## Supporting Information

## 3,5,7-Substituted Pyrazolo[4,3- $d$ ]pyrimidine Inhibitors of Cyclin-Dependent Kinases and

 Their Evaluation in Lymphoma ModelsRadek Jorda ${ }^{\text {a,\# }}$, Libor Havliček ${ }^{\text {b,\# }}$, Antonín Š̌turc ${ }^{\text {b }}$, Diana Tuškovác ${ }^{\text {c }}$ Lenka Daumovác, Mahmudul Alam ${ }^{\mathrm{c}}$, Jana Škerlovád ${ }^{\text {de }}$, Michaela Nekardová ${ }^{\text {d,f }}$, Miroslav Peřina ${ }^{\text {a }}$, Tomáš Pospíšil ${ }^{\text {a }}$, Jitka Širokáa ${ }^{\text {a }}$ Lubor Urbánek ${ }^{\text {a }}$, Petr Pachl ${ }^{\text {d }}$, Pavlína Řezáčová ${ }^{\text {d,e }}$, Miroslav Strnad ${ }^{\text {a }}$, Pavel Klener ${ }^{\text {c }}$, Vladimír Kryštof ${ }^{\text {a* }}$
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## Content

## 1. Computational Analysis

Supplementary Figure S1.
Supplementary Table 1.
Supplementary Figure S2.
Supplementary Figure S3.
Supplementary Figure S4.
Supplementary Figure S5.
2. Crystal parameters, data collection and refinement statistics

Supplementary Table 2.
3. Kinase selectivity profile

Supplementary Table 3.
4. Further in vitro data

Supplementary Table 4.
Supplementary Figure S6.
Supplementary Figure S7.
Supplementary Figure S8.
5. Physicochemical and pharmacological properties of $\mathbf{4 . 3 5}$

Pharmacokinetic analysis in mice
Supplementary Figure S9.
6. Further in vivo data

Supplementary Figure S10.
Supplementary Figure S11.
7. Additional Methods
8. NMR Spectra of Prepared Compounds
9. References

## 1. Computational Analysis

The theoretical study analyses the binding mode of compound $\mathbf{4 . 3 5}$ described in this work, and its nitrogen-containing pyrazolo[4,3- $d$ ]pyrimidine analog $\mathbf{5 i}$ which was described previously. ${ }^{1}$ We computationally investigated two conformers of each inhibitor. The conformers $\mathbf{4 . 3 5}$ calc and $\mathbf{5 i} \mathrm{i}_{\text {calc1 }}$ were identified by the computational procedure ${ }^{2}$ as the most stable geometries. For this purpose, we utilized the CR8 (PDB ID: 3DDP) ${ }^{3}$ molecule as the template. The $\mathbf{4 . 3 5}$ calc2 conformer is identical to the $\mathbf{4 . 3 5}$ geometry published here and $\mathbf{5 i}_{\text {calc2 }}$ was designed according to this conformation. As CDK2 model, we used the crystal structure 3DDQ which was computationally adjusted in our previously study ${ }^{2}$.


Supplementary Figure S1. Structures of CR8 (left), compound $\mathbf{5 i}$ (middle) and $\mathbf{4 . 3 5}$ (right). To assess the $\Delta \mathrm{G}^{\prime}$ int contributions of the inhibitor parts, the inhibitors were fragmented into four parts (the scaffold and three substituents).

We used computational methods and methodology validated by a representative dataset of CDK2 inhibitors. ${ }^{2}$ The approach is based on the hybrid quantum mechanics/semiempirical quantum mechanics (QM/SQM) method, ${ }^{4}$ which employs the DFT method with an empirical dispersion for QM and the PM6 method ${ }^{5}$ for SQM part. The solvent effect is described by the continuum solvation model COSMO $^{6}$ at the SQM level for the whole system. The protein-inhibitor binding affinities are expressed as the computational interaction "free" energies $\Delta \mathrm{G}^{\prime}$ int (in $\mathrm{kcal} \mathrm{mol}^{-1}$ ) of the protein-inhibitor complexes. They are fundamental for the description of protein-inhibitor interactions.

The fragmentation of the inhibitors enables the assessment of the $\Delta \mathrm{G}^{\prime}{ }_{\text {int }}$ contributions of the inhibitor parts (molecular fragments) in the complex with the whole protein. The inhibitors taken from the optimized protein-inhibitor complex were divided into four parts
(the scaffold and thee substituents), and the $\Delta \mathrm{G}^{\prime}{ }_{\text {int }}$ values are described at the $\mathrm{QM} / \mathrm{SQM}$ level as well. ${ }^{2}$

The computationally optimized protein-inhibitor complexes have the following binding modes. The inhibitors are bound by three hydrogen bonds between the scaffold and the protein hinge region (carbonyl of Leu83, backbone NH of Leu83, carbonyl of Glu81), similar to the binding modes of CR8. The compounds also create a hydrogen bond between Lys89 and the 4-(2-pyridyl)benzylamine moiety at position 7. The compounds are stabilized by dispersion interactions between the 3-isopropyl group and Phe80 as well. The inhibitors make other hydrogen bonds between the substituents at position 5 and the residues situated at the edge of the active site. The 2 -aminoethylthio moiety of $\mathbf{4 . 3 5}_{\text {calc } 1}$ is bound to carbonyl of Gln131 and the terminal amide of Asn132. The 2-aminoethylamino moiety of $\mathbf{5} \mathbf{i}_{\text {calc }}$ forms the hydrogen bonds with the same residues. An analogous 4-hydroxybutylamine group of CR8 is bound by two hydrogen bonds with the backbone NH and Glu12 carbonyl (Supplementary Figures S2 and S3).

