**Supporting Information**

**Transmembrane Anion Transport Mediated by Adamantyl-functionalized Imidazolium Salts**

Julien Gravel and Andreea R. Schmitzer

Département de Chimie, Université de Montréal, Montréal, Canada

Université de Montréal, 2900 Edouard Montpetit, CP 6128, succ. Centre ville, Montréal, Québec, Canada. E-mail: ar.schmitzer@umontreal.ca

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**Experimental**

## General procedures

All chemicals were purchased from Aldrich Chemicals and used without further purification. CDCl3 were purchased from CDN Isoptopes. NMR experiments were recorded on Bruker Avance 300, Bruker Avance 400 and Bruker Avance 500. Mass spectral data were obtained by the Université de Montréal Mass Spectrometry Facility and were recorded on a Mass spectrometer TSQ Quantum Ultra (Thermo Scientific) with accurate mass options instrument. Fluorimetric studies were performed on a Varian Cary Eclipse Fluorescence spectrophotometer equipped with a temperature controller.

## Synthesis

1,3-bis(adamantan-1-yl)imidazolium bromide **(1a)**: 

Adamantylamine (2 g, 13.2 mmol, 1 eq.) is solubilized in 20 mL of MeOH with formaldehyde 37% (1.04 mL, 14 mmol, 1.1 eq.) and agitated for 30 minutes at room temperature. Solubilized adamantylamine (2 g, 13.2 mmol, 1 eq.) in 20 mL of MeOH is slowly added over 1 hour. During this time, a solution of 20% HBr (3.84 mL, 14 mmol, 1.1 eq.) was cooled in the ice bath. Cold HBr 20% is added dropwise and the mixture is agitated 2 hours at room temperature. Glyoxal 40% (1.6 mL, 14 mmol, 1.1 eq.) is added dropwise and the reaction mixture is agitated overnight at reflux. The solvent are evaporated and the crude product is triturated with 20 mL of AcOEt and the precipitated solid is filtrated. The product is then treated with 250 mL of saturated NaHCO3 and extracted with 3 portions of 200 mL of dichloromethane. The product is concentrated and purified by chromatography column using a 90%/10% dichloromethane/MeOH eluent. Combined fractions of the product were evaporated and triturated with 20 mL of AcOEt. The precipitate was filtrated and rinsed with 2 portions of 20 mL of AcOEt and dried to yield a white solid (1.1 g, 19%).

1H NMR (CDCl3, 500MHz): δ = 10.32 (t, J=1.7 Hz, 1 H), 7.44 (d, J=1.8 Hz, 2 H), 2.32 - 2.38 (m, 18 H), 1.84 - 1.89 (m, 6 H), 1.76 - 1.81 ppm (m, 6 H)

13C NMR (CDCl3, 126MHz): δ = 134.3, 117.7, 61.2, 42.9, 35.3, 29.6 ppm

HRMS (ESI) for C23H33N2: Calc. (m/z): 337.2648; found (m/z): 337.2648.

1,3-bis(adamantan-1-yl)imidazolium bis(trifluoromethylsulfonyl)imide (**1b**):

1,3-bis(adamantan-1-yl)imidazolium bromide (200 mg, 0.48 mmol, 1 eq.) was solubilized in a minimum amount of MeOH and an excess of LiNTf2 (413mg, 1.44 mmol, 3 eq.) is added. The solution is heated to 50°C overnight. Solvent is evaporated and triturated in 50 mL of water. The precipitate is filtered to yield a white solid (223mg, 75%).

1H NMR (CDCl3, 300MHz): δ = 8.59 (t, J=1.7 Hz, 1 H), 7.54 (d, J=1.7 Hz, 2 H), 2.32 (br. s., 6 H), 2.17 (d, J=2.9 Hz, 12 H), 1.80 ppm (d, J=2.9 Hz, 12 H)

13C NMR (CDCl3, 75MHz): δ = 129.8, 119.9, 119.2, 60.9, 42.6, 35.2, 29.4 ppm

HRMS (ESI) for C23H33N2: Calc. (m/z): 337.2638; found (m/z): 337.2648.

HRMS (ESI) for C2F6NO4S2: Calc. (m/z): 279.9178; found (m/z): 279.9176.

1-(bromomethyl)adamantane (**2**): 

Adamantanemethanol (2 g, 12 mmol, 1 eq.) was introduced in 30 mL of concentrated hydrobromic acid and refluxed for 12 hours. The reaction mixture was cooled down to room temperature and was then diluted with 50 mL of distillated water. The aqueous layer is then extracted with dichloromethane (75 mL × 3). The combined organic extracts were washed with 100 mL of a saturated NaHCO3 solution, with 100 mL of distillated water, with 100 mL of Brine. The organic extracts were dried over MgSO4, filtered, and concentrated to yield a white powder (2.59 g, 94 %).

Characterization data are in accordance with literature.([*1*](#_ENREF_1))

1-(2-bromoethyl)adamantane (**3**):

Adamantaneethanol (2 g, 11 mmol, 1 eq.) was introduced in 30 mL of concentrated hydrobromic acid and refluxed for 12 hours. The solution was cooled down to room temperature and was then diluted with 50 mL of distillated water. The aqueous layer is then extracted with dichloromethane (75 mL × 3). The combined organic extracts were washed with 100 mL of a saturated NaHCO3 solution, with 100 mL of distillated water and with 100 mL of Brine. The organic extracts were dried over MgSO4, filtered and concentrated to yield a white powder (2.45 g, 91 %).

Characterization data are in accordance with literature.([*2*](#_ENREF_2))

1-(2-(adamantan-1-yl)methyl)imidazole (**4**):

Sodium hydroxide (312 mg, 7.8 mmol, 3 eq.) and imidazole were suspended in 20 mL of dry DMF and stirred for 2 hours under nitrogen atmosphere at room temperature. The resulting solution is added on 1-(bromomethyl)adamantane (600 mg, 2.6 mmol, 1 eq.) via a syringe and heated to 100°C overnight. The reaction mixture was cooled down to room temperature and quenched with methanol. Solvents were evaporated and the crude product was transferred in an Erlenmeyer using 100 mL of DCM and 150 mL of distillated water. The heterogeneous mixture was vigorously stirred for 20 min. and the organic layer was decanted, washed with 100 mL of Brine, dried over MgSO4, and concentrated to yield a white solid (429 mg, 76 %).

1H NMR (CDCl3, 400MHz): δ = 7.39 (s, 1 H), 7.05 (s, 1 H), 6.84 (s, 1 H), 3.59 (s, 2 H), 2.01 (br. s., 3 H), 1.67 - 1.78 (m, 3 H), 1.56 - 1.64 (m, 3 H), 1.50 ppm (d, J=2.2 Hz, 6 H)

13C NMR (CDCl3, 126MHz): δ = 138.3, 128.4, 120.6, 59.7, 40.3, 36.6, 34.1, 28.1 ppm

HRMS (ESI) for C14H21N2: Calc. (m/z): 217.1699; found (m/z): 217.1689.

