## **Supplementary Material**

# (±) Gancochlearols A and B: Cytotoxic and COX-2 inhibitory meroterpenoids from *Ganoderma cochlear*

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#### Abstract

(+)- and (–)-gancochlearols A (1) and B (2), two pairs of dimeric mertoterpenoid enantiomers were isolated from the fruiting bodies of *Ganoderma cochlear*. Their structures were identified by spectroscopic methods. Biological assessments show that the enantiomers of 1 and 2 are cytotoxic against three human cancer cell lines (A549, K562, Huh-7) and could inhibit COX-2 expression with IC<sub>50</sub> values less than 10  $\mu$ M.

Keywords: Ganoderma cochlear; meroterpenoids; cytotoxic activity; COX-2

Figure S1. The structures of compounds 1 and 2

Figure S2. Key HMBC , 1H-1H COSY and ROESY correlations of compounds 1 and 2

**Table S1.** <sup>1</sup>H (600 MHz) and <sup>13</sup>C NMR (150 MHz) data of **1** ( $\delta$  in ppm, J in Hz, Methanol-d<sub>4</sub>)

**Table S2.** <sup>1</sup>H (600 MHz) and <sup>13</sup>C NMR (150 MHz) data of **2** ( $\delta$  in ppm, J in Hz, CDCl<sub>3</sub>)

**Table S3.** Cytotoxic and COX-2 inhibitory activities of the isolates

**Figure S3.** <sup>1</sup>H NMR spectrum of **1** in methanol- $d_4$ 

**Figure S4.** <sup>13</sup>C NMR and DEPT spectra of 1 in methanol- $d_4$ 

**Figure S5.** HSQC spectrum of 1 in methanol- $d_4$ 

**Figure S6.** HMBC spectrum of 1 in methanol- $d_4$ 

**Figure S7.** <sup>1</sup>H-<sup>1</sup>H COSY spectrum of 1 in methanol- $d_4$ 

**Figure S8.** ROESY spectrum of 1 in methanol- $d_4$ 

Figure S9. HRESIMS of 1

**Figure S10.** Optical rotatory data of (+)-1

**Figure S11.** Optical rotatory data of (–)-1

Figure S12. CD spectrum of (+)-1

Figure S13. CD spectrum of (–)-1

**Figure S14.** <sup>1</sup>H NMR spectrum of **2** in CDCl<sub>3</sub>

Figure S15. <sup>13</sup>C NMR and DEPT spectra of 2 in CDCl<sub>3</sub>

Figure S16. HSQC spectrum of 2 in CDCl<sub>3</sub>

Figure S17. HMBC spectrum of 2 in CDCl<sub>3</sub>

**Figure S18.** <sup>1</sup>H-<sup>1</sup>H COSY spectrum of **2** in CDCl<sub>3</sub>

Figure S19. ROESY spectrum of 2 in CDCl<sub>3</sub>

Figure S20. HRESIMS of 2

Figure S22. Optical rotatory data of (+)-2

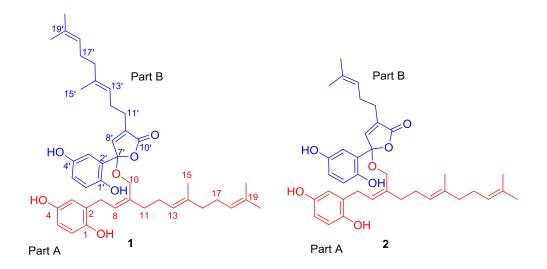


Figure S1. The structures of compounds 1 and 2

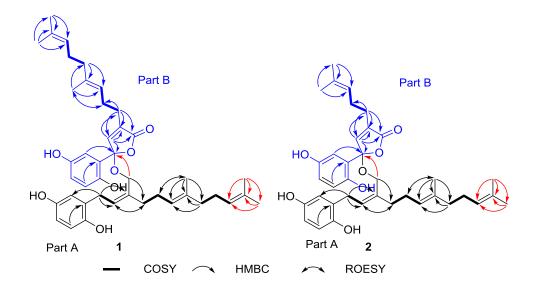


Figure S2. Key HMBC, <sup>1</sup>H-<sup>1</sup>H COSY and ROESY correlations of compounds 1 and 2

