Tuberculosis 98 (2016) 62-76

Contents lists available at ScienceDirect

Tuberculosis

journal homepage: http://intl.elsevierhealth.com/journals/tube

HIV-1 and the *Mycobacterium tuberculosis* granuloma: A systematic review and meta-analysis



^a Clinical Infectious Diseases Research Initiative Institute of Infectious Disease and Molecular Medicine, University of Cape Town, South Africa

^b Royal Hobart Hospital, Tasmania, Australia

^c Department of Medicine, University of Cape Town, South Africa

^d Francis Crick Institute Mill Hill Laboratory, London, United Kingdom

^e Department of Medicine, Imperial College London, W21PG, United Kingdom

ARTICLE INFO

Article history: Received 4 November 2015 Received in revised form 22 February 2016 Accepted 29 February 2016

Keywords: Mycobacterium tuberculosis Tuberculosis Granuloma HIV AIDS Tissue Histology

SUMMARY

Infection with HIV-1 greatly increases the risk of active tuberculosis (TB). Although hypotheses suggest HIV-1 disrupts Mycobacterium tuberculosis (Mtb) granuloma function, few studies have examined this directly. The objective of this study was to determine what evidence exists about the effect HIV-1 coinfection has upon Mtb granulomas. A systematic search of PubMed, Web of Science, and Medline up to 20 March 2015 was conducted, to identify studies comparing Mtb-infected tissue from HIV-1 infected and uninfected persons, or HIV-1 infected persons with stratified peripheral CD4 T cell (pCD4) counts. We summarized findings that focused on how HIV-1 changes granuloma formation, bacterial presence, cellular composition, and cytokine production. Nineteen studies with a combined sample size of 899 persons were included. Although studies frequently were limited by variable or inadequately described definitions of outcomes and analytical methods, HIV-1 was found to be associated with increased bacillary load within Mtb-infected tissue. Reductions in pCD4 counts within co-infected persons associated with both poorer granuloma formation and higher bacterial load. The high degree of heterogeneity among studies combined with experimental limitations made it difficult to conclusively support previously published and prevalent hypotheses about HIV-1/Mtb co-infection granulomas. To elucidate the validity of these hypotheses we have described areas that can be improved in future studies in order to clarify the influence HIV-1 co-infection has upon the Mtb granuloma.

© 2016 The Authors. Published by Elsevier Ltd. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

1. Introduction

Tuberculosis (TB) and HIV-1 are two of the leading infectious causes of death worldwide and TB is the leading cause of death among HIV-1 infected persons [1]. Once infected with *Mycobacterium tuberculosis* (Mtb), HIV-1 infected persons have increased morbidity and mortality due to TB compared to HIV-1 uninfected persons [2]. As peripheral CD4 T cell (pCD4) counts fall, susceptibility to active and disseminated TB increase, however, HIV-1 infected persons with relatively preserved pCD4 counts are also at increased risk [3]. It has been hypothesized that the primary

* Corresponding author. Current address: Rm7026, Biomedical Science Tower 3, University of Pittsburgh, 3501 Fifth Avenue, Pittsburgh, PA 15261, USA.

cause for increased TB susceptibility in HIV-1 infected persons is due to immunological disruptions of the Mtb granuloma [4,5].

The granuloma is the hallmark of TB. Granulomas consist of a collection of organized immunological cells that form in response to Mtb infection [6]. Granulomas commonly consist of infected and recruited macrophages, differentiated epithelioid cells, all surrounded by a lymphocyte layer. The relationship between the granuloma and TB is complex and not fully understood because granulomas can prevent dissemination and kill Mtb, but also allow persistence of Mtb and even be permissive to its growth [7,8]. This illustrates the delicate balance between bacterial growth and death within the microenvironment of the granuloma and that granulomas form an incompletely effective or even bacterium-permissive immunological response [7,9]. It is hypothesized that HIV-1 disrupts this balance by causing granulomas to be more disorganized, killing resident CD4 T cells, and dysregulating normal T cell, and macrophage function (Table 1) [4,5,10–14], leading to an

1472-9792/© 2016 The Authors. Published by Elsevier Ltd. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).



REVIEW



CrossMark

Tuberculosis

E-mail address: CRDiedrich.Publications@gmail.com (C.R. Diedrich).

¹ Diedrich CR and O'Hern J contributed equally to this publication.

Table 1		
Hypotheses how HIV-1 manip	oulates Mtb	granulomas.

Review (first author, year published)	Main hypotheses regarding granulomas in HIV-1/Mtb co-infected persons
Ledru, 1999 [10]	HIV-1's ability to manipulate cytokine and nitric oxide production that are recruited to the granuloma may play an important role in increasing bacterial dissemination.
Bocchino, 2000 [11]	Poor granuloma formation within co-infected persons most likely results from a disruption in the pro- and anti- inflammatory production of cytokines and an increase in cell death of CD4 T cells.
Lawn, 2002 [5]	Granulomas within co-infected persons will be poorly formed and contain increased bacterial growth through the impairment of cellular recruitment and cell-mediated granulomatous response.
Diedrich, 2011 [4]	Granulomas within co-infected persons will have impaired architecture, reduced CD4 T cell counts, impaired T cell and macrophage function, and increased cell death.
Kwan, 2011 [12]	HIV-1 replication may be induced by Mtb-infected macrophages that will indeed lead to HIV-1 infection of adjacent macrophages and CD4 T cells and activated CD4 T cells will increase HIV-1 replication
Geldmacher, 2012 [13]	Preferential depletion of Mtb-specific and total CD4 T cells may play a significant role in granuloma disruption within HIV-1 infected persons
Ansari, 2013 [14]	HIV-1 enters granulomas and causes CD4 T cell apoptosis, depletion, and disrupted recruitment of T cells, which leads to granuloma disorganization.

increase in susceptibility to both active and disseminated TB disease.

Many studies in humans that support these hypotheses have measured immunological responses within non-tissue resident cells: peripheral blood mononuclear cells (PBMC), bronchoalveolar fluid (BALF), and pericardial fluid (PCF) [15–18]. These studies have demonstrated impaired Mtb-specific T cell activity [15,16,18] and killing of Mtb-specific peripheral CD4 T cells [19], and total BAL CD4 T cells [17] in persons infected with HIV-1. Although these data are convincing, there are significant variations in cellular composition and Mtb-specific immunological responses within PBMC, BALF, PCF, and granulomas [20,21]. Extrapolating data from non-tissue resident cells to what is occurring within granulomas may not be appropriate [22]. This illuminates a need to study human granulomas directly in HIV-1/Mtb co-infected persons to better understand how HIV-1 manipulates the TB granuloma. Understanding how HIV-1 manipulates granulomas directly will elucidate the mechanistic cause HIV-1 exploits that increases TB susceptibility.

We systematically reviewed HIV-1/Mtb co-infection literature that examined Mtb granulomas directly. The objective of this systematic review and meta-analysis was to elucidate how Mtbinfected tissues differ between HIV-1 infected and uninfected persons. We focused on studies that reported how HIV-1 changed: granuloma formation [23-29]. granuloma organization [23.26–28.30–35], granuloma caseation [23.25.26.29–33], Mtb growth [23-25,27,31,32,35,36], cellular populations [23,26,30-33], cytokine expression [30,31,37] and HIV-1 virion presence within excised tissue [33,38]. To help reduce some of the variability observed within this literature our second objective of this review was to illuminate future strategies to study, analyze, and report granuloma-based data.

2. Methods

2.1. Search strategy and selection criteria

PubMed, Web of Science, and Medline were searched using predetermined combinations of terms (Supplemental Table 1) for relevant peer reviewed studies (through 20 March 2015) that reported histological data in TB diseased tissue from HIV-1/Mtb co-infected persons. We reviewed original articles published in all languages. In addition to the database search, we screened citations in the full-text articles reviewed here, published reviews, book chapters, and suggested papers from experts in the field.

The primary objective of this review was to identify how Mtb granulomas from HIV-1/Mtb co-infected persons differed from granulomas obtained from HIV-1 uninfected persons. Studies were

eligible for inclusion if they compared the histology of Mtb infected tissue from HIV-1 infected and uninfected persons, or from HIV-1 infected persons with stratified pCD4 counts. Studies were required to include an acceptable means of defining HIV-1-infected and uninfected groups (either HIV-1 serology or a documented past history of HIV-1 infection, or for studies prior to 1990, an acceptable HIV/AIDS diagnosis by World Health Organization criteria at the time of publication) and confirming TB diagnosis (microbiology or histology consistent with TB, with or without a consistent clinical picture including course of illness and response to treatment). Studies were excluded if the method of biopsy was only fine-needle aspiration (FNA) as this method of excision was unlikely to preserve granuloma architecture and may not capture entire granulomas within the target tissue. Where studies reported or appeared to report on the same or overlapping persons, the results from the earlier study were excluded. Reviewers independently assessed the eligible articles for inclusion and exclusion criteria; disagreements were resolved by consensus. The included studies were assessed for quality of study design and potential limitations to findings.

2.2. Data extraction

Results from the individual studies were categorized into the following outcomes for comparison between HIV-1/Mtb co-infected and HIV-1 uninfected persons, or HIV-1-infected with stratified pCD4 counts: 1) proportion of biopsied samples with granulomas present, 2) quality of granuloma formation (quality of granuloma formation was defined independently within each study), 3) cellular and cytokine presence, 4) proportion of biopsied samples containing Mtb (acid fast bacilli [AFB] or culture positivity [CFU], 5) bacillary load within AFB+ samples, and 6) HIV-1 virion presence.

2.3. Statistical analysis

For outcomes where studies reported individual quantitative or semi-quantitative results for the different outcomes HIV-1/Mtb coinfected and Mtb-only infected persons, a meta-analysis was performed using the Cochrane Database's RevMan program. We calculated summative risk ratios for changes in 1) granuloma presence, 2) quality of granuloma formation, 3) AFB presence 4) CFU in HIV-1/Mtb-co-infected versus Mtb-only infected persons and 5) AFB load. Where results were categorically scored and not simply dichotomous (for quality of granuloma formation and AFB load), the proportions of persons in each group with the highest scores (for well-formed granulomas and AFB load) or lowest scores (for poorly formed granulomas) were used in meta-analyses. Data sets were treated as dichotomous and risk ratio with 95% CIs were calculated using Mantel–Haenszel method with a random effects analysis model. Heterogeneity was assessed with I^2 statistics and defined as low ($I^2 \le 25\%$), moderate ($25\% < I^2 \le 75\%$), high ($I^2 > 75\%$).

