Supporting Information for

Recognition of HClO in Live Cells with Separate Signals Using a Ratiometric Fluorescent Sensor with Fast Response

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Materials and methods

All of the solvents used were of analytical grade. **BRCIO** was dissolved in dimethyl sulphoxide (DMSO) at a concentration of 5 mM as the stock solution. Slight pH variations in the solutions were achieved by adding the minimum volumes of NaOH or HCl (1 M). 1 H-NMR and 13 C-NMR spectra were recorded on a VARIAN INOVA-400 spectrometer with chemical shifts (δ) reported as ppm (in CDCl₃, TMS as the internal standard). Mass spectrometry data were obtained with an HP1100LC/MSD mass spectrometer and an LC/Q-TOF MS spectrometer. Fluorescence measurements were performed on a VARIAN CARY Eclipse fluorescence spectrophotometer (Serial No. FL0812-M018). Excitation and emission slit widths were modified to adjust the fluorescence intensity to a suitable range. Absorption spectra were measured on a Lambda 35 UV/VIS spectrophotometer (Perkin Elmer). All pH measurements were made with a Model PHS-3C meter.

Live cell imaging experiments

MCF-7 cells were cultured in Dulbecco's modified Eagle's medium (Invitrogen) supplemented with 10% fetal bovine serum (Invitrogen). Cells were seeded in 24-well flat-bottomed plates and incubated for 24 h at 37 °C under 5% CO₂. **BRCIO** (1 μM) was added (the concentration of DMSO was maintained to be less than 0.2%) and cells were further incubated for 20 min, followed by washing thrice with phosphate-buffered saline (PBS). After incubation with NaOCl (100 μM), the MCF-7 cells were rinsed three times with PBS and the fluorescence imaging was performed with OLYMPUS FV-1000 inverted fluorescence microscope with 100× objective lens. Under the confocal fluorescence microscope, **BRCIO** was excited at 488 nm and emission was collected at green channel (490-520 nm) and red channel (560-600 nm).

Computational calculation

All the quantum-chemical calculations were done with the Gaussian 09 suite¹. The parameter referred to the previous work²⁻³. The geometry optimizations of the dyes were performed using density functional theory (DFT) with Becke's three-parameter hybrid exchange function with Lee-Yang-Parr gradient-corrected correlation functional (B3-LYP functional) and 6-31G** basis set⁴. No constraints to bonds/angles/dihedral angles were applied in the calculations and all atoms were free to optimize.

Determination of the detection limit

The detection limit was calculated based on the fluorescence titration curve of **BRCIO** in the presence of NaClO (0-100 μ M). The fluorescence intensity of **BRCIO** was measured by three times and the standard deviation of blank measurement was achieved. The detection limit was calculated by using detection limit was calculated with the following equation:

Detection limit = $3\sigma/k$

Where σ is the standard deviation of the blank measurement, k is the slope between the fluorescence ratios versus NaClO concentration.

Determination of quantum yields

The fluorescence quantum yields of **BRCIO** and **BORCIO** were determined according to the method below⁵.

$$\varphi_u = \frac{(\varphi_s)(FA_u)(A_s)(\lambda_{exs})(\eta_u^2)}{(FA_s)(A_u)(\lambda_{exu})(\eta_s^2)}$$

Where φ is fluorescence quantum yield; FA is integrated area under the corrected emission spectra; A is the absorbance at the excitation wavelength; $\lambda_{\rm ex}$ is the excitation wavelength; η is the

refractive index of the solution; the subscripts u and s refer to the unknown and the standard, respectively. We chose Rhodamine B as standard, which has a fluorescence quantum yield of 0.49 in ethanol⁶.

Synthesis of BRClO

Pyrrole (1.36 g, 20 mmol) was dissolved in 5 mL CH₂Cl₂, and acetyl chloride (0.22 g, 3 mmol) was added. After the reaction mixture was stirred under N₂ atmosphere at rt for 6 h, triethylamine (2 mL) and BF₃·OEt₂ (4 mL) were added dropwise, and the mixture was stirred at 40 □ overnight. The solvent was removed under reduced pressure and the crude product was purified by column chromatography (silica, CH₂Cl₂/petroleum ether = 1/1, v/v) to obtain **BRCIO** (0.95 g, 17.3 %). ¹H NMR (400 MHz, CDCl₃), δ : 10.48 (s, 1H, NH), 7.60 (s, 1H, CH), 7.32 (s, 1H, CH), 7.16 (s, 1H, CH), 7.01 (s, 1H, CH), 6.90 (s, 1H, CH), 6.47 (s, 1H, CH), 6.38 (s, 1H, CH), 2.53 (s, 3H, CH₃). ¹³C NMR (400 MHz, CDCl₃), δ : 151.25, 137.90, 137.03, 136.20, 133.84, 125.80, 123.50, 121.86, 120.45, 118.07, 115.19, 111.44, 29.55, 15.83. TOF MS: m/z calcd for C₁₄H₁₃BN₃F₂⁺ [M+H]⁺ 272.1092, found: 272.1174.