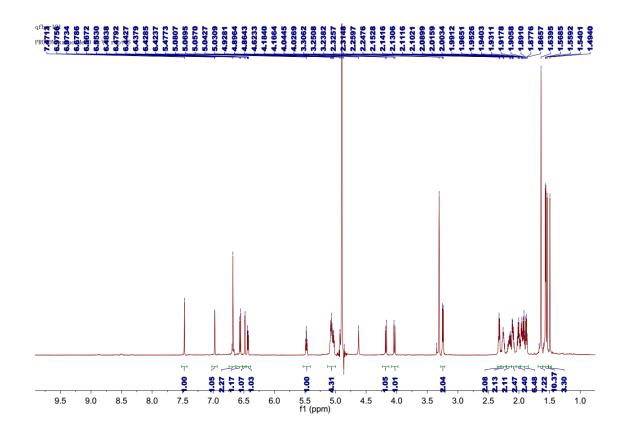
Position	$\delta_{\rm H}(J \text{ in Hz})$	$\delta_{ m C}$	Position	$\delta_{\rm H}(J \text{ in Hz})$	$\delta_{ m C}$
1		149.0 s	1′		149.5 s
2		118.0 s	2'		123.7 s
3	6.48 (d, 2.8)	117.4 d	3'	6.97 (d, 2.0)	114.7 d
4		151.05 s	4′		151.06 s
5	6.43 (dd, 8.5, 2.8)	114.2 d	5'	6.68 (overlap)	118.0 s
6	6.80 (d, 8.5)	116.5 d	6'	6.68 (overlap)	118.3 d
7	3.25 (m)	29.2 t	7′		108.6 s
8	5.48 (t)	129.6 s	8′	7.74 (s)	148.6 d
9		136.6 s	9'		135.5 s
10	4.18 (d, 10.6)	63.1 t	10′		173.7 s
	4.04 (d, 10.6)				
11	2.15 (m)	36.6 t	11′	2.33 (m)	26.0 t
12	2.10 (dd, 13.6, 6.3)	27.7 t	12'	2.26 (m)	26.7 t
13	5.08 (t, 6.0)	125.4 d	13′	5.07 (overlap)	123.7 d
14		137.9 s	14′		136.2 s
15	1. 54 (s)	16.2 q	15'	1.49 (s)	16.2 q
16	1.88 (m)	40.8 t	16′	1.92 (m)	40.8 t
17	1.95 (m)	27.7 t	17′	2.02 (m)	27.7 t
18	5.03 (t, 7.1)	125.5 d	18′	5.06 (overlap)	125.6 d
19		132.1 s	19′		132.0 s
20	1.56 (s)	17.8 q	20'	1.57 (s)	17.8 q
21	1.64 (s)	25.9 q	21'	1.64 (s)	25.9 q

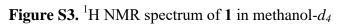
**Table S1.** <sup>1</sup>H (600 MHz) and <sup>13</sup>C NMR (150 MHz) data of **1** ( $\delta$  in ppm, J in Hz, Methanol- $d_4$ ).