The evaluation of the computational data (Supplementary Table 1) shows that the interaction between the protonated amino group of Lys89 and the free electron pair of the pyridine nitrogen substantially contributes to the total $\Delta \mathrm{G}^{\prime}{ }_{\text {int }}$ values of all the complexes. The $\Delta \mathrm{G}^{\prime}$ int of $\mathbf{4 . 3 5}_{\text {calc }}$ indicates that this inhibitor is the best. The $\Delta \mathrm{G}^{\prime}{ }_{\text {int }}$ contributions of the scaffolds denote the larger values of the pyrazolo[4,3-d]pyrimidine core ( $\mathbf{4 . 3 5}_{\text {calc } 1}$ and $\mathbf{5} \mathrm{i}_{\text {calc1 }}$ ) compared with the purine core (CR8), in line with our previous report. ${ }^{2}$ Importantly, the $\Delta \mathrm{G}^{\prime}$ int contribution of the 2 -aminoethylthio moiety of $\mathbf{4 . 3 5}_{\text {calc }}$ is significantly higher than that of the 2-aminoethylamino of $\mathbf{5 i}_{\text {calc } 1}$. In addition, the total $\Delta \mathrm{G}^{\prime}{ }_{\text {int }}$ of $\mathbf{4 . 3 5}_{\text {calcl }}$ (as well as the sum of the $\Delta G^{\prime}$ int of its fragments) indicates higher binging affinity than $\mathbf{5 i}_{\text {calcl }}$. Thus, the computational results indicate that the replacement of the alkylamino group at the 5-position of the heterocycle by the 5 -alkylthio group is completely relevant.

It should be noted that the computationally predicted conformer $\mathbf{4 . 3 5}_{\text {calc }}$ differs in the geometry of the 2-aminoethylthio moiety at position 5 from the conformer detected in the crystal structure. This substituent in $\mathbf{4 . 3 5}_{\text {calc }}$ and $\mathbf{5 i}$ calc is similarly oriented to the analogous moiety of CR8 at position 5 (Supplementary Figures S2, S3 and S5). In contrast, in the crystal structure, the substituent is oriented in the opposite direction. We evaluated the effect of this orientation in the $\mathbf{4 . 3 5}_{\text {calc2 }}$ and $\mathbf{5 i} \mathrm{i}_{\text {calc2 }}$ conformers. $\mathbf{4 . 3 5}_{\text {calc2 }}$ corresponds to the crystal structure and $\mathbf{5} \mathbf{i}_{\text {calc2 }}$ was modeled similarly. The amino group of the moiety at position 5 of $\mathbf{4 . 3 5}_{\text {calc2 }}$ is bound to the carboxylate of Asp86 and the backbone carbonyl of Gln131, and the
amino group of the moiety at position 5 of $\mathbf{5 i}_{\text {calcı }}$ forms the hydrogen bond with the backbone carbonyl of Gln131 (Supplementary Figures S2 and S3). The comparison of the total $\Delta \mathrm{G}^{\prime}{ }_{\text {int }}$ values of these conformers shows lower binding affinity than those of $\mathbf{4 . 3 5}_{\text {calc }}$ and $\mathbf{5 i}_{\text {calc }}$. The $\Delta \mathrm{G}^{\prime}$ int contributions of the moiety at position 5 are also lower than $\mathbf{4 . 3 5}_{\text {calc } 1}$ and $\mathbf{5 i}_{\text {calc }}$; the $5 \mathbf{i}_{\text {calc } 2}$ moiety even results in repulsion (Supplementary Table 1). We conclude that although the $\mathbf{4 . 3 5}_{\text {calc } 1}$ conformer is more stable than the $\mathbf{4 . 3 5}_{\text {calc2 }}$ conformer, the $\mathbf{4 . 3 5}_{\text {calc2 }}$ conformer (in complex with the protein) is favorable because the protein-inhibitor complex can be substantially stabilized by two crystal water molecule chains (Figure 2, Supplementary Figures S4 and S5). ${ }^{7}$

Supplementary Table 1. $\Delta \mathrm{G}^{\prime}{ }_{\text {int }}$ values of the protein-inhibitor complexes and $\Delta \mathrm{G}^{\prime}{ }_{\text {int }}$ of the protein-fragment complexes that describe the contributions of the molecular parts (the scaffold and the substituents, see Supplementary Figure 1) to the binding affinity.

| Inhibitor | $\Delta \mathrm{G}^{\text {int }}{ }^{\text {in }} \mathrm{kcal} \mathrm{mol}^{-1}$ |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Fragments of Inhibitors |  |  |  | Suma of Fragments | CDK2inhibitor complex |
|  | scaffold | R1 | R2 | R3 |  |  |
| CR8 | -11.69 | -17.92 | -8.24 | -3.81 | -41.66 | -44.37 |
| $4.35{ }_{\text {calc } 1}$ | -15.25 | -18.28 | -7.41 | -3.20 | -44.15 | -47.15 |
| $4.35{ }_{\text {calc2 }}$ | -13.74 | -17.43 | -5.34 | -3.55 | -40.07 | -44.19 |
| $5 \mathbf{5 i a l c}$ | -14.62 | -17.99 | -4.52 | -3.11 | -40.24 | -44.62 |
| $55_{\text {calc2 }}$ | -18.40 | -17.94 | 0.20 | -4.55 | -40.70 | -43.57 |



Supplementary Figure S2. The computationally optimized complexes of CDK2 with the inhibitors 4.35 and CR8. Only the key residues are depicted. The geometries of the $\mathbf{4 . 3 5}$ conformers ( $\mathbf{4 . 3 5}_{\text {calc1 }}-$ salmon color, $\mathbf{4 . 3 5}_{\text {calc2 }}$ - aquamarine color; $\mathrm{IC}_{50}=0.002$ ) in comparison with CR8 (gray color; $\mathrm{IC}_{50}=0.062$ ) are illustrated.


Supplementary Figure S3. The computationally optimized complexes of CDK2 with the inhibitors 5i and CR8. Only the key residues are depicted. The geometries of the 5i conformers ( $\mathbf{5 i} \mathrm{i}_{\text {calc } 1}$ - salmon color, $\mathbf{5} \mathbf{i}_{\text {calc } 2}$ - aquamarine color; $\mathrm{IC}_{50}=0.018$ ) in comparison with CR8 (gray color; $\mathrm{IC}_{50}=0.062$ ) are illustrated.


