

Transcriptional Profiling of Ivermectin Resistant Cooperia punctata Using Deep Sequencing

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Abstract

Intestinal parasites pose a burden to the livestock industry throughout the world. Since the discovery of macrocyclic lactones, Ivermectin has been a popular choice to combat nematode (roundworm) infectious due to its broad spectrum of action and high efficacy. Widespread use of this drug has led to the development of resistance in many parasitic nematodes, including the highly prevalent cattle intestinal parasite Cooperia punctata. Ivermectin is known to bind and activate glutamate-gated chloride channels, thereby disrupting neurotransmission and paralyzing the worms. We hypothesize that Ivermectin treatment of drug-resistant C. punctata will lead to changes in the expression pattern of genes related to the drugs mode of action. We used high throughput sequencing to study the adult transcriptome of drug-resistant C. punctata and to assess Ivermectin's impact on gene expression. RNAseq datasets from treated and untreated Ivermectin resistant adult worms were assembled into 110,772 unique transcripts corresponding to 66,788 genetic loci. 18,585 loci showed evidence of alternative splicing, with an average of 4 transcripts per alternatively spliced locus. Fragmentation rate was estimated at 18.1% by comparison with *C. elegans* CDSs. 101,411 CDSs, representing some 69% of the complete C. punctata gene repertoire, were predicted from our dataset. The predicted CDSs are associated with 4,738 unique InterPro protein domains, 3,013 unique Kyoto Encyclopedia of Gene and Genomes (KEGG) terms, and 524 unique Gene Ontology (GO) terms. 5,308 predicted CDSs were up-regulated in Ivermectin treated worms while 4,047 were down-regulated. Thirteen GO terms were statistically over-represented among up-regulated CDSs, including gated channels and metabolism. This suggests that functions related to Ivermectin's mode of action are up-regulated in response to treatment. These results provide insights into potential mechanisms of Ivermectin resistance and possible genetic markers that may be used to monitor the spread of resistant *C. punctata* in cattle and prevent treatment failures.

Introduction

Cooperia punctata

- Cooperia ssp. most prevalent internal parasite in U.S. cattle¹
- Free living (L1 L3i) and parasitic (L3act adult) stages
- Causes damage to intestines, decreased nutrient uptake

Ivermectin

- Broad spectrum anthelmintic drug
- Binds to glutamate-gated chloride channels (GluCl)
- Causes paralysis leading to starvation
- Widespread use in cattle since 1980's
- Ivermectin resistance is becoming a problem²

Study Design

- USDA juvenile cattle were infected with Ivermectin resistant *C. punctata*
- One calf treated with Ivermectin, the other left untreated
- Samples of *C. punctata* taken when cows reach adult stage
- Create Illumina RNA-Seq data

Specific Aims

- Assemble draft transcriptome for C. punctata
- Gene prediction and assessment of differential gene expression