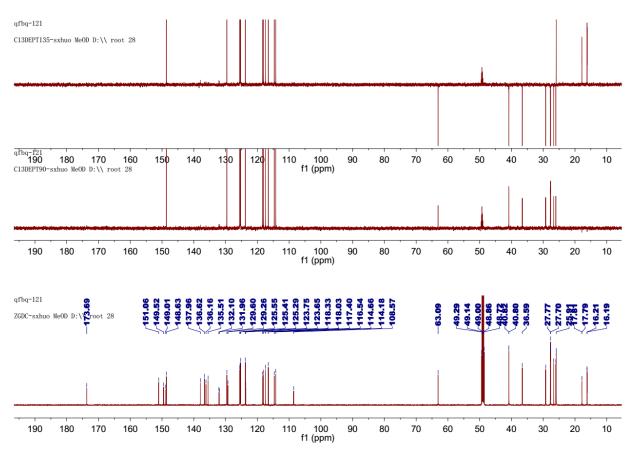
Position	$\delta_{\rm H}(J \text{ in Hz})$	$\delta_{ m C}$	Position	$\delta_{\rm H}(J \text{ in Hz})$	$\delta_{ m C}$
1		147.5 s	1′		148.2 s
2		127.2 s	2'		121.4 s
3	6.60 (d, 2.8)	116.9 d	3'	6.77 (d, 2.6)	112.9 d
4		149.5 s	4'		149.1 s
5	6.58 (d, 8.6, 2.8)	114.1 d	5'	6.79 (dd, 8.6, 2.8)	118.4 s
6	6.65 (d, 8.5)	116.7 d	6′	6.83 (d, 8.6)	118.9 d
7	3.26 (m)	29.1 t	7′		106.6 s
8	5.55 (t)	128.8 s	8′	7.12 (s)	145.6 s
9		135.2 s	9′		135.3 s
10	4.25 (d, 10.6)	62.7 t	10′		171.2 s
	4.12 (d, 10.6)				
11	2.12 (m)	35.6 t	11'	2.33 (m)	25.1 t
12	2.09 (m)	26.4 t	12'	2.26 (m)	25.7 t
13	5.07 (t, 6.9)	123.2 d	13'	5.03 (t, 7.1)	122.2 d
14		135.7 s	14'		133.6 s
15	1. 54 (s)	16.0 q	15'	1.53 (s)	17.7 q
16	1.94 (m)	39.6 t	16′	1.63 (s)	25.6 t
17	2.02 (m)	26.4 t			
18	5.04 (t, 7.1)	124.3 d			
19		131.4 s			
20	1.59 (s)	17.7q			
21	1.67 (s)	25.6 q			

## **Table S2.** <sup>1</sup>H (600 MHz) and <sup>13</sup>C NMR (150 MHz) data of **2** ( $\delta$ in ppm, J in Hz, CDCl<sub>3</sub>).

Compound	IC <sub>50</sub> (µM)						
	COX-2	A549	K562	Huh-7			
(+)-1	2.18	9.59	10.9	9.88			
(–)-1	4.24	8.86	12.8	18.4			
(+)-2	4.80	10.2	15.2	19.0			
(-)-2	4.86	10.4	13.1	7.60			
Positive cotontrol <sup><i>a</i></sup>	0.0171	0.00172	0.00324	0.00706			







