

## SUPPLEMENTARY MATERIAL

# Secondary metabolites from the fermented rice of the fungus *Monascus purpureus* and their bioactivities

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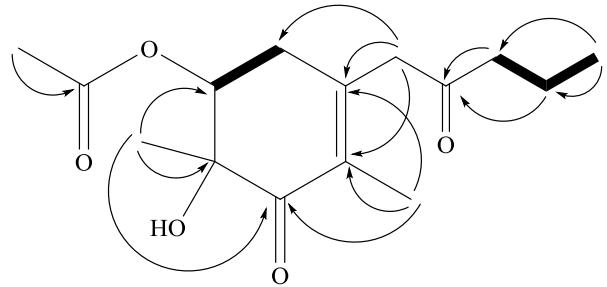
Phytochemical investigation of the EtOAc-soluble fraction of the ethanolic extract of a yellow mutant of the fungus *Monascus purpureus* BCRC 38110 (Eurotiaceae) grown on rice resulted in the isolation of one new azaphilone derivative, monapurpureusone (**1**), one acetophenone metabolite isolated for the first time from natural source, monapurpureusin (**2**), along with four known compounds, TW94a (**3**), ergosterol (**4**), monascin (**5**), and ankaflavin (**6**). The structures and relative configurations of these compounds were elucidated by spectroscopic analyses, including 1D- and 2D-NMR spectroscopy and mass spectrometry, and by the comparison of their NMR data with those of related compounds. Some phytochemicals were evaluated for both anti-inflammatory activity through the measurement of nitric oxide (NO) production levels in lipopolysaccharide (LPS)-stimulated murine-derived macrophages RAW264.7 cell lines and antioxidant activities.

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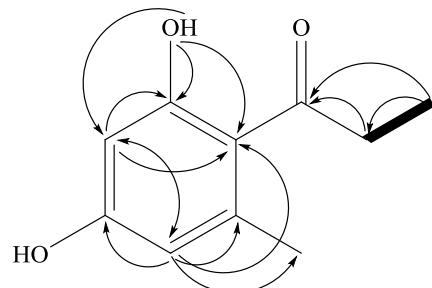
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**Table S1.**  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR Data ( $\text{CDCl}_3$ , 600 and 150 MHz, resp.) of compounds **1** and **2**.  $\delta$  in ppm,  $J$  in Hz.

	<b>1</b>		<b>2</b>	
	$\delta_{\text{C}}$	$\delta_{\text{H}}$	$\delta_{\text{C}}$	$\delta_{\text{H}}$
1	11.7	1.76 ( <i>q</i> , $J = 1.2$ )		166.3
2			101.8	6.26 ( <i>dd</i> , $J = 2.7, 0.6$ )
3	205.6		160.5	
4	48.8	3.33 ( <i>d</i> , $J = 16.8$ ), 3.50 ( <i>d</i> , $J = 16.8$ )	111.8	6.23 ( <i>dq</i> , $J = 2.7, 1.2$ )
4a	146.2			
5	37.9	2.49 ( <i>ddd</i> , $J = 18.0, 10.5, 1.2$ , $\text{H}_{\text{ax}}$ ), 2.53 ( <i>ddd</i> , $J = 18.0, 6.2, 1.2$ , $\text{H}_{\text{eq}}$ )		142.2
6	67.9	4.84 ( <i>dd</i> , $J = 10.5, 6.2$ )	115.6	11.49 ( <i>s</i> )
7	85.1		207.4	
8	195.1		37.3	2.92 ( <i>q</i> , $J = 7.2$ )
8a	132.1			
9	45.0	2.45 ( <i>t</i> , $J = 7.8$ )	8.9	1.21 ( <i>t</i> , $J = 7.2$ )
10	17.2	1.61 ( <i>sextet</i> , $J = 7.8$ )	25.1	2.56 ( <i>s</i> )
11	13.6	0.92 ( <i>t</i> , $J = 7.8$ )		
Me-12	15.8	1.39 ( <i>s</i> )		
14	170.2			
15	21.5	2.10 ( <i>s</i> )		
OH-1			13.2 ( <i>br s</i> )	
OH-3			5.60 ( <i>br s</i> )	
OH-7		2.50 ( <i>br s</i> )		

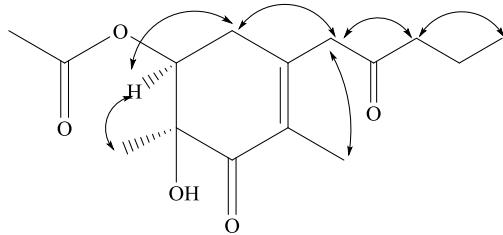


**1**

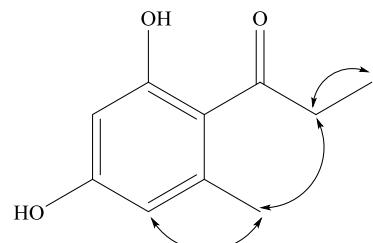


**2**

Figure S1. COSY (—) and HMBC (→) correlations of **1** and **2**.