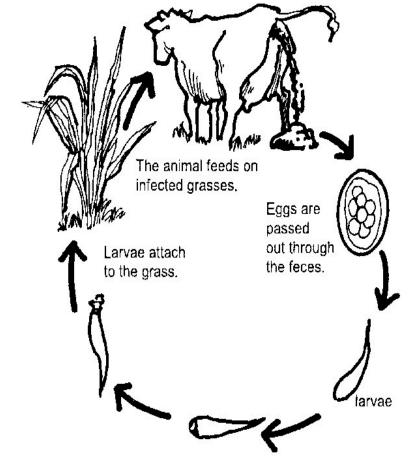


Figure 1: Lifecycle of *C. punctata*

Methods

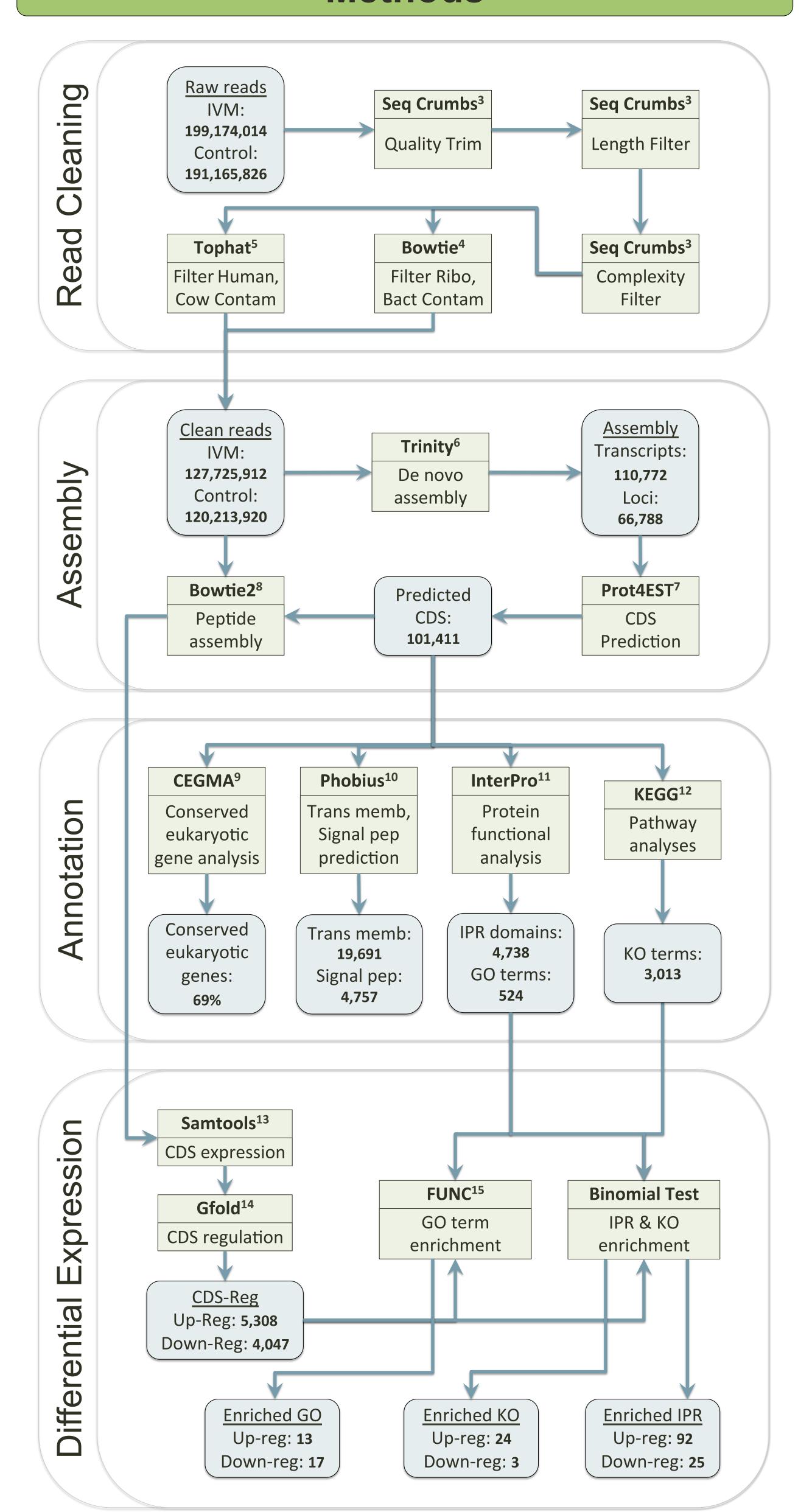


Figure 2: Bioinformatic workflow includes four main steps: read cleaning, *de novo* transcriptome assembly and CDS prediction, CDS annotation, and assessment of differential expression. Square tan boxes include software name and their function. Blue rounded boxes include type of data generated and relevant tallies.

