



Pivotal role of median eminence tanycytes for hypothalamic function and neurogenesis

Karine Rizzoti*, Robin Lovell-Badge**

The Francis Crick Institute, Mill Hill Laboratory, The Ridgeway, Mill Hill, London NW7 1AA, UK



ARTICLE INFO

Article history:

Received 30 June 2016

Accepted 11 August 2016

Available online 13 August 2016

Keywords:

Median eminence

Tanycyte

Neurogenesis

ABSTRACT

Along with the sub-ventricular zone of the forebrain lateral ventricles and the sub-granular zone of the dentate gyrus in the hippocampus, the hypothalamus has recently emerged as a third gliogenic and neurogenic niche in the central nervous system. The hypothalamus is the main regulator of body homeostasis because it centralizes peripheral information to regulate crucial physiological functions through the pituitary gland and the autonomic nervous system. Its ability to sense signals originating outside the brain relies on its exposure to blood-born molecules through the median eminence, which is localized outside the blood-brain barrier. Within the hypothalamus, a population of specialized radial glial cells, the tanycytes, control exposure to blood-born signals by acting both as sensors and regulators of the hypothalamic input and output. In addition, lineage-tracing experiments have recently revealed that tanycytes represent a population of hypothalamic stem cells, defining them as a pivotal cell type within the hypothalamus. Hypothalamic neurogenesis has moreover been shown to have an important role in feeding control and energy metabolism, which challenges previous knowledge and offers new therapeutic options.

Crown Copyright © 2016 Published by Elsevier Ireland Ltd. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

1. Introduction

The neuronal and gliogenic cell types of the mammalian central nervous system (CNS) are essentially generated during embryonic development, but new cells also emerge post-natally both in normal and pathological situations. These encompass cell turnover for specific neuronal populations, such as olfactory bulb neurons, and, to some degree, regeneration in response to injury. Adult-born cells are also thought to underlie aspects of brain plasticity. At least some of these new cells differentiate from neural stem cells (NSCs) found in restricted microenvironments, defined as niches, where their maintenance, proliferative and differentiation potential are tightly controlled (Dimou and Gotz, 2014). The two main niches in the mammalian brain are located in the sub-ventricular zone (SVZ) of the forebrain lateral ventricles and in the sub-granular zone (SGZ) of the dentate gyrus of the hippocampus. In these two regions extensive studies have described the components and architecture of the niche. NSCs of the SVZ and SGZ are astroglial. They

are mostly quiescent, while immediate progenitors, or transit-amplifying cells, are committed toward differentiation and possess a higher proliferative potential. Within the niche, interactions with neighbouring cells are important, but signals from the periphery are also sensed by NSCs as they are in direct contact with capillaries and, for SVZ NSCs, also with the cerebro-spinal fluid (CSF). In rodents and non-human primates, NSCs give rise to neurons, astrocytes and oligodendrocytes. In the SVZ of rodents and many other mammals the NSCs give rise to neuroblasts that form a rostral migratory stream (RMS) to give rise to olfactory bulb neurons. In humans, however, the adult SVZ generates mostly striatal interneurons. These therefore have a different identity compared to other mammals and they are hypothesized to underlie specific aspects of human neural plasticity (Ernst et al., 2014). In the hippocampus, many more neurons are generated than survive, but those that do integrate into existing circuits locally. Evidence suggests that neurogenesis in the dentate gyrus is important for certain types of learning and memory. The rates of neurogenesis in the dentate gyrus of human and rodents are comparable, but cell turnover appears more extensive in humans (Spalding et al., 2013).

Aside from these two regions, the hypothalamus has recently emerged as a third site of postnatal neurogenesis and gliogenesis. The hypothalamus is the central regulator of body homeostasis and

* Corresponding author.

** Corresponding author.

E-mail addresses: karine.rizzoti@crick.ac.uk (K. Rizzoti), robin.lovell-badge@crick.ac.uk (R. Lovell-Badge).

of several important processes such as feeding, growth, reproduction, stress and more generally metabolism (Saper and Lowell, 2014). It is organized in multiple nuclei, or groups of neurons, arranged around a small ventral region of the third ventricle. Each nucleus regulates different physiological functions, such as circadian rhythms by the supra-chiasmatic nucleus, or feeding behaviour by the arcuate nucleus. At the base of this ventricle, and therefore within the hypothalamus, the median eminence (ME) is an important site of information transfer because the blood-brain-barrier (BBB) is interrupted, defining the ME as a circumventricular organ (CVO) (Miyata, 2015). This implies a local transfer of molecules to and from the bed of fenestrated capillaries of the hypophyseal portal system located on the ventral-most aspect of ME. The hypothalamus can therefore sense and centralize information from the periphery, and also from other brain regions via neuronal connections, to regulate pituitary hormone secretions and to control other functions such as appetite, sleep and aging.

In contrast with the SVZ and the SGZ, we know very little about the hypothalamic NSC niche. As we will discuss here, lineage-tracing experiments have demonstrated that a population of specialized radial glial cells called tanyocytes (Rodriguez et al., 2005) have gliogenic and neurogenic properties. Tanyocyte cell bodies are located around the base of the third ventricle. These cells are morphologically defined by the presence of a single long basal process and are mostly devoid of cilia. Tanyocytes are a heterogeneous cell population, with the different sub-types designated according to their dorso-ventral location, and whose processes reach toward the hypothalamic parenchyma, or, ventrally, toward the fenestrated capillaries of ME (Fig. 1). These tanyocytes are therefore unique among other NSC populations because they have unrestricted access to blood-borne signals and are also in contact with the CSF. As we will review here, these features endow them with both unique and crucial properties as hypothalamic sensors and sentinels that distinguish them from other NSCs. Here we will first describe the ontogeny of hypothalamic tanyocytes and review their specific properties at the ME before describing their NSC potential,

and the significance of hypothalamic neurogenesis.

2. Embryonic origin of hypothalamic tanyocytes

The hypothalamus develops from the embryonic ventral forebrain (Ferran et al., 2015). During the specification of the neural plate, at 8 days post-coitum (dpc) in mice, the prospective hypothalamus is situated at the midline, in the rostral most position. It is in contact with the future pituitary, which is present as the hypophyseal placode at this stage, in the adjacent ectoderm. As the neural plate bends to close (McShane et al., 2015), increased proliferation of the dorsal telencephalic progenitors versus ventral ones induces an apparent shift of the prospective hypothalamus, which becomes localized posteriorly and ventrally to the telencephalic vesicles in 9.5dpc mouse embryos. At the midline of the hypothalamic neuroepithelium, right above the developing pituitary or Rathke's Pouch, the infundibulum becomes apparent from 9.5dpc. Morphologically this appears as a local extension of the neuroepithelium toward the developing pituitary that gives rise to the ME, to the pituitary stalk, which connects ME to the gland, and to the posterior lobe of the pituitary (Fig. 2). Tanyocytes also originate in the infundibulum, from which glial cell types will mostly differentiate in the embryo (Goto et al., 2015; Pearson and Placzek, 2013), however infundibular progenitors have the potential to generate neurons *in vitro* (Pearson et al., 2011).

