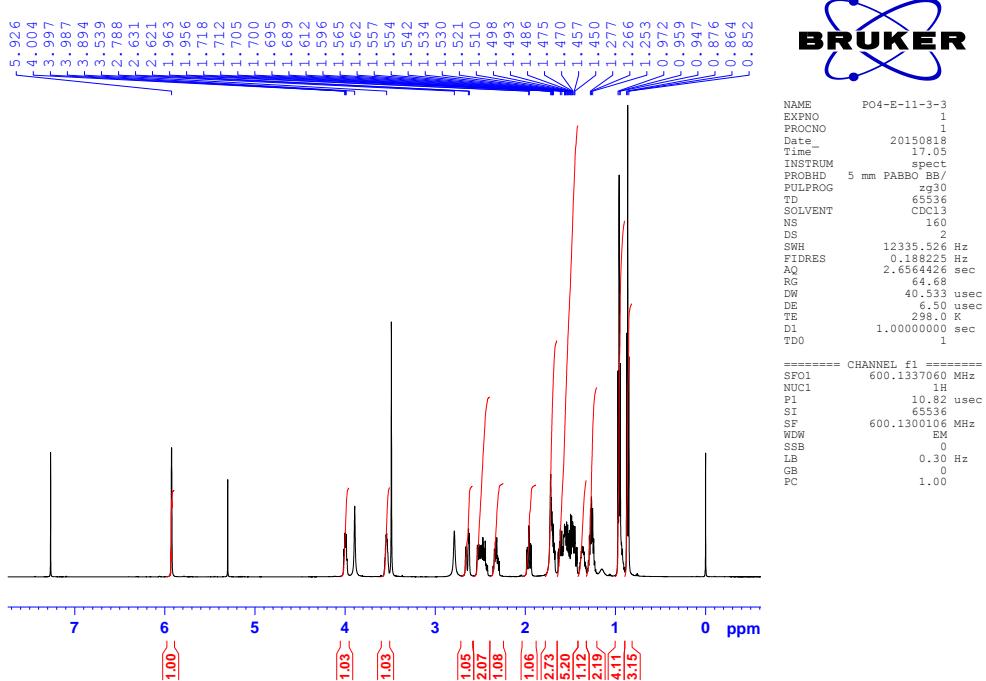
**Figure S29** Key  $^1\text{H}$ - $^1\text{H}$  COSY and HMBC correlations and MS/MS fragment ion peaks for **3/4**