1-(2-(adamantan-1-yl)ethyl)imidazole (**5**): 

Sodium hydroxide (493 mg, 12.3 mmol, 3 eq.) and imidazole were suspended in 20 mL of dry DMF and stirred for 2 hours under nitrogen atmosphere at room temperature. The resulting solution is added on 1-(2-bromoethyl)adamantane (1 g, 4.1 mmol, 1 eq.) via a syringe and heated to 40°C overnight. The reaction mixture was cooled down to room temperature and quenched with methanol. Solvents were evaporated and the crude product was transferred in an Erlenmeyer using 100 mL of DCM and 150 mL of distillated water. The heterogeneous mixture was vigorously stirred for 20 min. and the organic layer was decanted, washed with 100 mL of Brine, dried over MgSO4, and concentrated to yield a pale yellow solid (850 mg, 89 %).

1H NMR (CDCl3, 400MHz): δ = 7.49 (s, 1 H), 7.07 (s, 1 H), 6.93 (s, 1 H), 3.92 - 3.99 (m, 2 H), 2.01 (br. s., 3 H), 1.71 - 1.80 (m, 3 H), 1.58 - 1.71 (m, 5 H), 1.56 ppm (br. s., 6 H)

13C NMR (CDCl3, 101MHz): δ = 136.5, 128.9, 118.3, 45.0, 41.8, 41.7, 36.5, 31.5, 28.0 ppm

HRMS (ESI) for C15H23N2: Calc. (m/z): 231.1856; found (m/z): 231.1856.

1-(2-(adamantan-1-yl)methyl)-2-methylimidazole (**6**):

Sodium hydroxide (260 mg, 6.5 mmol, 3 eq.) and 2-methylimidazole (536 mg, 6.5 mmol, 3 eq.) were suspended in 20 mL of dry DMF and stirred for 2 hours under nitrogen atmosphere at room temperature. The resulting solution is added on 1-(bromomethyl)adamantane (500 mg, 2.2 mmol, 1 eq.) via a syringe and heated to 100°C overnight. The reaction mixture was cooled down to room temperature and quenched with methanol. Solvents were evaporated and the crude product was transferred in an Erlenmeyer using 100 mL of DCM and 150 mL of distillated water. The heterogeneous mixture was vigorously stirred for 20 min. and the organic layer was decanted, washed with 100 mL of Brine, dried over MgSO4, and concentrated to yield a white solid (443 mg, 88 %).

1H NMR (CDCl3, 300MHz): δ = 6.88 (s, 1 H), 6.74 (s, 1 H), 3.49 (s, 2 H), 2.36 (s, 3 H), 1.99 (br. s., 3 H), 1.65 - 1.76 (m, 3 H), 1.54 - 1.64 (m, 3 H), 1.51 ppm (d, *J*=2.0 Hz, 6 H)

13C NMR (CDCl3, 75MHz): δ = 145.3, 126.2, 121.2, 58.2, 40.6, 36.6, 35.2, 28.1, 13.7 ppm

HRMS (ESI) for C15H23N2: Calc. (m/z): 231.1856; found (m/z): 231.1866.

1-(2-(adamantan-1-yl)ethyl)-2-methylimidazole (**7**):

Sodium hydroxide (100 mg, 2.5 mmol, 3 eq.) and 2-methylimidazole (205 mg, 2.5 mmol, 3 eq.) were suspended in 15 mL of dry DMF and stirred for 2 hours under nitrogen atmosphere at room temperature. The resulting solution is added on 1-(2-bromoethyl)adamantane (205 mg, 0.82 mmol, 1 eq.) via a syringe and heated to 40°C overnight. The reaction mixture was cooled down to room temperature and quenched with methanol. Solvents were evaporated and the crude product was transferred in an Erlenmeyer using 100 mL of DCM and 150 mL of distillated water. The heterogeneous mixture was vigorously stirred for 20 min. and the organic layer was decanted, washed with 100 mL of Brine, dried over MgSO4, and concentrated to yield a pale brown solid (161 mg, 81 %).

1H NMR (CDCl3, 400MHz): δ = 6.92 (s, 1 H), 6.82 (s, 1 H), 3.77 - 3.89 (m, 2 H), 2.39 (s, 3 H), 2.02 (br. s., 3 H), 1.75 - 1.78 (m, 3 H), 1.64 - 1.71 (m, 3 H), 1.58 (d, *J*=2.0 Hz, 6 H), 1.47 - 1.54 ppm (m, 2 H)

13C NMR (CDCl3, 126MHz): δ = 127.1, 118.8, 45.1, 42.2, 41.1, 37.0, 31.9, 28.5, 29.6 13.0 ppm

HRMS (ESI) for C16H25N2: Calc. (m/z): 245.2012; found (m/z): 245.2003.

1,3-bis(2-(adamantan-1-yl)methyl)imidazolium bromide (**8a**):

1-(2-(adamantan-1-yl)methyl)imidazole (500 mg, 2.3 mmol, 1 eq.) is solubilized with 1-(2-bromomethyl)adamantane (583 mg, 2.5 mmol, 1.1 eq.) in 5 mL of NMP in a sealed vial and were heated using a Biotage® Initiator Classic Microwave Synthesizer at 200°C for 2 hours. The NMP is evaporated and the crude mixture is triturated with 20 mL AcOEt. The precipitate is filtrated and dried to yield a white solid (717 mg, 70 %).

1H NMR (CDCl3, 500MHz): δ = 10.51 (s, 1 H), 7.21 (d, *J*=1.5 Hz, 2 H), 4.11 (s, 4 H), 2.05 (br. s., 6 H), 1.70 – 1.78 (m, 6 H), 1.58 - 1.64 (m, 6 H), 1.57 ppm (d, *J*=2.6 Hz, 12 H)

13C NMR (CDCl3, 126MHz): δ = 139.0, 122.6, 61.4, 39.8, 36.3, 34.1, 27.9 ppm

HRMS (ESI) for C25H37N2: Calc. (m/z): 365.2951; found (m/z): 365.2944.

1,3-bis(2-(adamantan-1-yl)methyl)imidazolium bis(trifluorometylsulfonyl)imide (**8b**): 

1,3-bis(2-(adamantan-1-yl)ethyl)imidazolium bromide (150mg, 0.34 mmol, 1 eq.) is solubilized in a minimum amount of MeOH and an excess of LiNTf2 (290mg, 1.01 mmol, 3 eq.) is added. The solution is heated to 50°C overnight. Solvent is evaporated and triturated in 50 mL of water. The precipitate is filtered to yield a white solid (196mg, 90%).

1H NMR (CDCl3, 300MHz): δ = 8.75 (s, 1 H), 7.11 (d, *J*=1.7 Hz, 2 H), 3.93 (s, 4 H), 2.06 (br. s., 6 H), 1.69 - 1.81 (m, 6 H), 1.55 - 1.66 (m, 6 H), 1.50 ppm (d, *J*=2.7 Hz, 12 H)

13C NMR (CDCl3, 101MHz): δ = 137.2, 123.1, 119.8, 61.7, 39.7, 36.3, 34.0, 27.9 ppm

19F NMR (CDCl3, 282MHz): δ = -80.2 ppm

HRMS (ESI) for C25H37N2: Calc. (m/z): 365.2951; found (m/z): 365.2943.

HRMS (ESI) for C2F6NO4S2: Calc. (m/z): 279.9178; found (m/z): 279.9183.