Supplementary Figure S4. The nonoptimized complex of CDK2 with 4.35 (lime color) compared with the computed complex of CDK2 with the conformer $\mathbf{4 . 3 5}$ calc1 (salmon color). Only the key residues are depicted. This demonstrates how the less stable $\mathbf{4 . 3 5}$ conformer is substantially stabilized by the hydrogen bonds with two crystal water chains. The geometry of hydrogens of water molecules was modeled in PyMol. ${ }^{8}$


Supplementary Figure S5. The nonoptimized complex of CDK2 with 4.35 (lime color) compared with the nonoptimized complex of CDK2 with CR8 (blue-white color). Only the key residues are depicted. The inhibitors stabilized by hydrogen bonds with the crystal water chains are shown.

## 2. Crystal parameters

Supplementary Table 2. Crystal parameters, data collection and refinement statistics

## Data collection statistics

Space group
Cell parameters ( $\AA$; ${ }^{\circ}$ )
Number of molecules in
AU
Wavelength (Å)
Resolution (Å)
Number of unique
reflections
Multiplicity
Completeness (\%)
$\mathrm{R}_{\text {meas }}{ }^{\mathrm{a}}$
CC (1/2)
Average $I / \sigma(I)$
Wilson $B\left(\AA^{2}\right)^{b}$

C2221
$\mathrm{a}=71.0, \mathrm{~b}=112.6, \mathrm{c}=$
159.7; $\alpha=\beta=\gamma=90$

1
0.918
48.01-2.15 (2.28-2.15)
$34,773(5,088)$
6.2 (4.3)
98.3 (90.3)
21.2 (166.4)
99.4 (29.8)

Refinement statistics
Resolution range (A)
No. of reflections in
working set
No. of reflections in
test set
$R$ value (\%) ${ }^{\text {c }}$
$8.0(0.9)$
38.4
$R_{\text {free }}$ value (\%) ${ }^{\text {d }} 24.0$
RMSD bond length ( $\AA$ )
0.013

RMSD angle ( ${ }^{\circ}$ )
1.619

Number of atoms in AU 4,638
Number of protein atoms
in AU
4,394
Number of water
molecules in AU
160
Mean B value $\left(\AA^{2}\right) \quad 40.2$
Ramachandran plot statistics ${ }^{e}$
Residues in favored
regions (\%)
Residues in allowed
regions (\%)
${ }^{{ }^{2}} \mathrm{R}_{\text {meas }}$ defined in reference. ${ }^{\text {b }}{ }^{\text {b Wilson }} \mathrm{B}$ by Sfcheck program from CCP4 suite. ${ }^{10}{ }^{c}$ R-value $=\left|\left|F_{0}\right|-\left|F_{C}\right|\right| /\left|F_{0}\right|$, where $F_{0}$ and $F_{C}$ are the observed and calculated structure factors, respectively. ${ }^{d} R_{\text {free }}$ is equivalent to the $R$-value but is calculated for $5 \%$ of the reflections chosen at random and omitted from the refinement process. ${ }^{11}{ }^{e}$ As determined by MolProbity. ${ }^{12}$

## 3. Kinase selectivity profile

Supplementary Table 3. Kinase selectivity profile against a panel of 50 human kinases. The profiling was performed in duplicate at a compound concentration of $1 \mu \mathrm{M}$.

| kinase | Residual <br> activity (\%) | kinase | Residual <br> activity (\%) |
| :--- | :--- | :--- | :--- |
| AMPK (hum) | 34 | NEK6 | 89 |
| Aurora B | 55 | p38a MAPK | 92 |
| BTK | 95 | PAK4 | 33 |
| CAMK1 | 35 | PDK1 | 106 |
| CAMKKb | 58 | PIM1 | 116 |
| CK18 | 11 | PKA | 92 |
| CK2 | 75 | PKBa | 124 |
| DYRK1A | 63 | PKCa | 80 |
| EF2K | 99 | PKD1 | 44 |
| EPH-A2 | 102 | PLK1 | 94 |
| GSK3b | 99 | PRK2 | 106 |
| HER4 | 100 | RIPK2 | 103 |
| HIPK2 | 96 | ROCK 2 | 111 |
| CHK2 | 18 | RSK1 | 35 |
| IGF-1R | 116 | S6K1 | 93 |
| IRAK4 | 131 | SGK1 | 107 |
| JAK2 | 111 | SmMLCK | 45 |
| JNK1 | 94 | Src | 74 |
| Lck | 82 | SRPK1 | 86 |
| LKB1 | 86 | SYK | 117 |
| MARK3 | 32 | TAK1 | 97 |
| MKK1 | 48 | TBK1 | 98 |
| MLK3 | 97 | TrkA | 91 |
| MSK1 | 78 | TTK | 78 |
| MST2 | 96 | VEG-FR | 114 |

## 4. Additional in vitro data

Supplementary Table 4. In vitro testing of $\mathbf{4 . 3 5}$ on a panel of cancer cell lines (National Cancer Institute Developmental Therapeutics Program).