**Figure S4.**<sup>13</sup>C NMR and DEPT spectra of **1** in methanol- $d_4$ 

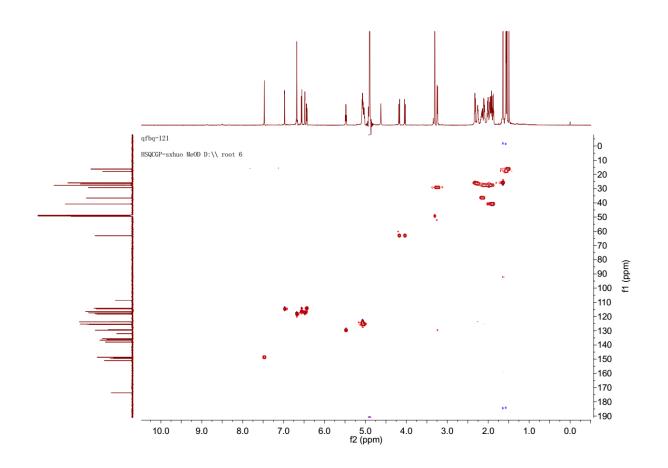


Figure S5. HSQC spectrum of 1 in methanol- $d_4$ 

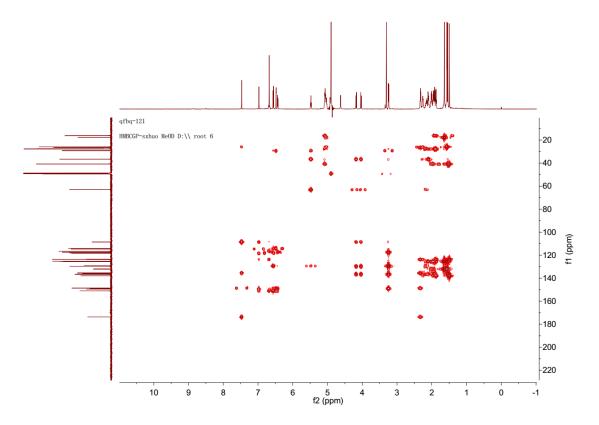
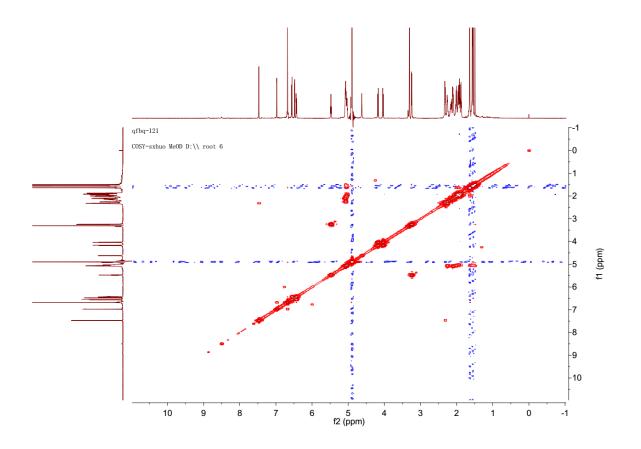


Figure S6. HMBC spectrum of 1 in methanol- $d_4$ 



**Figure S7.** <sup>1</sup>H-<sup>1</sup>H COSY spectrum of 1 in methanol- $d_4$ 

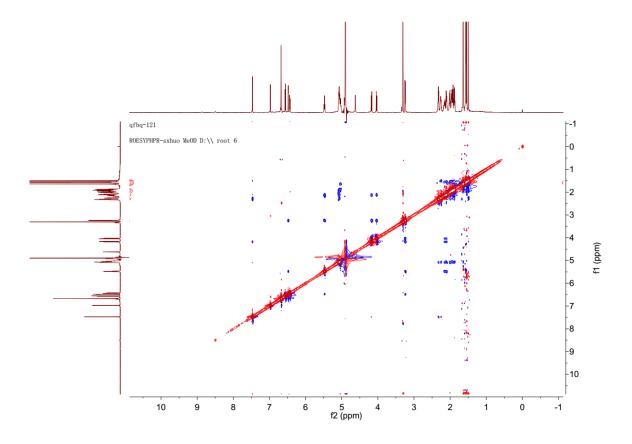
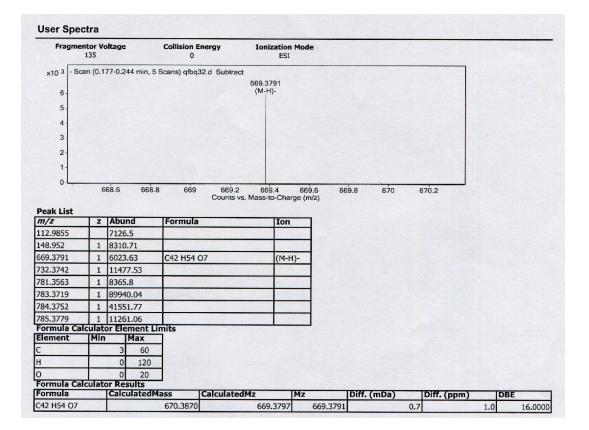
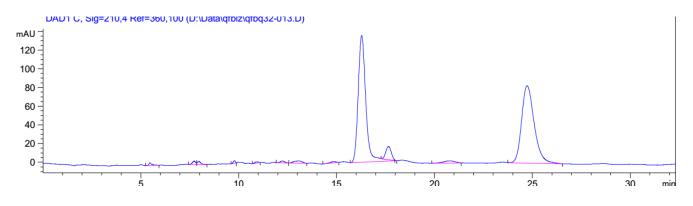