1,3-bis(2-(adamantan-1-yl)ethyl)imidazolium bromide (**9a**): 

1-(2-(adamantan-1-yl)ethyl)imidazole (500 mg, 2.2 mmol, 1 eq.) is solubilized with 1-(2-bromoethyl)adamantane (792 mg, 3.3 mmol, 1.5 eq.) in 20 mL of acetonitrile and were heated to reflux during 72 hours. The solvent is evaporated and the crude mixture is triturated with 20 mL AcOEt. The precipitate is filtrated, rinsed with 2 portions of 20 mL of AcOEt, and dried to yield a white solid (722 mg, 70 %).

1H NMR (CDCl3, 400MHz): δ = 10.80 (s, 1 H), 7.27 (s, 2 H), 4.33 - 4.43 (m, 4 H), 2.00 (br. s., 6 H), 1.56 - 1.78 ppm (m, 28 H)

13C NMR (CDCl3, 400MHz): δ = 213.1, 208.9, 189.9, 189.6, 189.0, 187.6, 186.5, 185.5 ppm

HRMS (ESI) for C27H41N2: Calc. (m/z): 393.3264; found (m/z): 393.3269.

1,3-bis(2-(adamantan-1-yl)ethyl)imidazolium tetrafluoroboride (**9b**):



1,3-bis(2-(adamantan-1-yl)ethyl)imidazolium bromide (50 mg, 0.11 mmol, 1 eq.) is solubilized in a minimum amount of MeOH and an excess of NaBF4 (35 mg, 0.32 mmol, 3 eq.) is added. The solution is heated to 50°C overnight. Solvents are evaporated and triturated in 50 mL of water. The precipitate is filtered to yield a white solid (47 mg, 89%).

1H NMR (CDCl3, 400MHz): δ = 8.98 (br. s., 1 H), 7.33 (s, 2 H), 4.15 - 4.30 (m, 4 H), 1.99 (br. s., 6 H), 1.53 - 1.80 ppm (m, 28 H)

13C NMR (CDCl3, 101MHz): δ = 135.7, 121.6, 45.4, 43.9, 41.6, 36.4, 31.7, 28.0 ppm

19F NMR (CDCl3, 282MHz): δ = -151.3 ppm

HRMS (ESI) for C27H41N2: Calc. (m/z): 393.3264; found (m/z): 393.3269.

1,3-bis(2-(adamantan-1-yl)ethyl)imidazolium hexafluorophosphate (**9c**):



1,3-bis(2-(adamantan-1-yl)ethyl)imidazolium bromide (50 mg, 0.11 mmol, 1 eq.) is solubilized in a minimum amount of MeOH and an excess of KPF6 (59mg, 0.32 mmol, 3 eq.) is added. The solution is heated to 50°C overnight. Solvent is evaporated and triturated in 50 mL of water. The precipitate is filtered to yield a white solid (54 mg, 92%).

1H NMR (CDCl3, 400MHz): δ = 9.08 (br. s., 1 H), 7.25 (br. s., 2 H), 4.23 (br. s., 4 H), 2.01 (br. s., 6 H), 1.50 - 1.81 ppm (m, 28 H)

13C NMR (CDCl3, 126MHz): δ = 135.7, 122.1, 45.8, 44.2, 41.9, 36.8, 32.0, 28.4 ppm

19F NMR (CDCl3, 282MHz): δ = -71.0, -73.5 ppm

31P NMR (CDCl3, 162MHz): δ = -143 ppm.

HRMS (ESI) for C27H41N2: Calc. (m/z): 393.3264; found (m/z): 393.3267.

HRMS (ESI) for F6P: Calc. (m/z): 144.9647; found (m/z): 144.9647.

1,3-bis(2-(adamantan-1-yl)ethyl)imidazolium bis(trifluorometylsulfonyl)imide (**9d**):



1,3-bis(2-(adamantan-1-yl)ethyl)imidazolium bromide (50 mg, 0.11 mmol, 1 eq.) is solubilized in a minimum amount of MeOH and an excess of LiNTf2 (92mg, 0.32 mmol, 3 eq.) is added. The solution is heated to 50°C overnight. Solvent is evaporated and triturated in 50 mL of water. The precipitate is filtered to yield a white solid (67 mg, 91%).

1H NMR (CDCl3, 400MHz): δ = 9.02 (s, 1 H), 7.22 (s, 2 H), 4.22 - 4.29 (m, 4 H), 2.02 (br. s., 6 H), 1.72 - 1.79 (m, 6 H), 1.64 - 1.71 (m, 10 H), 1.57 ppm (s, 10 H)

13C NMR (CDCl3, 126MHz): δ = 135.5, 122.2, 119.8, 45.9, 44.2, 41.9, 36.7, 32.0, 28.4 ppm

19F NMR (CDCl3, 282MHz): δ = -78.8 ppm

HRMS (ESI) for C27H41N2: Calc. (m/z): 393.3264; found (m/z): 393.3270.

HRMS (ESI) for C2F6NO4S2: Calc. (m/z): 279.9178; found (m/z): 279.9182.

1,3-bis(2-(adamantan-1-yl)methyl)-2-methylimidazolium bromide (**10a**):

1-(2-(adamantan-1-yl)methyl)-2-methylimidazole (80mg, 0.35 mmol, 1 eq.) is solubilized with 1-(2-bromomethyl)adamantane (160 mg, 2.5 mmol, 2 eq.) in 2 mL of NMP in a sealed vial and were heated using a Biotage® Initiator Classic Microwave Synthesizer at 200°C for 2 hours. The NMP is evaporated and the crude mixture is triturated with 20 mL AcOEt. The precipitate is filtrated and dried to yield a white solid (116 mg, 71 %).

1H NMR (CDCl3, 400MHz): δ = 7.38 (s, 2 H), 4.01 (s, 4 H), 2.81 (s, 3 H), 2.07 (br. s., 6 H), 1.72 - 1.80 (m, 6 H), 1.61 - 1.68 (m, 6 H), 1.59 ppm (s, 12 H)

13C NMR (CDCl3, 101MHz): δ = 144.9, 122.8, 60.6, 40.4, 36.3, 35.4, 27.9, 12.5 ppm

HRMS (ESI) for C26H39N2: Calc. (m/z): 379.3108; found (m/z): 379.3118.

1,3-bis(2-(adamantan-1-yl)methyl)-2-methylimidazolium bis(trifluorometylsulfonyl)imide (**10b**):



1,3-bis(2-(adamantan-1-yl)methyl)-2-methylimidazolium bromide (115mg, 0.25 mmol, 1 eq.) is solubilized in a minimum amount of MeOH and an excess of LiNTf2 (215mg, 0.75 mmol, 3 eq.) is added. The solution is heated to 50°C overnight. Solvent is evaporated and triturated in 50 mL of water. The precipitate is filtered to yield a white solid (143mg, 87%).

1H NMR (CDCl3, 300MHz): δ = 7.13 (s, 2 H), 3.83 (s, 4 H), 2.63 (s, 3 H), 2.07 (br. s., 6 H), 1.72 - 1.80 (m, 6 H), 1.58 - 1.66 (m, 6 H), 1.54 ppm (d, *J*=2.4 Hz, 12 H)

13C NMR (CDCl3, 101MHz): δ = 144.6, 122.6, 120.0, 60.4, 40.2, 36.2, 35.3, 27.8, 11.3 ppm

19F NMR (CDCl3, 282MHz): δ = -80.1 ppm

HRMS (ESI) for C26H39N2: Calc. (m/z): 379.3108; found (m/z): 379.3108

HRMS (ESI) for C2F6NO4S2: Calc. (m/z): 279.9178; found (m/z): 279.9175.