3. Results

Our initial search yielded 3645 abstracts and titles (Figure 1). After review of these abstracts, titles, and where appropriate, full studies, 60 studies were identified that examined the histopathology of Mtb infection in persons co-infected with HIV-1. Eight studies were excluded because, while they described the findings in tissue from HIV-1/Mtb co-infected persons, no comparison was made to either HIV-1 uninfected persons, or between different HIV-1 persons at different stages of HIV-1 infection or pCD4 count [39–46]. Sixteen studies were excluded, because their results were based on FNA [47–62]. Four studies did not adequately define HIV-1 [63–66] and one did not adequately define HIV-1 or TB status [67]. Four studies did not differentiate results between Mtb and other mycobacteria [68–71]. One study may have been biased by

persons having co-morbidities [72]. Two studies [31,33] reanalyzed, at least in part, some results from persons in previous studies [73,74]. For these overlapping studies, results from the later published study were included [31,33]. Five studies did not sufficiently examine tissue samples [75–79].

Nineteen unique studies [23-38,80-82] were included in this review (Table 2). A total of 413 Mtb-only infected persons (with a range between 3 and 108 persons, median n = 14) and 486 HIV-1/Mtb co-infected persons (range: 5-109, n = 13). The site of TB disease within each study was varied: five examined lvmph node [26,31,32,80,83], seven examined pleura [23,27,29,35–37,82], three examined lung [24,28,30], one each examined spine [33], pericardium [25], brain [81], and one study analyzed multiple tissues from different persons (lymph node, lung, pleura, bone marrow) [34]. pCD4 counts were stratified for comparison in HIV-1/Mtb co-infected persons in five studies [23,28,31,34,38]. One study described stratified pCD4 counts without presenting individual data [35]. Two of the pCD4-stratified studies examined only HIV-1/Mtb co-

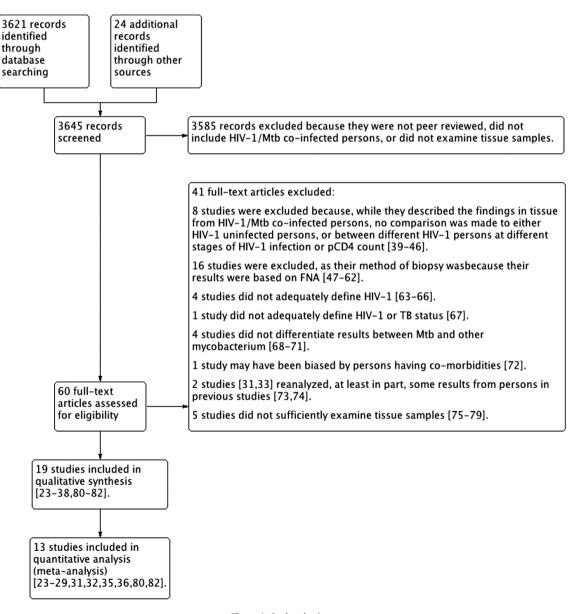


Figure 1. Study selection.

Table 2 Study summaries.

Study (first author, date, country)	Number of persons with Mtb-only (mean age)	Number of HIV-1 co-infected persons (mean age)	HIV-1 stage and mean pCD4	Biopsy site	Key measurements relevant to review	Main findings (co- infected vs Mtb- only)	Comments
Bezuidenhout 2009 South Africa [37]	6 (38)	6 (34·5)	Early stage- qualified by no OI	Pleura	- Granuloma necrosis and counts - IFNγ, TNF, IL12, IL4 mRNA	†- Granuloma necrosis †- TNF mRNA production ?- IFNγ, TNF, IL12, IL4 mRNA production	 166 granulomas from 12 patients but analyzed granulomas as independent entities, which may have biased results toward persons that contained more granulomas Methodology for examining cytokine production was difficult to interpret and compare among other studies Selection of region of interest (ROI) was not described Granuloma definitions were not described No mention of pathologists being
Conde 2003 Brazil [82]	71 (37·2)*	13	Not stated	Pleura	AFB and CFU	↑ AFB smear presence • No significant difference in CFU	blinded - The primary aim of this study was to investigate diagnostic yield of sputum induction in pleural TB. Pleural biopsy tissue, however, was examined as part of the study and the results from this incorporated to our study. - No blinding was described - *Mean age of all persons within
Danaviah 2013 South Africa [33]	9 (44·4)	13 (26·4)	544.6 +- 315	Spine	- Viral load (tissue homogenate and plasma) - CD3, CD4, CD8 markers - giant cell (GC) counts	 Same granuloma organization Same CD3% ↓ CD4% ↑ CD8% 	study - Further analysis of persons from previously published study [82] - Selection of <i>intact</i> granulomas may have biased results toward well- formed granulomas - Single CD4 and CD8 markers used may lead to non T cells being quantified - Selection of ROI not described. - Granuloma definitions were not described - No mention of pathologists being blinded
De Noronha 2007	4	5	Not stated	Lung	- Granuloma organization, architecture and	↑ Granuloma disorganization • Similar necrosis (co	blinded - Autopsies performed on persons that died of ontinued on next page)

Table 2 (continued)

Study (first author, date, country)	Number of persons with Mtb-only (mean age)	Number of HIV-1 co-infected persons (mean age)	HIV-1 stage and mean pCD4	Biopsy site	Key measurements relevant to review	Main findings (co- infected vs Mtb- only)	Comments
Brazil 30]					composition (from H&E), necrosis, bacilli load, TNF, TGF-β	 TNF production Similar TGFβ AFB presence stated (not presented individually) 	pulmonary TB may have biased results towards more severe TB disease. - 3 blinded pathologist were used - Selection of ROI was not described - Granuloma definitions were
Di Perri 1996 Italy [28]	16 (32·5)	16 (30·7)	>400:3 200-400: 3 100-200: 3 <100: 7	Lung	Granuloma organization, AFB load correlated with stratified pCD4 counts	↓ Granuloma counts ↑ Poorly formed granulomas ↓ Well formed granulomas ↑ AFB presence and load • pCD4 counts indirectly correlate to ↑ bacterial load, ↑ poorly formed granulomas, ↓ well formed granulomas	not described - All patients were sputum negative prior to bronchoscopy, which may have lead to selection bias. - Method of biopsy - from area of consolidation; or right middle lobe o lingual in case of no focal or only diffused consolidation may have biased results - Multiple pathologist reviewers, howeve no blinding detailed. - Granuloma and AFB load descriptions were available - Selection of ROI was not described
Elliott 1993 Zambia 36]	5	9	Not stated	Pleura	CFU	• No significant difference in CFU	 Study described Study described clinical and diagnostic differences between HIV-1 infected and uninfected persons with TB. No blinding was described
Heyderman 1998 Zimbabwe [23]	11- (33)	63 (23)	191 (0–2009) >500 11 200–490: 19 100–190: 6 <100: 14	Pleura	Granuloma counts, poorly formed, caseation, GC, AFB and CFU positive, AFB load, stratified pCD4	 No difference in granuloma counts, caseation, formation, giant cells, AFB+ or CFU+ samples, or scanty AFB † Tissue with numerous AFB+ pCD4 counts may correlate to increased bacterial load 	 - 63 persons in co- infected group, - pCD4 stratified to results. - Granuloma formation, scanty and numerous AFE loads were not described. - Selection of ROI was not described. - Blinded pathologists were not described.
Hochedez 2003 France [80]	19 (38·5)	13 (36·5)	Not stated	LN	AFB, CFU, Granuloma presence, Caseation	• No significant difference in granuloma presence, AFB, CFU, or caseation	- Method of selection of ROI no described
Jones 1993 USA [34]	NA	23	Stratified pCD4/ mm ³ from 21 persons: <101: 9	9 LN, 6 lung, 4 pleura, 4 bone marrow	Granulomatous changes stratified to pCD4 counts	• Granulomatous changes observed regardless of pCD4 counts.	- Granuloma formation and tissue AFB presence were not primary

Table 2 (continued)

Study (first author, date, country)	Number of persons with Mtb-only (mean age)	Number of HIV-1 co-infected persons (mean age)	HIV-1 stage and mean pCD4	Biopsy site	Key measurements relevant to review	Main findings (co- infected vs Mtb- only)	Comments
			101–200: 5 201–299: 3 >300: 4				aim of this study - Granulomatous changes were not defined.
Kennedy 1992 USA [24]	45	67	Not stated (likely many late due to year of study)	Lung	Granuloma counts, AFB and CFU positivity	• No difference in AFB smear or CFU ↓ Granuloma presence	 The primary aim of this study was t assess the utility of bronchoscopy in sputum negative persons in the diagnosis of pulmonary TB in HIV-1 infected patients, rather than to assess granuloma differences 67 co-infected an 45 HIV-1 uninfected person were in this study Granulomas wer not defined Potential sampling bias associated with bronchoscopic biopsy site selection techniqu may have occurree Selection of ROI was not described Blinded pathologists were not described
Luzze 2001 Uganda [35]	33 (33)	109 (34)	Not stated	Pleura	Granuloma formation, CFU, BACTEC culture, days to culture positivity	 ↓ Well formed granulomas ↑ CFU positivity • No significant difference in BACTEC culture positivity or days to positivity • Granuloma formation was not different in persons with fewer pCD4 counts 	The primary aim of study was to compare clinical, radiographic and diagnostic method in pleural TB between HIV infected and uninfected person - Method of granuloma assessment is not specifically described. - No blinding was described
Muller 1994 Germany [31]	8	8	Individual pCD4/ mm ³ : 100–200:4 200–300: 2 300-400: 2	Lymph node (LN)	Necrosis, MNGC, Epithelioid cell layer, CD68, lysozyme, α-1-anti- chymotryp., Mac387, Ki-M8, CD43, CD3, CD4, CD8, CD25, HLA- DR, Ki-67, CD22, IL- 1α, IL-1β, IL-6, TNF, IFN-α, IFN-β. Stratified to pCD4 counts		- All results excep cytokine production appeared to overla with a previously published paper [83] - Examined 22 histological outcomes and stratified pCD4 counts