Figure S8. ROESY spectrum of 1 in methanol- $d_4$ 









Optica	rotation mi	easurement						
Model	: P-1020 (A	060460638)						
No.	Sample	Mode	Data	Monitor Blank	Temp. Cell Temp Point	Date Comment Sample Name	Light Filter Operator	Cycle Time Integ Time
No.1	1 (1/3)	Sp.Rot	47.5160	0.0153 0.0000	22.7 10.00 Cell	Sat Oct 24 11:02:31 2015 0.00322g/mL Acetonitrile QFBQ32A	Na 589nm	2 sec 10 sec
No.2	1 (2/3)	Sp.Rot	48.1370	0.0155 0.0000	22.7 10.00 Cell	Sat Oct 24 11:02:44 2015 0.00322g/mL Acetonitrile QFBQ32A	Na 589nm	<sup>2 sec</sup> <sup>10 sec</sup> + 47, 95 96
No.3	1 (3/3)	Sp.Rot	48.1370	0.0155 0.0000	22.7 10.00 Cell	Sat Oct 24 11:02:58 2015 0.00322g/mL Acetonitrile QFBQ32A	Na 589nm	2 sec 10 sec

Figure S11. Optical rotatory data of (+)-1

Mode	: P-1020 (A	060460638)						
No.	Sample	Mode	Data	Monitor Blank	Temp. Cell Temp Point	Date Comment Sample Name	Light Filter Operator	Cycle Time Integ Time
No.1	2 (1/3)	Sp.Rot	-50.6170	-0.0164 0.0000	23.1 10.00 Cell	Sat Oct 24 11:17:46 2015 0.00324g/mL Acetonitrile QFBQ32B	Na 589nm	2 sec 10 sec
No.2	2 (2/3)	Sp.Rot	-49.3830	-0.0160 0.0000	23.1 10.00 Cell	Sat Oct 24 11:17:59 2015 0.00324g/mL Acetonitrile QFBQ32B	Na 589nm	2 sec 10 sec - TO: 4117
No.3	2 (3/3)	Sp.Rot	-51.2350	-0.0166 0.0000	23.1 10.00 Cell	Sat Oct 24 11:18:12 2015 0.00324g/mL Acetonitrile OFBQ32B	Na 589nm	2 sec 10 sec

Figure S12. Optical rotatory data of (–)-1

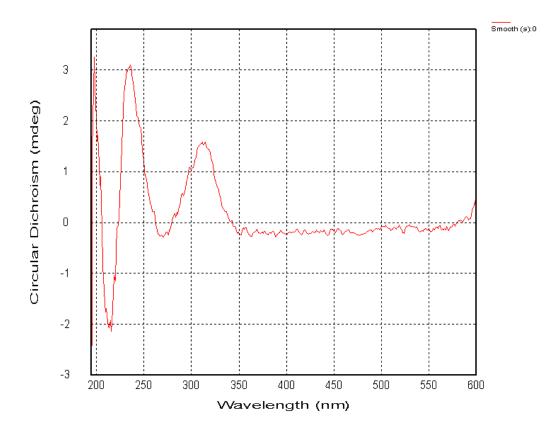
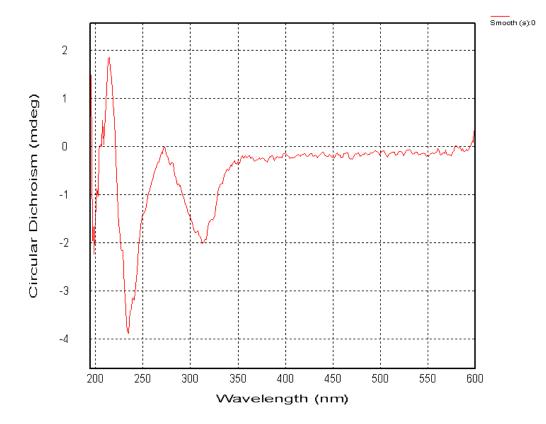
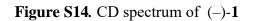