1,3-bis(2-(adamantan-1-yl)ethyl)-2-methylimidazolium bromide (**11a):**



1-(2-(adamantan-1-yl)ethyl)-2-methylimidazole (500mg, 2.1 mmol, 1 eq.) is solubilized with 1-(2-bromoethyl)adamantane (747 mg, 3.1 mmol, 1.5 eq.) in 20 mL of acetonitrile and were heated to reflux 72 hours. The solvent is evaporated and the crude mixture is triturated with 20 mL AcOEt. The precipitate is filtrated, rinsed with 2 portions of 20 mL of AcOEt, and dried to yield a white solid (804 mg, 67 %).

1H NMR (CDCl3, 500MHz): δ = 7.52 (s, 2 H), 4.20 - 4.25 (m, 4 H), 2.79 (s, 3 H), 2.01 (br. s., 6 H), 1.71 - 1.77 (m, 6 H), 1.63 - 1.68 (m, 6 H), 1.55 - 1.61 ppm (m, 16 H)

13C NMR (CDCl3, 126MHz): δ = 142.8, 121.5, 44.6, 44.1, 42.1, 36.7, 32.1, 28.4, 11.0 ppm

HRMS (ESI) for C28H43N2: Calc. (m/z): 407.3421; found (m/z): 407.3418.

1,3-bis(2-(adamantan-1-yl)ethyl)-2-methylimidazolium bis(trifluorometylsulfonyl)imide (**11b**):



1,3-bis(2-(adamantan-1-yl)ethyl)-2-methylimidazolium bromide (200 mg, 0.41 mmol, 1 eq.) is solubilized in a minimum amount of MeOH and an excess of LiNTf2 (353mg, 1.23 mmol, 3 eq.) is added. The solution is heated to 50°C overnight. Solvent is evaporated and triturated in 50 mL of water. The precipitate is filtered to yield a white solid (246mg, 87%).

1H NMR (CDCl3, 300MHz): δ = 7.19 (s, 2 H), 4.04 - 4.12 (m, 4 H), 2.60 - 2.64 (m, 3 H), 2.02 (br. s., 6 H), 1.71 - 1.81 (m, 6 H), 1.62 - 1.71 (m, 6 H), 1.50 - 1.61 ppm (m, 16 H)

13C NMR (CDCl3, 101MHz): δ = 142.8, 121.0, 120.0, 44.4, 43.8, 42.0, 36.7, 32.0, 28.4, 9.7 ppm

19F NMR (CDCl3, 282MHz): δ = -80.1 ppm

HRMS (ESI) for C28H43N2: Calc. (m/z): 407.3421; found (m/z): 407.3435

HRMS (ESI) for C2F6NO4S2: Calc. (m/z): 279.9178; found (m/z): 279.9171.

## Lucigenin-based fluorescence assays

### Preparation of EYPC liposomes for lucigenin-based fluorescence assays

A lipid film was formed by evaporating a chloroform solution containing 50 mg or 25 mg of EYPC lipids (Egg Yolk Phosphatidyl Choline) under reduced pressure at 25°C for at least 2 hours. The lipid film was then hydrated with 500 μL of a 2 mM lucigenin solution containing 100 mM NaCl and 10 mM NaH2PO4/Na2HPO4 buffer (pH = 6.2). The resulting suspension was then subjected to at least 10 freeze/thaw cycles (1 minute at -78°C followed by 1 minute at 35°C) and vortexed for 30 seconds between each cycles. The suspension is then extruded through a 100 nm polycarbonate membrane 21 times. The liposomes are purified through a Sephadex G-25 column to remove the extravesicular lucigenin dye using a 100 mM NaNO3 and 10 mM NaH2PO4/Na2HPO4 buffer (pH = 6.2) eluent. The isolated liposomes were diluted to 10 mM (50 mg of EYPC) or 5 mM (25 mg of EYPC) relative to the lipid, assuming all EYPC was conserved through manipulations.

### Preparation of EYPC:Cholesterol liposomes for lucigenin-based fluorescence assays

The procedures are the same as the *Preparation of EYPC liposomes for lucigenin-based fluorescence assays*, except that cholesterol was added in proportion of 7:3 EYPC:Cholesterol in the chloroform solution.

### Preparation of DPPC liposomes for lucigenin-based fluorescence assays

The procedures are the same as the *Preparation of EYPC liposomes for lucigenin-based fluorescence assays*, except that EYPC is replaced by Dipalmitoylphosphatidylcholine (DPPC).

### Lucigenin-based fluorescence assays with EYPC liposomes

A 50 μL (10 mM EYPC liposomes stock solution) or 100 μL (5 mM EYPC liposomes stock solution) aliquot of liposomes solution was added to a 2.5 mL solution containing 10 mM NaH2PO4/Na2HPO4 buffer (pH = 6.2) and 100 mM salt (salt = NaCl, NaNO3, NaHCO3, Na2SO4, NaClO4). The lucigenin fluorescence was monitored by excitation at λex = 372 nm and the emission was recorded at λem = 503 nm. At t = 50 s, 50 μL of solution of transporter at different concentrations in MeOH were added, and at t = 300 s, 100 μL of a Triton-X 5% (V/V) solution were added in order to lyse the liposomes and observe maximal lucigenin fluorescence in solution. The temperature was set to 25°C. Experiments were repeated in triplicate and all traces reported are the average of the three trials.

### Lucigenin-based fluorescence assays with EYPC:Cholesterol liposomes

The procedures are the same as the *Lucigenin-based fluorescence assays with EYPC liposomes*, except that it was a EYPC:Cholesterol 7:3 liposomes stock solution.

### Lucigenin-based fluorescence assays with DPPC liposomes

The procedures are the same as the *Lucigenin-based fluorescence assays with EYPC liposomes*, except that it was a DPPC liposomes stock solution with the temperature set to 30°C or 40°C.

### Tranport in EYPC:Cholesterol liposomes

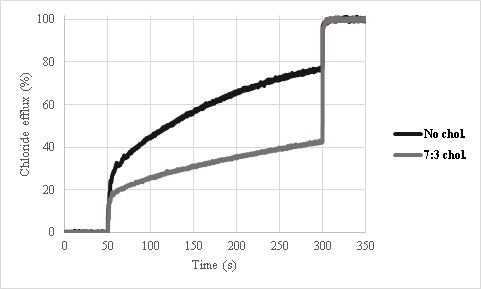


Figure S1. Relative chloride transport activity of imidazolium salt 9a at 20 mol% relative to EYPC or EYPC:Cholesterol (7:3). Intravesicular: 2 mM lucigenin, 100 mM NaCl, 10 mM phosphate buffer (pH=6.2). Extravesicular: 100 mM NaNO3, 10 mM phosphate buffer (pH=6.2).