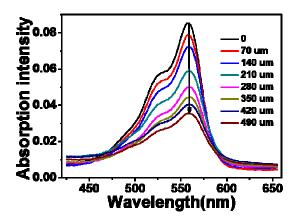


Figure S1 Absorption spectra of **BRCIO** (1 μ M) upon the titration of NaClO (0-490 μ M) in PBS (0.01 M) solution (ethanol/water = 1/9, v/v, pH 7.4).

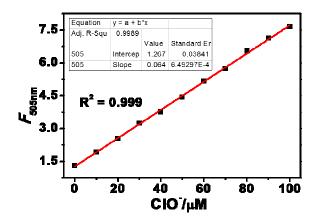


Figure S2 Fluorescence intensity at 505 nm of BRCIO (1 μ M) change as a function of NaClO (0-100 μ M) in PBS (0.01 M) solution (ethanol/water = 1/9, v/v, pH 7.4). λ_{ex} = 480 nm.

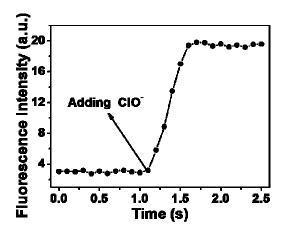


Figure S3 The time courses of Fluorescence intensity at 505 nm of BRCIO (1 μ M) after adding 300 μ M NaClO in PBS (0.01 M) solution (ethanol/water = 1/9, v/v, pH 7.4). Time range: 0-2.5 s. $\lambda_{ex} = 480$ nm.

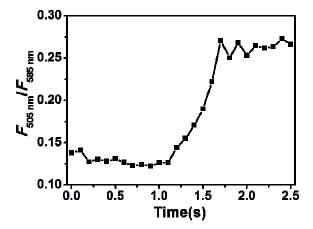


Figure S4 The time courses of Fluorescence ratio ($F_{505 \text{ nm}}/F_{585 \text{ nm}}$) of **BRCIO** (1 µM) after adding 10 µM NaClO in PBS (0.01 M) solution (ethanol/water = 1/9, v/v, pH 7.4). Time range: 0-2.5 s. $\lambda_{\text{ex}} = 480 \text{ nm}$.

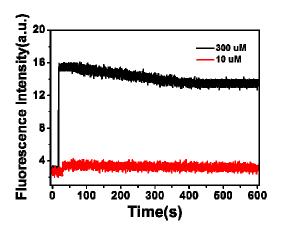


Figure S5 The time courses of Fluorescence intensity at 505 nm of BRCIO (1 μ M) after adding 300 and 10 μ M NaClO in PBS (0.01 M) solution (ethanol/water = 1/9, v/v, pH 7.4). Time range: 0-600 s. λ_{ex} = 480 nm.

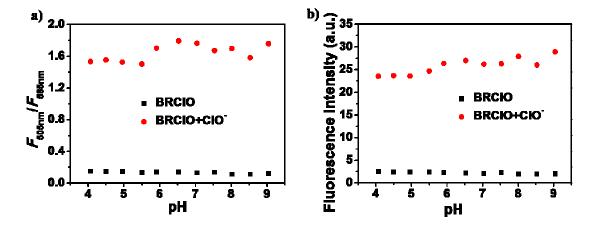


Figure S6 Effect of pH on fluorescence ratios ($F_{505 \text{ nm}}/F_{585 \text{ nm}}$) (a) and intensities at 505 nm (b) of **BRCIO** and **BORCIO** in PBS (0.01 M) solution (ethanol/water = 1/9, v/v, pH 7.4). Fluorescence intensity was measured at $\lambda_{\text{ex}} = 480 \text{ nm}$.

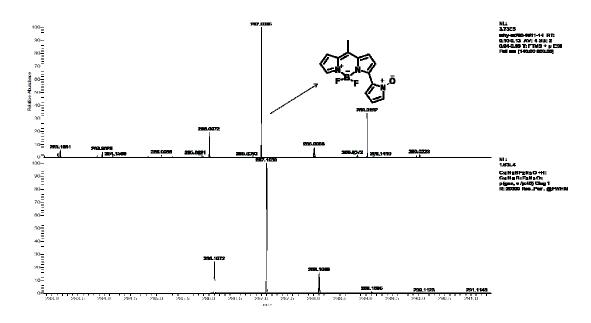


Figure S7 HRMS spectra of BORCIO.