67

Table 2 (continued)

Study (first author, date, country)	Number of persons with Mtb-only (mean age)	Number of HIV-1 co-infected persons (mean age)	HIV-1 stage and mean pCD4	Biopsy site	Key measurements relevant to review	Main findings (co- infected vs Mtb- only)	Comments
Ngilimana 1996 Rwanda	3 (27)	12 (32)	Not stated	LN	Langhan cells, Epithelioid cells, lymphocytic cuff,	↓* Well formed granulomas ↑ AFB load	* Some conclusions were difficult to determine because of qualitative nature of measurements - Description of granulomas was detailed in
[32]					foamy macrophages, necrosis type, AFB load, epithelioid venule, plasmocytes, granuloma formation	†* Epithelioid macrophages ↓*Lymphocytes and Lanhan cells	previously published study [51] - Examined a 9 histological outcomes in 12 co- infected persons and only 3 Mtb only persons-Specific LN type was not discussed, - Blinding and ROI selected were not described * All measurement
Perfura-Yone 2011 Cameroon [29]	108 with pleural biopsy	81 with pleural biopsy	Not stated	Pleura	Granuloma counts and caseum.	 No difference in granuloma presence or necrosis † Necrosis within persons with <200 pCD4/mm³ 	were categorical - Study focused on clinical, radiological, hematological as well as histological picture in a high number of persons with tuberculous pleural effusion. - 108 Mtb-only and 81 co-infected persons were examined in this study. - The method for selection of ROI noi described - There was not mention of blinding of assessing pathologists
Reuter 2006 South Africa [25]	20	5	Not stated	Pericardium	Granuloma counts, central necrosis, CFU positivity	• No significant difference in granuloma counts, necrosis or AFB presence	 Sub dividing 25 persons into 4 groups and 7 histopathological outcomes resulted in small 8 outcome containing only 1 person. Blinding and ROI selected were not described Granulomas were not defined
Shen 1988 USA [26]	3	5	AIDS by CDC 1985 criteria (Ol)	LN	AFB, brief granuloma description, epithelioid macrophage counts, CD3, CD4, CD8, CD26, HLA- DR, CD14 percentages	 No difference in granuloma formation AFB load stated without data CD3 and CD4% No difference in CD8%, but CD8 T cells are distributed throughout granulomas No difference in other markers 	 Only 3 persons ir Mtb-only group Problematic definition for 2 persons in HIV-1 group because they were defined as risi factors and symptoms indicative of AIDS related complex without defining what satisfied this these patients

Study (first author, date, country)	Number of persons with Mtb-only (mean age)	Number of HIV-1 co-infected persons (mean age)	HIV-1 stage and mean pCD4	Biopsy site	Key measurements relevant to review	Main findings (co- infected vs Mtb- only)	Comments
Trajman 1997 Brazil [27]	27	9	Not specified	Pleura	Granuloma presence, formation, caseous necrosis, epithelioid cells, GC	• No difference in granuloma counts or formation • No difference in caseous necrosis, epithelioid cells or GC presence	were deleted from our analysis. - 2 independent observers were used - ROI and blinding were not described - Precise granulom descriptions were not provided. - Outcomes assessed by impression of one pathologist, but with no clearly described criteria for outcomes. - ROI and blinding were not described
Tripathi	5	5	Not stated	Brain	Granuloma	↓ Well formed	 Precise granulom descriptions were not provided. Granuloma
2014 India [81]	(27.8)	(31.2)			formation	granulomas in HIV- 1 co-infected persons	descriptions were not a primary goal of study - Autopsies were performed betwee 2 and 13.45 h pos mortem, which may have biased results - Assessment of IL
							6, MIP-1 α , IL-8, an TNF- α described in methods but not
Van Der Ende 1999 Germany [38]	NA	9	Group 1: >150 pCD4/mm ³ Group 2 <50 pCD4/ mm ³	LN	HIV-1 RNA, granuloma presence and formation from stratified pCD4 counts	 Group 1- inflammation was disorganized with no well-formed granuloma † HIV-1 RNA in and near granulomas than non- granulomatous areas (data not presented in study). CD4 T cells remained the main source of HIV RNA in both groups. 	reported in results - Specific LN excised not identified - Compared co- infected patients a different periphera CD4 counts. - Only study to directly examine HIV-1 presence within granuloma at different pCD4 counts - 2 persons with <50 pCD4/mm ³ an <i>M. avium</i> complex were excluded from our analysis - ROI was not described - Precise granulom

infected persons [34,38]. These studies typically included few persons and techniques for defining and assessing outcomes such as granuloma presence and formation, AFB load and cellular presence were variable and sometimes not defined at all. Histological analyses were also not always performed in a blinded fashion and the methods for identifying regions of interest (ROI) examined were not defined. These limitations may have biased results within each of the studies.

3.1. Does HIV-1 co-infection reduce the granuloma presence in Mtbinfected persons?

HIV-1 has been hypothesized to reduce Mtb granuloma formation [5]. If an infected person cannot form granulomas they may be more susceptible to Mtb dissemination. To determine if this hypothesis was true we examined the proportion of Mtb-infected persons with granulomas visible in excised tissue was identified

	HIV T	ΓВ	TB alc	one		Risk Ratio	Risk Ratio	
Study or Subgroup	Events	Total	Events	Total	Weight	M-H, Random, 95% CI	M–H, Random, 95	% CI
Kennedy 1992	11	59	16	37	8.2%	0.43 [0.23, 0.82]		
Reuter 2006	2	5	13	20	3.6%	0.62 [0.20, 1.89]		
Di Perri 1996	10	16	16	16	14.3%	0.64 [0.43, 0.93]		
Heyderman 1998	32	63	7	11	11.0%	0.80 [0.48, 1.33]		
Hochedez, 2003	5	6	13	13	13.8%	0.81 [0.55, 1.22]		
Trajman 1997	7	9	22	27	14.0%	0.95 [0.64, 1.41]		
Shen 1988	5	5	3	3	12.7%	1.00 [0.64, 1.56]	_ + _	
Perfura-Yone 2011	71	81	89	108	22.4%	1.06 [0.94, 1.20]	•	
Total (95% CI)		244		235	100.0%	0.82 [0.65, 1.03]	•	
Total events	143		179					
Heterogeneity: Tau ² =	= 0.06; Cl	$hi^2 = 1$	8.48, df =	= 7 (P =	= 0.010);	$l^2 = 62\%$	0.05 0.2 1	5 20
Test for overall effect	: Z = 1.68	8 (P = 0).09)				0.05 0.2 1	5 20

Figure 2. Mantel-Haenszel random effects risk ratio for Mtb granuloma presence in HIV-1 co-infected persons.

and compared between HIV-1-infected and -uninfected persons in eight studies [23–29,80]. Granuloma presence was calculated using the proportion of excised tissue containing at least one granuloma. One study was excluded from this analysis as granuloma presence was a criteria for tissue selection [33] while another was excluded because it reported total Mtb granulomas observed [37] and not granulomas per excised tissue. The lowest granuloma score in one study represented non-organized inflammatory patterns [28], which were not counted as granulomas. For all these studies, granuloma presence was a dichotomous variable, without quantifving granulomas in each excised sample.

HIV-1 did not change the likelihood of excised tissue containing granulomas. The proportion of persons with tissue that contained granulomas varied widely for both groups across the studies. In tissue from HIV-1 infected persons, Mtb granuloma presence ranged from 19% [24] to 100% [26], and from HIV-1 uninfected persons from 43% [24] to 100% [26]. All studies apart from two [26], reported at least a slight reduction in granuloma presence within HIV-1 infected persons. However, only one study reached statistical significance [24]. The probability ratio calculated based on forest plot did not identify a significant difference for granuloma presence between the two groups (Figure 2, RR: 0.82, 0.65–1.03). Varying pCD4 counts correlated to a difference in granuloma presence within HIV-1/Mtb co-infected persons in one study [28] but not in another [23].

3.2. Does HIV-1 co-infection change the quality of Mtb granuloma formation?

The ability of granulomas to kill Mtb and prevent its dissemination, in part, relies on how well they are organized [6]. It has been hypothesized that HIV-1 disrupts normal granuloma formation [4,5], which could increase TB susceptibility in HIV-1 infected persons. To determine if HIV-1 changes Mtb granuloma formation quality we identified eight studies that reported the presence of well- or poorly-formed granulomas [23,26–28,30–32,35]. Techniques used to score granuloma organization were highly variable and included: an assessment of the presence of epithelioid cells, lymphocytes and caseation [28,30], or focused on the organization of the epithelioid cells alone [31]1. The remaining studies relied on pathologist impressions only without providing a description [23,26,27,32,35].

No quantifiable difference in the appearance of well- or poorlyformed Mtb granulomas were observed in HIV-1 co-infected and HIV-1 uninfected persons. Four studies presented findings for welland poorly-formed granulomas [27–29,32], while one presented results for poorly-formed granulomas only [23] or well-formed granulomas only [35] in HIV-1 co-infected and uninfected persons. Four of the five studies that examined the presence of wellformed granulomas identified a non-statistically significant [28,31,32] or significant [35] reduction in HIV-1 infected persons, while the remaining study identified no difference [27]. Similarly, three of five studies that quantified poorly-formed granulomas identified a non-statistically significant increase within co-infected persons [28,31,32], while the remaining two identified no difference [23,27]. Forest plots for both well- and poorly-formed granulomas did not meet statistical significance (Figure 3A: well-formed: RR 0.33 [0.08, 1.26], 3B: poorly-formed: RR 2.63 [0.24, 28.91]).

Oualitative assessments of Mtb granuloma formation were highly variable and inconclusive in both HIV-1 infected and HIVuninfected persons. Other studies described granuloma formation quality between the two groups without presenting results from individual persons [26,30,33,81]. Shen et al. [26], made an overall statement that all five persons with AIDS and TB lymphadenitis contained caseous granulomas similar to those in the HIV-1 uninfected persons. Danaviah et al. [33], also reported no difference in granuloma formation between co-infected and singly infected persons with spinal TB, but their method stated intact granulomas were chosen for examination, a strategy that may have biased results toward well-formed granulomas. Conversely, De Noronha et al. [30], reported three well-defined zones (caseation, epithelioid cell layer and lymphocytic cuff) within the granulomas of persons with pulmonary TB without HIV-1 compared to atypical disorganized arrangement of the granulomas located in co-infected persons. Tripathi et al. [81], described the presence of well-formed granulomas in the brain of HIV-1 uninfected persons with TB meningitis but noted their absence in HIV-1 co-infected persons. While it is widely believed the increased susceptibility of HIV-1 infected persons to disseminated TB is caused by impaired granuloma formation, we found the results reported in this area to be conflicting and thus the evidence to support this hypothesis lacking.