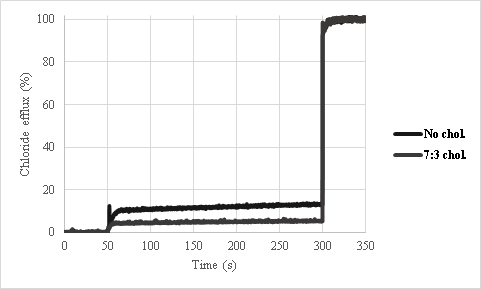


Figure S2. Relative chloride transport activity of imidazolium salt **11b** at 20 mol% relative to EYPC or EYPC:Cholesterol (7:3). Intravesicular: 2 mM lucigenin, 100 mM NaCl, 10 mM phosphate buffer (pH=6.2). Extravesicular: 100 mM NaNO3, 10 mM phosphate buffer (pH=6.2).

### Transport in DPPC liposomes

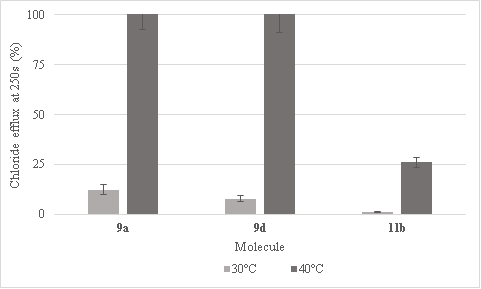
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Figure S3. Relative chloride transport activity of imidazolium salts at 15 mol% relative to DPPC. Intravesicular: 2 mM lucigenin, 100 mM NaCl, 10 mM phosphate buffer (pH=6.2). Extravesicular: 100 mM NaNO3, 10 mM phosphate buffer (pH=6.2).

## NMR Job’s plots

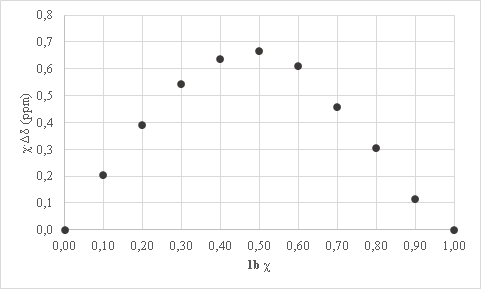
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Figure S4. 1b NMR Job’s plot with TBAC salt in CDCl3. Total concentration = 12,5mM. Effect on H2 proton signal.

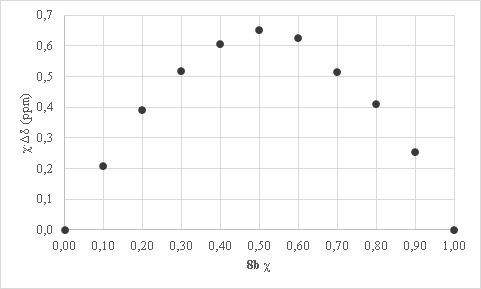


Figure S5. 8b NMR Job’s plot with TBAC salt in CDCl3. Total concentration = 12,5mM. Effect on H2 proton signal.

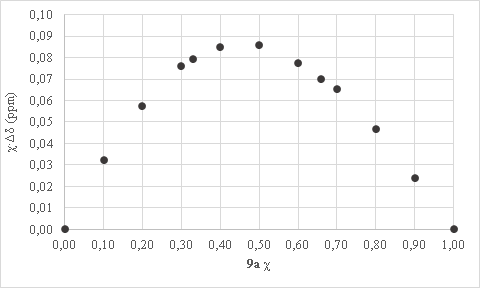
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Figure S6. 9a NMR Job’s plot with TBAC salt in CDCl3. Total concentration = 12.5mM (effect on the H2 proton).

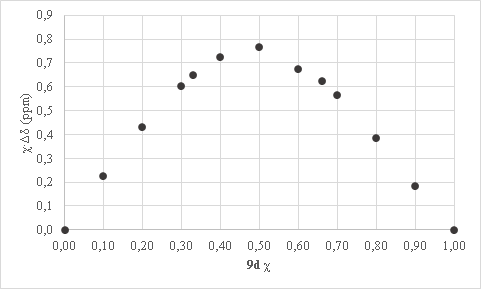


Figure S7. 9d NMR Job’s plot with TBAC salt in CDCl3. Total concentration = 12.5mM (effect on the H2 proton).

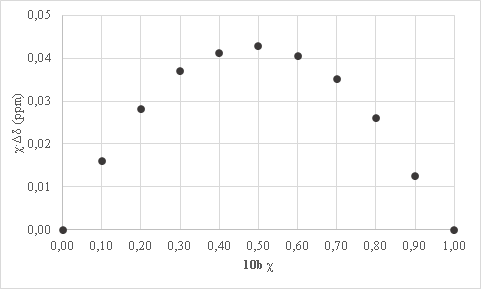


Figure S8. 10b NMR Job’s plot with TBAC salt in CDCl3. Total concentration = 12.5mM (effect on the H2 proton).

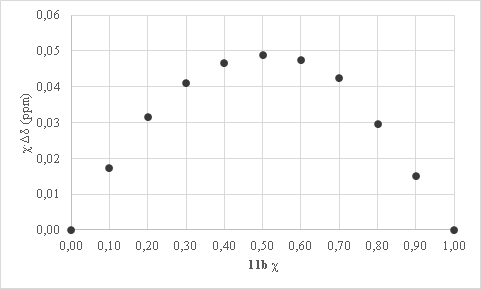
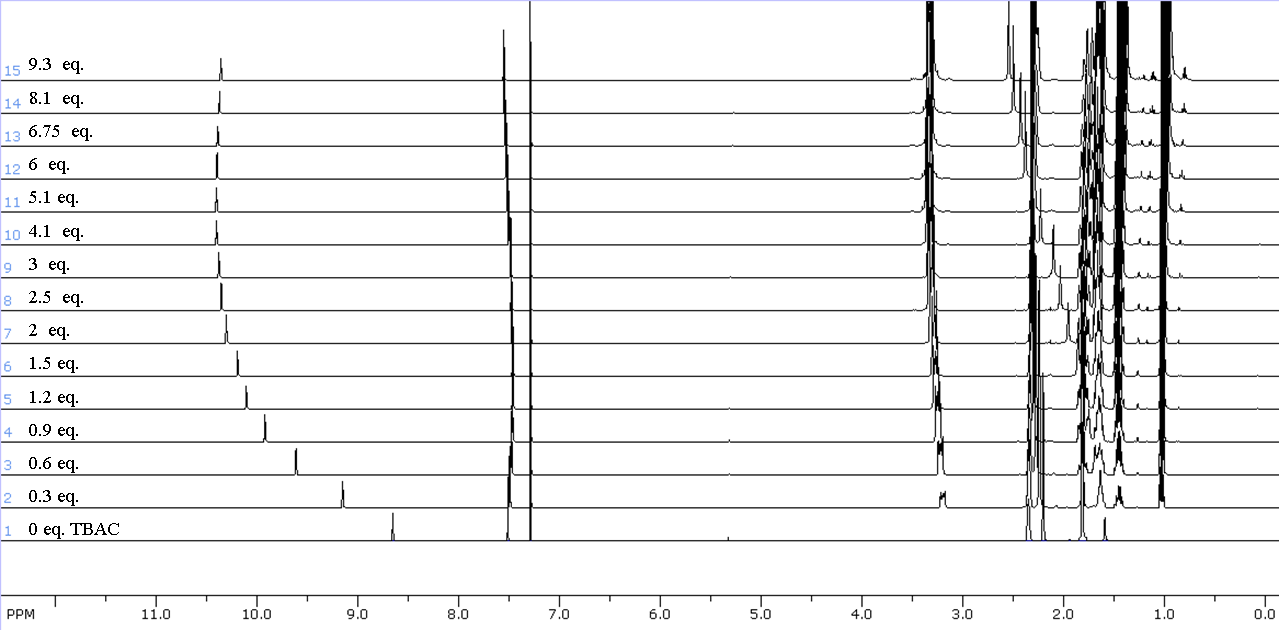


Figure S9. 11b NMR Job’s plot with TBAC salt in CDCl3. Total concentration = 12.5mM (effect on the H2 proton).