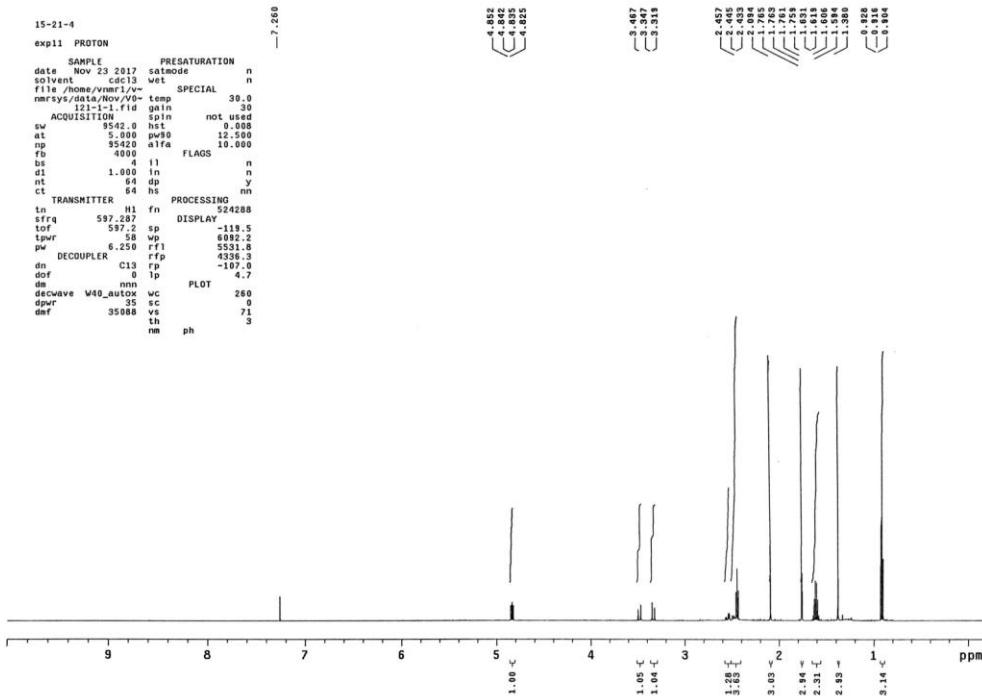


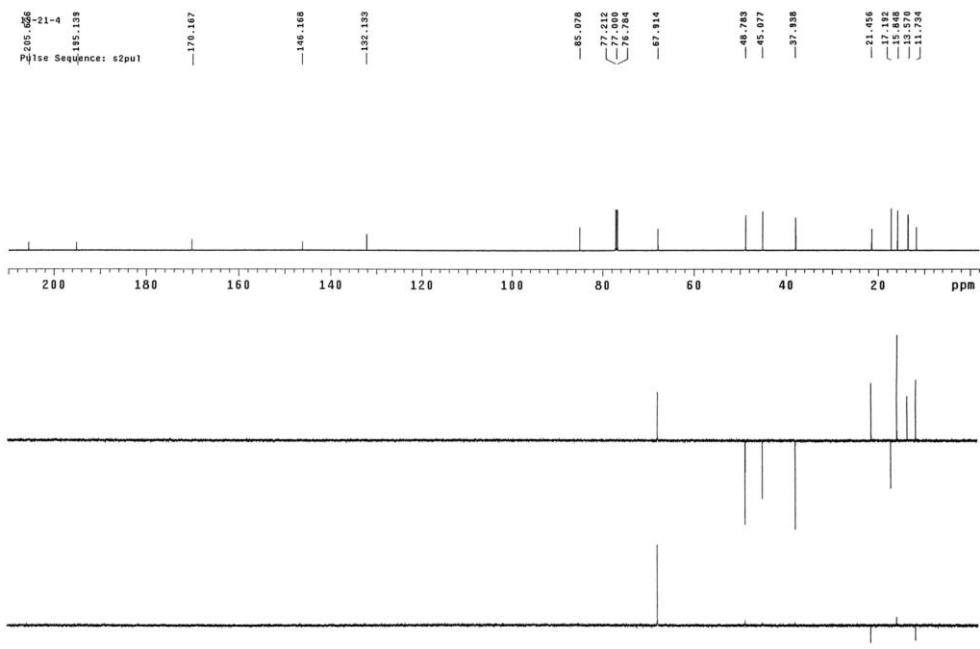
**1**



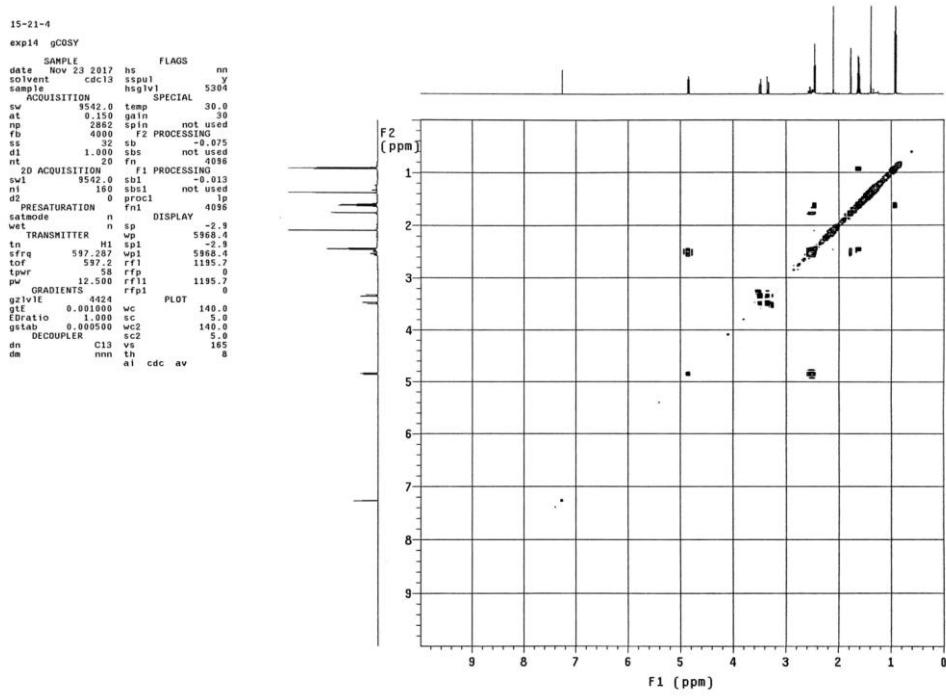
**2**

Figure S2. NOESY correlations of **1** and **2**.





**Figure S5.** DEPT spectrum of **1** (150 MHz in  $\text{CDCl}_3$ )

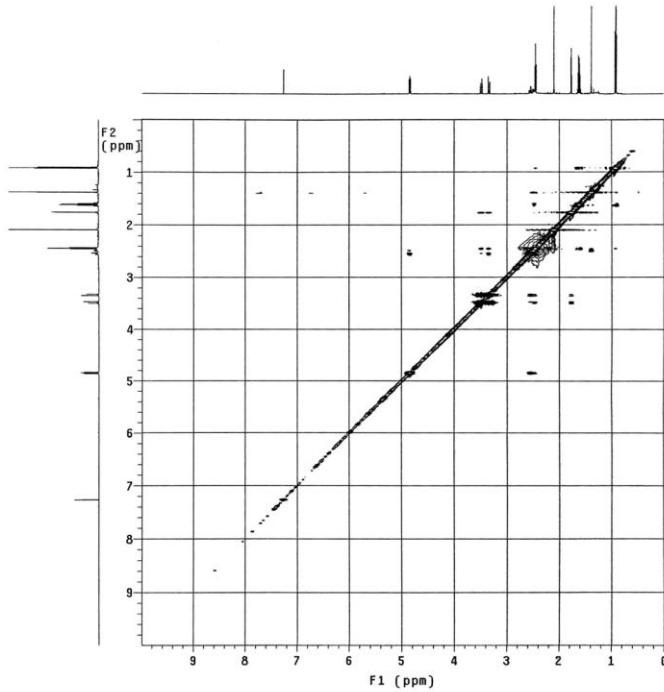


**Figure S6.**  $^1\text{H}$ - $^1\text{H}$  COSY spectrum of **1**

```

15-21-4
exp15 NOESY
      SAMPLE   FLAGS
date Nov 23 2017 hs   nn
solvent cdc13 sspl   y
sample   hsg1g  y
      ACQUISITION hsg1g  5384
sw1  9542.0 SPECIAL
at   0.150 temp  30.0
tp  2802.5 spin   26
fb  4000 spin   not used
ss  32
d1  1.500 gfs2 PROCESSING
nt  20 gfs   not used
dt  0.069
sw2  9542.0 F1 PROCESSING
f1  4096
ni  160 gfs1  0.015
tn  TRANSMITTER H1 proc1 ip
sfrq 597.281 f1n1 4096
tot  3.000 sp   DISPLAY -2.9
tpwr 58 s   5968.4
pw  12.500 wp   5968.4
mixN NOESY  0.600 wpl1  5968.4
      PRESATURATION rfp1  1195.7
satmode n   fp
wetmode n   rfp1  1195.7
      DECOUPLER C13 PLOT
dn  nnn wc   140.0
sc  vco  140.0
wc2 140.0
sc2 5.0
ve  1172
th  2
al  cdc ph