HIV-1 did not change the presence of caseous granulomas. A hallmark of TB is the caseous granuloma [6], and to determine if HIV-1 changes caseation necrosis we identified studies that qualitatively [30,31,33] or quantitatively [23,25,26,29,32,37] reported caseous granuloma presence in co-infected and singly infected persons. Most studies identified no difference in granulomatous necrosis or caseation between co-infected and singly infected persons [23,26,29,31]. Two studies reported less necrosis in co-infected persons without statistical analysis [25,32]. Only one study reported a statistically significant increase in necrotic lung granulomas in co-infected persons [37]. These data suggested that HIV-1 co-infection did not change the likelihood of granulomas becoming caseous.

pCD4 depletion associated with poorly formed Mtb granulomas in HIV-1 co-infected persons. To determine if HIV-1 progression

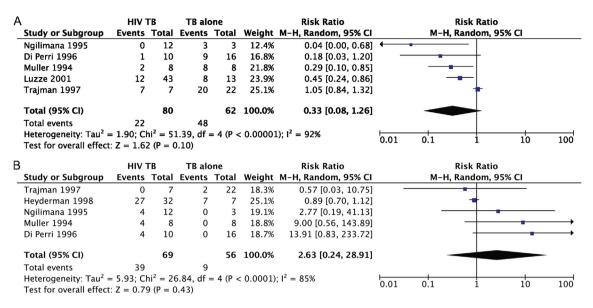


Figure 3. Mantel-Haenszel random effects risk ratio for well-formed- (A) and poorly-formed (B) granulomas in HIV-1 co-infected persons.

associated with disrupted granuloma formation we examined five studies that correlated pCD4 counts with granuloma formation in co-infected persons [28,31,34,35,38]. Three studies reported more poorly formed granulomas in those with lower pCD4 counts [28,31,34]. One study described no correlation between pCD4 counts and granuloma appearance [35]. Di Perri et al. [28], demonstrated that six persons with the lowest categorically defined granuloma scores (unorganized inflammation) had a mean pCD4 of 49.2 pCD4/mm³ compared to the remaining ten persons with a mean pCD4 of 238.6 pCD4/mm³. Similarly, Muller et al. [31], reported well-formed granulomas in two persons with >300 pCD4/ mm³, a combination of well- and poorly-formed granulomas in persons with 200-300 pCD4/mm³ and only necrosis and foamy macrophages within lymph nodes of persons with <200 pCD4/ mm³. Van Der Ende et al. [38], reported an absence of well-formed granulomas within the two HIV-1/Mtb co-infected persons with <50 pCD4s/mm³, while the five persons with >170 pCD4/mm³ contained multiple well-formed granulomas. Granulomatous changes were observed but not defined, in 75% (9/12) of excised tissues from HIV-1 co-infected persons with <101 pCD4/mm³, although this reduction was not significantly different from other co-infected persons with higher pCD4 counts [34]. Taken together, lower pCD4 counts in HIV-1 infected persons generally correlated to poorer granuloma formation, particularly as pCD4 counts fall below 50 pCD4/mm³.

3.3. Does HIV-1 change the cellular composition of Mtb granulomas?

It is hypothesized that T cells and macrophages, essential components for normal granuloma activity, are preferentially killed or manipulated by HIV-1 at the site of TB disease [4,5,13]. To determine if cellular composition of granulomas from HIV-1 co-infected persons differ from HIV-1 uninfected persons we identified three studies that compared the presence of T cells in two groups. The studies that quantified T cell presence used single CD3, CD4, or CD8 antibodies for identification [26,31,33], so the presence of CD4+CD3- (macrophages, dendritic cells) and CD8+CD3- (NK cells) non-T cells may have biased results by staining false positives.

CD3. CD4. or CD8 T cell counts within granulomas were highly variable. Two studies qualitatively identified fewer lymphocytes within granulomas from co-infected persons [30,32], but did not specifically identify the types of lymphocytes involved because the assessments were made on hematoxylin and eosin stained slides. Findings for CD3 cells were conflicting. Two studies identified no difference in CD3 T cell presence within co-infected granulomas [31,33], while a third reported a reduction [26]. CD8 T cell presence was also variable. One study demonstrated an increase in CD8 T cells within granulomas from co-infected persons [33], while two identified no difference [26,31]. A more detailed examination of CD8 T cell localization within the granuloma found that, although the number of CD8 T cells were the same, they were more widely dispersed throughout granulomas of HIV-1 infected persons, while they were confined to the lymphocytic mantle surrounding the epithelioid cell layer in HIV-1 uninfected persons [26]. Two studies reported a statistically significant reduction in CD4 T cells in granulomas of co-infected persons [26,33] while a third identified fewer CD4 T cells in only three of eight persons with HIV-1 co-infection [31] compared to the HIV-1 uninfected group. The three persons with fewer CD4 T cells all had <200 pCD4/mm³. The high variability in T cell staining make it difficult to determine how HIV-1 changes cellular composition.

Macrophage presence within Mtb granulomas were highly variable. Macrophages are also an essential component of granulomas, forming the epithelioid cell layer. Findings from studies in this area were also conflicting [23,31–33]. Two studies described a disruption of the epithelioid laver within granulomas in HIV-1 infected persons without defining those disruptions [31,32]. The presence of Langhan's giant cells, which form when macrophages fuse and are commonly seen in Mtb granulomas, were also variable. Results from two studies demonstrated no difference in count [23,33], while another study found fewer Langhan's giant cells in those co-infected with HIV-1 [31]. Muller et al. [31], determined that HIV-1 uninfected persons and those with >200 pCD4/mm³ contained more Langhan's giant cells within their granulomas than HIV-1 uninfected persons. In contrast, granulomas from those with <200 pCD4/mm³ primarily contained foamy macrophages. Due to the high variability in macrophage and Langhan giant cells counts,

А		HIV 1	ГВ	TB alo	ne		Risk Ratio			Risk Ratio		
	Study or Subgroup	Events	Total	Events	Total	Weight	M-H, Random, 95% Cl		М-Н,	Random, 959	% CI	
	Kennedy 1992	1	56	4	25	10.2%	0.11 [0.01, 0.95]					
	Hochedez, 2003	1	6	3	13	10.9%	0.72 [0.09, 5.59]			-	_	
	Muller 1994	4	8	5	8	24.3%	0.80 [0.33, 1.92]					
	Reuter 2006	1	5	4	21	11.5%	1.05 [0.15, 7.47]					
	Heyderman 1998	12	63	1	11	11.7%	2.10 [0.30, 14.53]		-			
	Conde 2003	5	13	9	71	23.7%	3.03 [1.21, 7.61]					
	Ngilimana 1995	9	12	0	3	7.6%	5.85 [0.43, 79.80]				•	
	Total (95% CI)		163		152	100.0%	1.19 [0.52, 2.71]			-		
	Total events	33		26								
	Heterogeneity: Tau ² =	0.52; Cl	$ni^2 = 11$	L.38, df =	= 6 (P =	= 0.08); I ²	= 47%	0.02	0.1		10	50
	Test for overall effect:	Z = 0.42	2 (P = 0)	.68)				0.02	0.1	1	10	50
D							Biel, Betie			Diel: Detie		
В	Church and Curchanness	HIV T		TB alo		W	Risk Ratio			Risk Ratio		
В	Study or Subgroup		Total	Events	Total	-	M-H, Random, 95% Cl		М-Н,	Risk Ratio Random, 959	% CI	
B _	Elliott 1993	Events 1	Total 9	Events 1	Total 5	1.4%	M-H, Random, 95% Cl 0.56 [0.04, 7.09]		М-Н,		% CI	
B _	Elliott 1993 Reuter 2006	Events 1 2	Total 9 5	Events 1 13	Total 5 21	1.4% 6.6%	M-H, Random, 95% Cl 0.56 [0.04, 7.09] 0.65 [0.21, 1.99]		М-Н,		% CI	
B _	Elliott 1993 Reuter 2006 Hochedez, 2003	Events 1 2 3	Total 9 5 6	Events 1 13 10	Total 5 21 13	1.4% 6.6% 10.7%	M-H, Random, 95% Cl 0.56 [0.04, 7.09] 0.65 [0.21, 1.99] 0.65 [0.28, 1.53]		М-Н, 		<u>% CI</u>	
B _	Elliott 1993 Reuter 2006 Hochedez, 2003 Kennedy 1992	Events 1 2 3 29	Total 9 5 6 56	Events 1 1 1 1 1 1 1 1 5 1 1 1 1 1 1 1 1 1 1	Total 5 21 13 34	1.4% 6.6% 10.7% 26.7%	M-H, Random, 95% Cl 0.56 [0.04, 7.09] 0.65 [0.21, 1.99] 0.65 [0.28, 1.53] 1.17 [0.74, 1.85]		М-Н, 		<u>% CI</u>	
B _	Elliott 1993 Reuter 2006 Hochedez, 2003 Kennedy 1992 Heyderman 1998	Events 1 2 3 29 16	Total 9 5 6 56 38	Events 1 13 10 15 3	Total 5 21 13 34 9	1.4% 6.6% 10.7% 26.7% 8.2%	M-H, Random, 95% Cl 0.56 [0.04, 7.09] 0.65 [0.21, 1.99] 0.65 [0.28, 1.53] 1.17 [0.74, 1.85] 1.26 [0.47, 3.42]		<u>М-Н,</u> 		<u>% CI</u>	
B _	Elliott 1993 Reuter 2006 Hochedez, 2003 Kennedy 1992 Heyderman 1998 Conde 2003	Events 1 2 3 29 16 10	Total 9 5 6 56 38 13	Events 1 1 3 10 15 3 42	Total 5 21 13 34 9 71	1.4% 6.6% 10.7% 26.7% 8.2% 34.7%	M-H, Random, 95% Cl 0.56 [0.04, 7.09] 0.65 [0.21, 1.99] 0.65 [0.28, 1.53] 1.17 [0.74, 1.85] 1.26 [0.47, 3.42] 1.30 [0.91, 1.85]		<u>М-Н,</u> —		<u>% CI</u>	
В _	Elliott 1993 Reuter 2006 Hochedez, 2003 Kennedy 1992 Heyderman 1998	Events 1 2 3 29 16	Total 9 5 6 56 38	Events 1 13 10 15 3	Total 5 21 13 34 9	1.4% 6.6% 10.7% 26.7% 8.2%	M-H, Random, 95% Cl 0.56 [0.04, 7.09] 0.65 [0.21, 1.99] 0.65 [0.28, 1.53] 1.17 [0.74, 1.85] 1.26 [0.47, 3.42]		<u>M-H,</u> 		<u>~ CI</u>	
B	Elliott 1993 Reuter 2006 Hochedez, 2003 Kennedy 1992 Heyderman 1998 Conde 2003	Events 1 2 3 29 16 10	Total 9 5 6 56 38 13	Events 1 1 3 10 15 3 42	Total 5 21 13 34 9 71 24	1.4% 6.6% 10.7% 26.7% 8.2% 34.7%	M-H, Random, 95% Cl 0.56 [0.04, 7.09] 0.65 [0.21, 1.99] 0.65 [0.28, 1.53] 1.17 [0.74, 1.85] 1.26 [0.47, 3.42] 1.30 [0.91, 1.85]	-	<u>М-Н,</u> 		<u>~ CI</u>	
B _	Elliott 1993 Reuter 2006 Hochedez, 2003 Kennedy 1992 Heyderman 1998 Conde 2003 Luzze 2001	Events 1 2 3 29 16 10	Total 9 5 6 56 38 13 73	Events 1 1 3 10 15 3 42	Total 5 21 13 34 9 71 24	1.4% 6.6% 10.7% 26.7% 8.2% 34.7% 11.7%	M-H, Random, 95% Cl 0.56 [0.04, 7.09] 0.65 [0.21, 1.99] 0.65 [0.28, 1.53] 1.17 [0.74, 1.85] 1.26 [0.47, 3.42] 1.30 [0.91, 1.85] 2.70 [1.20, 6.03]		<u>М-Н,</u> — –		<u>~ CI</u>	
B _	Elliott 1993 Reuter 2006 Hochedez, 2003 Kennedy 1992 Heyderman 1998 Conde 2003 Luzze 2001 Total (95% CI)	Events 1 2 3 29 16 10 41 102	Total 9 5 6 56 38 13 73 200	Events 1 1 1 3 10 15 3 42 5 89	Total 5 21 13 34 9 71 24 177	1.4% 6.6% 10.7% 26.7% 8.2% 34.7% 11.7% 100.0%	M-H, Random, 95% Cl 0.56 [0.04, 7.09] 0.65 [0.21, 1.99] 0.65 [0.28, 1.53] 1.17 [0.74, 1.85] 1.26 [0.47, 3.42] 1.30 [0.91, 1.85] 2.70 [1.20, 6.03] 1.21 [0.89, 1.63]		<u>М-Н,</u> 		<mark>% CI</mark>	