| Log10 Concentration |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Time |  |  | Mean | Optical | Densitic |  |  |  | arcent | rowth |  |  |  |  |  |  |
| Panelicell Line | Zero | Crs | -9.0 | $-8.0$ | $-7.0$ | -60 | -5.0 | $-9.0$ | $-8.0$ | -7.0 | -6.0 | $-5.0$ | Gats |  | TGt |  | LC50 |
| Leukennia |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| CCRF-CEM | 0.778 | 3.313 | 3.365 | 2.033 | 0.872 | 0856 | 0.838 | 102 | 49 | 4 | 3 | 2 | 9788-9 |  | $100 \mathrm{E}-5$ | $\geqslant$ | $1,00 \mathrm{E}-5$ |
| HL-60(TB) | 1.240 | 3.162 | 3.334 | 1.645 | 1.097 | 0.962 | 0.794 | 109 | 21 | $-12$ | 22 | -36 | 469 E .9 |  | 4.43E-8 |  | $1.00 \mathrm{E}-5$ |
| K-562 | 0.312 | 2.191 | 2.423 | 1.474 | 0.726 | 0.540 | 0.480 | 112 | 62 | 22 | 12 | 9 | $198 \mathrm{E}-8$ |  | 1.00E-5 |  | 100E-5 |
| MOLT-4 | 0.907 | 3.294 | 3.358 | -1.571 | 0.940 | 0.865 | 0.860 | 103 | 28 | 1 | -5 | -5 | $5.05 \mathrm{E}-9$ |  | $169 \mathrm{E}-7$ |  | $1.00 \mathrm{E}-5$ |
| RPMM-8226 | 0.943 | 3.133 | 3.120 | 2.301 | 0.898 | 0.898 | 0.796 | 99 | 62 | -5 | -5 | $-16$ | $151 \mathrm{E}-8$ |  | 8 47E-8 |  | $1.00 \mathrm{E}-5$ |
| SR | 0.298 | 1.456 | 1.512 | 0.925 | 0.453 | 0.417 | 0.411 | 106 | 54. | 13 | 10 | 10 | 1.26E-8 |  | 1.00E-5 | $\geqslant$ | 1.00E-5 |
| Non-Small Cell Lung Cancer |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| A549/ATCC | 0.434 | 2214 | 2.285 | 1,954 | 0.653 | 0.699 | 0.622 | 104 | 85 | 13 | 15 | 11 | $306 E-8$ | $>$ | 100E-5 | $>$ | 1.00E-5 |
| EKVX | 1.041 | 2888 | 2.679 | 2.587 | 1:390 | 1.051 | 1.017 | 89 | 84 | 18 | 1 | $-2$ | $328 \mathrm{E}-8$ |  | $1.52 \mathrm{E}-6$ |  | $1.00 \mathrm{E}-5$ |
| HOP 62 | 0.905 | 2.433 | 2230 | 2.049 | 1.083 | 0.877 | 0.788 | 87 | 75 | 12 | -3 | . 13 | 247 E 8 |  | 6.04 E .7 |  | 1.00E-5 |
| HOP-92 | 1.115 | 1.768 | 1.753 | 1.540 | 1.099 | 1.058 | 1.095 | 98 | 65 | -1 | -4 | -2 | 1.69E-8 |  | 9.50E-8 |  | 1.00E-5 |
| $\mathrm{NCH}-\mathrm{H} 226$ | 0.953 | 2.553 | 2440 | 2.487 | 1.276 | 0.981 | 0.971 | 93 | 96 | 20 | 2 | 1 | $4.04 \mathrm{E}-8$ |  | 1.00E-5 |  | $1.00 \mathrm{E}-5$ |
| $\mathrm{NCH}-\mathrm{H} 23$ | 0.661 | 2293 | 2.203 | 1.796 | 0.820 | 0.654 | 0.592 | 94 | 70 | 10 | -1 | -10 | $2.12 \mathrm{E}-8$ |  | 797E.7 |  | 1.00 E 5 |
| NCH H 322 M | 0.782 | 2.031 | 2.002 | 1.536 | 0.924 | 0.782 | 0.827 | 98 | 60 | 11 | $-3$ | 4 | $1.63 \mathrm{E}-8$ |  |  |  | 1,00E-5 |
| $\mathrm{NCl}-\mathrm{H} 460$ | 0.245 | 2.430 | 2.541 | 2.589 | 0.435 | 0.352 | 0.219 | 105 | 107 | 9 | 5 | -11 | $3.81 \mathrm{E}-8$ |  | 205E-6 |  | $1.00 \mathrm{E}-5$ |
| NCl-H522 | 0.929 | 2408 | 2393 | 0.979 | 0.955 | 0.849 | 0840 | 99 | 3 | 2 | -9 | $-10$ | 326E-9 |  | $147 \mathrm{E}-7$ | > | $1.00 \mathrm{E}-5$ |
|  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| COLO 205 | 0.638 | 2249 | 2.175 | 1629 | 0.387 | 0.286 | 0.324 | 95 | 62 | . 39 | . 55 | 49 | $130 \mathrm{E}-8$ |  | 4.07E-8 |  |  |
| HCC-2998 | 0.800 | 2837 | 2808 | 2.253 | 1.163 | 0.978 | 0.887 | 99 | 71 | 18 | 9 | 4 | $250 \mathrm{E}-8$ |  | 1.00E-5 | $\geqslant$ | 1.00E-5 |
| HCT-116 | 0.274 | 2.476 | 2.149 | 1.938 | 0.406 | 0.395 | 0.379 | 85 | 76 | 6 | 5 | 5 | $233 \mathrm{E}-8$ | $>$ | 1.00 E 5 |  | $1.00 \mathrm{E}-5$ |
| HCT-15 | 0.347 | 2139 | 2.081 | 2.155 | 2.056 | 1.074 | 0.420 | 97 | 101 | 96 | 41 | 4 | $6.72 \mathrm{E}-7$ | > | 1.00E-5 | > | 1,00E-5 |
| HT29 | 0.322 | 2030 | 2.061 | 0.800 | 0.405 | 0.368 | 0.324 | 102 | 26 | 5 | 3 |  | $5.03 \mathrm{E}-9$ | ? | 100E-5 |  | $1.00 \mathrm{E}-5$ |
| KM12 | 0.457 | 2664 | 2.632 | 1.840 | 0,695 | 0.644 | 0.554 | 99 | 63 | 10 | 8 | 4 | $174 \mathrm{E}-8$ |  | $\geq 1.00 \mathrm{E}-5$ | > | $1.00 \mathrm{E}-5$ |
| SW-620 | 0.386 | 2356 | 2.377 | 1.441 | 0,564 | 0.500 | 0.391 | 101 | 54 | 9 | 6 |  | $120 \mathrm{E}-8$ | $\bigcirc$ | 1.00E-5 | $>$ | 1.00E-5 |
| CNS Cancer |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| SF-268 | 0.646 | 2043 | 2.006 | 1.214 | 0.783 | 0.734 | 0.639 | 97 | 41 | 10 | 6 | -1 | 6.84E-9 |  | 7.12E-6 | $>$ | $1.00 \mathrm{E}-5$ |
| SF-295 | 0.819 | 2532 | 2318 | 2.368 | 1.200 | 0.788 | 0.733 | 97 | 90 | 22 | -4 | -11 | 391E-8 |  | $7.12 \mathrm{E}-7$ |  | 1.00E-5 |
| SF-539 | 1.125 | 3.054 | 2.960 | 3.059 | 1.115 | 0.693 | 0.433 | 96 | 100 |  | -38 | -62 | 3.14E-8 |  | 979E-8 |  | $3.17 \mathrm{E}-6$ |
| SNE-19 | 0.980 | 2.703 | 2.643 | 2.047 | 1.185 | 1.134 | 1.102 | 97 | 62 | 12 | 9 | 7 | $1.73 \mathrm{E}-8$ | $\geqslant$ | -100E-5 | $\geqslant$ | 1.00E-5 |
| SNB-75 | 0.929 | 1.736 | 1571 | 1.261 | 0.577 | 0.260 | 0.077 | 80 | 41 | . 38 | - 72 | -92 | 5.87E-9 |  | $331 \mathrm{E}-8$ |  | $2.26 \mathrm{E}-7$ |
| U251 | 0.476 | 2238 | 2.228 | 1.872 | 0.720 | 0.680 | 0.622 | 99 | 79 | 14 | 12 | 8 | 280E-8 | $\cdots$ | - 100E-5 | $\bigcirc$ | 1.00E-5 |