```

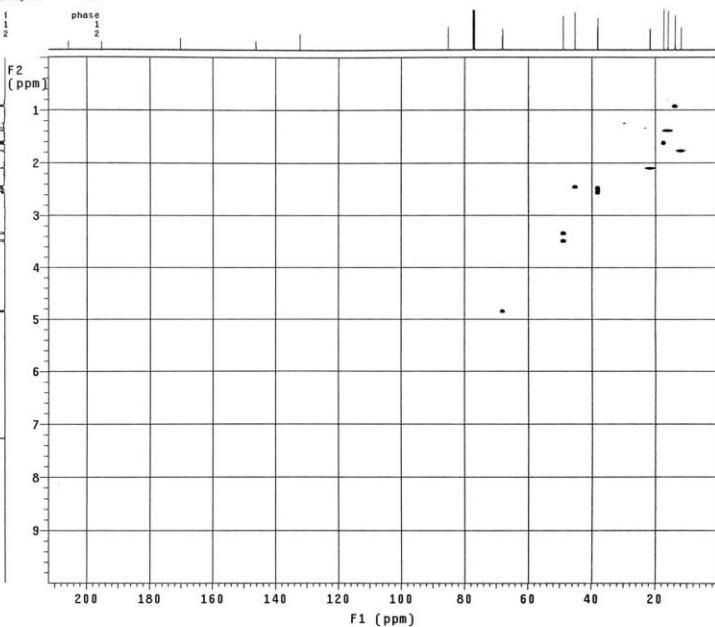


**Figure S7.** NOESY spectrum of **1**

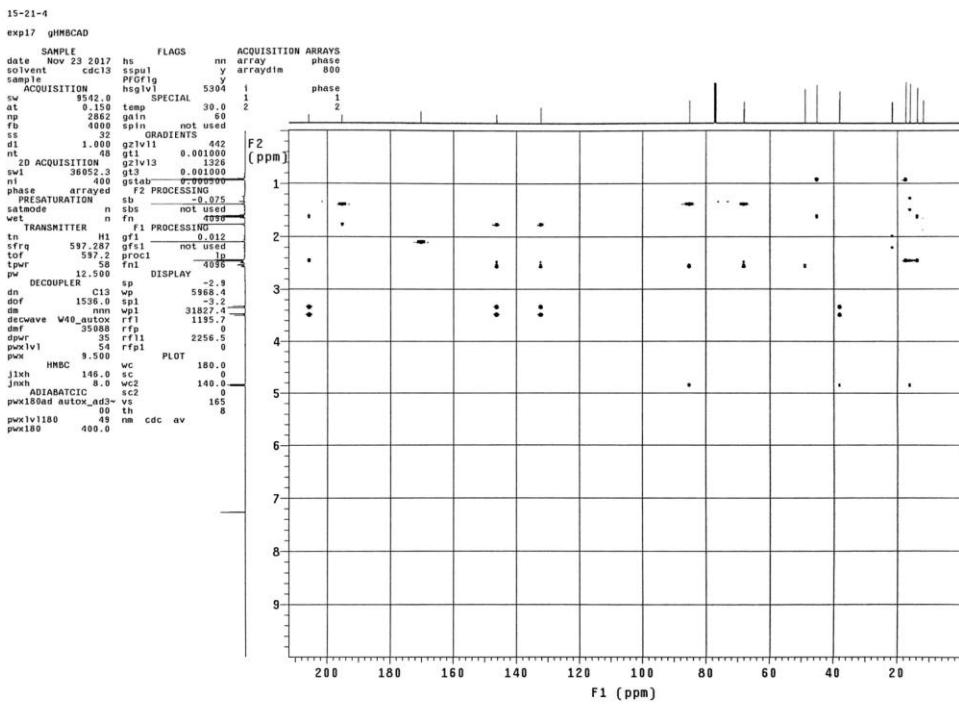
```

15-21-4
exp16 gHSQCAD
      SAMPLE   FLAGS
date Nov 23 2017 hs   nn
solvent cdc13 sspl   y
arraydim 320
sample   hsg1g  y
      ACQUISITION hsg1g  5304
sw1  9542.0 SPECIAL
at   0.150 temp  30.0
tp  2802.5 spin   26
fb  4000 spin   not used
ss  32 GRADIENTS
d1  1.00 g2v1c  4424
nt  24 g1t   0.002000
      2D ACQUISITION Edratio  3.977
sw2  3062.3 F2 PROCESSING
ni  160 gfs1  0.002000
phase arrayed gf   0.059
      PRESATURATION rfp1  1195.7
satmode n   fp
wetmode n   rfp1  1195.7
      TRANSMITTER H1 gfs1  0.005
tn  597.281 proc1 ip