## NMR Titrations

A)



B)

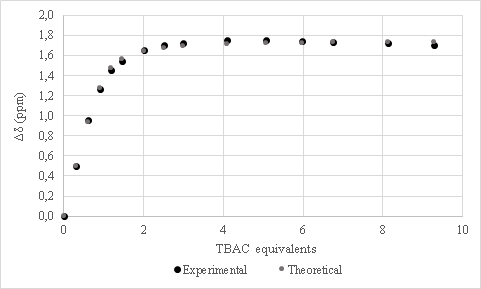
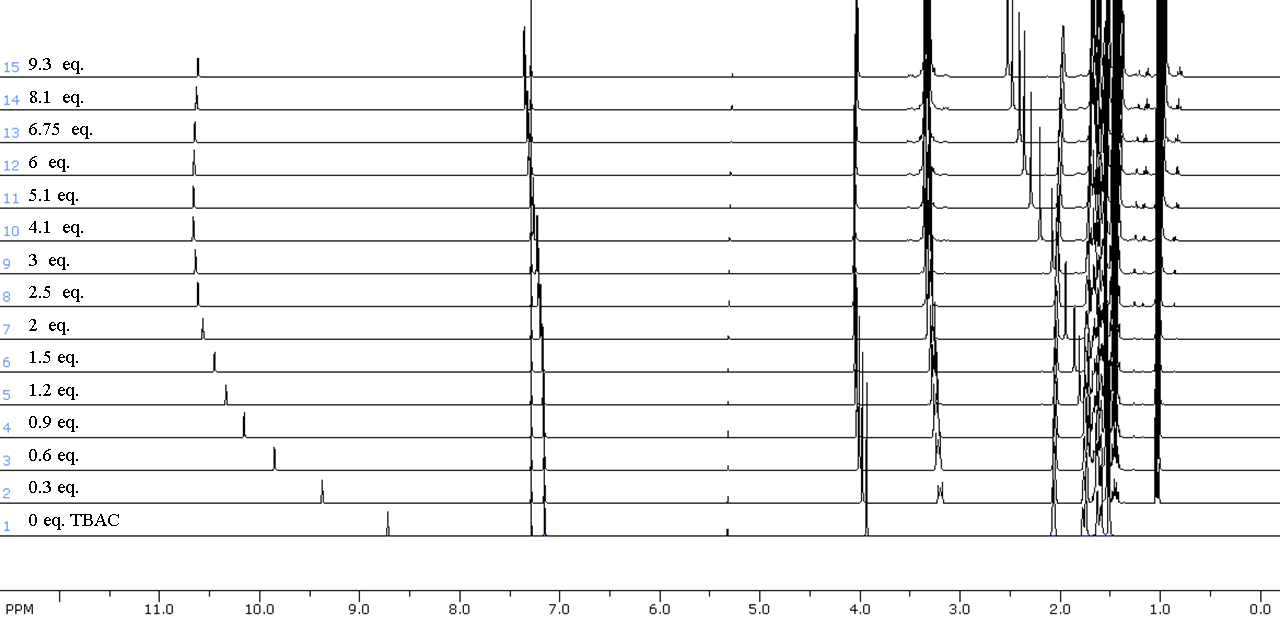


Figure S10. Stacked 1H NMR spectra (A) and titration curves (B) of [**1b**] = 12.5mM with TBAC (400 MHz, CDCl3, 298 K).

A)



B)

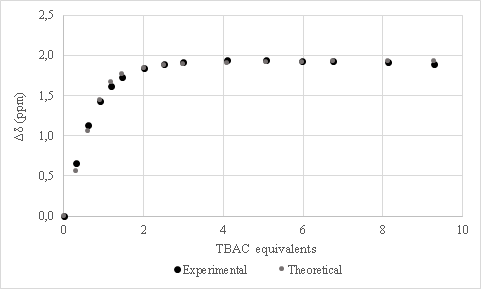
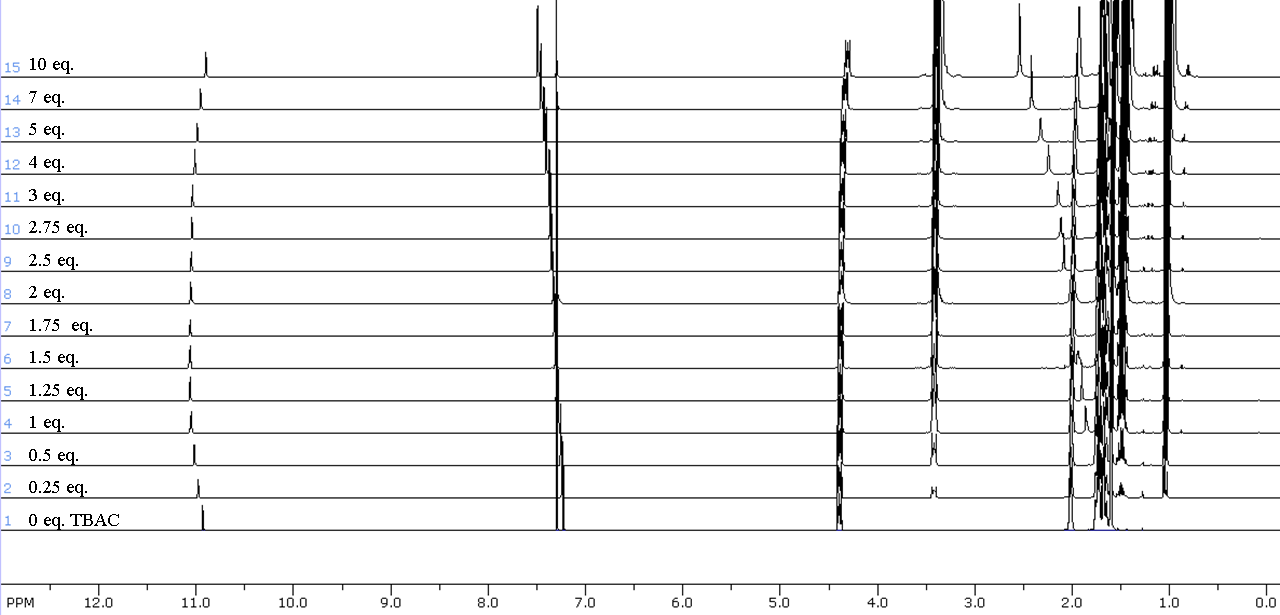


Figure S11. Stacked 1H NMR spectra (A) and titration curves (B) of [**8b**] = 12.5mM with TBAC (400 MHz, CDCl3, 298 K).