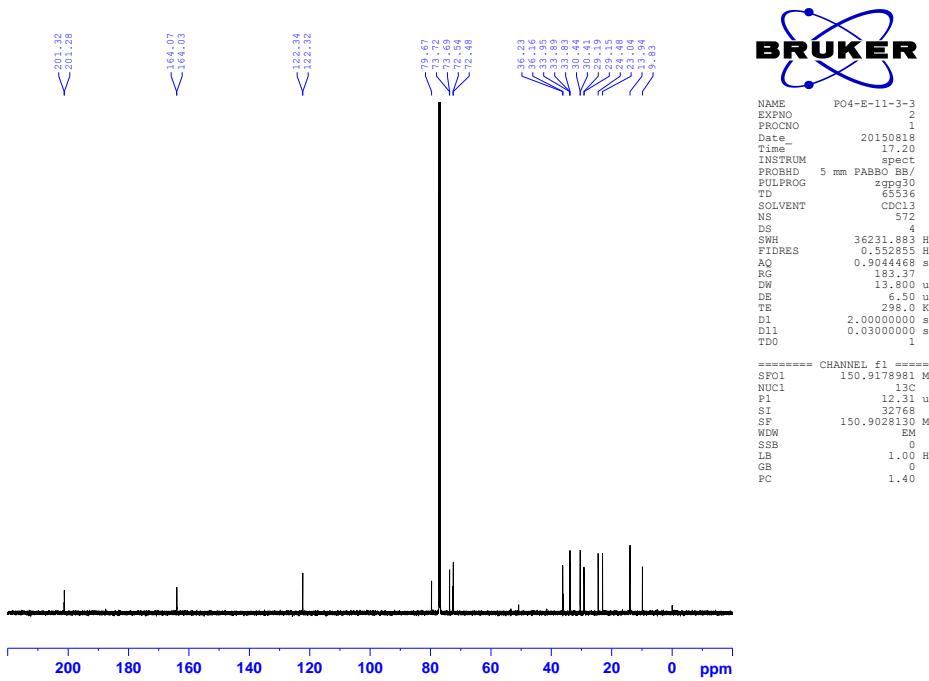
**Figure S30** IR spectrum of **3/4**



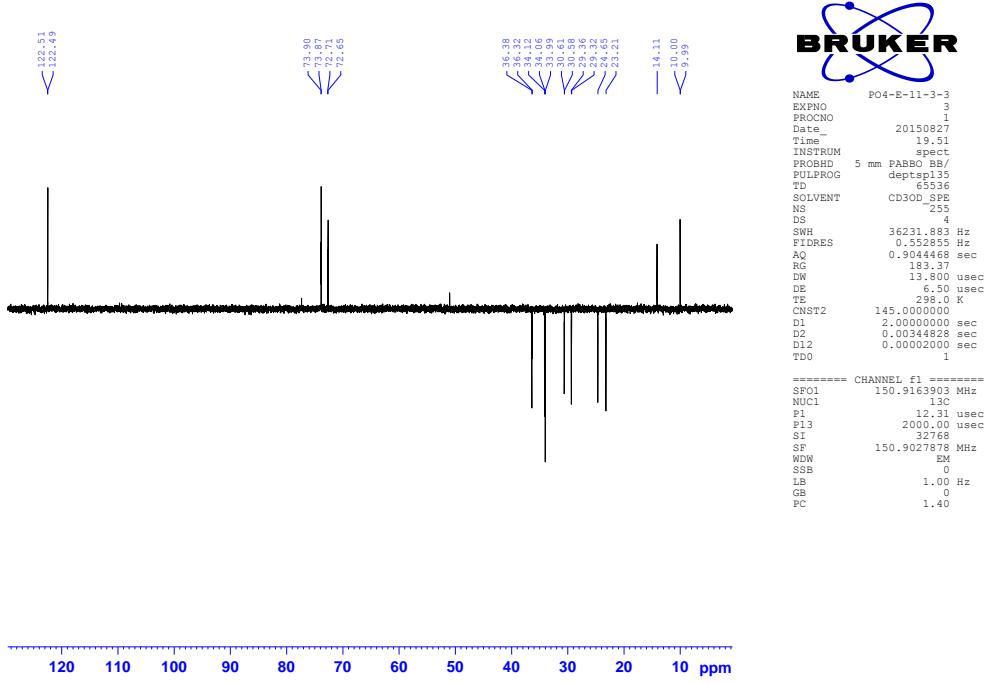
**Figure S31** MS/MS of **5/6** (a: MS<sup>1</sup>, b: MS<sup>2</sup>)



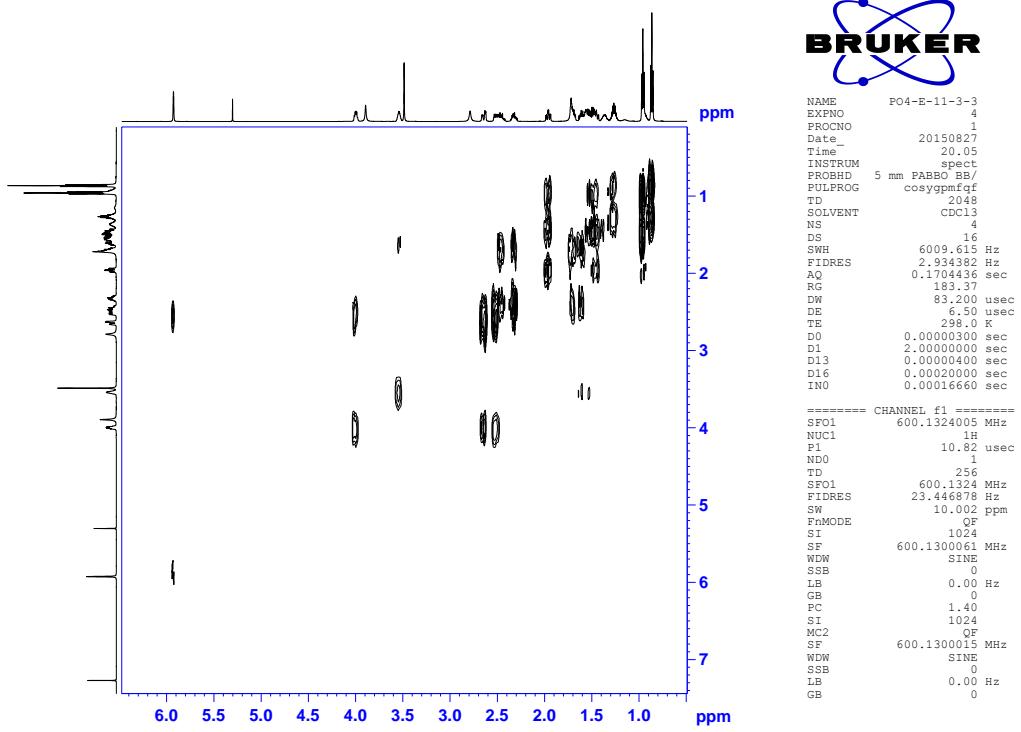
**Figure S32**  $^1\text{H}$  NMR spectrum of **5/6** in  $\text{CDCl}_3$