tot  58.2 s   4096
tpwr 58 s   DISPLAY -2.9
pw  12.500 sp
      DECOUPLER C13 sp1  5968.4
dn  nnn wc   180.0
sc  vco  140.0
wc2 140.0
sc2 5.0
ve  47
      ADIABATIC th
multif1g 0 vs
multif2g 47
mult 0 th
      psw1ad autow1ad nm cdc ph
psw18ad autow1ad 0
psw18ad 300
psw18 400.0
psw1180 48
psw18ref autow1ref r200
psw18r 1998.0
psw1v180r 40

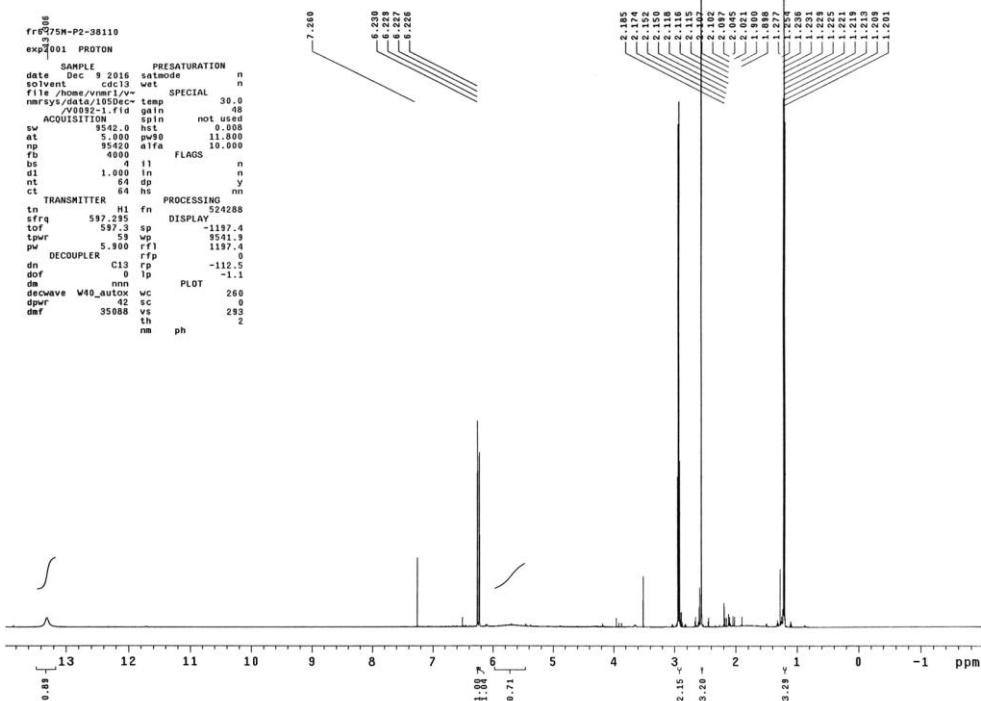
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**Figure S8.** HSQC spectrum of **1**



**Figure S9.** HMBC spectrum of **1**

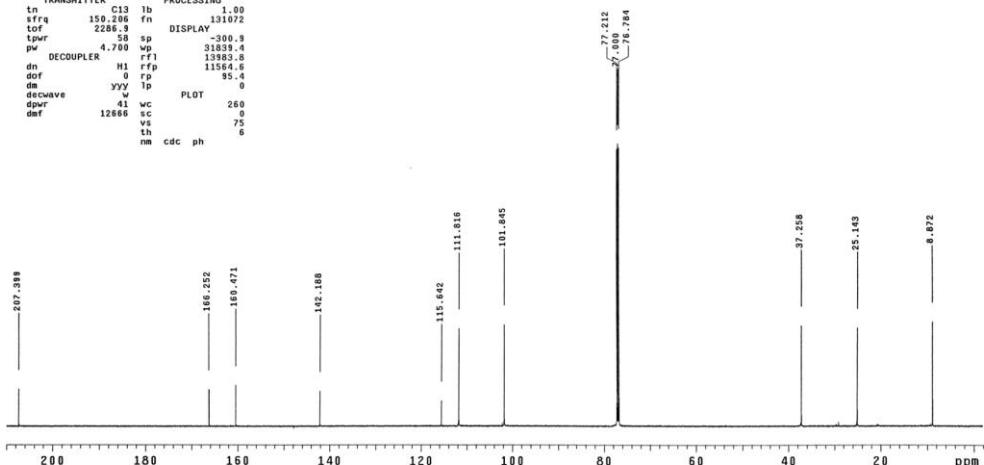


**Figure S10.**  $^1\text{H}$  NMR spectrum of **2** (600 MHz in  $\text{CDCl}_3$ )

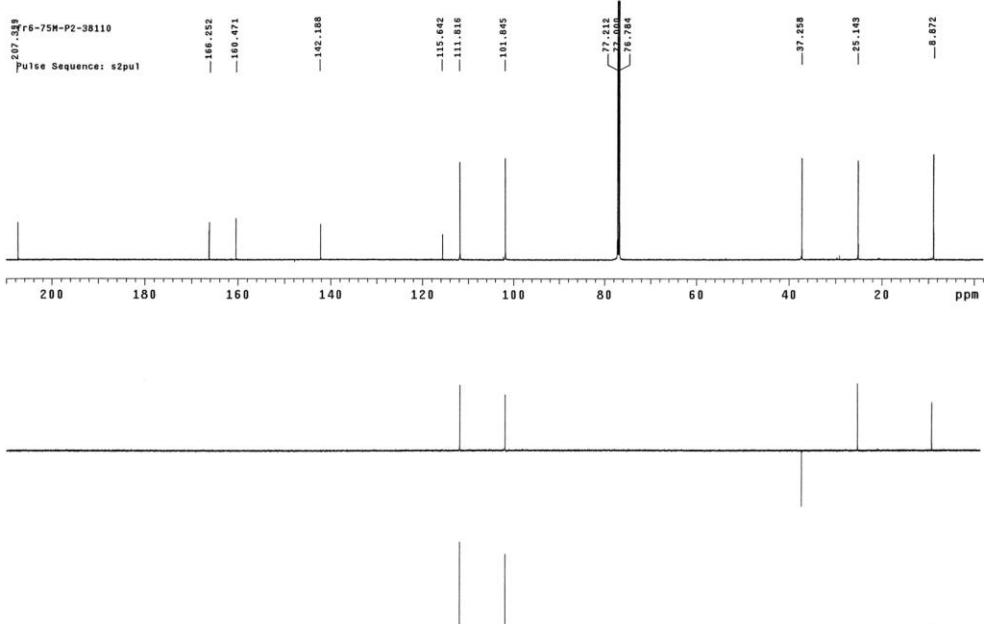
```