| Melanoma |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| LOX MY/ | 0.174 | 1.338 | 1284 | 0.573 | 0.209 | 0.234 | 0.162 | 95 | 34 | 3 | 5 | 77 | 5.53 E .9 |  | $262 \mathrm{E}-6$ | $>$ | 1.00E 5 |
| MALME-3M | 0.845 | 1.579 | 1.517 | 1339 | 0.377 | 0.462 | 0.276 | 92 | 67 | -55 | -45 | -67 | $138 \mathrm{E}-8$ |  | $3.54 \mathrm{E}-8$ |  |  |
| M14 | 0.439 | 1716 | 1.605 | 1213 | 0.502 | 0.339 | 0.129 | 91 | 61 | 5 | -23 | -71 | $155 \mathrm{E}-8$ |  | $151 \mathrm{E}-7$ |  | $370 \mathrm{E}-6$ |
| MDA-MB-435 | 0.479 | 2411 | 2.336 | 0.911 | 0.485 | 0.519 | 0.197 | 96 | 22 |  | 2 | -69 | $4.22 \mathrm{E}-9$ |  | 1.00E-6 |  | 7.13E-6 |
| SK-MEL-2 | 1.025 | 2471 | 2.535 | 2.420 | 1.039 | 0.581 | 0.345 | 104 | 96 | 1 | -43 | -66 | $3.06 \mathrm{E}-8$ |  | $1.05 \mathrm{E}-7$ |  | 1.94E-6 |
| SK-MEL-28 | 0.800 | 2572 | 2.508 | 2.022 | 1.017 | 0.647 | 0552 | 96 | 67 | 8 | -27 | -37 | $1.97 \mathrm{E}-8$ |  | 1.71E-7 | $>$ | 1.00E-5 |
| SK-MEL-5 | 0.795 | 3.278 | 3.264 | 3.195 | 0.775 | 1.228 | 0.653 | 99 | 97 | -3 | 17 | -18 | $295 E-8$ |  |  | $\cdots$ | 1.00E-5 |
| UACC-257 | 0.955 | 2200 | 2.197 | 2.183 | 1,267 | 1.007 | 1.017 | 94 | 93 | 24 | 8 | 5 | 4.14E-8 | $>$ | $>100 \mathrm{E}-5$ | > | 100E-5 |
| UACC-62 | 0.894 | 3.023 | 2.974 | 2.698 | 0.549 | 0.417 | 0.280 | 98 | 85 | -39 | $-53$ | -69 | $1.91 \mathrm{E}-8$ |  | 4.86E-8 |  | 5.87E-7 |
| Ovarian Cancer |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| IGROV1 | 0.693 | 2284 | 2.254 | 1.830 | 0.804 | 0.822 | 0.725 | 98 | 71 | 7 | 8 | 2 | 2.15E-8 | $\geqslant$ | 100E-5 | $\geqslant$ | $1.00 \mathrm{E}-5$ |
| OVCAR-3 | 0.434 | 1.602 | 1.653 | 1213 | 0.509 | 0.489 | 0.469 | 104 | 67. | 6 | 5 | 3 | $1.89 \mathrm{E}-8$ |  | 1.00E-5 |  | 1.00E-5 |
| OVCAR-A | 0700 | 1.565 | 1.491 | 1024 | 0.740 | 0.678 | 0525 | 91 | 37 | 5 | -3 | -25 | 585E-9 |  | $394 \mathrm{E}-7$ | $>$ | 1.00E-5 |
| OUCAR-5 | 0.681 | 1.905 | 1.854 | 1.889 | 1.005 | 0.950 | 0.899 | 96 | 99 | 26 | 22 | 18 | 4. $72 \mathrm{E}-8$ |  | 1.00E-5 | $>$ | $1.00 \mathrm{E}-5$ |
| OVCAR-8 | 0.675 | 2598 | 2.648 | 2.416 | 1.091 | 0.909 | 0.917 | 103 | 91 | 22 | 12 | 13 | 387E-8 |  | 1.00E-5 |  | 1,00E-5 |
| NCUADR-RES | 0.562 | 2.124 | 2.100 | 2.136 | 2.068 | 1.836 | 1.112 | 98 | 101 | 96 | 82 | 36 | 4.87E-6 |  | 1.00E-5 |  | $1.00 \mathrm{E}-5$ |
| SK-OV-3 | 0.888 | 2227 | 2.133 | 2.141 | 1.224 | 1.084 | 1.136 | 93 | 94 | 25 | 15 | 19 | $433 \mathrm{E}-8$ |  | $1.00 \mathrm{E}-5$ | $>$ | 1.00E-5 |
| Renal Cancer |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| 785-0 | 0.886 | 2620 | 2.438 | 2491 | 1.913 | 0.891 | 0.713 | 90 | 93 | 59 |  | -20 | $1.43 \mathrm{E}-7$ |  | 1.03E-6 | $\geqslant$ | $1.00 \mathrm{E}-5$ |
| A498 | 1.496 | 2523 | 2.498 | 2.596 | 1,352 | 0.870 | 0.630 | 98 | 106 | - 10 | 42 | -58 | 306E-8 |  | 8. $26 \mathrm{E}-8$ |  | 3.22 E 6 |
| ACHN | 0.462 | 2.009 | 1.944 | 2032 | $1: 832$ | 0.465 | 0.536 | 96 | 102 | 69 |  | 5 | 273E-7 |  | -1.00E-5 |  | $1.00 \mathrm{E}-5$ |
| CAKK-1 | 0.593 | 3040 | 2.928 | 2.957 | 2.910 | 2430 | 0.868 | 95 | 97 | 95 | 75 | 11 | $247 \mathrm{E}-6$ | > | $>1.00 \mathrm{E}-5$ | $>$ | 1.00 E 5 |
| RXX 393 | 0.632 | 1304 | 1.274 | 1.121 | 0.459 | 0.337 | 0.279 | 96 | 73 | $-27$ | -47 | -56 | $1.69 \mathrm{E}-8$ |  | $5.32 \mathrm{E}-8$ |  | $227 \mathrm{E}-6$ |
| SN12C | 1.145 | 3322 | 3.361 | 2.653 | 1.540 | 1.433 | 1.253 | 102 | 69 | 18 | 13 | 5 | 238 E 8 |  | 100E-5 |  | $1.00 \mathrm{E}-5$ |
| U0-31 | 0.822 | 2371 | 2290 | 2.265 | 2.078 | 0.539 | 0.650 | 95 | 93 | 81 | $-34$ | -21 | $1.86 \mathrm{E}-7$ |  | 503E-7 | $>$ | 1.00E-5 |
| Prostate Cancer |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| PC 3 | 0.504 | 1789 | 1.793 | 1501 | 0.638 | 0.547 | 0.494 | 100 | 78 | 10 | 3 | 2 | 257 E 8 |  | 4.11E. 6 | $>$ | 1.00E-5 |
| DU-145 | 0.490 | 1.963 | 2.001 | 1,883 | 0.770 | 0.610 | 0.568 | 103 | 95 | 19 | 8 | 5 | 3.09E-8 | $\bigcirc$ | -1.00E-5 | $>$ | 1.00E-5 |
| Breast Cancer |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| MCF7 | 0.427 | 2233 | 2.073 | 0.841 | 0,567 | 0.473 | 0.408 | 91 | 23 | 8 | 3 | -5 | 4.01E-9 |  | 228E-6 | > | $1.00 \mathrm{E}-5$ |
| MDA-ME-231/ATCC | 0.877 | 1.930 | 1.951 | 1,160 | 0.929 | 0.781 | 0.756 | 102 | 27 | 5 | -11 | -14 | 4 92E-9 |  | $205 \mathrm{E}-7$ | > | $1.00 \mathrm{E}-5$ |
| HS 6781 | 1.166 | 2116 | 2.068 | 1729 | 1.240 | 1.209 | 1.094 | 95 | 59 | 8 | 4 | -6 | $151 \mathrm{E}-8$ |  | 262E-6 | 3 | $1.00 \mathrm{E}-5$ |
| BT-549 | 1.282 | 2428 | 2.292 | 2.334 | 1391 | 0.868 | 0.488 | 88 | 92 | 9 | 32 | -62 | $322 \mathrm{E}-8$ |  | $1.68 \mathrm{E}-7$ |  | $395 \mathrm{E}-6$ |
| T-47D | 1.047 | 2.069 | 1.880 | 1.718 | 1.103 | 1.063 | 1.058 | 82 | 66 | 5 | 2 | 1 | $182 \mathrm{E}-8$ | $>$ | $\rightarrow 1.00 \mathrm{E}-5$ | $\geqslant$ | 1,00E-5 |
| MDA MB-468 | 0.746 | 1.455 | 1459 | 0.776 | 0.473 | 0.436 | 0.470 | 101 | 4 | -37 | 42 | -37 | $335 \mathrm{E}-9$ |  | $127 \mathrm{E}-8$ |  | $1.00 \mathrm{E}-5$ |