Results

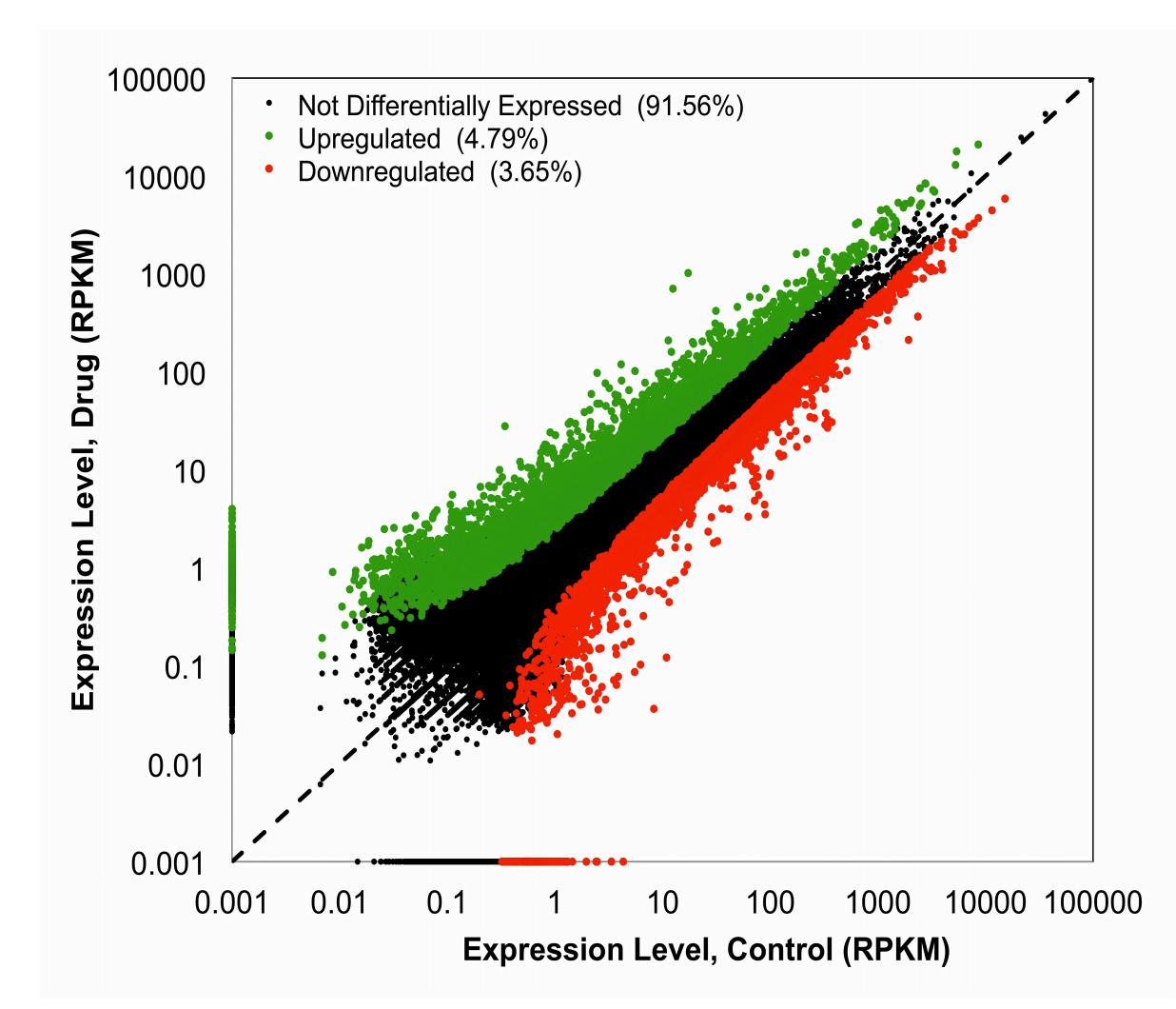


Figure 3: Differential expression of *C. punctata* CDSs in response to drug treatment. Genes were chosen based on significant (p<10⁻⁵) Gfold values, with a threshold cut off 1 standard deviation above or below the mean.

(A) InterPro Domains (B) KEGG Orthologies Low density lipoprotein-related protein 2 <1.00E-15 ATP-binding cassette, subfamily D 1.06E-12 (ALD), member 3 4.41E-12 Myosin heavy chain ninopeptidase/leukotriene A4 hydrolase 1.98E-10 JDP-glucuronosyl/UDI <1.00E-1 2-oxoglutarate dehydrogenase E1 3.47E-09 <1.00E-1 Voltage-dependent calcium channel alpha munoglobulin subtype 2 <1.00E-1 Rap guanine nucleotide exchange factor Concanavalin A-like lectin/glucanase Immunoglobulin-like fold 1.07E-06 6.34E-13 Transcription factor SOX1/3/14/21 (SOX GPCR, rhodopsin-like, 7TM omain of unknown function DUF3358 Polypyrimidine tract-binding protein eurotransmitter-gated ion-channel C-type lectin fold <1.00E-1 Erythrocyte membrane protein band 4.1 Inositol 1,4,5-triphosphate receptor type 1 1.98E-06 <1.00E-1 Mitogen-activated protein kinase kinase <1.00E-1 Peptidase S1A, chymotrypsin-type Peptidase S1/S6, chymotrypsin/Hap, 1.40E-06 Nicotinic acetylcholine receptor Peptidase cysteine/serine, trypsin-like 2.09E-06 Leukotriene-A4 hydrolase 3.32E-06 Ca2+ transporting ATPase, plasma 5.55E-06 5.66E-06 Laminin, alpha 1/2 3.52E-06 Nucleolar protein 56 3.53E-06 Splicing factor, arginine/serine-rich 1/9 9.14E-06 Microspherule protein 1 Collagen triple helix repeat

Figure 4: (A) InterPro domains (IPR) and **(B)** KEGG Orthologies (KO) significantly (p<10⁻⁵, FDR-corrected binomial dist. test) enriched among genes up/down-regulated in Ivermectin resistant *C. punctata* in response to drug treatment. Green=up-regulated; red=down-regulated.

