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Concentration-Response Screening of Environmental Chemicals for In Vitro Point-of-Departure Determination using a Targeted RNA-Seq Whole Transcriptome Assay

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1 Background and Objectives

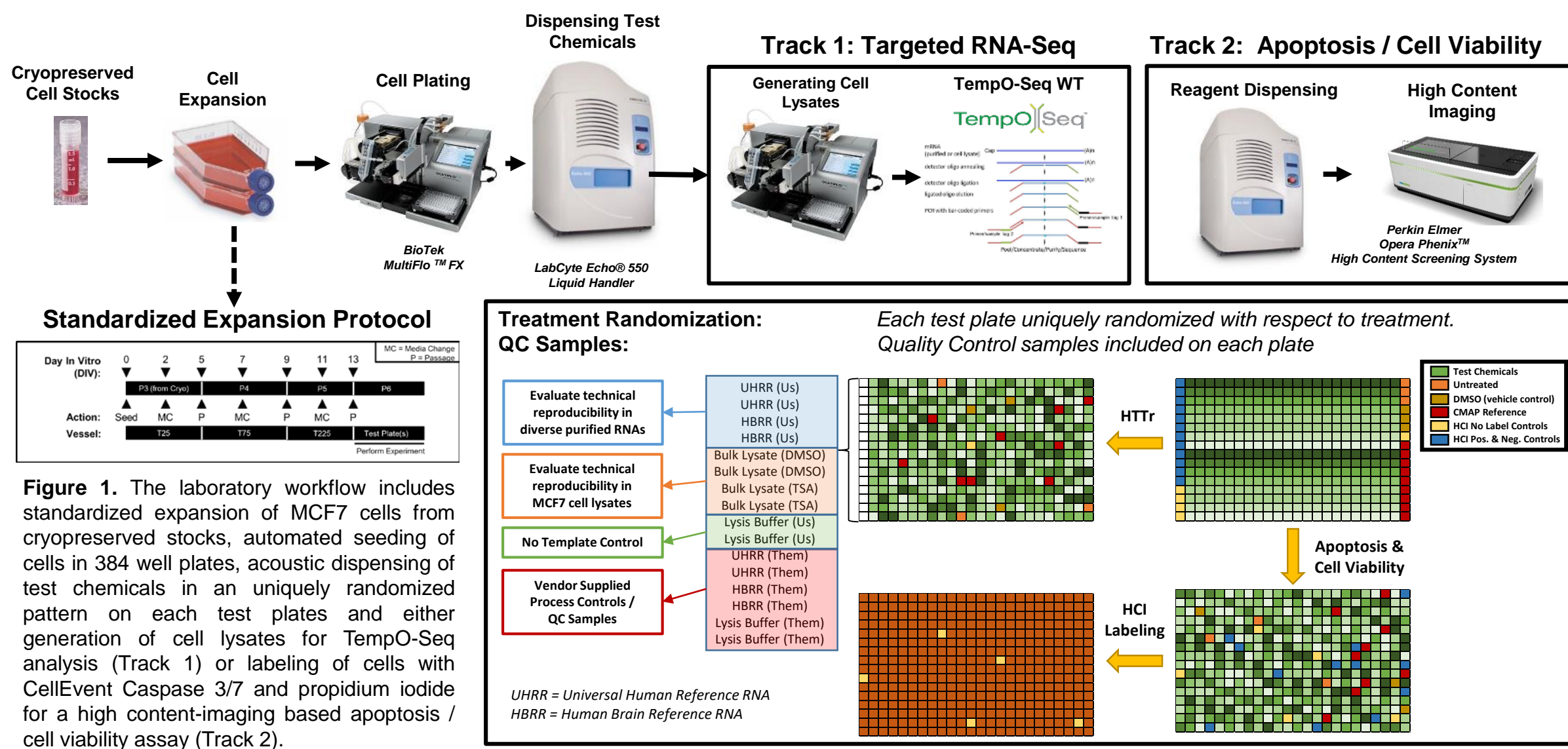
Background

- Current initiatives in environmental toxicology include integrating toxicity testing data from *in vitro* and alternative methods into risk assessment practices.
- Previously, *in vitro* point-of-departure (PODs) estimates suitable for use in screening-level risk assessments have been based on panels of *in vitro* assays that report a limited number of endpoints and that may not provide comprehensive coverage of all biological pathways.
- Increasing efficiency and declining cost of generating whole transcriptome profiles has made high-throughput transcriptomics (HTTr) a practical option for determining *in vitro* bioactivity thresholds and increasing coverage of biological space in chemical screening.