Supplementary Figure S6: Effect of $\mathbf{4 . 3 5}$ on apoptosis in lymphoma cell lines. Annexin V/PI assay was carried out as described in Methods.


Supplementary Figure S7: Fluorometric assay of caspase-3 and caspase-7 activity in lysates prepared from treated cells. The activities were measured in the presence of the fluorogenic substrate Ac-DEVD-AMC (gray columns) or the fluorogenic caspase inhibitor Ac-DEVDCHO (white columns) as a control and normalized to an untreated control.


Supplementary Figure S8: Induction of apoptosis in UPF1H and MAVER-1 cancer cells treated with pyrazolo[4,3- $d$ ] pyrimidine $\mathbf{4 . 3 5}$ for 24 h . Actin level is included as a loading control.

## 5. Physicochemical and pharmacological properties of $\mathbf{4 . 3 5}$

| Aqueous solubility <br> $(200 \mu \mathrm{M}$ in PBS, pH 7.4$)$ | $91 \%$ |
| :--- | :--- |
| Aqueous solubility <br> $(200 \mu \mathrm{M}$ in simulated intestinal fluid $)$ | $91 \%$ |
| Aqueous solubility <br> $(200 \mu \mathrm{M}$ in simulated gastric fluid) | $91 \%$ |
| Plasma protein binding <br> $\left(10 \mu \mathrm{M}, 4\right.$ h at $\left.37{ }^{\circ} \mathrm{C}\right)$ | $94 \%$ |
| In vitro absorption Caco-2 <br> (A-B permeability @ $10 \mu \mathrm{M})$ | $0.0\left(10^{-6} \mathrm{~cm} / \mathrm{s}\right)$ |
| In vitro absorption Caco-2 <br> (B-A permeability @ $10 \mu \mathrm{M})$ | $15.8\left(10^{-6} \mathrm{~cm} / \mathrm{s}\right)$ |
| Intrinsic clearance | 0 min |
| $(0.1 \mu \mathrm{M}$ with human liver microsomes) | 15 min |
|  | 30 min |
|  | 45 min |
|  | 60 min |

Physicochemical and pharmacological properties were determined by Eurofin Panlabs.
Aqueous solubility. The solubility was determined by comparing the peak area of the compound standard solution ( $200 \mu \mathrm{M}$, methanol/water, $60 / 40, \mathrm{v} / \mathrm{v}$ ) with the peak area of the compound in a buffer sample. In addition, chromatographic purity (\%) was defined as the peak area of the principal peak relative to the total integrated peak area in the HPLC chromatogram of the calibration standard.