fr6-75M-P2-38110
exp2002 CARBON
      SAMPLE          PRESATURATION
      date Dec 9 2010 satmode    n
      solvent   cdc13 vetcoupling n
      file /home/vmervi/vet          SPECIAL
      nmrys/data/105sec< temp      30.0
      nmrys/data/105sec< f1dec    5.00
      ACQUISITION spin      not used
      sw       37875.8 hz      0.000
      al      0.000000      0.000
      np      65536 alfa      10.000
      fb      178000          FLAGS
      bs      8 t1      n
      d1      1,000 in      n
      nt      128000 dp      y
      ct      128000 hs      nn
      TRANSMITTER C13 lb      PROCESSING
      tn      C13 lb      1.00
      sfrf      151.000 fn      131072
      tof      2286.9 DISPLAY
      tpur      58 sp      -300.9
      pw      4.700 us      313.14
      DECOUPLER H1 rfp      13983.8
      dn      H1 rfp      11564.6
      off      0      95.4
      dm      VVY ip      0
      decwave      w      PLOT
      dpwr      41 wc      260
      dswr      128000 sc      0
      vs      75
      tc      6
      nm      cdc ph

```



**Figure S11.**  $^{13}\text{C}$  NMR spectrum of **2** (150 MHz in  $\text{CDCl}_3$ )



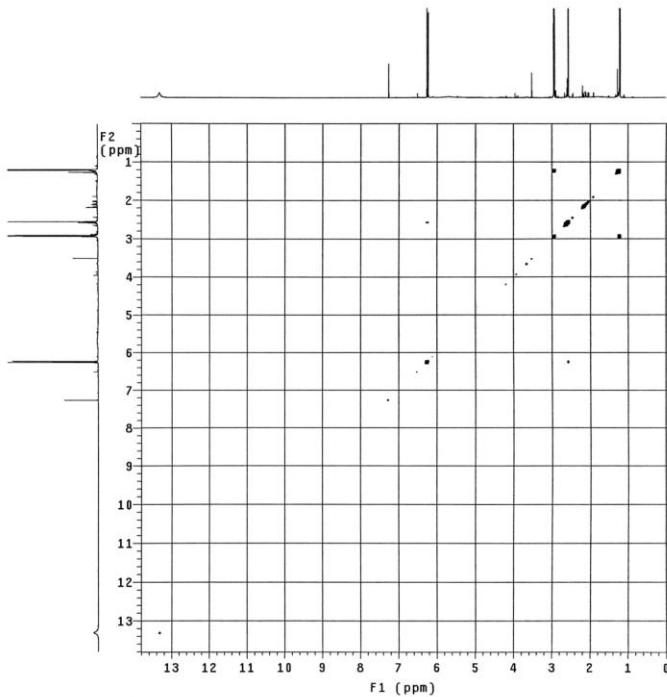
**Figure S12.** DEPT spectrum of **2** (150 MHz in  $\text{CDCl}_3$ )