A)



B)

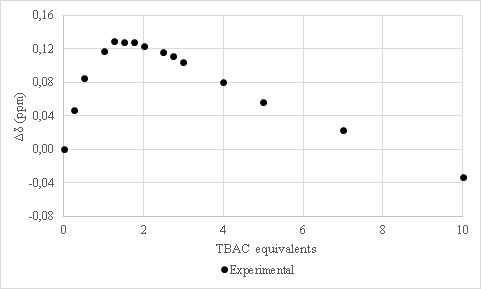
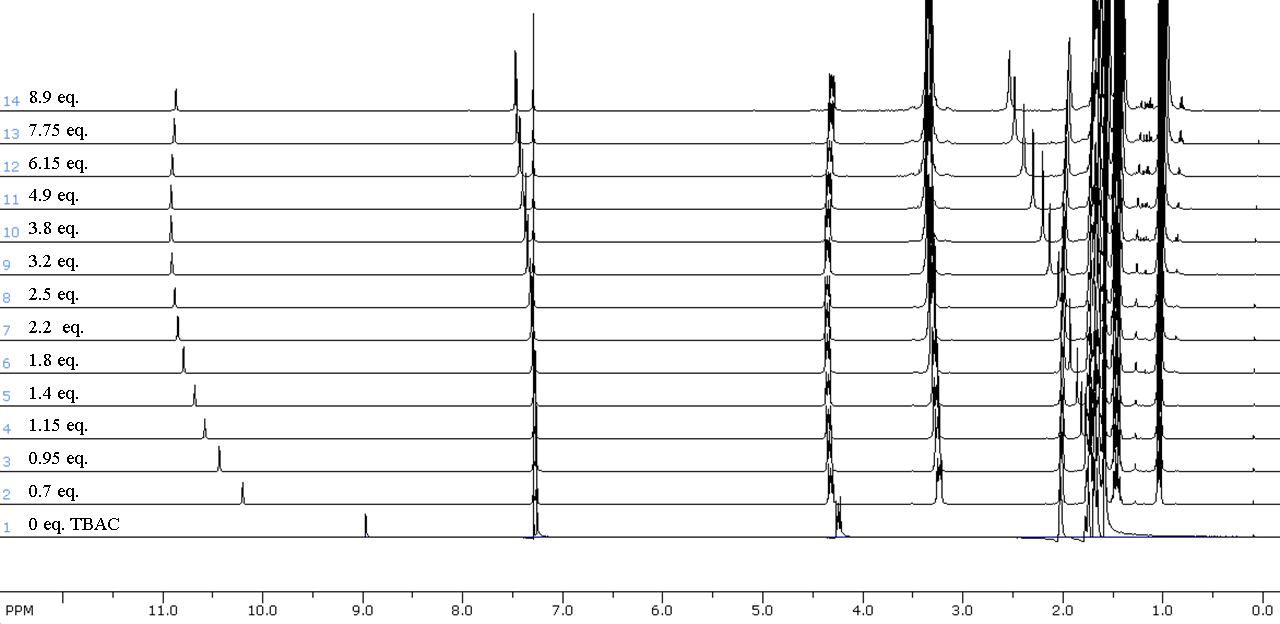


Figure S12. Stacked 1H NMR spectra (A) and titration curves (B) of [**9a**] = 12.5mM with TBAC (400 MHz, CDCl3, 298 K).

A)



B)

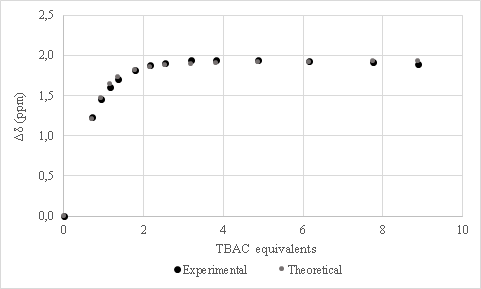
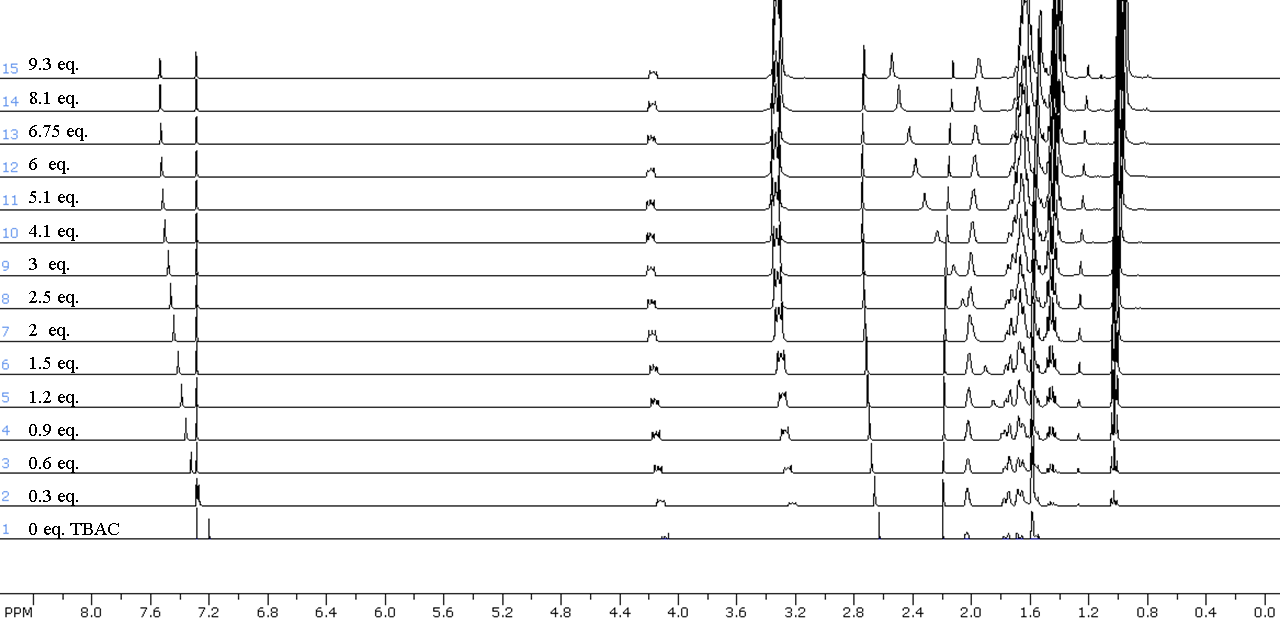


Figure S13. Stacked 1H NMR spectra (A) and titration curves (B) of [**9d**] = 12.5mM with TBAC (400 MHz, CDCl3, 298 K).