Plasma protein binding. The peak areas of the test compound in the buffer and test samples were used to calculate percent binding.

Caco-2 permeability. The apparent permeability coefficient (Papp) of the test compound was determined. Fluorescein permeability assessment (in the A-B direction at pH 7.4 on both sides) was performed after the permeability assay for the test compound. The percent of inhibition was calculated by subtracting the percent of control from 100.

Intrinsic Clearance. Intrinsic clearance was determined with human liver microsomes.

Metabolic stability, expressed as percent of the parent compound remaining, was calculated by comparing the peak area of the compound at the time point relative to that at time- 0 .


Supplementary Figure S9: Mean plasma concentration-time profile ( $\pm$ SD) of 4.35 in mice following $10 \mathrm{mg} / \mathrm{kg}$ i.v. administration ( $\mathrm{n}=3$ ). See Methods for experimental details

## 6. Additional in vivo data



Supplementary Figure S10. Inhibitory effect of pyrazolo[4,3-d]pyrimidine $\mathbf{4 . 3 5}$ on apoptosis in lymphoma xenografts derived from the UPF1H cell line. Tumor-bearing mice were intravenously injected with 4.35. Twenty-four hours after injection, the mice were euthanized, and the tumors were removed and analyzed. Tubulin level is included as a loading control.


Supplementary Figure S10. Inhibitory effect of pyrazolo[4,3-d]pyrimidine $\mathbf{4 . 3 5}$ on some proteins in lymphoma xenografts derived from MINO-1 and UPF1H cell lines. Tumorbearing mice were intravenously injected with 4.35. Twenty-four hours after injection, the mice were euthanized, and the tumors were removed and analyzed. Tubulin level is included as a loading control.

## 7. Additional Methods

## Annexin V/PI Assay

Number of apoptotic and/or necrotic cells was determined by flow cytometry (BD FACS Canto II) using Annexin V FITC (Apronex, Czech Republic) and propidium iodide (Sigma). Percentage of cell death was calculated from the proportion of the living cells only using the following formula: \% cell death $=(($ Agent induced apoptosis - basal apoptosis $) /(100-$ basal apoptosis))* 100

## Caspase activity assay

The cells were homogenized in an extraction buffer ( $10 \mathrm{mM} \mathrm{KCl}, 5 \mathrm{mM}$ HEPES, 1 mM EDTA, 1 mM EGTA, $0.2 \%$ CHAPS, inhibitors of proteases, pH 7.4 ) on ice for 20 min . The homogenates were clarified by centrifugation at 10000 xg for 30 min at $4{ }^{\circ} \mathrm{C}$, and then the proteins were quantified and diluted to equal concentrations. Lysates were then incubated for 4 h with 100 mM Ac-DEVD-AMC (Enzo Life Sciences) as a substrate of caspases 3 and 7 in
 fluorescence of the product was measured using a Fluoroskan Ascent microplate reader (Labsystems) at 355/460 nm (excitation/emission).

## Pharmacokinetic analysis in mice

Compound 4.35 was dissolved in DMSO at 100 mM . Mice were i.v. administered $10 \mathrm{mg} / \mathrm{kg}$ compound 4.35. Three mice were sacrificed at each time point, and blood samples were collected by cardiac puncture into heparinized tubes at $15,30,60,120$ and 240 min after drug administration. Blood samples were centrifuged immediately after collection at 1000 g and 4 ${ }^{\circ} \mathrm{C}$ for 5 min , and the supernatants were collected and frozen at $-20^{\circ} \mathrm{C}$. Before analysis, the plasma samples were thawed at room temperature, precipitated in methanol (plasma/methanol, $1 / 3, \mathrm{v} / \mathrm{v}$ ), spiked with the internal standard (IS, roscovitine), vortexed and then centrifuged ( $\left.4{ }^{\circ} \mathrm{C}, 9000 \mathrm{~g}, 10 \mathrm{~min}\right)$. The supernatants were transferred to Eppendorf tubes with microspin filters ( $0.2 \mu \mathrm{~m}, \mathrm{NY}$ ) and centrifuged $\left(4^{\circ} \mathrm{C}, 9000 \mathrm{~g}, 5 \mathrm{~min}\right)$. The filtrates were evaporated under nitrogen and reconstructed in $50 \%$ methanol. A total of $10 \mu \mathrm{~L}$ of the filtrate was injected in an Acquity BEH C18 column ( $100 \times 2.1 \mathrm{~mm}, 1.7 \mu \mathrm{~m}$ ) (Waters, Ireland) maintained at $40{ }^{\circ} \mathrm{C}$. The analyses were performed on a liquid chromatography system (Acquity UPLC System, Waters, Milford, MA, USA) coupled to a Micromass Quattro Micro ${ }^{\text {TM }}$ API (Waters MS Technologies, Manchester, UK) detector. A gradient of 15 mM ammonium formate at pH 4.0 (A) and methanol (B) at a flow rate of $0.25 \mathrm{~mL} / \mathrm{min}$ was
employed ( $0-6$ min linear gradient $35-100 \%$ B, 6-6.5 min $100 \%$ B, 6.5-7 min linear gradient $100-35 \%, 7-8 \mathrm{~min} 35 \% \mathrm{~B}$ ). The mass spectrometry system was operated in positive ionization mode using multiple reaction monitoring and scanning a transition of $420.2>403$ (collision energy 18 V ) for $\mathbf{4 . 3 5}$ and a transition of $355.5>90.86$ (collision energy 40 V ) for IS. The source and analyzer parameters were set as follows: capillary voltage 1000 V ; cone voltage 30 V; extractor 2 V ; RF lens 0.2 V ; cone and desolvation gas was nitrogen at 100 and $350{ }^{\circ} \mathrm{C}$ and flow rates 2.0 and $550 \mathrm{~L} / \mathrm{h}$; LM/HM resolution 12.5 ; ion energy $1,0.3 \mathrm{~V}$; ion energy $2,1.5 \mathrm{~V}$; entrance, exit and multiplier voltages, $2.0 \mathrm{~V}, 2.0 \mathrm{~V}$ and 650 V , respectively. Argon was used as the collision gas. The quantification of the analyte in mouse plasma samples was based on a subsequently measured matrix-matched calibration curve in the range of $0.025-25 \mu \mathrm{~mol} / \mathrm{L}$ (7 calibration concentration levels) constructed by plotting the analyte response (analyte/IS peak area ratio multiplied by IS concentration) against the analyte concentration by applying a logarithmic transformation. The obtained slope, intercept and $\mathrm{r}^{2}$ of the calibration curve were $0.8726,0.1938$ and 0.9979 , respectively. More than $75 \%$ of the back calculated concentrations of the calibration standards were within the $\pm 15 \%$ of the nominal concentration.
8. NMR Spectra of Prepared Compounds