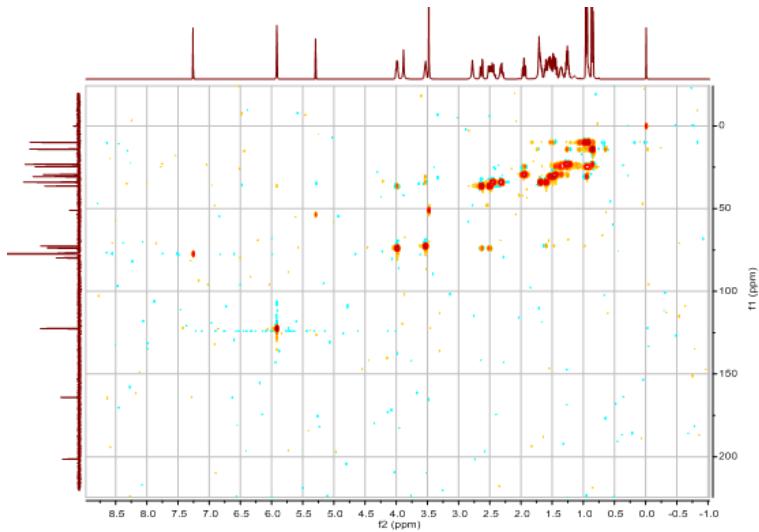
**Figure S33**  $^{13}\text{C}$  NMR spectrum of **5/6** in  $\text{CDCl}_3$



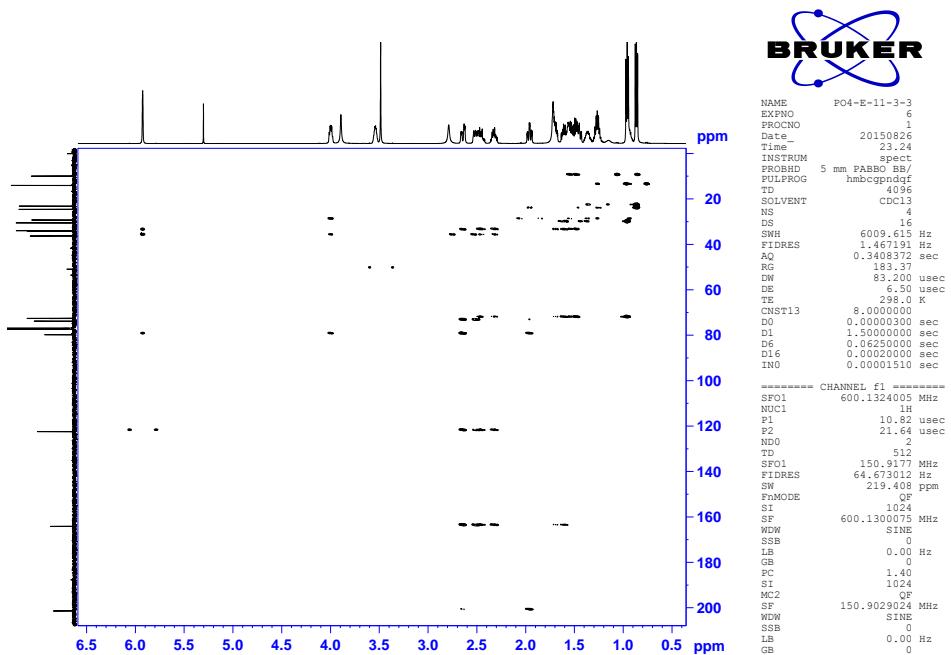
**Figure S34** DEPT spectrum of **5/6** in  $\text{CDCl}_3$