Results

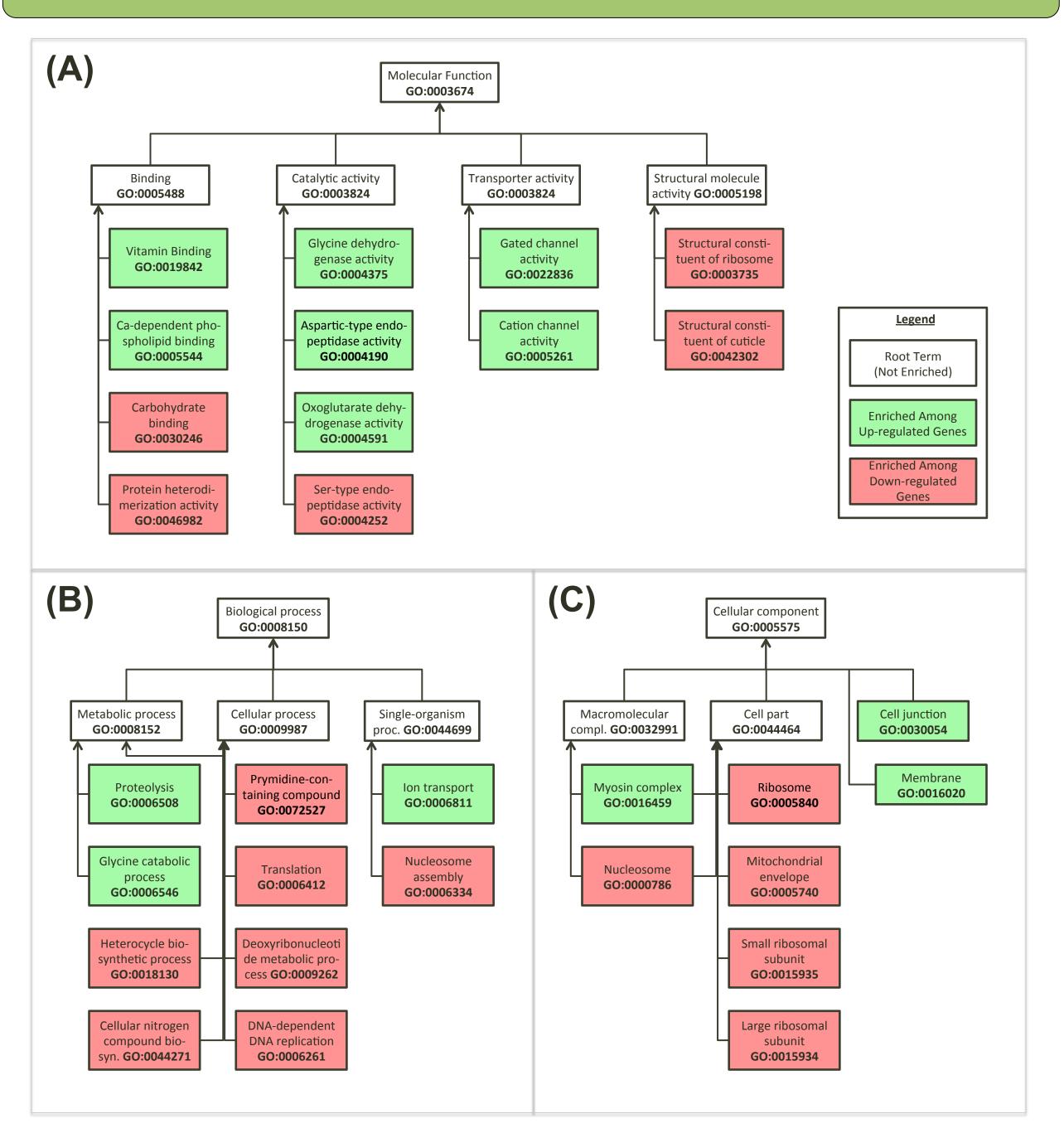


Figure 5: Gene Ontology (GO) terms significantly (p<10⁻⁵, using FUNC, FDR-corrected) enriched among genes up/down-regulated related to **(A)** Molecular Functions, **(B)** Biological Processes, and **(C)** Cellular Components in Ivermectin resistant *C. punctata* in response to drug treatment.

Discussion

- Transporter activity related to Ivermectin's mode of action was up-regulated. This also may be related to P-glycoprotein up-regulation, shown to facilitate Ivermectin resistance in nematodes¹⁶
- Metabolism was up-regulated, possibly due to withstanding starvation during paralysis (also observed in *Caenorhabditis* elegans)¹⁷

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