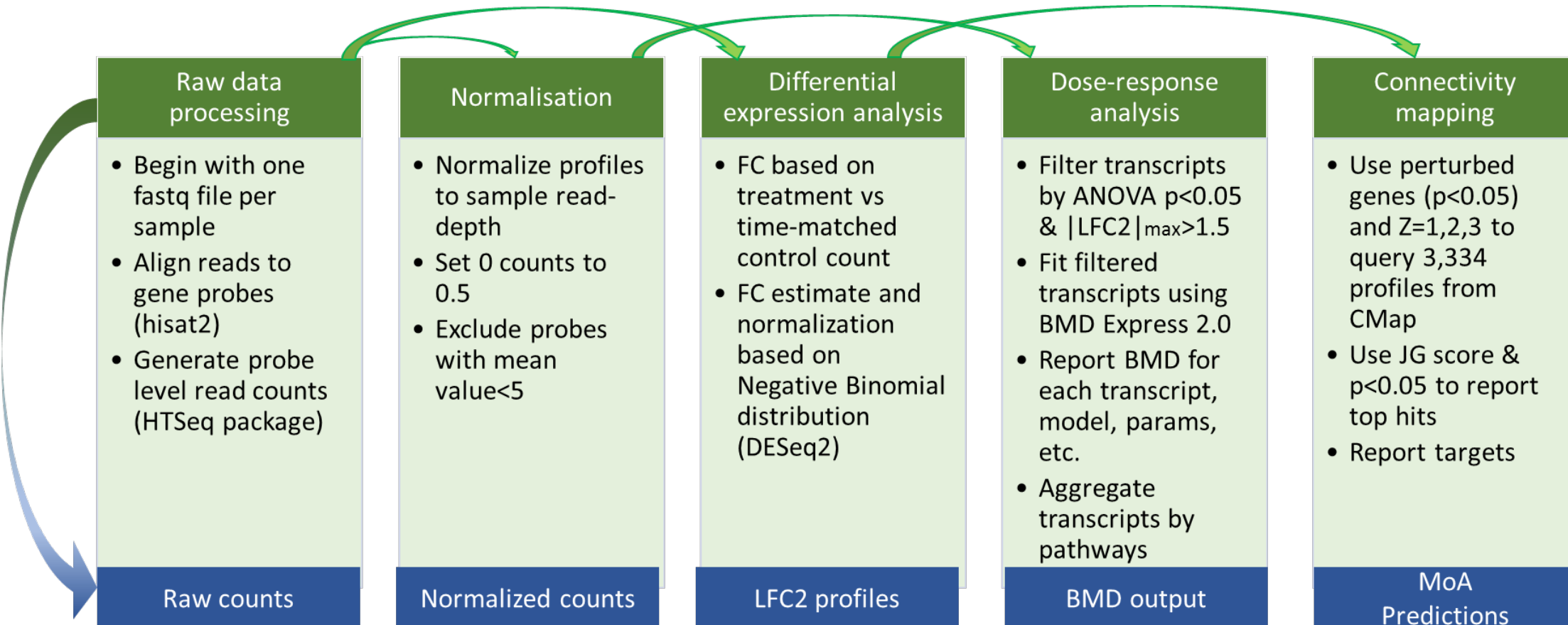
Objectives

- Develop a laboratory workflow for concentration-response screening of environmental chemicals using a targeted RNA-Seq human whole transcriptome assay (TempO-Seq).
- Perform concentration-response screening of a large number of chemicals (n=2,200) from the USEPA Toxicity Forecaster (ToxCast) compound library in MCF7 cells at a single time point (6 hours post-exposure).
- Evaluate the performance of the targeted RNA-Seq assay across plates and screening blocks using reference samples (i.e. commercially-available purified RNAs & laboratory generated bulk cell lysates) and treatment with reference chemicals.
- Establish a data processing pipeline for data normalization, differentially expressed gene (DEG) identification and determination of *in vitro* PODs.