**Figure S35**  $^1\text{H}$ - $^1\text{H}$  COSY spectrum of **5/6** in  $\text{CDCl}_3$



**Figure S36** HMQC spectrum of **5/6** in  $\text{CDCl}_3$



**Figure S37** HMBC spectrum of **5/6** in  $\text{CDCl}_3$

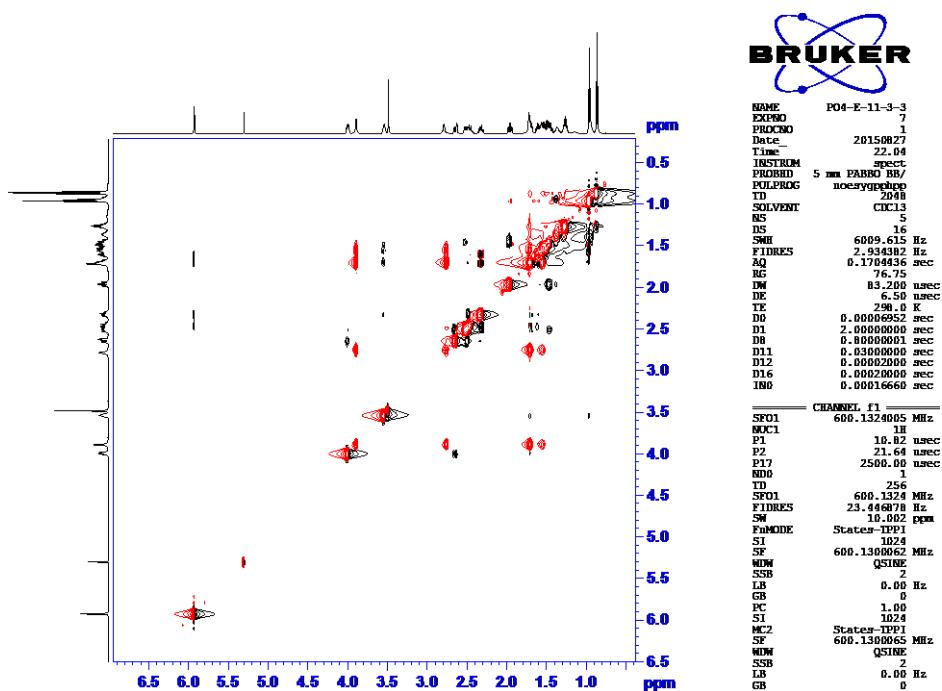


Figure S38 NOESY spectrum of **5/6** in CDCl<sub>3</sub>

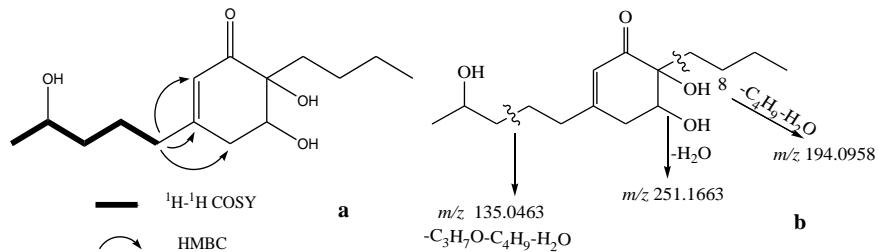


Figure S39 Key <sup>1</sup>H-<sup>1</sup>H COSY and HMBC correlations and MS/MS fragment ion peaks for **5/6**