Figure 4. Mantel-Haenszel random effects risk ratio for AFB (A) and CFU (B) presence within excised tissues of HIV-1 co-infected persons.

nothing conclusive can be stated about how HIV-1 changes their presence within granulomas.

3.4. Does HIV-1 change cytokine expression within Mtb granulomas?

Granulomas contain a delicate balance of pro- and antiinflammatory cytokine responses [84]. If HIV-1 disrupts this balance, then Mtb dissemination may occur. Although HIV-1 reduces IFN- γ and TNF production by Mtb-specific T cells in BAL, PF, and PCF [15–19], it is not known how HIV-1 changes cytokine expression in granulomas.

Cytokine expression in co-infected granulomas was highly variable and inconclusive. Cytokine expression was also analyzed within granulomas of co-infected and singly infected persons [30,31,37] and findings were again conflicting. TNF production within granulomas was highly variable in co-infected persons. More granulomatous cells were positive for TNF mRNA in coinfected persons than HIV-1 uninfected persons [37]. Likewise, a qualitative trend of more overall TNF production in co-infected persons with <160 pCD4/mm³ was also observed [31]. De Noronha et al. [30] reported less total TNF produced within granulomas of HIV-1 co-infected persons with TB lymphadenitis, without quantification. No difference in expression of IL-1 α , IL-1 β , IFN- α and IFN- β [31] or the mRNA presence of IFN- γ , IL-4 and IL-12p40 [37] were identified between the two groups. Foamy macrophages that replaced epithelioid macrophages in granulomas of four coinfected persons with $<200 \text{ pCD4/mm}^3$, were more intensely stained for IL-1 α , IL-1 β , IFN- α and IFN- β [31]. The lack of studies and high variability in staining makes it difficult to quantify how HIV-1 changes cytokine production within Mtb granulomas.

3.5. Does HIV-1 increase the Mtb content of granulomas?

The primary function of a granuloma is to kill Mtb and prevent its dissemination. HIV-1 increases the risk of extrapulmonary TB [1]. The increased risk of Mtb dissemination is hypothesized to be caused by reduced Mtb killing within the granulomas. To determine if Mtb granulomas from co-infected persons were more likely to contain Mtb than HIV-1 uninfected persons, we analyzed studies that specifically quantified either AFB+ or CFU+ samples out of the total number of excised tissue from both co-infected and singly infected persons. One study could not be included in this meta-analysis because their lowest categorical score was defined as *no more than a single bacillary unit was seen in ten microscopic fields* [28], which could technically represent zero bacilli within ten fields (AFB-) or one bacilli (AFB+).

pCD4 depletion increases the likelihood of Mtb granulomas containing at least one bacillus, while HIV does not increase bacilli presence. Of the seven studies that reported the proportion of AFB+ biopsied tissue within HIV-1 co-infected and uninfected persons [23–25,31,32,80,82] a high variability of AFB+ rates existed in both groups with HIV-1 co-infected persons ranging from 1.8 [24] to 75% [32] and HIV-1 uninfected persons from 0 [32] to 62.5% [31]. HIV-1 did not change AFB+ presence (Figure 4A. RR 1.19 [0.52, 2.71]). Likewise, the seven studies that identified CFU+ [23–25,35,36,80,82] did not suggest a difference in bacterial content (Figure 4B. RR 1.021 [0.89, 1.63]).

As pCD4 decreased the likelihood of granulomas containing at least one bacillus increased. Two studies found AFB+ presence inversely correlated with pCD4 counts in HIV-1 infected persons [23,28]. Biopsies from co-infected persons with <100 pCD4/mm³ were more likely to be AFB+ (43%, 6/14) than co-infected persons with >100 pCD4/mm³ (8%, 3/36) [23]. The depletion of pCD4 T cells led to increases in Mtb presence within granulomas.

HIV-1 increased the amount of Mtb in granulomas. Three studies quantified AFB load in excised tissues in each group [23,28,32]. One study quantified AFB load between a low \leq 1 AFB to a high of >10 AFB per 100× ROI [28], another identified the number of tissue samples with either numerous vs. scanty AFB [23], and the final study categorically assigned absent to abundant AFB [32] without a description. The forest plot and subsequent calculated risk ratio for HIV-1 infected persons identified an increased AFB load within AFB+ tissue (Figure 5. RR 4.30 [1.07, 17.18]). Di Perri et al. [28], identified an inverse correlation between pCD4 count and AFB load within excised tissue from co-infected persons.

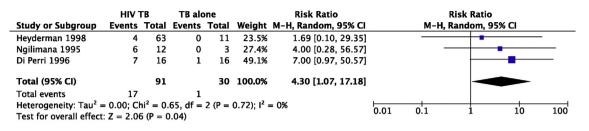


Figure 5. Mantel-Haenszel random effects risk ratio for abundant AFB load in HIV-1 co infected persons.

Conversely, HIV-1 infection status did not change days to positive BACTEC culture in excised pleural tissue [35]. HIV-1 infection appears to increase Mtb abundance within individual granulomas.

3.6. Does HIV-1 preferentially infect TB diseased tissue?

TB granulomas have been hypothesized to be preferential sites of HIV-1 replication (Table 1) [4,5,12]. If HIV-1 preferentially localizes within granulomas then the virus would have a better chance of killing Mtb-specific T cells and Mtb-infected macrophages because of their high prevalence within granulomas. This specific killing could lead to increased Mtb growth.

HIV-1 infects Mtb granulomas. Van der Ende et al. [38] gualitatively examined HIV-1 RNA presence within T cells and macrophages in lymph nodes from five persons with >150 pCD4/mm³ to two persons with <50 pCD4/mm³. In the former group, there were well-organized granulomas and the number of HIV-1 RNA+ cells was 10-fold higher in or near granulomas compared to other nongranulomatous areas. In those persons with pCD4 counts <50 pCD4/mm³, HIV-1 RNA+ cells were distributed more evenly throughout the lymph node, independent of the inflammatory infiltrate. CD4 T cells were the dominant source of HIV-1 RNA within most of the lymph nodes. These findings were based on a small sample size and were only presented as an overall impression rather than by individual findings. HIV-1 viral RNA was also identified with PCR in homogenized spinal tissue that contained granulomas [33], but it was not possible to determine if virus originated in the granuloma or surrounding tissue. HIV-1 does localize within Mtb granulomas, but more studies need to be performed to confirm what cells are preferentially infected.

4. Discussion

It has been hypothesized that HIV-1 increases TB susceptibility by reducing the formation and function of Mtb granulomas. Studies that have examined Mtb granulomas directly in HIV-1 co-infected persons have not been collated until now. We performed this systematic review and meta-analysis to determine which hypotheses could be confirmed by the human literature and which ones need more research. Despite the strong epidemiological association between TB and HIV-1s [12], research into how HIV-1 manipulates the Mtb granuloma was conflicting and quite limited.