```

fr6-75M-P2-38110
exp2004 gCOSY

SAMPLE           FLAGS      nn
date Dec 9 2016 hs          y
solvent        cdc13 sspp1   y
sample          hs1v1      5352
ACQUISITION    SPECIAL
sw      9542.0 temp 30.0
at      0.150      30.0
np      2862 spin  not used
fb      4000 f2 PROCESSING
st      32 f2      0.075
d1      1.000 sbs  not used
nt      32 fn      4096
d2      32 f1      0.075
PRESATURATION  f1 PROCESSING
sw1     9542.0 sb1      -0.017
n1      160 sb1      not used
d2      0 f1c1      1 p
satmode       n sp      DISPLAY
wetmode       n sp      -0.017
TRANSMITTER   wp      8242.1
tn      H1 sp1      0
sfrq    597.295 wp      8242.1
t0f     597.3 rfi1    1197.4
tpwr    50 rfi1    0
pw      11.800 rfi1    1197.4
GRADIENTS    4464 PLOT
g2v1e   0.001000 wc      140.0
Edratio  1.000 sc      5.0
getrau  0.000500 sc      140.0
DECOUPLER    sc2      5.0
dn      C13 vs      17
dm      mnn w      2
ai      cdc av

```



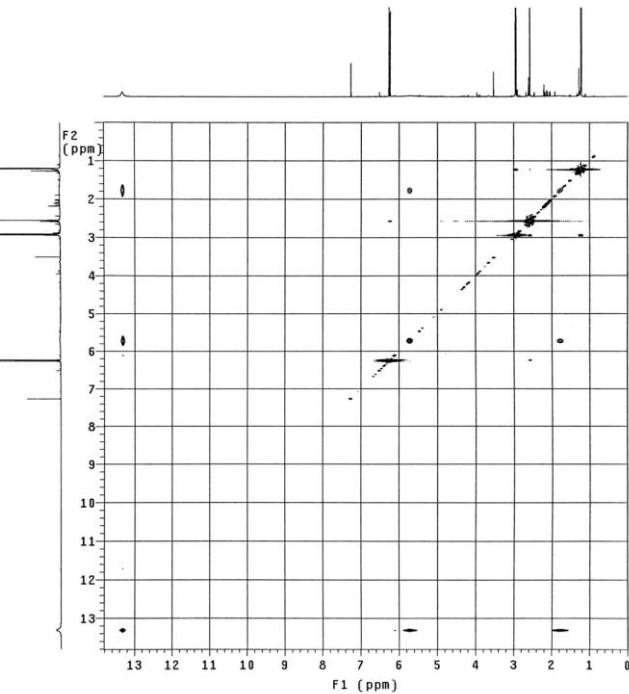
**Figure S13.**  $^1\text{H}$ - $^1\text{H}$  COSY spectrum of 2

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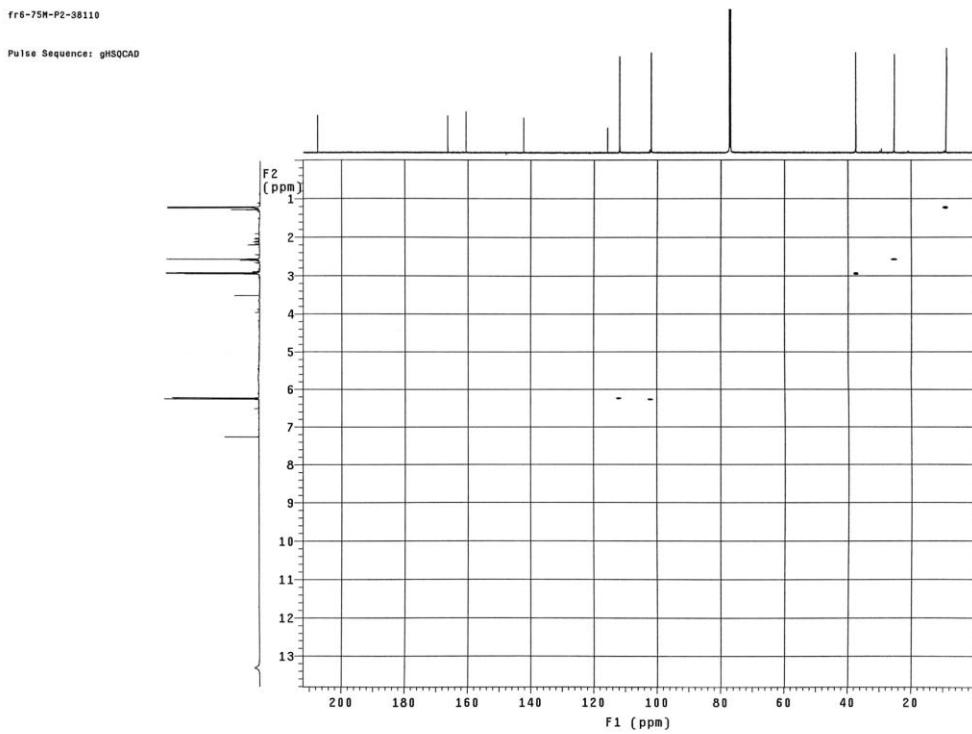