The secreted molecule Sonic Hedgehog (SHH) is crucial early on for specification, and later for regionalisation of the hypothalamus (Manning et al., 2006; Szabo et al., 2009; Zhao et al., 2012; Trowe et al., 2013). Emergence of the infundibulum has been shown to rely on an antagonism between members of the bone morphogenic protein (BMP) family and SHH, which is excluded from the infundibulum (Zhao et al., 2012; Trowe et al., 2013). Members of the fibroblast growth factor family (FGFs) are present in the infundibulum, and required for infundibular cell expansion in chick (Pearson et al., 2011). The NOTCH pathway is also necessary for infundibulum formation as deletion of the NOTCH effectors *Hes1*

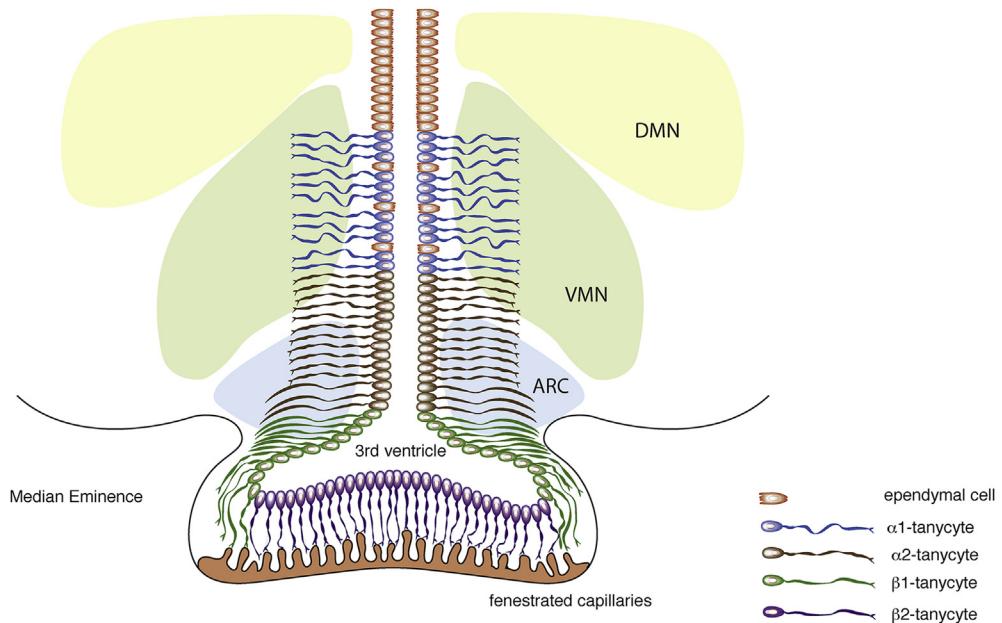


Fig. 1. Distribution of different tanyocyte subtypes along the third ventricular surface. The dorso-ventral organization of tanyocytes and ependymal cells in relation to the median eminence (ME) is illustrated here. β 2-tanyocytes are the most ventral tanyocytes, they are in contact with the fenestrated capillaries of ME and the third ventricle CSF that they isolate from free diffusion of blood-borne signals (Mullier et al., 2010). Just dorsal to these, β 1-tanyocytes perform the same barrier function for the arcuate nucleus (Rodriguez et al., 2005). α 1 and α 2 tanyocytes are present dorsally to β cells. ARC = arcuate nucleus, VMN = ventro-medial nucleus, DMN = dorso-medial nucleus.

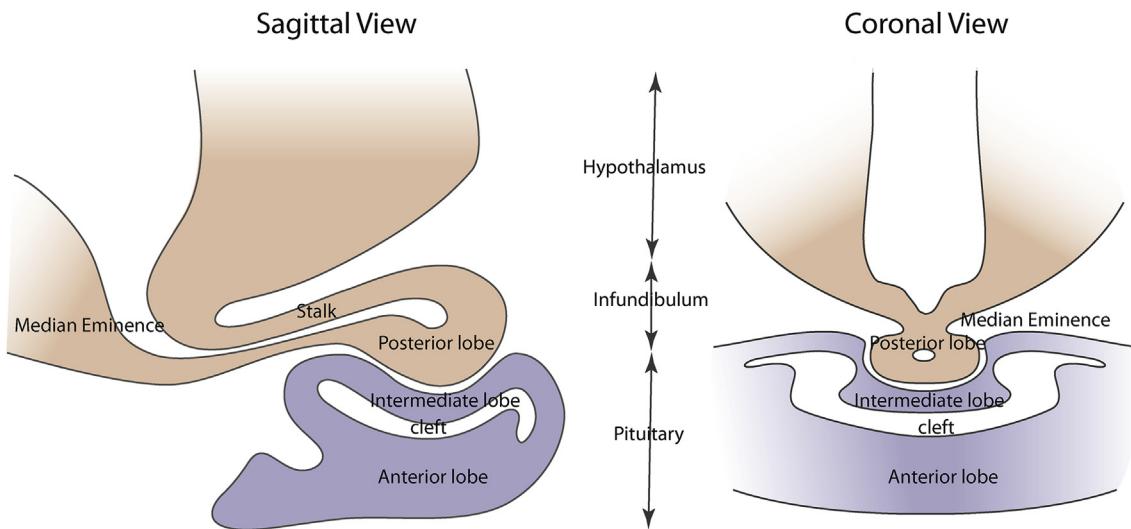


Fig. 2. Appearance of the infundibulum at 16.5dpc in the mouse. On a sagittal view the infundibulum is clearly observed as an extension from the ventral floor of the third ventricle. The distal most region in close apposition with the future intermediate lobe of the pituitary, will become the posterior lobe of the gland, containing specialized glial cells, termed pituicytes, and axons terminals from oxytocin and vasopressin neurons. The infundibulum will also give rise to the pituitary stalk, the physical link between the median eminence (ME) and the pituitary. Proximally, it will form ME itself, comprising tanyctyes, axons terminals from hypothalamic neurons and a bed of fenestrated capillaries. Vascularization of mesodermal and neural crest origin gradually develops in the embryonic pituitary and median eminence (Etchevers et al., 2001).

and *Hes5* result in premature cell cycle exit in this region, preventing evagination (Goto et al., 2015). In contrast, the failures in infundibular formation observed in embryos deleted for the transcription factor LHX2 (Zhao et al., 2010) and also TBX3 (Trove et al., 2013) are thought to result from hyperproliferation, demonstrating the requirement for a delicate balance between cell proliferation and migration for proper infundibular morphogenesis (Pearson et al., 2011). Proper infundibular morphogenesis is important because both induction and maintenance of Rathke's pouch rely on infundibular signals (Takuma et al., 1998).