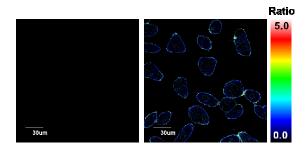


Figure S8 Confocal ratio images ($I_{490-520\text{nm}}/I_{560-600\text{nm}}$) of without (left) and with (right) exogenous NaClO (100 μ M) in MCF-7 cells using sensor **BRClO** (1 μ M) for 30 min at 37 °C. λ_{ex} = 488 nm. Scale bar = 30 μ m.

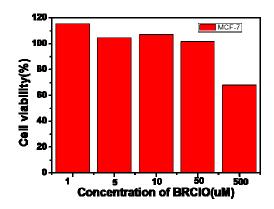


Figure S9 Cytotoxicity assays of BRClO at 1 $\,^{\circ}$ 5 $\,^{\circ}$ 10 $\,^{\circ}$ 50 and 500 μM against MCF-7 cells for 24

h.

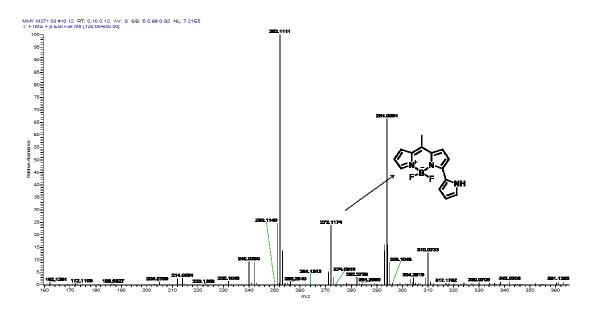


Figure S10 HRMS spectra of BRCIO.

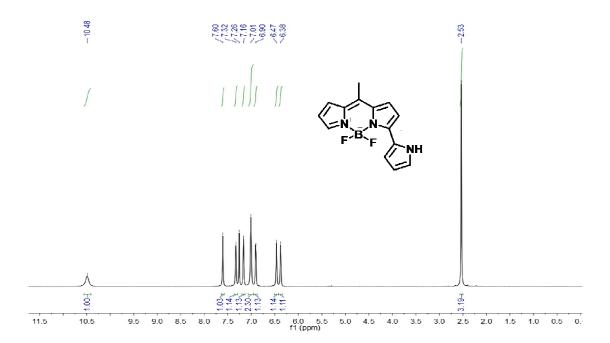


Figure S11 ¹HNMR of **BRCIO**.

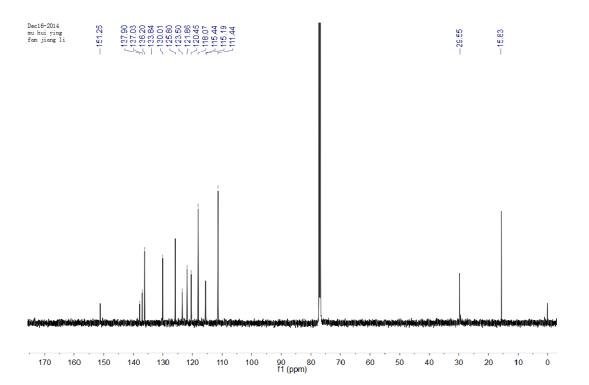


Figure S12 ¹³CNMR of BRCIO.

Reference

- (1) Firisch, M. J. e. a. Gaussian 09, Revision A.02 2009.
- (2) Zhou, L.-C.; Zhao, G.-J.; Liu, J.-F.; Han, K.-L.; Wu, Y.-K.; Peng, X.-J.; Sun, M.-T., The charge transfer mechanism and spectral properties of a near-infrared heptamethine cyanine dye in alcoholic and aprotic solvents. *J. Photochem. Photobiol. A* **2007**, *187*, 305-310.
- (3) Cao, J.; Wu, T.; Hu, C.; Liu, T.; Sun, W.; Fan, J.; Peng, X., The nature of the different environmental sensitivity of symmetrical and unsymmetrical cyanine dyes: an experimental and theoretical study. *Phys. Chem. Chem. Phys.* **2012**, *14*, 13702-13708.
- (4) Dreizler, M. R. G., E. K. U. Density Functional Theory 1990.
- (5) Karstens, T.; Kobs, K., Rhodamine B and rhodamine 101 as reference substances for fluorescence quantum yield measurements. *J. Phys. Chem.* **1980**, *84*, 1871-1872.
- (6) Casey, K. G.; Quitevis, E. L., Effect of solvent polarity on nonradiative processes in xanthene dyes: Rhodamine B in normal alcohols. *J. Phys. Chem.* **1988**, *92*, 6590-6594.