

Retrofitting an Estrogen Receptor Transactivation Assay with Metabolic Competence Using Alginate Immobilization of Metabolic Enzymes (AIME)

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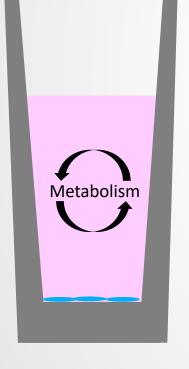
Evaluating Xenobiotic Metabolism in High-Throughput Chemical Screening

- The US EPA's ToxCast program consists of approximately 600 assay endpoints that are comprised of both cell-free and cell-based technologies which are run in high-throughput screening (HTS) platforms.
- Problem: Uncertainty regarding the effects of liver- and tissue-specific xenobiotic metabolism on these
 assay endpoints.
- **Objective**: Incorporate metabolic capabilities onto existing HTS assay platforms to provide a more comprehensive evaluation of potential toxicological hazards.



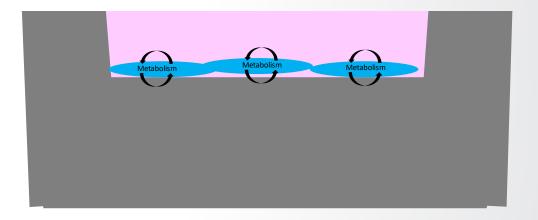
Technical Strategies for Retrofitting High-throughput Assays with Metabolic Capabilities

Extracellular/Cell-free Method



- Capable of metabolizing chemicals in the medium of both cell-based assays <u>and</u> cellfree assays
- More closely models hepatic metabolism and effects of circulating metabolites

Cell-based Method

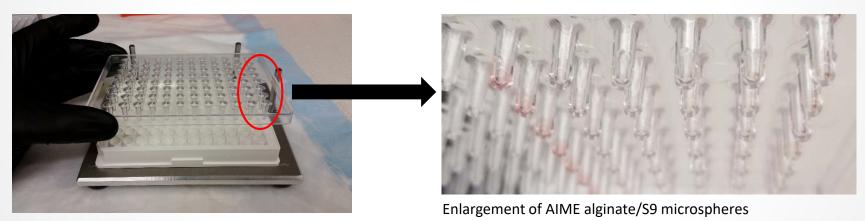


- Capable of metabolizing chemicals inside the cell, but only for cell-based assays
- More closely models effects of directacting metabolites



Alginate Immobilization of Metabolic Enzymes (AIME)

The AIME platform retrofits existing HTS assays with metabolic competence by encapsulating and attaching induced rat liver homogenate (S9) to solid supports extending from custom microplate lids.



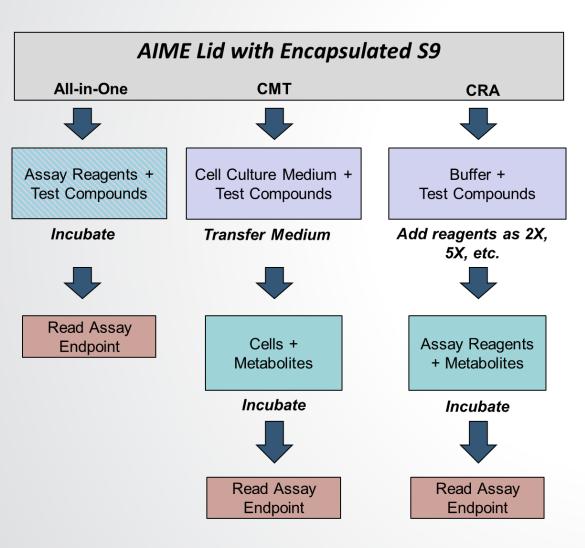
AIME Lid & 96-Well Assay Plate



Cross section of AIME Lid & Assay Plate



AIME Deployment Strategies for Cell-Based & Cell-Free Assays



<u>All-in-One Method:</u> Metabolism of test compounds & assay are run simultaneously.

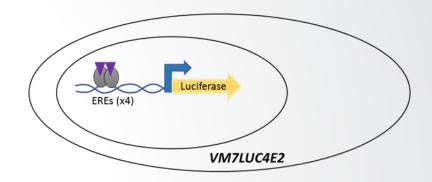
<u>Conditioned Medium Transfer (CMT):</u> Test compound metabolism occurs separately & prior to assay. Medium containing metabolites is then transferred to the assay plate.

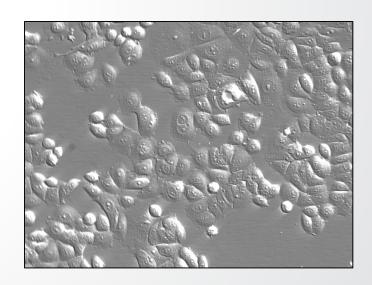
<u>Concentrated Reagent Addition (CRA):</u> Test compound metabolism occurs in the assay plate in conditions favorable to enzymatic activity. Following metabolite generation, concentrated reagents are added to initiate the assay.