Tanyctyes emerge late during gestation and terminal differentiation is completed post-natally, within a month in rats (Rodriguez et al., 2005). The transcription factors LHX2 and RAX are important regulators of ventral hypothalamic development and tanyctye specification and differentiation (Zhao et al., 2010; Salvatierra et al., 2014; Lu et al., 2013; Shimogori et al., 2010). They are both expressed in the developing hypothalamus and maintained post-natally in tanyctyes (Shimogori et al., 2010; Salvatierra et al., 2014). Embryonic deletion of *Lhx2* prevents proper tanyctye specification in the embryo where an expansion of ependymal cell fate marker expression is observed (Salvatierra et al., 2014). Post-natal terminal differentiation of α - and β -tanyctyes is impaired in these animals; morphology is affected with acquisition of multiple cilia, characteristic of ependymal cells, but the phenotypic conversion is incomplete because retention of some tanyctytic features is observed. Early post-natal deletion of *Lhx2* in tanyctyes does not result in expansion of ependymal cell marker expression, but it still prevents proper tanyctye differentiation. Lack of RAX expression, a direct target of LHX2, is thought to explain this phenotype (Salvatierra et al., 2014). Altogether these data suggest that LHX2 is required for tanyctye specification in the embryo, and for their differentiation post-natally. Moreover, while there is a relative flexibility between tanyctytic and ependymal cell fates in the embryo, once the cells have been specified post-natally this plasticity appears to be lost. In addition, manipulation of the WNT signalling pathway in the ventro-medial hypothalamus suggests that it is involved in post-natal regulation of tanyctye numbers (Wang et al., 2012). In conclusion, hypothalamic adult SCs originate from foetal infundibular progenitors, in parallel with what has been

demonstrated in the SVZ where slowly dividing embryonic neural progenitors give rise to adult NSCs (Furutachi et al., 2015), and suggested in the pituitary where Rathke's Pouch progenitors were shown to generate adult SCs (Rizzoti et al., 2013).

3. Tanyctyes act as hypothalamic sentinels, sensors, and neuroendocrine output modulators

3.1. Tanyctyes regulate diffusion of blood-borne molecules

The blood-brain barrier (BBB) allows a restricted and regulated access of blood borne molecules to the brain. It is characterized by the presence of tight junctions between endothelial cells, preventing free diffusion of molecules across this layer. The seven CVOs of the brain, including the ME, are defined as areas where the BBB is interrupted: capillaries in these regions are fenestrated (Miyata, 2015). These CVOs are also characterized by the presence of tanyctyes that are in contact with both endothelial cells and the ventricle. Examination of cell junctions and tissue permeability have suggested that, in the absence of BBB, tanyctyes restrict the diffusion of blood-borne signals to protect CSF integrity, acting therefore as ventricular barriers (Langlet et al., 2013a).

In the ME, this barrier function is restricted to ventral $\beta 2$ tanyctyes (Mullier et al., 2010), while $\beta 1$ tanyctyes are proposed to limit parenchymal diffusion of blood-borne molecules to the arcuate nucleus (Rodriguez et al., 2005). Permeability of the ME is modulated according to changing physiological situations, such as fasting. Metabolic signals need to reach feeding control circuits in the arcuate nucleus rapidly and fenestration of the capillaries is consequently increased. Tanyctyes that have been demonstrated to act as glucose sensors are involved in these changes (Bolborea and Dale, 2013). In response to fasting, and a consequent a drop in blood glucose, they can induce an increased vascular permeability through enhanced secretion of VEGFA (Langlet et al., 2013b). Leptin is a crucial peptide regulating food intake that is mostly secreted by adipocytes. It activates leptin receptors in the brain to regulate food intake, and resistance to leptin is associated with obesity (Roh et al., 2016). Mechanisms underlining resistance are unclear but a defective transport across the BBB has been proposed (El-Haschimi

et al., 2000). Tanyocytes have been proposed to represent intermediates for its diffusion in the medio-basal hypothalamus (MBH), suggesting that they regulate the hypothalamic response to leptin (Balland et al., 2014). Tanyocytes may not be the only cell type to regulate this response as a very recent study demonstrated that ME oligodendrocyte precursor cells are required for maintenance of leptin receptor positive dendrites, and therefore hypothalamic leptin sensing (Djogo et al., 2016).

Alterations in hypothalamic glucose sensing have been reported in Alzheimer disease (AD) patients and transgenic mouse model (Niwa et al., 2002). In addition, alterations in leptin levels and hypothalamic dysfunction are increasingly implicated in the weight loss characterizing AD (Ishii and Iadecola, 2015). It would therefore be of interest to investigate the functionality of tanyocytes and a possible link with AD.

3.2. Tanyocytes control neuroendocrine output at the ME

Fasting has pleiotropic physiological consequences, initially a drop in blood glucose as mentioned earlier, that tanyocytes and some hypothalamic neurons sense. It also induces a transient reduction in activity of the hypothalamo-pituitary-thyroid (HPT) axis, a crucial regulator of metabolism (Joseph-Bravo et al., 2015), and this contributes to reduced energy expenditure when calorific intake is low. On top of the HPT axis, hypothalamic Thyrotropin Releasing Hormone (TRH) is collected by ME capillaries and transported to the pituitary where it stimulates the secretion of Thyroid Stimulating Hormone (TSH) and prolactin. TSH induces secretion of the thyroid pro-hormone T₄ that must be converted to T₃ by the deiodinases Dio1 and 2 to be active. In turn, TH exerts a negative feedback on TRH synthesis and secretion. In the brain, Dio2 is predominantly expressed by ME tanyocytes. These are therefore important regulators of hypothalamic TH levels (Bolborea and Dale, 2013). TRH levels are also finely regulated, in particular by an ectopeptidase, the pyroglutamyl peptidase II (PPII) that hydrolyses the neuropeptide. In the hypothalamus, PPII is present in tanyocytes, particularly of the β 2 type, where its expression and activity are regulated positively by TH (Sanchez et al., 2009). β 2-tanyocytes are also associated with TRH neuron termini at the ME. Inhibition of PPII *in vivo* results in more TRH being secreted at the ME, strongly suggesting that β 2-tanyocytes regulate TRH levels and that they participate in the negative feedback action of TH on TRH secretion (Sanchez et al., 2009). Finally, tanyctic PPII levels increase transiently during fasting, implying that tanyocytes are involved in the reduction of TRH levels induced by caloric restriction (Lazcano et al., 2015). Moreover, regulation of the HPT axis by tanyocytes is proposed to be involved in weight fluctuations observed in some seasonal mammals (Ebling, 2015).