fr6-75M-P2-38110
exp2005 NOESY

SAMPLE           FLAGS      nn
date Dec 9 2016 hs          y
solvent        cdc13 sspp1   y
sample          hs1v1      5352
ACQUISITION    SPECIAL
sw      9542.0 temp 30.0
at      0.150      30.0
np      2862 spin  not used
fb      4000 f2 PROCESSING
st      32 f2      0.068
d1      1.000 sbs  not used
nt      32 gfs      not used
sw1     9542.0 f1 PROCESSING
n1      160 gfi1    0.019
TRANSMITTER   H1 f1c1      1 p
tn      H1 sp1      0
sfrq    597.295 fn1c1    4096
t0f     597.3 sp      DISPLAY
tpwr    50 sp      -0.0
pw      11.800 wp      8242.1
mixN   0.600 sp1      0
NOESY   0.600 sp1      0
PRESATURATION  rfi1    1197.4
satmode       n rfi1    1197.4
wetmode       n rfi1    1197.4
DECOUPLER    rfp1      0
dn      C13 vs      PLOT
dm      mnn w      2
ai      cdc ph

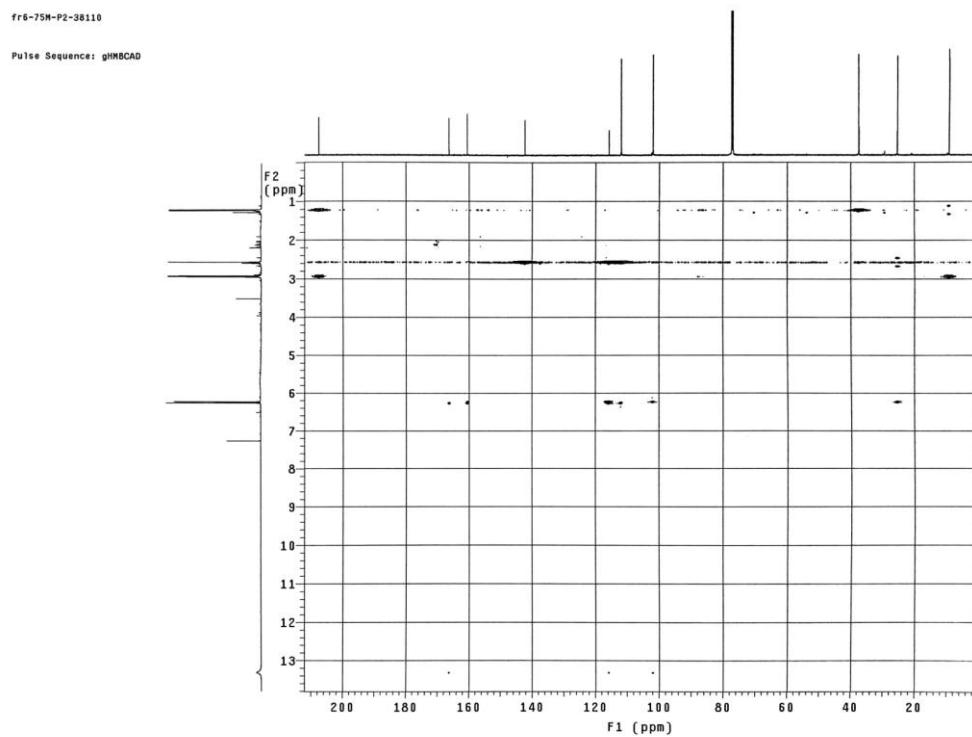
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**Figure S14.** NOESY spectrum of 2



**Figure S15.** HSQC spectrum of **2**



**Figure S16.** HMBC spectrum of **2**