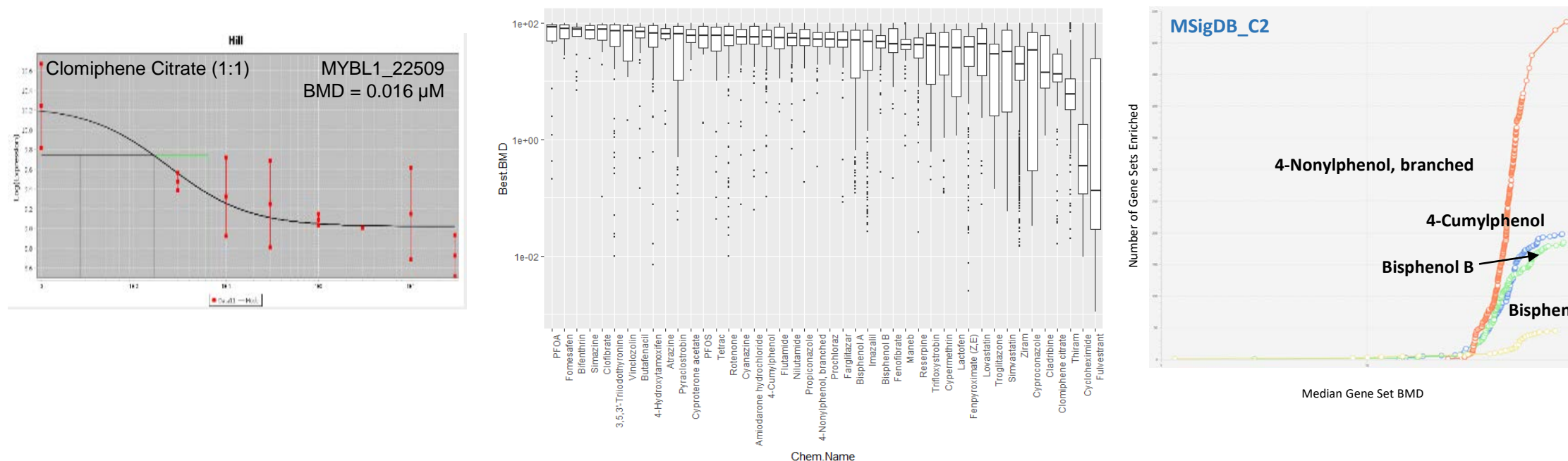
2 Laboratory Workflow



3 HTTr Data Analysis Pipeline



5 POD Determination



4 HTTr Assay Performance

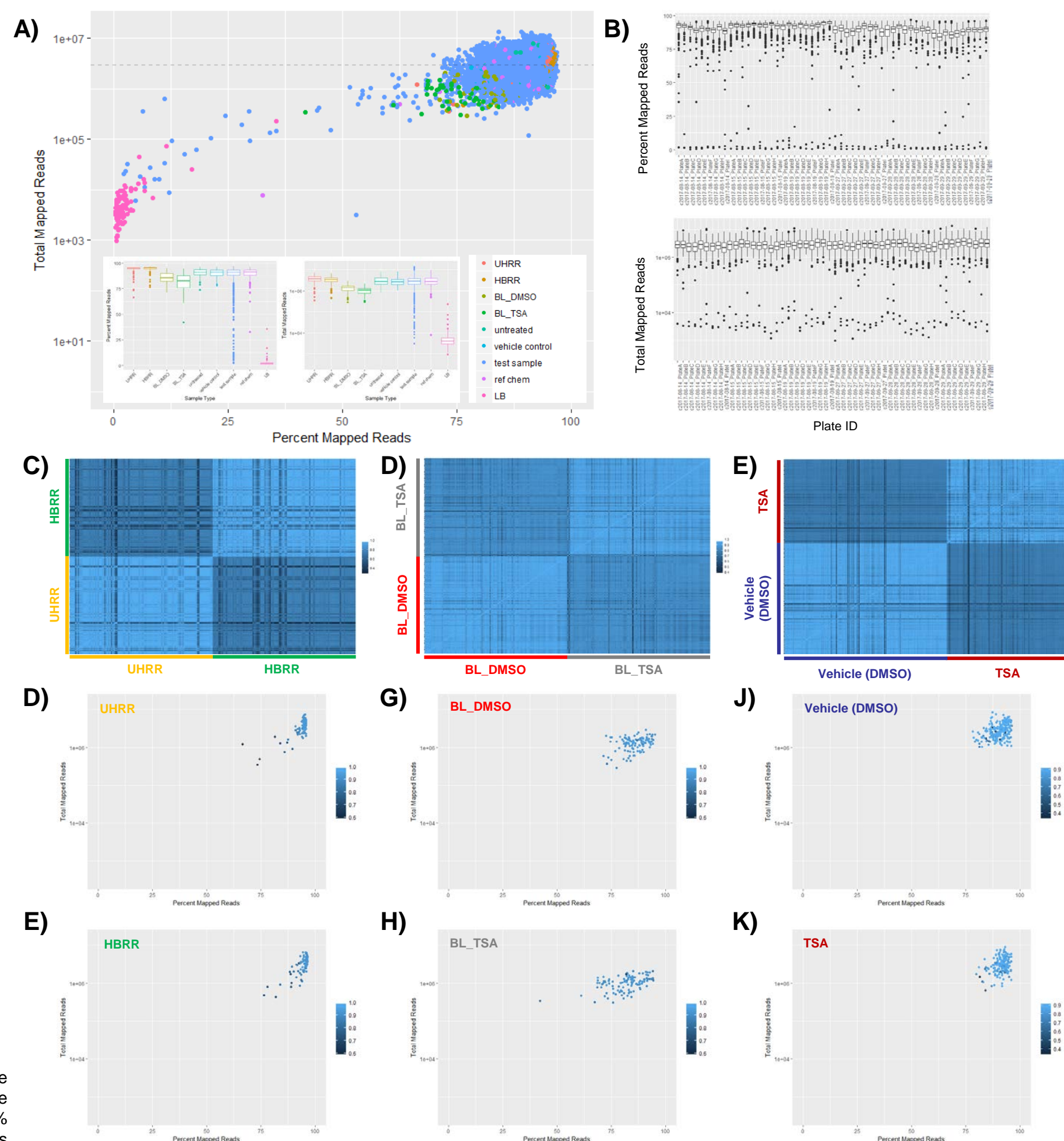


Figure 2. Summary of results from n=54 TempO-Seq assay plates. A majority of samples had total mapped reads greater than 1x10⁶ (mean ~ 3x10⁶) and percent mapped reads > 75% (mean ~90%) (A). The distribution of total mapped reads and percent mapped reads were consistent across assay plates (B). Correlations of like QC samples across plates was high (>0.9) in most cases. Paired QC samples (i.e. UHRR vs. HBRR, BL_DMSO vs BL_TSA, DMSO vs TSA) showed distinct differences in the strength of correlations (C-D). Poor correlations with like samples observed in a minority of QC samples had no apparent relationship with total mapped reads or percent mapped reads (D-K).