Gonadotrophin Releasing Hormone (GnRH) controls reproductive function. It is released at the ME and induces secretion of pituitary luteinising and folliculo-stimulating hormones; these in turn stimulate production of steroids within the gonads (Herbison, 2016). In addition, while decreased GnRH levels had been associated with aging (Yin et al., 2009), a causative link has been demonstrated where reduction in GnRH levels initiate systemic aging (Zhang et al., 2013). GnRH secretion is tightly regulated and tanyocytes, along with ME astrocytes and endothelial cells, play an important role (Prevot et al., 2010). Their cytoplasmic processes engulf GnRH axon termini at the ME, and this ensheathment is modulated according to the phase of the oestrous cycle (Prevot et al., 1999). Ensheatment is associated with a restricted access of axon termini to the perivascular space, while tanyocyte endfeet retraction allows access to blood vessels and leads to increased GnRH release (Prevot et al., 2003). The secreted molecules TGF α , β and Semaphorin7A, a chemorepulsive axon guidance molecule,

regulate the morphological changes observed in tanyocytes during the oestrous cycle (Prevot et al., 2010; Parkash et al., 2015). Sema7A is expressed by tanyocytes and has a dual role at the ME: it induces GnRH axon termini retraction and tanyocyte endfeet engulfment, resulting in decreased GnRH secretion (Parkash et al., 2015). All together these data show that tanyocytes have an important role in the control of GnRH secretion and therefore reproduction. Moreover, the causative effect of reduced GnRH levels during aging (Zhang et al., 2013) may suggest a role for tanyocyte during this process (see below).

4. Tanyocytes comprise a population of hypothalamic stem cells

Cell division had been detected in the post-natal hypothalamus, particularly in the ventral region surrounding the third ventricle, and in rodents this can be stimulated by infusion of different growth factors, such as BDNF (Pencea et al., 2001), EFG and FGF (Xu et al., 2005), IGF (Perez-Martin et al., 2010) and CNTF (Kokoeva et al., 2005). The presence of hypothalamic progenitors was suggested by experiments *in vitro* (Markakis et al., 2004), while active neurogenesis was proposed to occur in the hypothalamus (Xu et al., 2005; Perez-Martin et al., 2010; Kokoeva et al., 2005; Kokoeva et al., 2007; Batailler et al., 2014). The physiological relevance of this was first suggested by Kokoeva et al., in 2005, on the basis of the link that was seen between newly generated neurons and long-term weight loss observed in CNTF treated animals (Kokoeva et al., 2005). Thanks to the availability of different relevant Cre strains, cell lineage tracing experiments have now firmly established the existence of active hypothalamic neurogenesis and gliogenesis, as well as the SC potential of tanyocytes (Lee et al., 2012; Li et al., 2012; Robins et al., 2013; Haan et al., 2013; Robins et al., 2013a; Chaker et al., 2016). However there is still debate about the type(s) of tanyocyte that really represents hypothalamic NSCs (Fig. 3). In addition, more remains to be known about the physiological significance of hypothalamic neurogenesis. For example, in seasonally breeding mammals, the rate of hypothalamic cell proliferation has been shown to vary according to day length, with increased levels in the short photoperiod in sheep (Batailler et al., 2015), and similar seasonal changes relating to food intake correlate with changes in tanyocyte function and neurogenesis in Syrian hamsters (Ebling, 2015; Samms et al., 2015). However, while the correlations are good, functional data is lacking.

4.1. Characterization of hypothalamic NSCs

Several observations including morphological features pointed toward α 2-tanyocytes as potential NSCs (Rodriguez et al., 2005), and this was recently confirmed (Robins et al., 2013a). More precisely, GLAST^{CreERT2} lineage tracing experiments first showed that α -tanyocytes are the only subtype to express GLAST. These α -tanyocytes give rise to β 1-tanyocytes, suggesting that the former may represent NSCs while the latter are committed progenitors. In addition, precise dissection of the third ventricle sub-ventricular zone further revealed that α 2 are the only tanyocytes with neurosphere forming ability, implying that they are NSCs (Robins et al., 2013a). Consistent with the biology of other NSC populations, α -tanyocyte proliferation is stimulated by FGF2 (Robins et al., 2013a) and IGF (Perez-Martin et al., 2010). However, in the adult, GLAST^{CreERT2} lineage tracing analyses have revealed that α -tanyocytes mostly give rise to parenchymal astrocytes while very few neurons are generated (Robins et al., 2013a).

The secreted factor FGF10 is expressed selectively in some β -tanyocytes, revealing previously unsuspected cell heterogeneity in this population (Haan et al., 2013; Hajhosseini et al., 2008). Lineage

tracing experiments using FGF10^{CreERT2} show that β -tanyocytes differentiate more during the early post-weaning period and, in contrast with α -tanyocytes (Robins et al., 2013a), and that their progeny is predominantly neuronal. Newly generated neurons integrate in the arcuate and ventromedial nuclei (Haan et al., 2013).

An additional site of neurogenesis was detected using Nestin-CreERT2 (Lee et al., 2012). Early post-natally, β 2 cells are the most proliferative among tanyocytes. Lineage tracing using Nestin-CreERT2 reveals that these cells are also the most neurogenic in young animals (Lee et al., 2012). Both in young animals and in adults, newly generated neurons remain within the ME (Lee et al., 2012; Lee et al., 2014). In addition, lineage tracing in the adult using an independently generated strain of Nestin-CreERT2 recently showed that neurogenesis persists as the animal is aging, and that new neurons are produced in all regions of the hypothalamus (Chaker et al., 2016).

In conclusion, while it is difficult to compare lineage-tracing experiments using different drivers, especially when inducible recombinases are used and varying degrees of mosaicism obtained, tanyocyte sub-types appear to differ significantly both in their potential and contribution to hypothalamic cell-turnover. Importantly, a parenchymal NSC population has also been proposed to exist in the hypothalamus (Robins et al., 2013b; Li et al., 2012). Regardless of their ventricular or parenchymal origin, the newly generated neurons appear to be predominantly associated with feeding control.

4.2. Physiological relevance of hypothalamic neurogenesis

There is significant cell turn-over in the arcuate nucleus postnatally (McNay et al., 2012) and, in both young and adult animals, new hypothalamic neurons are responsive to signals related to feeding control (Kokoeva et al., 2005; Lee et al., 2012; Li et al., 2012; Haan et al., 2013). In addition, hypothalamic neurogenesis is modulated in response to diet, where high fat diet (HFD) has been reproducibly shown to inhibit neurogenesis within the MBH and increase progenitor apoptosis (Li et al., 2012; Lee et al., 2014; McNay et al., 2012). In sharp contrast, neurogenesis in the ME is increased in response to HFD, specifically in females (Lee et al., 2012, 2014).