A)



B)

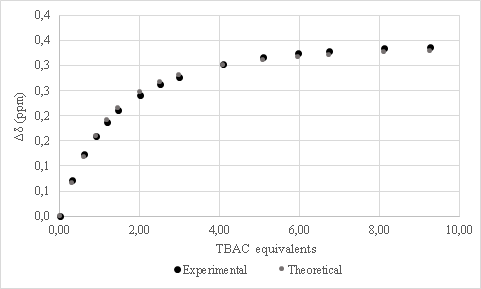
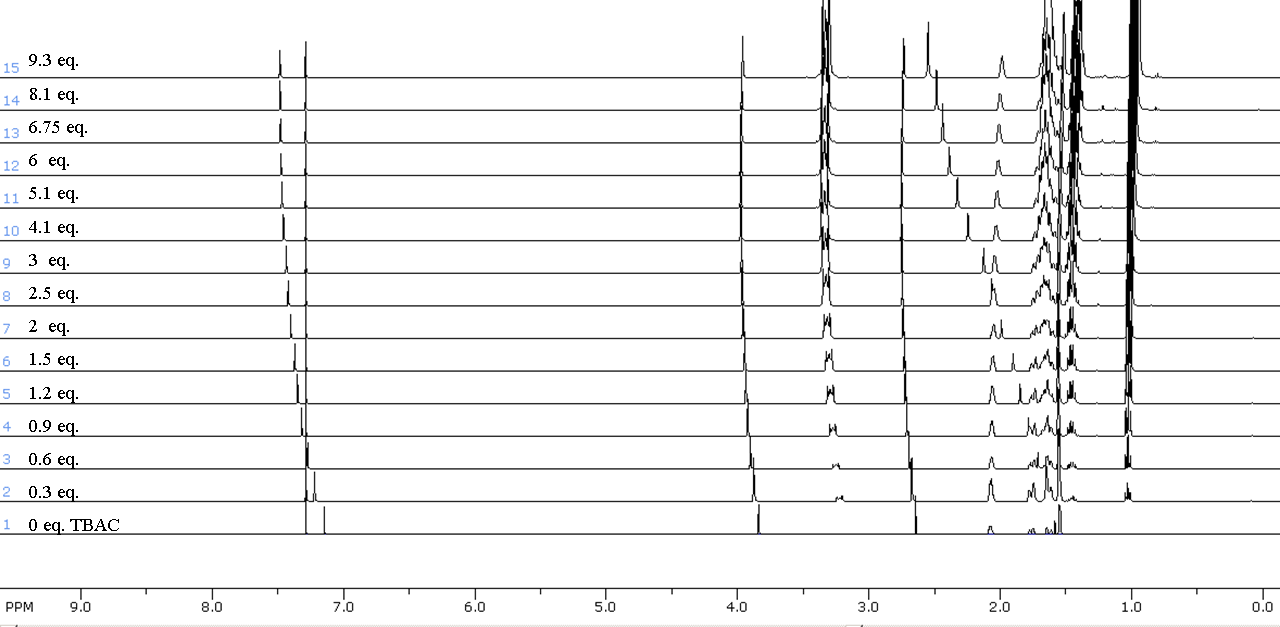


Figure S14. Stacked 1H NMR spectra (A) and titration curves (B) of [**10b**] = 12.5mM with TBAC (400 MHz, CDCl3, 298 K).

A)



B)

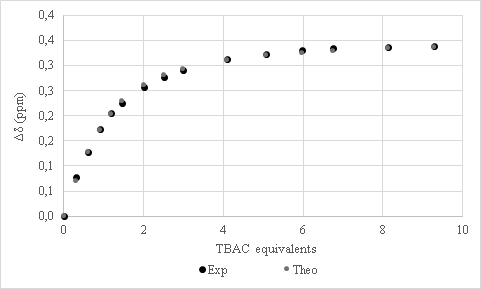


Figure S15. Stacked 1H NMR spectra (A) and titration curves (B) of [11b] = 12.5mM with TBAC (400 MHz, CDCl3, 298 K).

## HPTS-based fluorescence assays

### Preparation of EYPC liposomes for HPTS-based fluorescence assays

The procedures are the same as the *Preparation of EYPC liposomes for lucigenin-based fluorescence assays*, except that the lucigenin is replaced by a HPTS solution of 0.1 mM containing 100 mM of NaCl or NaClO4.

### HPTS-based fluorescence assays with EYPC liposomes

The procedures are the same as the *Lucigenin-based fluorescence assays with EYPC liposomes*, except that it was with the liposomes containing the HPTS probe.

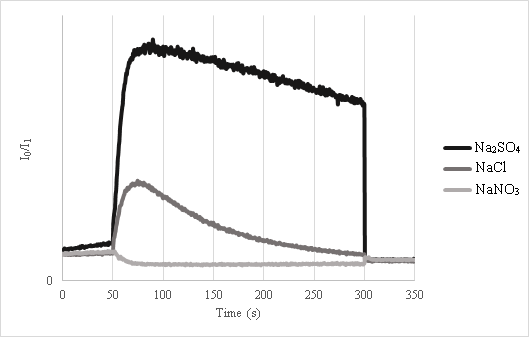


Figure S16. HPTS-based transport assay of imidazolium salt **9d** at 15 mol% relative to EYPC. Intravesicular: 0.1 HPTS, 100 mM NaOCl4, 10 mM phosphate buffer (pH=6.2). Extravesicular: 100 mM NaX or Na2X, 10 mM phosphate buffer (pH=6.2).

## U-tube experiments

U-tube experiment is performed in a U-shaped glass tube in which two aqueous phases are separated by a bulk hydrophobic solvent, such as chloroform (Figure S17).*(3)* U-tube experiment was performed with transporter **9d** in order to confirm the carrier transport mechanism.

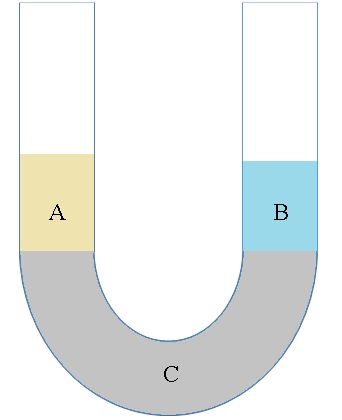


Figure S17. U-tube experiment scheme. A: Aqueous phase containing 0.1 mM Lucigenin (water, 5 mL). B: Receiving phase (water, 5 mL). C: Bulk organic phase (CHCl3, 10 mL) with transporter **9d** (1mM) or without.

If transporter **9d** is able to carry NO3- anions that are paired with the lucigenin dication across the bulk organic solvent, fluorescence should increase over time.

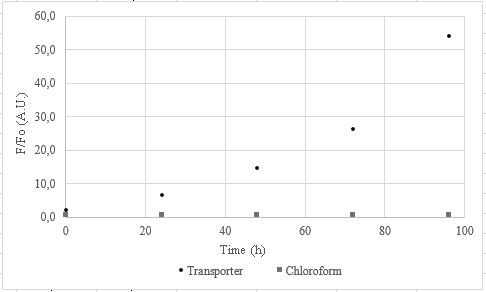


Figure S18. U-tube experiment results. Experiment conditions are shown in Figure S17.

U-tube experiments are conclusive with compound **9d** transporting NO3- using a carrier mechanism. Paired with EYPC:Cholesterol and DPPC studies, we believe it is strong evidence that molecule **9d** transport anion following a carrier mechanism.

## References

(1) Shahane, S.; Toupet, L.; Fischmeister, C.; Bruneau, C. *Eur. J. Inorg. Chem.* **2013,** 54.

(2) Belser, T.; Stöhr, M.; Pfaltz, A. *J. Am. Chem. Soc.* **2005,** *127*, 8720.

(3) Gokel, G. W.; Daschbach, M. M. Coord. Chem. Rev. 2008, 252, 886.