One outcome with consistent findings and a significant difference between the HIV-1 infected and uninfected groups was bacillary load. In the three studies that examined this outcome, HIV-1 co-infected persons consistently demonstrated greater bacillary load [23,28,32]. This was the only finding that provided some potential explanation for the increased susceptibility to more severe TB disease in HIV-1 infected persons. However, when studies were stratified by pCD4 counts [23,28,31,35,38], results were more consistent. Lower pCD4 counts were associated with increased bacillary presence [23,28], inflammatory cytokine production by foamy macrophages [31], and poorer granuloma formation [28,31,38]. These data suggest that as HIV-1 disease progresses, the ability to maintain normal granuloma function becomes impaired. These findings also illuminate the importance of stratifying pCD4 counts when analyzing outcomes from HIV-1 infected persons to achieve more accurate and consistent findings. Consideration of other clinical characteristics including duration of TB disease or symptoms, gender, age, HIV-1 and TB treatment status, and previous TB disease may also help to improve the consistency of findings.

In all other outcomes, there was high variation between studies, both in terms of methods used to assess outcomes, and also their results. Heterogeneity scores for all measured outcomes were moderate to high, and the summative risk ratios had very wide confidence intervals for the outcomes of granuloma presence, formation quality and AFB presence when comparing TB diseased tissue from HIV-1 infected and HIV-1 uninfected persons. In terms of studies' outcomes such as cellular composition of granulomas and cytokine expression, findings were also conflicting and no trend could be extracted. There was also a surprising lack of data that focused on the effect Mtb bacilli had upon the distribution of HIV-1 transcripts. Studies that only described findings from TB

Table	3
-------	---

Guidelines for future granuloma-based studies.

Category	Recommendations for future granuloma-based studies
Granuloma descriptions	 Include detailed descriptions of all granuloma scores for individual types of granulomas with reference images (if space is limited then the authors should provide example images upon request).
descriptions	 Detail how scores were tabulated. Were all granulomas counted and averaged? Do certain granulomas skew results because of their size?
	 Detail now scores were tabulated, were an granulonas counted and averaged? Do certain granulonas skew results because of their size? Present the range of granuloma scores within each individual as well as each group.
	• Explain how each granuloma or region of interest was chosen to be scored (random selection?)
	 Only compare granulomas that reside in the same tissue type among persons to reduce variability.
Study groups	HIV-1 and TB diagnoses should be included for all patients
	HIV-1 studies should include pCD4 counts
	Drug treatment should be stated
Study design	Were the observer's blinded?
	• Describe how each parameter was specifically quantified instead of using vague subjective terms such as "abundant" or "scanty".
	Were their enough persons to justify conclusions?
	• Autopsy studies should state how long the subject was dead prior to autopsy and studies that focus on excised tissue should state the time it required to fix the tissue.

Table 4	
Questions that still need to be answered within the HI	V-1/Mtb co-infection literature.

Category	Questions
Compositional	Does HIV-1 significantly change granuloma formation?
changes	Do HIV-1 and AFB presence correlate in infected tissue?
	Does HIV-1 significantly reduce CD4 T cell counts within granulomas?
Functional	• Does HIV-1 significantly reduce Mtb-specific T cell functionality (chemokine and cytokine production or cytolytic activity) within granulomas
changes	more than within the periphery?
-	Does pCD4 T cell depletion correlate to increased changes within granulomas?
	• How do functional T cell responses correlate among PBMC, BALF, PF, and within granulomas and does this correlate to disease status?
HIV-1 and ART	• Does HIV-1 preferentially replicate within granulomas? Which cells are preferentially infected within granulomas and the surrounding tissue?
focused	Do HIV-1 and Mtb co-localize within the same cells or within close proximity inside of granulomas?
research	Does anti-viral penetration of granulomas correlate to improved granuloma function, organization, reduced viral titers, or patient outcome?
Identifying	Does anti-viral penetration into granulomas correlate to pharmacokinetics observed within the plasma?
biomarkers	What blood biomarkers correlate to significant changes in granuloma function?

diseased tissue from HIV-1 infected persons that did not stratify pCD4 counts or make comparisons to HIV-1 uninfected persons were excluded from our review. However, data from the studies we excluded [39–46] revealed similar variability in findings, particularly in regards to granuloma formation and AFB presence (Supplemental Table 2). Heterogeneity and the high variability among techniques used may have reduced consistency in findings among the studies examined in this review. Some of the variability among these studies may have been reduced if they had been better powered.

High variability in methods used for each study may have limited our findings. As previously mentioned, where semiquantified scoring systems were used to quantify outcomes such as granuloma formation or AFB load, methods either varied or were not described. The methods scoring outcomes varied considerably: *poorly- or well-formed granulomas* [23,27,31], *well-formed with giant cells* [35], *scanty or numerous AFB* [23], *rare or abundant AFB* [32], +++ *or* + *AFB* [30] and other semi-quantified scores [28,30–32,81]. These terms were often applied without specific definition and as such were open to subjective interpretation and observer bias [85]. A significant variation likely existed in how individual tissues were scored among the studies and it was difficult to identify the effect this had on our overall results.

A second concern was that for semi-quantified variables, in particular quality of granuloma formation, results were represented by a single score. This score may have failed to represent the range of granulomas or unorganized inflammation that was likely located within a single tissue section. Granulomas are highly variable in terms of size, histological type [6], formation quality [31], cytokine expression [84], and bacterial load [86]. It was not clear in these studies whether this variability was assessed or how it was handled when choosing how to score the tissues. Conceivably, a number of strategies may have been adopted if such variations were present, including scoring by the largest granulomas, the most numerous, a *random* selection within the tissue, or an average of all granulomas present. Without knowing which methods were used to provide an overall score for tissues, the implication for potential bias was difficult to ascertain. The high variability in findings may have been reduced if all granulomas identified within each tissue was represented, along with exactly how each granuloma was selected and scored. Only one study specifically stated that the assessing pathologists were blinded [30], which could have reduced the risk of observer bias [85].

While the studies we reviewed assessed the effect of HIV-1 on Mtb granulomas, we have demonstrated there was little that could be conclusively stated about how HIV-1 changes the site of TB disease. The primary issue we identified was that findings from different studies could not always be compared, despite having examined the same outcomes. To increase comparability of future studies we have developed a basic guideline for research on the pathology of HIV-1/Mtb co-infection (Table 3) because important unanswered questions need further exploration (Table 4).

The objective of our guidelines are to ensure that future studies provide enough detail through a reduced-biased methodology that will allow other researchers to compare results. We also encouraged open access to examined images of Mtb-infected tissues within these studies. This would facilitate researchers' ability to evaluate their peers' analyses of these tissues and minimize redundant research effort. Implementing both these guidelines and open access to pathology images in future work would help build consensus on their criteria needed to properly assess granulomas, even if the number of available samples are small in individual studies. Increasing the cohesiveness of data will heavily contribute to the overall understanding of how HIV-1 alters immunity at the site of TB disease. We hope this will provide a greater understanding of the dynamics of this co-infection at the local level, which in turn may lead to novel interventions and therapies targeting the microenvironment of granulomas.

Acknowledgments

The authors would like to thank Philana Ling Lin and Joshua T. Mattila for their input on this manuscript.

Appendix A. Supplementary data

Supplementary data related to this article can be found at http://dx.doi.org/10.1016/j.tube.2016.02.010.

Funding: This research received funding from the European Community's Seventh Framework Program [FP7-2007-2013] under grant agreement HEALTH-F3-2012-305578. This study was supported by Wellcome Trust (084323, 104803), European Union (FP7-PEOPLE-2011) and Claude Leon Foundation. The funders of this review had no role in study design, data collection, data analysis, data interpretation, or writing of the report. RJW receives support from the Medical Research Council of the UK (U1175.02.002.00014.01) and from the National Research Foundation of South Africa (96841).

Competing interests: The authors do not have any conflicts of interest to disclosure.

Ethical approval: Not required.

References

- [1] World Health Organization. Global tuberculosis report 2014. World Health Publications; 2015.
- [2] Marcy O, Laureillard D, Madec Y, Chan S, Mayaud C, Borand L, et al. Causes and determinants of mortality in HIV-infected adults with tuberculosis: an

analysis from the CAMELIA ANRS 1295-CIPRA KH001 randomized trial. Clin Infect Dis 2014;59:435-45. http://dx.doi.org/10.1093/cid/ciu283.