VM7Luc4E2 Estrogen Receptor Transactivation Assay

- Developed by Jane Rogers and Michael Denison (In Vitro & Molecular Toxicology, 2000)
 - Human breast carcinoma cells (MCF-7 variant) containing a stably integrated ER-responsive luciferase reporter gene
 - Originally designated as BG1Luc4E2
 - Endogenously expresses ERα
 - Little to no expression of ERβ
- OECD approved method for the detection of ER agonists and antagonists (TG455/457)
- Part of theTox21 high-throughput screening portfolio

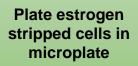






Retrofitting the VM7Luc4E2 ER Transactivation Assay with AIME

VM7Luc4E2 Estrogen Receptor TA





Recover 24 hours & remove medium



Medium containing metabolites



Determine luciferase activity





Dose test compounds into medium

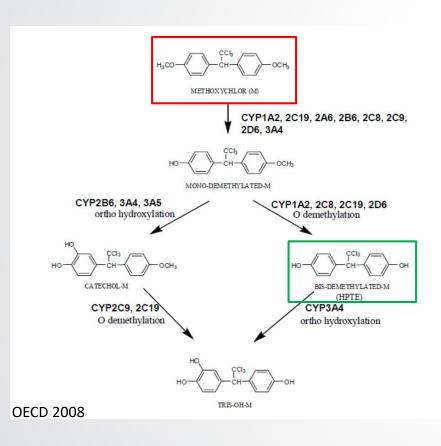


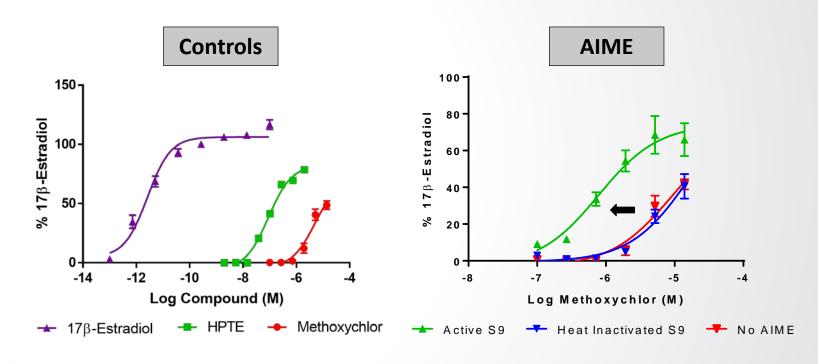
Incubate test compounds with AIME lid





Methoxychlor – A Reference Chemical for the Influence of Metabolism on Estrogen Receptor Activity

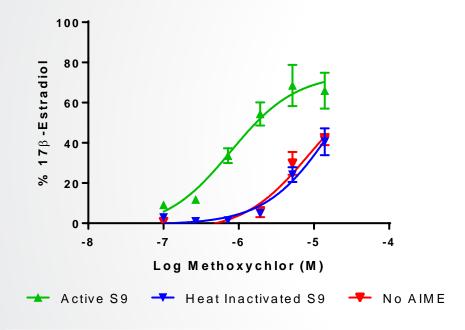




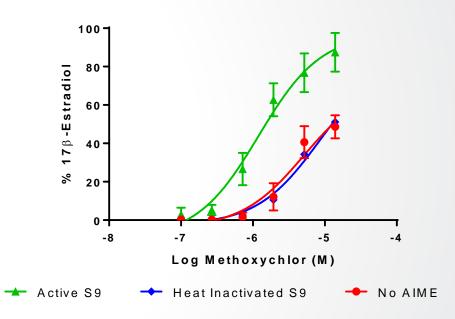
Potency shift between methoxychlor and HPTE defines assay window for estrogenic metabolites



The AIME Platform Can Be Successfully Scaled to a 384-Well Format



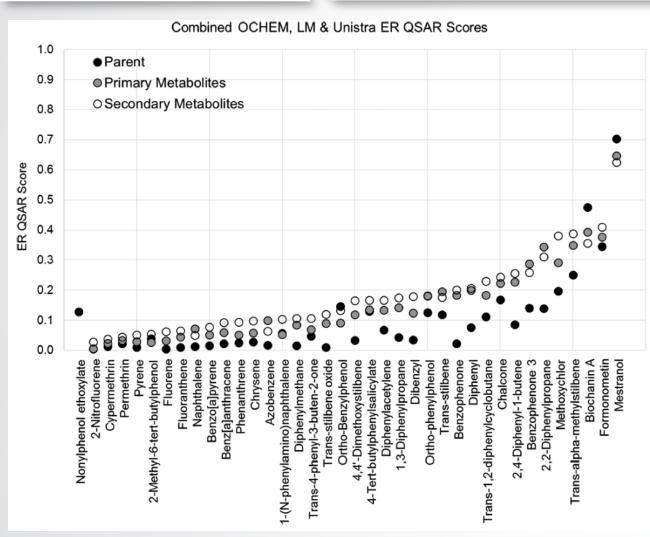
96-Well	EC50 (μM)	AUC
Active S9	0.78	88.20
Heat Inactivated S9	16.86	22.82
No AIME	8.44	24.91



384-Well	EC50 (μM)	AUC
Active S9	1.20	92.87
Heat Inactivated S9	8.40	37.10
No AIME	5.07	33.86



Screening for Predicted Estrogenic Metabolites in the AIME-coupled VM7Luc4E2 Assay



Proof-of-Concept - Screening of "Pinto Library"

- 38 chemicals with reported estrogenic metabolites (predicted true positive)
- 12 chemicals with no predicted estrogenic metabolites (predicted true negative)
- 20 additional chemicals of interest (VM7Luc4E2 assay positive & negative controls)

Pinto et al. (2016). Prediction of Estrogenic Bioactivity of Environmental Chemical Metabolites. Chemical Research in Toxicology



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