The physiological significance of this effect is suggested by a reduction in weight gain when ME neurogenesis is prevented (Lee et al., 2012). In addition, caloric restriction is associated with reduced proliferation and a tendency toward reduced neurogenesis within the ME (Lee et al., 2014).

Leptin deficiency, which is associated with obesity, also results in loss of hypothalamic SCs and impaired MBH neurogenesis in mice (McNay et al., 2012). The association between HFD, impaired hypothalamic neurogenesis and weight gain is further strengthened by the observation that hypothalamic inflammation also affects hypothalamic SCs (Li et al., 2012). Obesity is associated with hypothalamic inflammation (Cai and Liu, 2011). As this inflammation precedes obesity onset, it is increasingly suspected to be the cause, rather than the consequence, of diet-induced metabolic disease (Valdearcos et al., 2015). Hypothalamic microglia mediate inflammation induced by HFD (Li et al., 2012) and this affects hypothalamic SCs, which display increased apoptosis, reduced proliferation and inhibition of neural differentiation (Li et al., 2012). Therefore, in response to HFD and as a target of hypothalamic inflammation, reduced MBH neurogenesis is clearly associated with weight gain.

Microglia mediated hypothalamic inflammation also appears to have a crucial role in initiating systemic aging (Zhang et al., 2013). In this context, the relevant targets of the inflammatory cascade are the GnRH neurons, causing them to secrete less GnRH, and this results in systemic aging (Zhang et al., 2013). Decreased neurogenesis is observed in both the hypothalamus and hippocampus. GnRH administration can slow the ageing process and rescues neurogenesis in old mice; while a causative effect is not demonstrated, both at least correlate (Zhang et al., 2013). Interestingly aging is associated with a tendency to gain weight, in both humans and rodents. It would be of interest to examine a potential connection between decreased hypothalamic neurogenesis and weight gain in this context.

Hypothalamic neurogenesis is clearly associated with feeding control and energy metabolism. The tanyocytes are increasingly appearing as a pivotal hypothalamic cell type regulating peripheral input, neuroendocrine output and to generate new hypothalamic cells. It is now crucial to understand how signals are integrated and

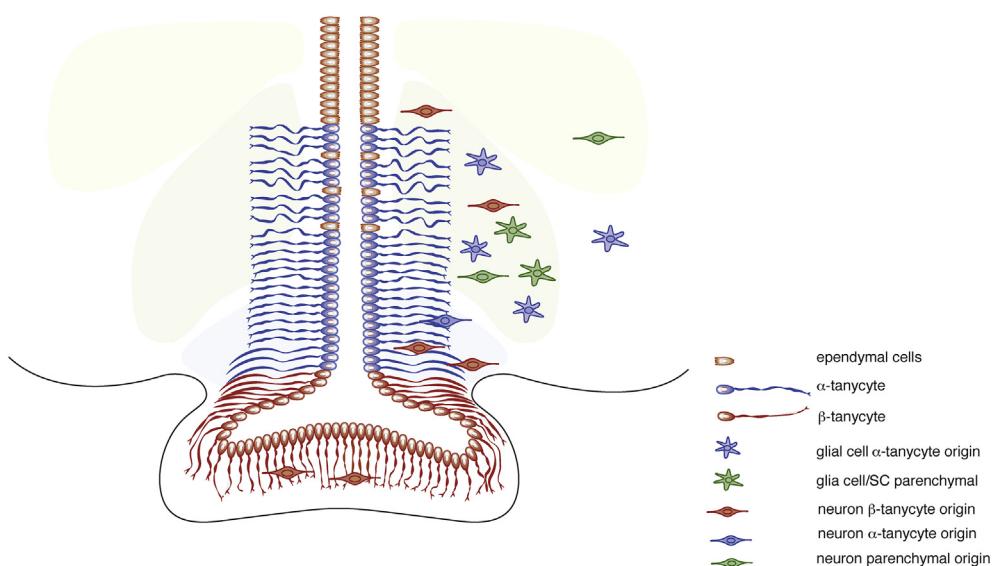


Fig. 3. Hypothalamic SCs and their progeny. β -tanyocytes can generate neurons that populate ME (Lee et al., 2012), and the mediobasal hypothalamus (MBH) (Haan et al., 2013). α -tanyocytes mostly generate parenchymal astrocytes and some rare neurons (Robins et al., 2013a). α -tanyocytes have also been proposed to give rise to β -tanyocytes defining the former as SCs, and the latter as progenitors (Robins et al., 2013a). Finally a parenchymal population of progenitors has been proposed to be present in the MBH (Li et al., 2012; Robins et al., 2013b).

the degree to which heterogeneity exists within this remarkable cell population.

5. Conclusion

Plasticity is an important aspect of hypothalamic function because constant adaptation to changing conditions is required to maintain homeostasis and to release appropriate signals, such as satiety after feeding. The recent demonstration that neurogenesis occurs in this region, and is altered in response to diet modification, suggests that modulation of the number of hypothalamic neurons may represent another way to adapt in response to changing physiological situations. This also implies that therapeutically modulating hypothalamic neurogenesis and/or neuronal populations may be beneficial, particularly in the context of metabolic syndromes. The development of induced Pluripotent SCs (iPSCs) ten years ago (Takahashi and Yamanaka, 2006) has been followed by remarkable progress toward regenerative medicine, disease modelling and drug screening. We can now generate many differentiated cell types from embryonic and/or iPSC, comprising hypothalamic neurons (Wataya et al., 2008; Wang et al., 2015; Merkle et al., 2015). In addition, hypothalamic cell transplantation can partially restore leptin responsiveness in leptin receptor-deficient mice (Czuprynski et al., 2011), demonstrating that cell manipulation in the hypothalamus has therapeutic benefits. Finally, it now appears essential to examine whether hypothalamic neurogenesis is involved in other physiological processes, such as puberty, pregnancy and lactation, where the organism needs to adapt to and trigger, especially in the case of puberty, a new physiological status. It would also be important to ask how neurogenesis relates functionally to seasonal changes in physiology and behaviour, associated with feeding or reproduction, and to aging.

Acknowledgements

We would like to thank members of the Lovell-Badge's lab for help and support, and the Research Illustration team at the Francis Crick Institute for assistance with the figures. This work was supported by the Francis Crick Institute, grant number 10107.