Figure S13. CD spectrum of (+)-1





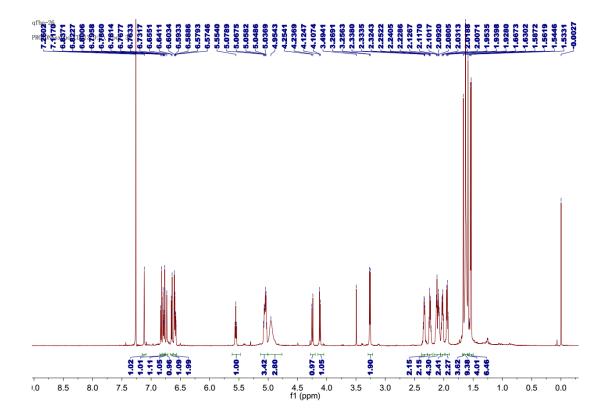


Figure S15. <sup>1</sup>H NMR spectrum of 2 in CDCl<sub>3</sub>

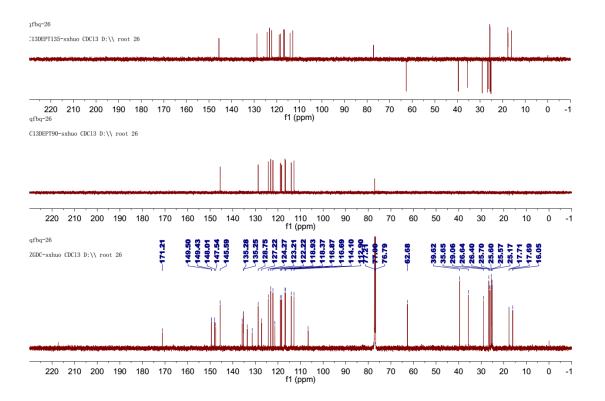


Figure S16. <sup>13</sup>C NMR and DEPT spectrum of 2 in CDCl<sub>3</sub>

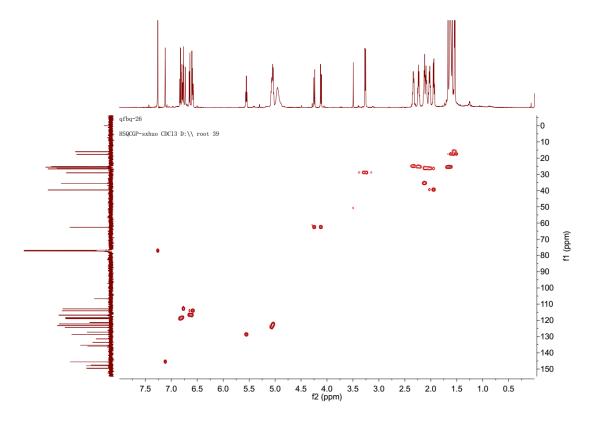


Figure S17. HSQC spectrum of 2 in CDCl<sub>3</sub>

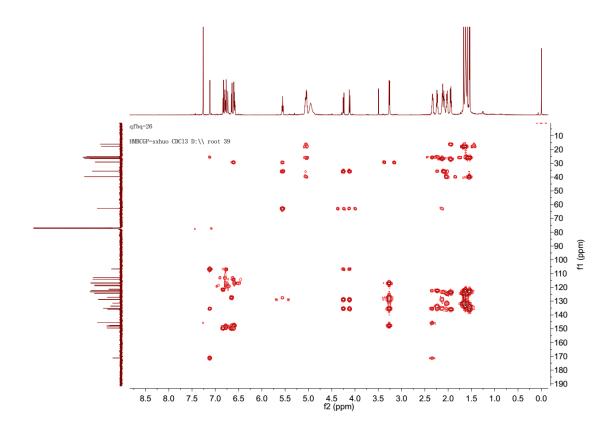


Figure S18. HMBC spectrum of 2 in CDCl<sub>3</sub>

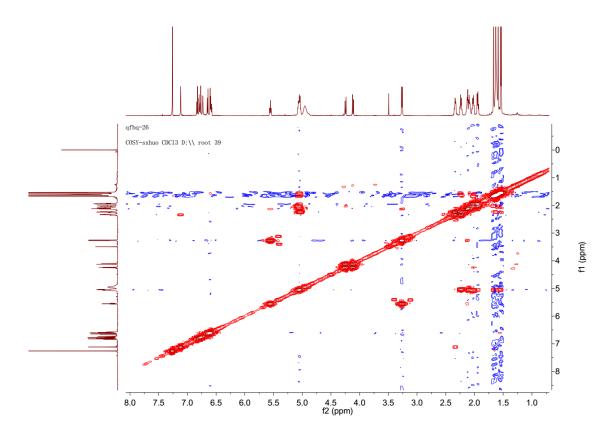


Figure S19. <sup>1</sup>H-<sup>1</sup>H COSY spectrum of 2 in CDCl<sub>3</sub>

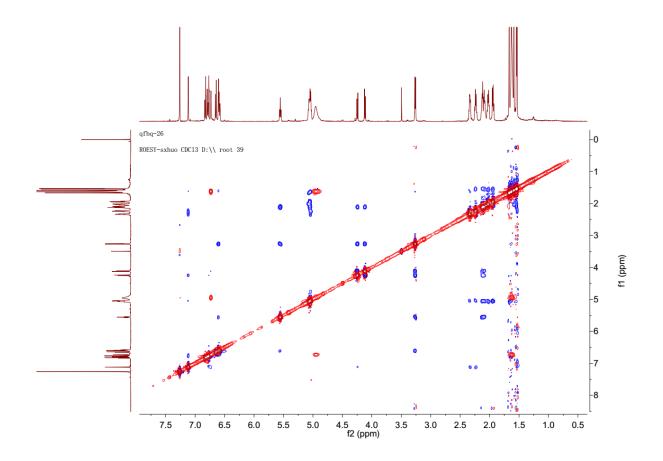


Figure S20. ROESY spectrum of 2 in CDCl<sub>3</sub>

### **Qualitative Analysis Report**

Data Filename	171206ESINA2.d	Sample Name	qfbq26a
Sample Type	Sample	Position	
Instrument Name	Agilent G6230 TOF MS	User Name	KIB
Acq Method	ESIN.m	Acquired Time	12/6/2017 10:15:38 AM
IRM Calibration Status	Success	DA Method	ESI.m