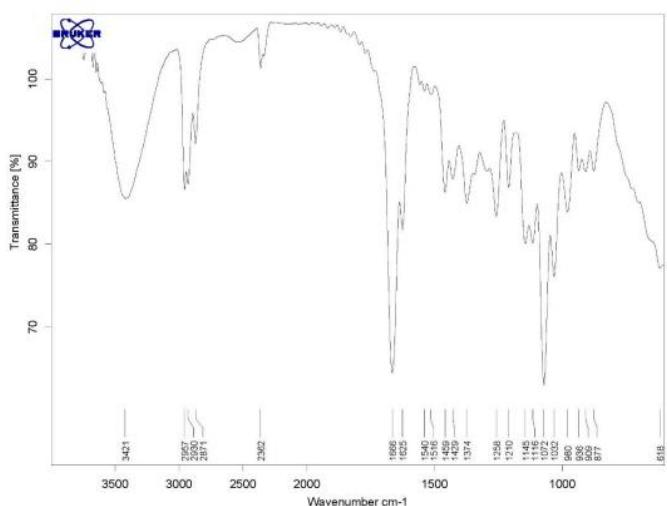
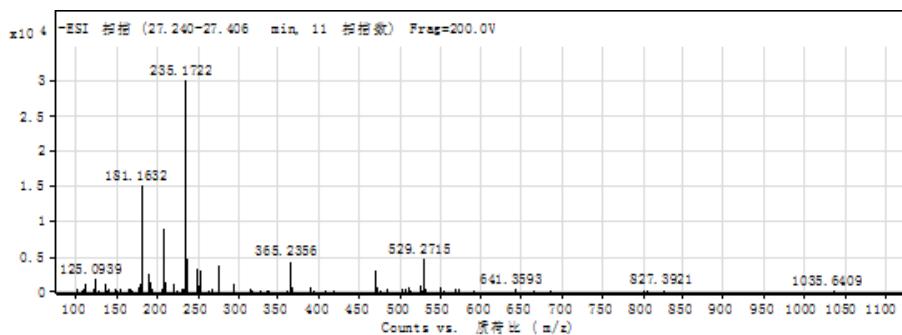
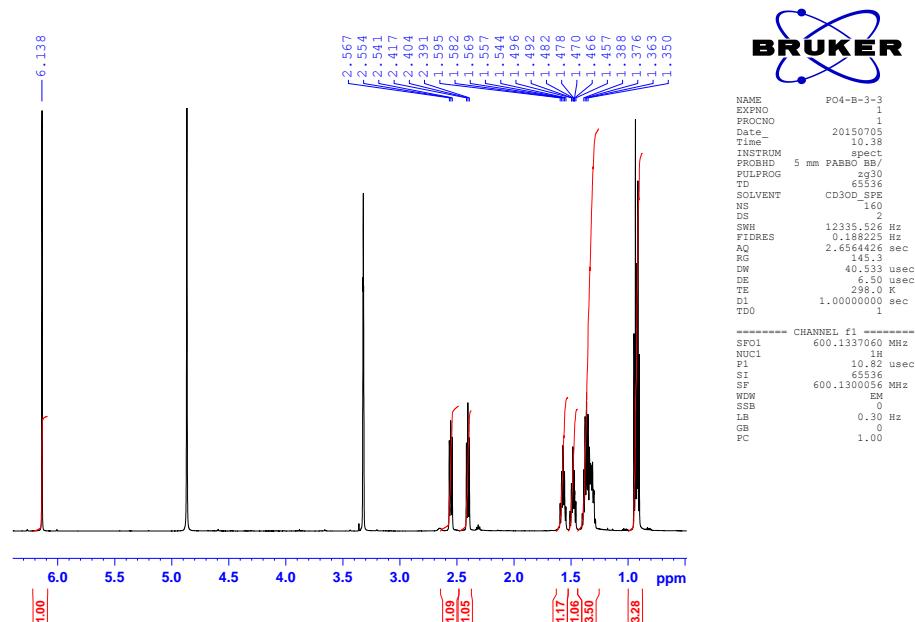