- [3] Gupta RK, Lawn SD, Bekker LG, Caldwell J, Kaplan R, Wood R. Impact of human immunodeficiency virus and CD4 count on tuberculosis diagnosis: analysis of city-wide data from Cape Town, South Africa. Int J Tuberc Lung Dis 2013;17: 1014–22. http://dx.doi.org/10.5588/ijtld.13.0032.
- [4] Diedrich CR, Flynn JL. HIV-1/Mycobacterium tuberculosiscoinfection immunology: how does HIV-1 exacerbate tuberculosis? Infect Immun 2011;79: 1407–17. http://dx.doi.org/10.1128/IAI.01126-10.
- [5] Lawn SD, Butera ST, Shinnick TM. Tuberculosis unleashed: the impact of human immunodeficiency virus infection on the host granulomatous response to Mycobacterium tuberculosis. Microbes Infect 2002;4:635–46.
- [6] Flynn JL, Klein E. "Pulmonary tuberculosis in monkeys" in a color atlas of comparative pathology of pulmonary tuberculosis. Boca Raton: CRC Press, Taylor & Francis Publishers; 2011.
- [7] Ramakrishnan L. Revisiting the role of the granuloma in tuberculosis. Nat Rev Immunol 2012;12:352–66. http://dx.doi.org/10.1038/nri3211.
- [8] Ozsoy S, Demirel B, Albay A, Kisa O, Dinc AH, Safali M. Tuberculosis prevalence in forensic autopsies. Am J Forensic Med Pathol 2010;31:55–7. http:// dx.doi.org/10.1097/PAF.0b013e3181c215f9.
- [9] Silva Miranda M, Breiman A, Allain S, Deknuydt F, Altare F. The tuberculous granuloma: an unsuccessful host defence mechanism providing a safety shelter for the bacteria? Clin Dev Immunol 2012;2012:139127. http:// dx.doi.org/10.1155/2012/139127.
- [10] Ledru E, Ledru S, Zoubga A. Granuloma formation and tuberculosis transmission in HIV-infected patients. Immunol Today 1999;20:336–7.
- [11] Bocchino M, Sanduzzi A, Bariffi F. Mycobacterium tuberculosis and HIV-1coinfection in the lung: synergic immune dysregulation leading to disease progression. Monaldi Arch Chest Dis 2000;55:381–8.
- [12] Kwan CK, Ernst JD. HIV-1and tuberculosis: a deadly human syndemic. Clin Microbiol Rev 2011;24:351–76. http://dx.doi.org/10.1128/CMR.00042-10.
- [13] Geldmacher C, Zumla A, Hoelscher M. Interaction between HIV-1and Mycobacterium tuberculosis: HIV-1-induced CD4 T-cell depletion and the development of active tuberculosis. Curr Opin HIV-1 AIDS 2012;7:268–75. http:// dx.doi.org/10.1097/COH.0b013e3283524e32.
- [14] Ansari AW, Kamarulzaman A, Schmidt RE. Multifaceted impact of host C-C chemokine CCL2 in the immuno-pathogenesis of HIV-1/M. tuberculosis coinfection. Front Immunol 2013;4:312. http://dx.doi.org/10.3389/ fimmu.2013.00312.
- [15] Sutherland R, Yang H, Scriba TJ, Ondondo B, Robinson N, Conlon C, et al. Impaired IFN-gamma-secreting capacity in mycobacterial antigen-specific CD4 T cells during chronic HIV-1 infection despite long-term HAART. Aids 2006;20:821-9. http://dx.doi.org/10.1097/01.aids.0000218545.31716.a4.
- [16] Matthews K, Ntsekhe M, Syed F, Scriba T, Russell J, Tibazarwa K, et al. HIV-1 infection alters CD4+ memory T-cell phenotype at the site of disease in extrapulmonary tuberculosis. Eur J Immunol 2012;42:147–57. http:// dx.doi.org/10.1002/eji.201141927.
- [17] Law KF, Jagirdar J, Weiden MD, Bodkin M, Rom WN. Tuberculosis in HIVpositive patients: cellular response and immune activation in the lung. Am J Respir Crit Care Med 1996;153:1377–84. http://dx.doi.org/10.1164/ ajrccm.153.4.8616569.
- [18] Kalsdorf B, Scriba TJ, Wood K, Day CL, Dheda K, Dawson R, et al. HIV-1 infection impairs the bronchoalveolar T-cell response to mycobacteria. Am J Respir Crit Care Med 2009;180:1262–70. http://dx.doi.org/10.1164/ rccm.200907-10110C.
- [19] Geldmacher C, Schuetz A, Ngwenyama N, Casazza JP, Sanga E, Saathoff E, et al. Early depletion of *Mycobacterium tuberculosis*-specific T helper 1 cell responses after HIV-1 infection. J Infect Dis 2008;198:1590–8. http://dx.doi.org/ 10.1086/593017.
- [20] Herzmann C, Ernst M, Ehlers S, Stenger S, Maertzdorf J, Sotgiu G, et al. Increased frequencies of pulmonary regulatory T-cells in latent Mycobacterium tuberculosis infection. Eur Respir J 2012;40:1450-7. http://dx.doi.org/ 10.1183/09031936.00214611.
- [21] Mattila JT, Diedrich CR, Lin PL, Phuah J, Flynn JL. Simian immunodeficiency virus-induced changes in T cell cytokine responses in cynomolgus macaques with latent *Mycobacterium tuberculosis* infection are associated with timing of reactivation. J Immunol 2011;186:3527–37. http://dx.doi.org/10.4049/ jimmunol.1003773.
- [22] Brighenti S, Andersson J. Local immune responses in human tuberculosis: learning from the site of infection. J Infect Dis 2012;205(Suppl. 2):S316–24. http://dx.doi.org/10.1093/infdis/jis043.
- [23] Heyderman RS, Makunike R, Muza T, Odwee M, Kadzirange G, Manyemba J, et al. Pleural tuberculosis in Harare, Zimbabwe: the relationship between human immunodeficiency virus, CD4 lymphocyte count, granuloma formation and disseminated disease. Trop Med Int Health 1998;3:14–20. http:// dx.doi.org/10.1046/J.1365-3156.1998.00167.X.
- [24] Kennedy DJ, Lewis WP, Barnes PF. Yield of bronchoscopy for the diagnosis of tuberculosis in patients with human immunodeficiency virus infection. Chest 1992;102:1040–4.
- [25] Reuter H, Burgess LJ, Schneider J, Van Vuuren W, Doubell AF. The role of histopathology in establishing the diagnosis of tuberculous pericardial effusions in the presence of HIV. Histopathology 2006;48:295–302. http:// dx.doi.org/10.1111/j.1365-2559.2005.02320.x.
- [26] Shen JY, Barnes PF, Rea TH, Meyer PR. Immunohistology of tuberculous adenitis in symptomatic HIV-1infection. Clin Exp Immunol 1988;72:186–9.

- [27] Trajman A, Neto EB, Belo MT, Teixeira EG, Selig L, Ferrari G, et al. Pleural tuberculosis and human immunodeficiency virus co-infection. Int J Tuberc Lung Dis December 1997;1(6). 498–501(4) 1997;1:498–501.
- [28] Di Perri G, Cazzadori A, Vento S, Bonora S, Malena M, Bontempini L, et al. Comparative histopathological study of pulmonary tuberculosis in human immunodeficiency virus-infected and non-infected patients. Tuber Lung Dis 1996;77:244–9.
- [29] Pefura Yone EW, Kuaban C, Simo L. Tuberculous pleural effusion in Yaounde, Cameroon: the influence of HIV-1infection. Rev Des Mal Respir 2011;28: 1138–45. http://dx.doi.org/10.1016/j.rmr.2011.05.008.
- [30] de Noronha ALL, Báfica A, Nogueira L, Barral A, Barral-Netto M. Lung granulomas from Mycobacterium tuberculosis/HIV-1 co-infected patients display decreased in situ TNF production. Pathol Res Pract 2008;204:155–61. http:// dx.doi.org/10.1016/j.prp.2007.10.008.
- [31] Muller H, Kruger S. Immunohistochemical analysis of cell composition and in situ cytokine expression in HIV- and non-HIV-associated tuberculous lymphadenitis. Immunobiology 1994;191:354–68. http://dx.doi.org/10.1016/ S0171-2985(11)80441-9.
- [32] Ngilimana PJ, Metz T, Munyantore S, Mureganshuro JM, Noel H, Roels H. Lymph node tuberculosis in HIV-1 seropositive patients in Central Africa. A characteristic histopathologic picture. Ann De Pathol 1995;15:38–44.
- [33] Danaviah S, Sacks JA, Kumar KP, Taylor LM, Fallows DA, Naicker T, et al. Immunohistological characterization of spinal TB granulomas from HIVnegative and -positive patients. Tuberculosis 2013;93:432–41. http:// dx.doi.org/10.1016/j.tube.2013.02.009.
- [34] Jones BE, Young SM, Antoniskis D, Davidson PT, Kramer F, Barnes PF. Relationship of the manifestations of tuberculosis to CD4 cell counts in patients with human immunodeficiency virus infection. Am Rev Respir Dis 1993;148: 1292–7. http://dx.doi.org/10.1164/ajrccm/148.5.1292.
- [35] Luzze H, Elliott AM, Joloba ML, Odida M, Oweka-Onyee J, Nakiyingi J, et al. Evaluation of suspected tuberculous pleurisy: clinical and diagnostic findings in HIV-1-positive and HIV-negative adults in Uganda 2001;5:746–53.
- [36] Elliott AM, Halwiindi B, Hayes RJ, Luo N, Tembo G, Machiels L, et al. The impact of human immunodeficiency virus on presentation and diagnosis of tuberculosis in a cohort study in Zambia. J Trop Med Hyg 1993;96:1–11.
- [37] Bezuidenhout J, Roberts T, Muller L, van Helden P, Walzl G. Pleural tuberculosis in patients with early HIV-1infection is associated with increased TNFalpha expression and necrosis in granulomas. PLoS One 2009;4:e4228. http://dx.doi.org/10.1371/journal.pone.0004228.
- [38] van der Ende ME, Schutten M, Raschdorff B, Grossschupff G, Racz P, Osterhaus AD, et al. CD4 T cells remain the major source of HIV-1 during end stage disease. AIDS 1999;13:1015–9.
- [39] Karunatilake H, Thamilvannan N, Wimalaratna H. Lack of granuloma formation in tuberculous lymphadenitis—clue to the diagnosis of human immunodeficiency virus infection. Ceylon Med J 2002;47:37.
- [40] Fischl MA, Daikos GL, Uttamchandani RB, Poblete RB, Moreno JN, Reyes RR, et al. Clinical presentation and outcome of patients with HIV-1infection and tuberculosis caused by multiple-drug-resistant bacilli. Ann Intern Med 1992;117:184–90.
- [41] Baba K, Dyrhol-Riise AM, Sviland L, Langeland N, Hoosen AA, Wiker HG, et al. Rapid and specific diagnosis of tuberculous pleuritis with immunohistochemistry by detecting Mycobacterium tuberculosis complex specific antigen MPT64 in patients from a HIV-1endemic area. Appl Immunohistochem Mol Morphol 2008;16:554–61. http://dx.doi.org/10.1097/PAI.0b013e31816c3f79.
- [42] Piratvisuth T, Siripaitoon P, Sriplug H, Ovartlarnporn B. Findings and benefit of liver biopsies in 46 patients infected with human immunodeficiency virus. J Gastroenterol Hepatol 1999;14:146–9.
- [43] Bhoopat L, Thamprasert K, Chaiwun B, Attasiri C, Vithayasai P, Chaimongkol B, et al. Histopathologic spectrum of AIDS-associated lesions in Maharaj Nakorn Chiang Mai Hospital. Asian Pac J Allergy Immunol 1994;12:95–104.
- [44] Yang GC, Schinella RA. The histopathology of tuberculosis in the acquired immunodeficiency syndrome: a study of nine cases. Prog AIDS Pathol 1990;2: 103–10.
- [45] Nambuya A, Sewankambo N, Mugerwa J, Goodgame R, Lucas S. Tuberculous lymphadenitis associated with human immunodeficiency virus (HIV) in Uganda. J Clin Pathol 1988;41:93–6.
- [46] Jagadha V, Andavolu RH, Huang CT. Granulomatous inflammation in the acquired immune deficiency syndrome. Am J Clin Pathol 1985;84:598–602.
- [47] Bem C, Patil PS, Elliott AM, Namaambo KM, Bharucha H, Porter JD. The value of wide-needle aspiration in the diagnosis of tuberculous lymphadenitis in Africa. Aids 1993;7:1221–5.
- [48] Finfer M, Perchick A, Burstein DE. Fine needle aspiration biopsy diagnosis of tuberculous lymphadenitis in patients with and without the acquired immune deficiency syndrome. Acta Cytol 1991;35:325–32.
- [49] Nayak S, Puranik SC, Deshmukh SD, Mani R, Bhore AV, Bollinger RC. Fineneedle aspiration cytology in tuberculous lymphadenitis of patients with and without HIV-1infection. Diagn Cytopathol 2004;31:204–6. http://dx.doi.org/ 10.1002/dc.20072.
- [50] Rajasekaran S, Gunasekaran M, Jayakumar DD, Jeyaganesh D, Bhanumathi V. Tuberculous cervical lymphadenitis in HIV-1 positive and negative patients. Indian J Tuberc 2001;48:201–4.
- [51] Schubert PT, Cotton MF, Wright CA. Cytomorphological patterns of M. bovis BCG and M. tuberculosis on fine needle aspiration biopsies: does HIV-1 make a difference? Diagn Cytopathol 2011;39:264–9. http://dx.doi.org/10.1002/ dc.21378.