Sample Group	Info.		

6200 series TOF/6500 series Q-TOF B.05.01 (B5125.2)

User Spectra

Acquisition SW Version

20	00 00	ltage	Collision E	nergy	Ionization Mode ESI		
x10 3 - Sca	an (0.	.327 min) 1712	06ESINA	2.d	601.3158		
					001.0156		
1.2-							
1-							
0.8-							
0.6-							
0.4							
0.2							
o							
- F		601	.31575	60	)1.3158	601.31585	
				Counts vs. Ma	ass-to-Charge (r	n/z)	
Peak List							
m/z	z	Abund	Formula		Ion		
112.9856	<u> </u>	1482.05	<b></b>				
227.1076	<u> </u>	858.24	<b></b>				
245.1186	<u> </u>	971.9					
271.0972		833.48					
303.124	1	9851.41					
325.1843		1276.8					
339.2001		1125.29					
601.3158	1	1390.31	C37 H45	07	M-		
1033.9881	1	180450.33					
1034.9898	1	21115.04					
Formula Calc Element	Min	or Element Lin	nics				
C		0 200	1				
н	-	0 200	1				
п О	<u> </u>	3 10	1				
O Formula Calc	ulate		1				
Formula		CalculatedMa	ass	Mz	Diff.(mDa)	Diff. (ppm)	DBE
C37 H45 O7			601.3165	601.3158		0.7	1.2 15.5