References

- Balland, E., Dam, J., Langlet, F., Caron, E., Steculorum, S., Messina, A., Rasika, S., Falluel-Morel, A., Anouar, Y., Dehouck, B., Trinquet, E., Jockers, R., Bouret, S.G., Prevot, V., 2014. Hypothalamic tanyocytes are an ERK-gated conduit for leptin into the brain. *Cell Metab.* 19, 293–301.
- Battailler, M., Droguerre, M., Baroncini, M., Fontaine, C., Prevot, V., Migaud, M., 2014. DCX-expressing cells in the vicinity of the hypothalamic neurogenic niche: a comparative study between mouse, sheep, and human tissues. *J. Comp. Neurol.* 522, 1966–1985.
- Battailler, M., Derouet, L., Butruille, L., Migaud, M., 2015. Sensitivity to the photoperiod and potential migratory features of neuroblasts in the adult sheep hypothalamus. *Brain Struct. Funct.* 221 (6), 3301–3314.
- Bolborea, M., Dale, N., 2013. Hypothalamic tanyocytes: potential roles in the control of feeding and energy balance. *Trends Neurosci.* 36, 91–100.
- Cai, D., Liu, T., 2011. Hypothalamic inflammation: a double-edged sword to nutritional diseases. *Ann. N. Y. Acad. Sci.* 1243, E1–E39.
- Chaker, Z., George, C., Petrovska, M., Caron, J.B., Lacube, P., Caille, I., Holzenberger, M., 2016. Hypothalamic neurogenesis persists in the aging brain and is controlled by energy-sensing IGF-I pathway. *Neurobiol. Aging* 41, 64–72.
- Czuprynski, A., Zhou, Y.D., Chen, X., McNay, D., Anderson, M.P., Flier, J.S., Macklis, J.D., 2011. Transplanted hypothalamic neurons restore leptin signaling and ameliorate obesity in db/db mice. *Science* 334, 1133–1137.
- Dimou, L., Gotz, M., 2014. Glial cells as progenitors and stem cells: new roles in the healthy and diseased brain. *Physiol. Rev.* 94, 709–737.
- Djogo, T., Robins, S.C., Schneider, S., Kryzskaya, D., Liu, X., Mingay, A., Gillon, C.J., Kim, J.H., Storch, K.F., Boehm, U., Bourque, C.W., Stroh, T., Dimou, L., Kokoeva, M.V., 2016. Adult NG2-Glia are required for median eminence-mediated leptin sensing and body weight control. *Cell Metab.* 23, 797–810.
- Ebling, F.J., 2015. Hypothalamic control of seasonal changes in food intake and body weight. *Front. Neuroendocrinol.* 37, 97–107.
- El-Haschimi, K., Pierroz, D.D., Hileman, S.M., Bjorbaek, C., Flier, J.S., 2000. Two defects contribute to hypothalamic leptin resistance in mice with diet-induced obesity. *J. Clin. Invest.* 105, 1827–1832.
- Ernst, A., Alkass, K., Bernard, S., Salehpour, M., Perl, S., Tisdale, J., Possnert, G., Druid, H., Frisen, J., 2014. Neurogenesis in the striatum of the adult human brain. *Cell* 156, 1072–1083.
- Etchevers, H.C., Vincent, C., Le Douarin, N.M., Couly, G.F., 2001. The cephalic neural crest provides pericytes and smooth muscle cells to all blood vessels of the face and forebrain. *Development* 128, 1059–1068.
- Ferran, J.L., Puelles, L., Rubenstein, J.L., 2015. Molecular codes defining rostrocaudal domains in the embryonic mouse hypothalamus. *Front. Neuroanat.* 9, 46.
- Furutachi, S., Miya, H., Watanabe, T., Kawai, H., Yamasaki, N., Harada, Y., Imayoshi, I., Nelson, M., Nakayama, K.I., Hirabayashi, Y., Gotoh, Y., 2015. Slowly dividing neural progenitors are an embryonic origin of adult neural stem cells. *Nat. Neurosci.* 18, 657–665.
- Goto, M., Hojo, M., Ando, M., Kita, A., Kitagawa, M., Ohtsuka, T., Kageyama, R., Miyamoto, S., 2015. Hes1 and Hes5 are required for differentiation of pituicytes and formation of the neurohypophysis in pituitary development. *Brain Res.*
- Haan, N., Goodman, T., Najdi-Samiei, A., Stratford, C.M., Rice, R., El Agha, E., Bellusci, S., Hajhosseini, M.K., 2013. Fgf10-expressing tanyocytes add new neurons to the appetite/energy-balance regulating centers of the postnatal and adult hypothalamus. *J. Neurosci.* 33, 6170–6180.
- Hajhosseini, M.K., De Langhe, S., Lana-Eloa, E., Morrison, H., Sparshott, N., Kelly, R., Sharpe, J., Rice, D., Bellusci, S., 2008. Localization and fate of Fgf10-expressing cells in the adult mouse brain implicate Fgf10 in control of neurogenesis. *Mol. Cell Neurosci.* 37, 857–868.
- Herbison, A.E., 2016. Control of puberty onset and fertility by gonadotropin-releasing hormone neurons. *Nat. Rev. Endocrinol.*