- [52] Shriner KA, Mathisen GE, Goetz MB. Comparison of mycobacterial lymphadenitis among persons infected with human immunodeficiency virus and seronegative controls. Clin Infect Dis 1992;15:601–5.
- [53] Sridhar CB, Kini U, Subhash K. Comparative cytological study of lymph node tuberculosis in HIV-infected individuals and in patients with diabetes in a developing country. Diagn Cytopathol 2002;26:75–80.
- [54] Llatjos M, Romeu J, Clotet B, Sirera G, Manterola JM, Pedro-Botet ML, et al. A distinctive cytologic pattern for diagnosing tuberculous lymphadenitis in AIDS. J Acquir Immune Defic Syndr 1993;6:1335–8.
- [55] Kamana NK, Wanchu A, Sachdeva RK, Kalra N, Rajawanshi A. Tuberculosis is the leading cause of lymphadenopathy in HIV-infected persons in India: results of a fine-needle aspiration analysis. Scand J Infect Dis 2010;42:827–30. http://dx.doi.org/10.3109/00365548.2010.498016.
- [56] Kumar P, Shashikala P, Chandrashekar HR, Alva NK. Acquired immunodeficiency syndrome presenting as testicular tuberculosis. J Assoc Physicians India 2000;48:1111–2.
- [57] Hemalatha AN, Manjunath YA, Prakash CJ. Fine needle aspiration biopsy diagnosis of tuberculous lymphadenitis in patients with and without acquired immune deficiency. Natl Tuberc Inst Bull 2000;36:5–6.
- [58] Pithie AD, Chicksen B. Fine-needle extrathoracic lymph-node aspiration in HIV-associated sputum-negative tuberculosis. Lancet 1992;340:1504–5.
- [59] Wamala D, Asiimwe B, Kigozi E, Mboowa G, Joloba M, Källenius G. Clinicopathological features of tuberculosis due to *Mycobacterium tuberculosis* Uganda genotype in patients with tuberculous lymphadenitis: a cross sectional study. BMC Clin Pathol 2014;14:14. http://dx.doi.org/10.1186/1472-6890-14-14.
- [60] Tirumalasetti N, Latha PP. Lymph nodes cytology in HIV-1 seropositive cases with haematological alterations. Indian J Med Res 2014;139:301–7.
- [61] Ablanedo-Terrazas Y, Alvarado-de la Barrera C, Ruiz-Cruz M, Reyes-Terán G. Mycobacterial cervicofacial lymphadenitis in human immunodeficiency virusinfected individuals after antiretroviral therapy initiation. Laryngoscope 2015. http://dx.doi.org/10.1002/lary.25470.
- [62] Van Rie A, Page-Shipp L, Mellet K, Scott L, Mkhwnazi M, Jong E, et al. Diagnostic accuracy and effectiveness of the Xpert MTB/RIF assay for the diagnosis of HIV-associated lymph node tuberculosis. Eur J Clin Microbiol 2013;32: 1409–15. http://dx.doi.org/10.1007/s10096-013-1890-0.
- [63] Miro AM, Gibilara E, Powell S, Kamholz SL. The role of fiberoptic bronchoscopy for diagnosis of pulmonary tuberculosis in patients at risk for AIDS. Chest 1992;101:1211–4.
- [64] Hill AR, Premkumar S, Brustein S, Vaidya K, Powell S, Li PW, et al. Disseminated tuberculosis in the acquired immunodeficiency syndrome era. Am Rev Respir Dis 1991;144:1164–70.
- [65] Relkin F, Aranda CP, Garay SM, Smith R, Berkowitz KA, Rom WN. Pleural tuberculosis and HIV-1 infection. Chest 1994;105:1338–41.
- [66] Vilaichone RK, Vilaichone W, Tumwasorn S, Suwanagool P, Wilde H, Mahachai V. Clinical spectrum of hepatic tuberculosis: comparison between immunocompetent and immunocompromised hosts. J Med Assoc Thai 2003;86(Suppl. 2):S432–8.
- [67] Govender S, Annamalai K, Kumar KP, Govender UG. Spinal tuberculosis in HIV-1positive and negative patients: immunological response and clinical outcome. Int Orthop 2000;24:163–6. http://dx.doi.org/10.1007/ s002640000125.
- [68] Leong AS, Wannakrairot P, Leong TY. Apoptosis is a major cause of so-called "caseous necrosis" in mycobacterial granulomas in HIV-infected patients. J Clin Pathol 2008;61:366–72. http://dx.doi.org/10.1136/jcp.2007.050690.
- [69] Orenstein MS, Tavitian A, Yonk B, Dincsoy HP, Zerega J, Iyer SK, et al. Granulomatous involvement of the liver in patients with AIDS. Gut 1985;26: 1220–5.

- [70] Wannakrairot P, Leong TY, Leong AS. The morphological spectrum of lymphadenopathy in HIV-1infected patients. Pathology 2007;39:223–7. http://dx.doi.org/10.1080/00313020701230674.
- [71] Chensue SW, Warmington KS, Berger AE, Tracey DE. Immunohistochemical demonstration of Interleukin-1 receptor antagonist protein and interleukin-1 in human lymphoid-tissue and granulomas. Am J Pathol 1992;140: 269–75.
- [72] Gutierrez EB, Zanetta DMT, Saldiva PHN, Capelozzi VL. Autopsy-proven determinants of death in HIV-infected patients treated for pulmonary tuberculosis in Sao Paulo, Brazil. Pathol Res Pract 2002;198:339–46.
- [73] Danaviah S, Govender S, Cassol S. Histopathology and genotyping in infectious spondylitis of HIV- and HIV+ patients. Clin Orthop Relat Res 2007;460:50–5. http://dx.doi.org/10.1097/BLO.0b013e31806a9147.
- [74] Muller H, Takeshita M. In situ immunophenotype of macrophages and lymphocytes in granuloma formation of tuberculous lymphadenitis in HIVinfected and immunocompetent patients. Res Virol 1991;142:159–72.
- [75] Babaeva II. Immunomorphologic features in HIV-infected patients with tuberculosis. Zhurnal Mikrobiol Epidemiol I Immunobiol 2008:101–3.
- [76] Babaeva I, Zemskova ZS, Gedymin LE, Demikhova OV. Pathomorphological features of pulmonary tuberculosis at different stages of HIV-1infection: autopsy data. Probl Tuberk I Bolezn Legk 2007:38–42.
- [77] Rana F, Hawken MP, Meme HK, Chakaya JM, Githui WA, Odhiambo JA, et al. Autopsy findings in HIV-1-infected adults in Kenya. J Acquir Immune Defic Syndr Hum Retrovirol 1997;14:83–5.
- [78] Abdel-Dayem HM, Naddaf S, Aziz M, Mina B, Turoglu T, Akisik MF, et al. Sites of tuberculous involvement in patients with AIDS. Autopsy findings and evaluation of gallium imaging. Clin Nucl Med 1997;22:310–4.
- [79] Harkin TJ, Ciotoli C, Addrizzo-Harris DJ, Naidich DP, Jagirdar J, Rom WN. Transbronchial needle aspiration (TBNA) infected with HIV. Am J Respir Crit Care Med 1998;157:1913–8.
- [80] Hochedez P, Zeller V, Truffot C, Ansart S, Caumes É, Tubiana R, et al. Lymphnode tuberculosis in patients infected or not with HIV: general characteristics, clinical presentation, microbiological diagnosis and treatment. Pathol Biol 2003;51:496–502.
- [81] Tripathi S, Patro I, Mahadevan A, Patro N, Phillip M, Shankar SK. Glial alterations in tuberculous and cryptococcal meningitis and their relation to HIV-1co-infection – a study on human brains. J Infect Dev Ctries 2014;8:1–23. http://dx.doi.org/10.3855/jidc.3894.
- [82] Conde MB, Loivos AC, Rezende VM, Soares SL, Mello FC, Reingold AL, et al. Yield of sputum induction in the diagnosis of pleural tuberculosis. Am J Respir Crit Care Med 2003;167:723–5. http://dx.doi.org/10.1164/rccm.2111019.
- [83] van der Ende ME, Schutten M, Racz P, Tanner-Racz K, Osterhaus AD. CD4+-Tcells are the predominant source of HIV-1 and HIV-2 production in asymptomatic people and AIDS patients with opportunistic infections. Nederl Tijdschr Voor Geneeskd 1998;142:2230.
- [84] Gideon HP, Phuah J, Myers AJ, Bryson BD, Rodgers MA, Coleman MT, et al. Variability in tuberculosis granuloma T cell responses exists, but a balance of pro- and anti-inflammatory cytokines is associated with sterilization. PLoS Pathog 2015;11:e1004603. http://dx.doi.org/10.1371/journal.ppat.1004603.
- [85] Guillery RW. On counting and counting errors. J Comp Neurol 2002;447:1–7. http://dx.doi.org/10.1002/cne.10221.
- [86] Lin PL, Ford CB, Coleman MT, Myers AJ, Gawande R, loerger T, et al. Sterilization of granulomas is common in active and latent tuberculosis despite within-host variability in bacterial killing. Nat Med 2014;20:75–9. http:// dx.doi.org/10.1038/nm.3412.