--- End Of Report ---

Figure S21. HRESIMS of 2

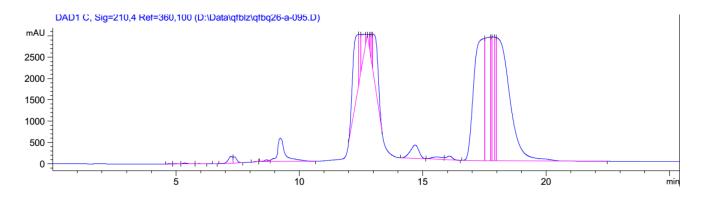


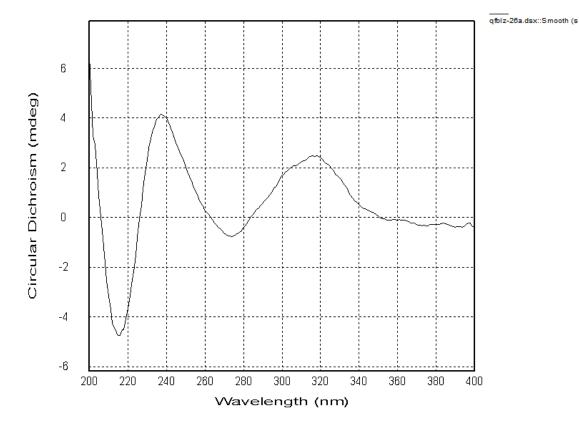
Figure S22. The chiral HPLC chromatogram of 2

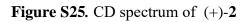
Optica	I rotation m	easurement						
Model	: P-1020 (A	060460638)						
No.	Sample	Mode	Data	Monitor Blank	Temp. Cell Temp Point	Date Comment Sample Name	Light Filter Operator	Cycle Time Integ Time
No.1	6 (1/3)	Sp.Rot	41.6670	0.0175 0.0000	23.2 50.00 Cell	Fri Oct 30 18:07:23 2015 0.00084g/mL MeOH QFBQ26A	Na 589nm	2 sec 10 sec
No.2	6 (2/3)	Sp.Rot	42.8570	0.0180 0.0000	23.3 50.00 Cell	Fri Oct 30 18:07:36 2015 0.00084g/mL MeOH QFBQ26A	Na 589nm	2 sec 10 sec + 42, 3016
No.3	6 (3/3)	Sp.Rot	42.3810	0.0178 0.0000	23.3 50.00 Cell	Fri Oct 30 18:07:50 2015 0.00084g/mL MeOH QFBQ26A	Na 589nm	2 sec 10 sec

Figure S23. Optical rotatory data of (+)-2

Optic	al rotation m	easurement						
Mode	I : P-1020 (/	4060460638)						
No.	Sample	Mode	Data	Monitor Blank	Temp. Cell Temp Point	Date Comment Sample Name	Light Filter Operator	Cycle Time Integ Time
No.1	8 (1/3)	Sp.Rot	-59.4440	-0.0214 0.0000	23.3 50.00 Cell	Fri Oct 30 18:16:36 2015 0.00072g/mL MeOH QFBQ26B	Na 589nm	2 sec 10 sec
No.2	8 (2/3)	Sp.Rot	-61.1110	-0.0220 0.0000	23.3 50.00 Cell	Fri Oct 30 18:16:50 2015 0.00072g/mL MeOH QFBQ26B	Na 589nm	2 sec 10 sec -60.64£1°
No.3	8 (3/3)	Sp.Rot	-61.3890	-0.0221 0.0000	23.3 50.00 Cell	Fri Oct 30 18:17:03 2015 0.00072g/mL MeOH QFBQ26B	Na 589nm	2 sec 10 sec

Figure S24. Optical rotatory data of (–)-2





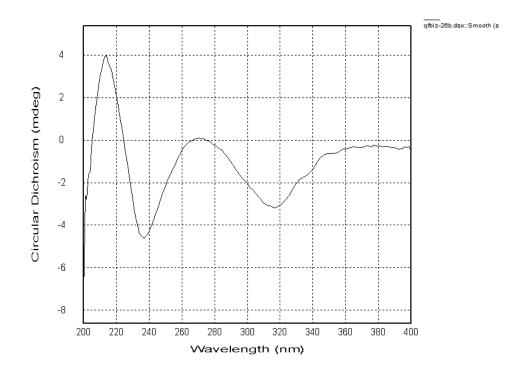


Figure S26. CD spectrum of (-)-2