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[3-(Dimehylamino)-1-propyl]thio-3-isopropyl-7-[4-(2-pyridyl)benzyl]amino-1(2)H-pyrazolo[4,3-d]pyrimidine (4.8)


 30
30

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5-(Methoxymethyl)thio-3-isopropyl-7-[4-(2-pyridyl)benzyl]amino-1(2)H-pyrazolo[4,3-d]pyrimidine (4.11)


(
(2-Aminocyclohexyl)thio-3-isopropyl-7-[4-(2-pyridyl)benzyl]amino-1(2)H-pyrazolo[4,3-d]pyrimidine (4.13)














| 0 | G.0 0'L | G'L | 02 | G'ర | 0 \% | §'E | O't | S't | O'S | 0.9 | ¢'9 |  |  | 0 '8 | S'8 |  | 6 | $0^{\prime} \mathrm{OL}$ |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
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5-(2-Amino-2-methyl-1-propyl)thio-3-isopropyl-7-[4-(2-pyridyl)benzyl]amino-1(2)H-pyrazolo[4,3-d]pyrimidine (4.24)









(2-Hydroxy-1-propyl)thio-3-isopropyl-7-[4-(2-pyridyl)benzyl]amino-1(2)H-pyrazolo[4,3-d]pyridmidine (4.29)






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08
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5-(3-Hydroxy-2-butyl)thio-3-isopropyl-7-[4-(2-pyridyl)benzyl]amino-1(2)H-pyrazolo[4,3-d]pyrimidine (4.34)



-160.31

- 155.75
$-149.44$
139.95
-138.55
-137.48
-137.11
$-137.11$
127.82
-126.52
-122.43
-120.03
$-120.03$
$-79.14 \mathrm{CHCL}$
5-(2-Amino-1-ethyl)thio-3-isopropyl-7-[4-(2-pyridyl)benzyl]amino-1(2)H-pyrazolo[4,3-d]pyrimidine (4.35)


## 9. References

(1)Vymetalova, L.; Havlicek, L.; Sturc, A.; Skraskova, Z.; Jorda, R.; Pospisil, T.; Strnad, M.; Krystof, V. 5-Substituted 3-isopropyl-7-[4-(2-pyridyl)benzyl]amino-1(2)H-pyrazolo[4,3-d]pyrimidines with anti-proliferative activity as potent and selective inhibitors of cyclin-dependent kinases. Eur. J. Med. Chem. 2016, 110, 291-301.
(2)Nekardova, M.; Vymetalova, L.; Khirsariya, P.; Kovacova, S.; Hylsova, M.; Jorda, R.; Krystof, V.; Fanfrlik, J.; Hobza, P.; Paruch, K. Structural basis of the interaction of cyclin-dependent kinase 2 with roscovitine and its analogues having bioisosteric central heterocycles. Chemphyschem. 2017, 18, 785-795.
(3)Bettayeb, K.; Oumata, N.; Echalier, A.; Ferandin, Y.; Endicott, J. A.; Galons, H.; Meijer, L. CR8, a potent and selective, roscovitine-derived inhibitor of cyclin-dependent kinases. Oncogene 2008, 27, 5797-5807.
(4)Fanfrlik, J.; Bronowska, A. K.; Rezac, J.; Prenosil, O.; Konvalinka, J.; Hobza, P. A reliable docking/scoring scheme based on the semiempirical quantum mechanical PM6-DH2 method accurately covering dispersion and H-bonding: HIV-1 protease with 22 ligands. J Phys. Chem B 2010, 114, 12666-12678.
(5)Stewart, J. J. Optimization of parameters for semiempirical methods V: modification of NDDO approximations and application to 70 elements. J Mol. Model. 2007, 13, 1173-1213.
(6)Klamt, A.; Schüürmann, G. COSMO: a new approach to dielectric screening in solvents with explicit expressions for the screening energy and its gradient. J. Chem. Soc., Perkin Trans. 2 1993, 0, 799-805.
(7)Schiebel, J.; Gaspari, R.; Wulsdorf, T.; Ngo, K.; Sohn, C.; Schrader, T. E.; Cavalli, A.; Ostermann, A.; Heine, A.; Klebe, G. Intriguing role of water in protein-ligand binding studied by neutron crystallography on trypsin complexes. Nat. Commun. 2018, 9, 3559.
(8)Schrodinger, LLC. [Version 1.3r1 in]. 2010. The PyMOL Molecular Graphics System.
(9)Diederichs, K.; Karplus, P. A. Improved R-factors for diffraction data analysis in macromolecular crystallography. Nat. Struct. Biol. 1997, 4, 269-275.
(10) The CCP4 suite: programs for protein crystallography. Acta Crystallogr. D. Biol. Crystallogr. 1994, 50, 760-763.
(11)Brunger, A. T. Free R value: a novel statistical quantity for assessing the accuracy of crystal structures. Nature 1992, 355, 472-475.
(12)Chen, V. B.; Arendall, W. B., III; Headd, J. J.; Keedy, D. A.; Immormino, R. M.; Kapral, G. J.; Murray, L. W.; Richardson, J. S.; Richardson, D. C. MolProbity: all-atom structure validation for macromolecular crystallography. Acta Crystallogr. D. Biol. Crystallogr. 2010, 66, 12-21.