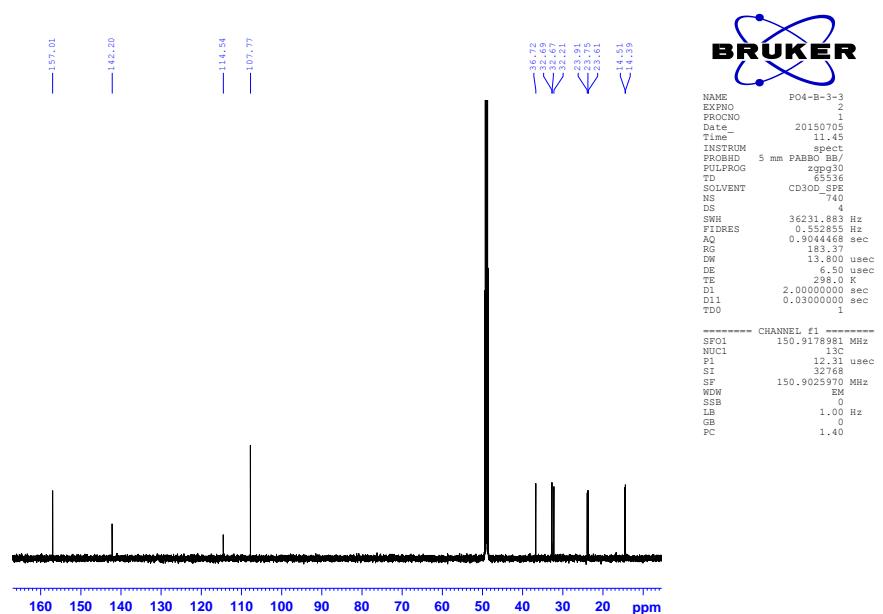
Figure S40 IR spectrum of **5/6**



**Figure S41** HRESI mass spectrum of **7**



**Figure S42**  $^1\text{H}$  NMR spectrum of **7** in  $\text{CD}_3\text{OD}$



**Figure S43**  $^{13}\text{C}$  NMR spectrum of **7** in  $\text{CD}_3\text{OD}$

**Table S1**<sup>1</sup>H NMR data for **1-6**(600 MHz, in CDCl<sub>3</sub>)

No.	<b>1</b>	<b>2</b>	<b>3/4<sup>a</sup></b>	<b>5/6</b>
3	3.99 ( dd, 10.2, 5.8)	3.99 (dd, 10.4, 5.8 Hz)	3.99 (dd, 10.5, 5.9 Hz)	3.92 (dd, 9.8, 5.7 Hz)
4	2.59 (dd, 18.5, 5.8 Hz)	2.81 (dd, 18.7, 5.8 Hz)	2.64 (ddd, 18.6, 5.9, 2.6 Hz)	2.48 (m), 2.63 (ddd, 18.6, 5.7, 3.0 Hz)
6	2.47 (ddd, 18.5, 10.2, 2.6 Hz)	2.38 (ddd, 18.7, 10.4, 2.6 Hz)	2.51 (m)	5.84 (bs)
6	6.15 (dd, 2.6, 1.3 Hz)	6.08 (d, 2.4 Hz)	5.92 (bs)	
7	1.98 (dt, 13.2, 4.2 Hz)	1.98 (dt, 13.2, 4.2 Hz)	1.96 (dt, 13.2, 4.2 Hz)	1.88 (dt, 12.0, 4.2 Hz)
7	1.47 (dt, 13.2, 4.2 Hz)	1.47 (dt, 12.0, 4.2 Hz)	1.47 (m)	1.53 (m)
8	0.95 (m), 1.34 (m)	0.95 (m), 1.37 (m)	1.47 (m), 1.36 (m)	0.99 (m), 1.43 (m)
9	1.27 (m)	1.26 (m)	1.26 (m)	1.28 (m)
10	0.86 (t, 7.3 Hz)	0.86 (t, 7.3 Hz)	0.86 (t, 7.3 Hz)	0.88 (t, 7.3 Hz)
11	4.26 (bt, 5.4 Hz)	4.23 (t, 6.4 Hz)	2.44 (m), 2.31 (m)	2.28 (m)
12	1.68 (m), 1.59 (m)	1.61 (m)	1.69 (m), 1.60 (m)	1.67 (m), 1.57 (m)
13	1.34 (m)	1.36 (m), 1.25 (m)	3.54 (m)	1.43 (m)
14	1.34 (m)	1.36 (m)	1.54 (m)	3.73 (q, 6.0)
15	0.91 (t, 7.2 Hz)	0.91 (t, 7.1 Hz)	0.96 (t, 7.2 Hz)	1.15 (d, 6.0 Hz)

<sup>a</sup> Measured in CD<sub>3</sub>OD**Table S2**<sup>13</sup>C NMR data for **1-6** (150 MHz, in CDCl<sub>3</sub>)

No.	<b>1</b>	<b>2</b>	<b>3/4<sup>a</sup></b>	<b>5/6</b>
1	201.5	201.9	201.46 (201.43)	203.1
2	80.4	80.4	79.84 (79.83)	80.7
3	74.1	73.7	73.88 (73.85)	74.2
4	32.8	31.5	36.39 (36.32)	37.9
5	164.7	164.8	164.22 (164.18)	165.39 (165.34) <sup>b</sup>
6	120.9	121.7	122.50 (122.48)	124.11 (124.09)
7	29.2	29.1	29.35 (29.32)	30.5
8	24.6	24.6	24.6	25.8
9	23.2	23.1	23.2	24.2
10	14.1	14.1	14.12 (14.11)	14.4
11	74.0	74.1	34.12 (34.06)	38.4
12	35.0	35.1	34.0	24.37 (24.34)
13	27.2	27.7	72.70 (72.63)	39.55 (39.52)
14	22.6	22.6	30.60 (30.57)	68.16 (68.10)
15	14.1	14.1	10.00 (9.99)	23.6

<sup>a</sup> Measured in CD<sub>3</sub>OD; <sup>b</sup> data in () for stereoisomer with different orientation of hydroxyl group in amyl chain.