- Ishii, M., Iadecola, C., 2015. Metabolic and non-cognitive manifestations of Alzheimer's disease: the hypothalamus as both culprit and target of pathology. *Cell Metab.* 22, 761–776.
- Joseph-Bravo, P., Jaimes-Hoy, L., Maria Uribe, R., Charli, J.L., 2015. 60 YEARS OF NEUROENDOCRINOLOGY: TRH, the first hypophysiotropic releasing hormone isolated: control of the pituitary-thyroid axis. *J. Endocrinol.* 226, T85–T100.
- Kokoeva, M.V., Yin, H., Flier, J.S., 2005. Neurogenesis in the hypothalamus of adult mice: potential role in energy balance. *Science* 310, 679–683.
- Kokoeva, M.V., Yin, H., Flier, J.S., 2007. Evidence for constitutive neural cell proliferation in the adult murine hypothalamus. *J. Comp. Neurol.* 505, 209–220.
- Langlet, F., Mullier, A., Bouret, S.G., Prevot, V., Dehouck, B., 2013a. Tanyocyte-like cells form a blood-cerebrospinal fluid barrier in the circumventricular organs of the mouse brain. *J. Comp. Neurol.* 521, 3389–3405.
- Langlet, F., Levin, B.E., Luquet, S., Mazzone, M., Messina, A., Dunn-Meynell, A.A., Balland, E., Lacombe, A., Mazur, D., Carmeliet, P., Bouret, S.G., Prevot, V., Dehouck, B., 2013b. Tanycytic VEGF-A boosts blood-hypothalamus barrier plasticity and access of metabolic signals to the arcuate nucleus in response to fasting. *Cell Metab.* 17, 607–617.
- Lazcano, I., Cabral, A., Uribe, R.M., Jaimes-Hoy, L., Perello, M., Joseph-Bravo, P., Sanchez-Jaramillo, E., Charli, J.L., 2015. Fasting enhances pyroglutamyl peptidase II activity in tanyocytes of the mediobasal hypothalamus of male adult rats. *Endocrinology* 156, 2713–2723.
- Lee, D.A., Bedont, J.L., Pak, T., Wang, H., Song, J., Miranda-Angulo, A., Takiar, V., Charubhumi, V., Balordi, F., Takebayashi, H., Aja, S., Ford, E., Fishell, G., Blackshaw, S., 2012. Tanyocytes of the hypothalamic median eminence form a diet-responsive neurogenic niche. *Nat. Neurosci.* 15, 700–702.
- Lee, D.A., Yoo, S., Pak, T., Salvatierra, J., Velarde, E., Aja, S., Blackshaw, S., 2014. Dietary and sex-specific factors regulate hypothalamic neurogenesis in young adult mice. *Front. Neurosci.* 8, 157.
- Li, J., Tang, Y., Cai, D., 2012. IKKbeta/NF-kappaB disrupts adult hypothalamic neural stem cells to mediate a neurodegenerative mechanism of dietary obesity and pre-diabetes. *Nat. Cell Biol.* 14, 999–1012.
- Lu, F., Kar, D., Gruenig, N., Zhang, Z.W., Cousins, N., Rodgers, H.M., Swindell, E.C., Jamrich, M., Schuurmans, C., Mathers, P.H., Kurrasch, D.M., 2013. Rax is a selector gene for mediobasal hypothalamic cell types. *J. Neurosci.* 33, 259–272.
- Manning, L., Ohyama, K., Saeger, B., Hatano, O., Wilson, S.A., Logan, M., Placzek, M., 2006. Regional morphogenesis in the hypothalamus: a BMP-Tbx2 pathway coordinates fate and proliferation through Shh downregulation. *Dev. Cell.* 11, 873–885.
- Markakis, E.A., Palmer, T.D., Randolph-Moore, L., Rakic, P., Gage, F.H., 2004. Novel neuronal phenotypes from neural progenitor cells. *J. Neurosci.* 24, 2886–2897.
- McNay, D.E., Briancon, N., Kokoeva, M.V., Maratos-Flier, E., Flier, J.S., 2012. Remodeling of the arcuate nucleus energy-balance circuit is inhibited in obese mice. *J. Clin. Invest.* 122, 142–152.
- McShane, S.G., Mole, M.A., Savery, D., Greene, N.D., Tam, P.P., Copp, A.J., 2015. Cellular basis of neuroepithelial bending during mouse spinal neural tube closure. *Dev. Biol.* 404, 113–124.
- Merkle, F.T., Maroof, A., Wataya, T., Sasai, Y., Studer, L., Eggan, K., Schier, A.F., 2015. Generation of neuropeptidergic hypothalamic neurons from human pluripotent stem cells. *Development* 142, 633–643.
- Miyata, S., 2015. New aspects in fenestrated capillary and tissue dynamics in the sensory circumventricular organs of adult brains. *Front. Neurosci.* 9, 390.
- Mullier, A., Bouret, S.G., Prevot, V., Dehouck, B., 2010. Differential distribution of tight junction proteins suggests a role for tanyocytes in blood-hypothalamus barrier regulation in the adult mouse brain. *J. Comp. Neurol.* 518, 943–962.
- Niwa, K., Kazama, K., Younkin, S.G., Carlson, G.A., Iadecola, C., 2002. Alterations in cerebral blood flow and glucose utilization in mice overexpressing the amyloid precursor protein. *Neurobiol. Dis.* 9, 61–68.

- Parkash, J., Messina, A., Langlet, F., Cimino, I., Loyens, A., Mazur, D., Gallet, S., Balland, E., Malone, S.A., Pralong, F., Cagnoni, G., Schellino, R., De Marchis, S., Mazzone, M., Pasterkamp, R.J., Tamagnone, L., Prevot, V., Giacobini, P., 2015. Semaphorin7A regulates neuroglial plasticity in the adult hypothalamic median eminence. *Nat. Commun.* 6, 6385.
- Pearson, C.A., Placzek, M., 2013. Development of the medial hypothalamus: forming a functional hypothalamic-neurohypophyseal interface. *Curr. Top. Dev. Biol.* 106, 49–88.
- Pearson, C.A., Ohyama, K., Manning, L., Aghamohammadzadeh, S., Sang, H., Placzek, M., 2011. FGF-dependent midline-derived progenitor cells in hypothalamic infundibular development. *Development* 138, 2613–2624.
- Pencea, V., Bingaman, K.D., Wiegand, S.J., Luskin, M.B., 2001. Infusion of brain-derived neurotrophic factor into the lateral ventricle of the adult rat leads to new neurons in the parenchyma of the striatum, septum, thalamus, and hypothalamus. *J. Neurosci.* 21, 6706–6717.
- Perez-Martin, M., Cifuentes, M., Grondona, J.M., Lopez-Avalos, M.D., Gomez-Pinedo, U., Garcia-Verdugo, J.M., Fernandez-Llubres, P., 2010. IGF-I stimulates neurogenesis in the hypothalamus of adult rats. *Eur. J. Neurosci.* 31, 1533–1548.
- Prevot, V., Croix, D., Bouret, S., Dutoit, S., Tramu, G., Stefano, G.B., Beauvillain, J.C., 1999. Definitive evidence for the existence of morphological plasticity in the external zone of the median eminence during the rat estrous cycle: implication of neuro-glio-endothelial interactions in gonadotropin-releasing hormone release. *Neuroscience* 94, 809–819.
- Prevot, V., Rio, C., Cho, G.J., Lomniczi, A., Heger, S., Neville, C.M., Rosenthal, N.A., Ojeda, S.R., Corfas, G., 2003. Normal female sexual development requires neuregulin-erbB receptor signaling in hypothalamic astrocytes. *J. Neurosci.* 23, 230–239.
- Prevot, V., Bellefontaine, N., Baroncini, M., Sharif, A., Hanchate, N.K., Parkash, J., Campagne, C., de Seranno, S., 2010. Gonadotrophin-releasing hormone nerve terminals, tanycytes and neurohaemal junction remodelling in the adult median eminence: functional consequences for reproduction and dynamic role of vascular endothelial cells. *J. Neuroendocrinol.* 22, 639–649.
- Rizzoti, K., Akiyama, H., Lovell-Badge, R., 2013. Mobilized adult pituitary stem cells contribute to endocrine regeneration in response to physiological demand. *Cell Stem Cell.* 13, 419–432.
- Robins, S.C., Stewart, I., McNay, D.E., Taylor, V., Giachino, C., Goetz, M., Ninkovic, J., Briancon, N., Maratos-Flier, E., Flier, J.S., Kokoeva, M.V., Placzek, M., 2013a. alpha-Tanycytes of the adult hypothalamic third ventricle include distinct populations of FGF-responsive neural progenitors. *Nat. Commun.* 4, 2049.
- Robins, S.C., Trudel, E., Rotondi, O., Liu, X., Djogo, T., Kryzskaya, D., Bourque, C.W., Kokoeva, M.V., 2013b. Evidence for NG2-glia derived, adult-born functional neurons in the hypothalamus. *PLoS One* 8, e78236.
- Rodriguez, E.M., Blazquez, J.L., Pastor, F.E., Pelaez, B., Pena, P., Peruzzo, B., Amat, P., 2005. Hypothalamic tanycytes: a key component of brain-endocrine interaction. *Int. Rev. Cytol.* 247, 89–164.
- Roh, E., Song do, K., Kim, M.S., 2016. Emerging role of the brain in the homeostatic regulation of energy and glucose metabolism. *Exp. Mol. Med.* 48, e216.
- Salvatierra, J., Lee, D.A., Zibetti, C., Duran-Moreno, M., Yoo, S., Newman, E.A., Wang, H., Bedont, J.L., de Melo, J., Miranda-Angulo, A.L., Gil-Perotin, S., Garcia-Verdugo, J.M., Blackshaw, S., 2014. The LIM homeodomain factor Lhx2 is required for hypothalamic tanycyte specification and differentiation. *J. Neurosci.* 34, 16809–16820.
- Samms, R.J., Lewis, J.E., Lory, A., Fowler, M.J., Cooper, S., Warner, A., Emmerson, P., Adams, A.C., Luckett, J.C., Perkins, A.C., Wilson, D., Barrett, P., Tsintzas, K., Ebling, F.J., 2015. Antibody-mediated inhibition of the FGFR1c isoform induces a catabolic lean state in siberian hamsters. *Curr. Biol.* 25, 2997–3003.
- Sanchez, E., Vargas, M.A., Singru, P.S., Pascual, I., Romero, F., Fekete, C., Charli, J.L., Lechan, R.M., 2009. Tanyocyte pyroglutamyl peptidase II contributes to regulation of the hypothalamic-pituitary-thyroid axis through glial-axonal associations in the median eminence. *Endocrinology* 150, 2283–2291.
- Saper, C.B., Lowell, B.B., 2014. The hypothalamus. *Curr. Biol.* 24, R1111–R1116.
- Shimogori, T., Lee, D.A., Miranda-Angulo, A., Yang, Y., Wang, H., Jiang, L., Yoshida, A.C., Kataoka, A., Mashiko, H., Avetisyan, M., Qi, L., Qian, J., Blackshaw, S., 2010. A genomic atlas of mouse hypothalamic development. *Nat. Neurosci.* 13, 767–775.
- Spalding, K.L., Bergmann, O., Alkass, K., Bernard, S., Salehpour, M., Huttner, H.B., Bostrom, E., Westerlund, I., Vial, C., Buchholz, B.A., Possnert, G., Mash, D.C., Druid, H., Frisen, J., 2013. Dynamics of hippocampal neurogenesis in adult humans. *Cell* 153, 1219–1227.
- Szabo, N.E., Zhao, T., Cankaya, M., Theil, T., Zhou, X., Alvarez-Bolado, G., 2009. Role of neuroepithelial Sonic hedgehog in hypothalamic patterning. *J. Neurosci.* 29, 6989–7002.
- Takahashi, K., Yamanaka, S., 2006. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell* 126, 663–676.
- Takuma, N., Sheng, H.Z., Furuta, Y., Ward, J.M., Sharma, K., Hogan, B.L., Pfaff, S.L., Westphal, H., Kimura, S., Mahon, K.A., 1998. Formation of Rathke's pouch requires dual induction from the diencephalon. *Development* 125, 4835–4840.
- Trowe, M.O., Zhao, L., Weiss, A.C., Christoffels, V., Epstein, D.J., Kispert, A., 2013. Inhibition of Sox2-dependent activation of Shh in the ventral diencephalon by Tbx3 is required for formation of the neurohypophysis. *Development* 140, 2299–2309.
- Valdearcos, M., Xu, A.W., Koliwad, S.K., 2015. Hypothalamic inflammation in the control of metabolic function. *Annu. Rev. Physiol.* 77, 131–160.
- Wang, X., Kopinke, D., Lin, J., McPherson, A.D., Duncan, R.N., Otsuna, H., Moro, E., Hoshijima, K., Grunwald, D.J., Argenton, F., Chien, C.B., Murtaugh, L.C., Dorsky, R.I., 2012. Wnt signaling regulates postembryonic hypothalamic progenitor differentiation. *Dev. Cell.* 23, 624–636.
- Wang, L., Meece, K., Williams, D.J., Lo, K.A., Zimmer, M., Heinrich, G., Martin Carli, J., Leduc, C.A., Sun, L., Zeltser, L.M., Freeby, M., Goland, R., Tsang, S.H., Wardlaw, S.L., Egli, D., Leibel, R.L., 2015. Differentiation of hypothalamic-like neurons from human pluripotent stem cells. *J. Clin. Invest.* 125, 796–808.
- Wataya, T., Ando, S., Muguruma, K., Ikeda, H., Watanabe, K., Eiraku, M., Kawada, M., Takahashi, J., Hashimoto, N., Sasai, Y., 2008. Minimization of exogenous signals in ES cell culture induces rostral hypothalamic differentiation. *Proc. Natl. Acad. Sci. U. S. A.* 105, 11796–11801.
- Xu, Y., Tamamaki, N., Noda, T., Kimura, K., Itokazu, Y., Matsumoto, N., Dezawa, M., Ide, C., 2005. Neurogenesis in the ependymal layer of the adult rat 3rd ventricle. *Exp. Neurol.* 192, 251–264.
- Yin, W., Wu, D., Noel, M.L., Gore, A.C., 2009. Gonadotropin-releasing hormone neuroterminals and their microenvironment in the median eminence: effects of aging and estradiol treatment. *Endocrinology* 150, 5498–5508.
- Zhang, G., Li, J., Purkayastha, S., Tang, Y., Zhang, H., Yin, Y., Li, B., Liu, G., Cai, D., 2013. Hypothalamic programming of systemic ageing involving IKK-beta, NF-kappaB and GnRH. *Nature* 497, 211–216.
- Zhao, Y., Mailloux, C.M., Hermesz, E., Palkovits, M., Westphal, H., 2010. A role of the LIM-homeobox gene Lhx2 in the regulation of pituitary development. *Dev. Biol.* 337, 313–323.
- Zhao, L., Zevallos, S.E., Rizzoti, K., Jeong, Y., Lovell-Badge, R., Epstein, D.J., 2012. Disruption of SoxB1-dependent sonic hedgehog expression in the hypothalamus causes septo-optic dysplasia. *Dev. cell* 22, 585–596.