Supporting Information. Part 2.

Efficient Functionalization of Oligonucleotides by New Achiral Nonnucleosidic Monomers

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Content

Page

3	Notes
5	NMR spectra

NMR spectra.

Notes

It has been observed that in ¹H NMR spectra of compounds **4a-g** the expected signals in some cases were accompanied by additional signals of lesser intensity, or the expected signals had more complex structure (higher multiplicity). Careful analysis showed that in all cases the ratio between the expected and the additional signals was constant and equal to 5:3. In the cases of more complex structure (higher multiplicity) the additional signal overlapped with the expected signal. Integration of the spectra by combining the expected and the additional peaks produced values that correlate well with the expected structure of the compound.

¹³C NMR spectra demonstrated similar splitting of the signals; the same ratio of 5:3 was maintained. We ascribe such a splitting of the signals to the presence of two rotamers in the ratio of 5:3 due to restricted rotation around a rigid oxalamide fragment in the presence of bulky substituents such as dimethoxytrityl group. In phosphoramidites **5a-g** the effect is even more pronounced due to the presence of another bulky group *N*,*N*-diisopropylamino-2-cyanoethoxyphosphinyl.

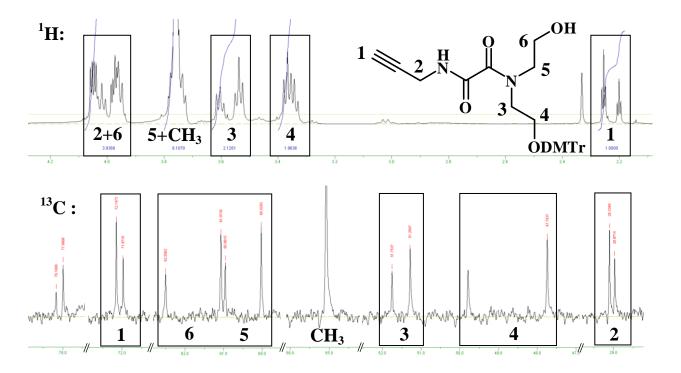


Fig. 1. Splitting of signals in ¹H (top) and ¹³C (bottom) NMR spectra of 4a.

An example of such a splitting is shown in Fig. 1. In the ¹H NMR spectrum of 4a (top) the peaks 1 and 3 double up; in the peak 4 there is overlap of two signals resulting in higher multiplicity. In the ¹³C spectrum (bottom) all the main signals are accompanied by additional minor peaks.

Analysis of the reaction mixtures of early oligonucleotide syntheses has revealed the presence of truncated sequences with the 5'-terminal ethyl phosphate group. The same byproducts were present in the reaction mixtures from different phosphoramidites such as **5b**, **5c** or **5g**. In ³¹P NMR spectra of phosphoramidites **5b**, **5c** and **5g** an additional peak at 140.0-146.6 ppm was observed that we have identified as *N*,*N*-diisopropyl-*O*-ethyl phosphoramidite. In some initial batches of the phosphoramidites the content of the byproduct was quite substantial and this could have resulted in the formation of oligonucleotide byproducts as ethyl phosphoramidite coelutes with the monomer on column. We have traced the origin of the admixture to the work-up of phosphitylation reaction with dichloromethane stabilized with 0.05% ethanol. After we have uncovered the fact, we have introduced quenching of phosphitylation reaction with small volume of 0.3 M aq KH₂PO₄ followed by extraction with dichloromethane. After quenching the amount of ethyl phosphoramidite in ³¹P spectra (e.g. **5a**, **5d** and **5e**) was significantly reduced, which has resulted in elimination of oligonucleotide byproducts with ethyl phosphate group.

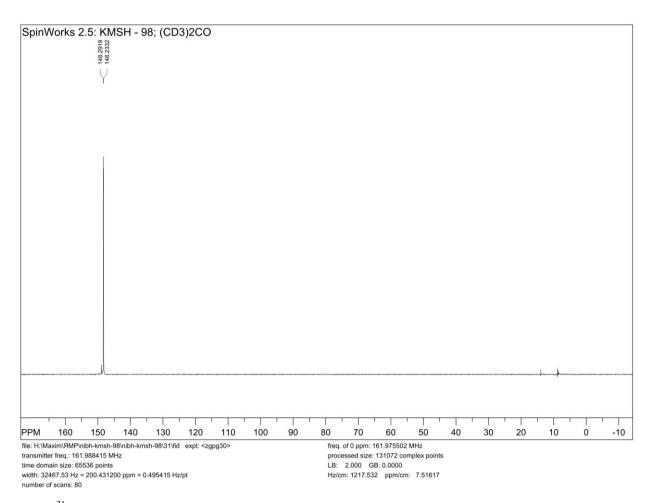
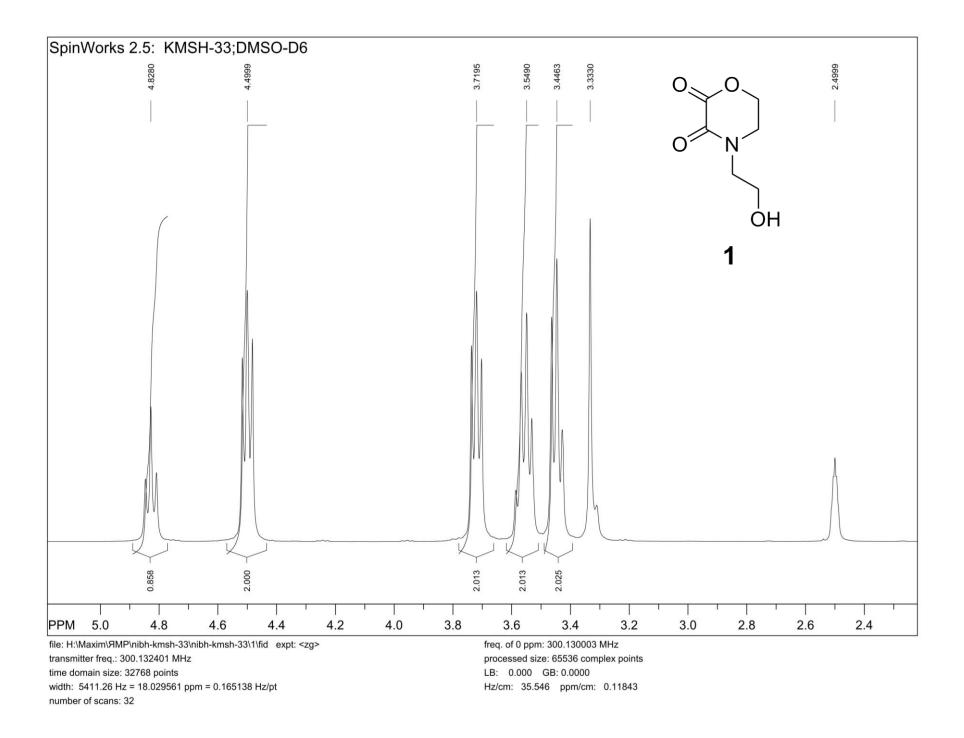
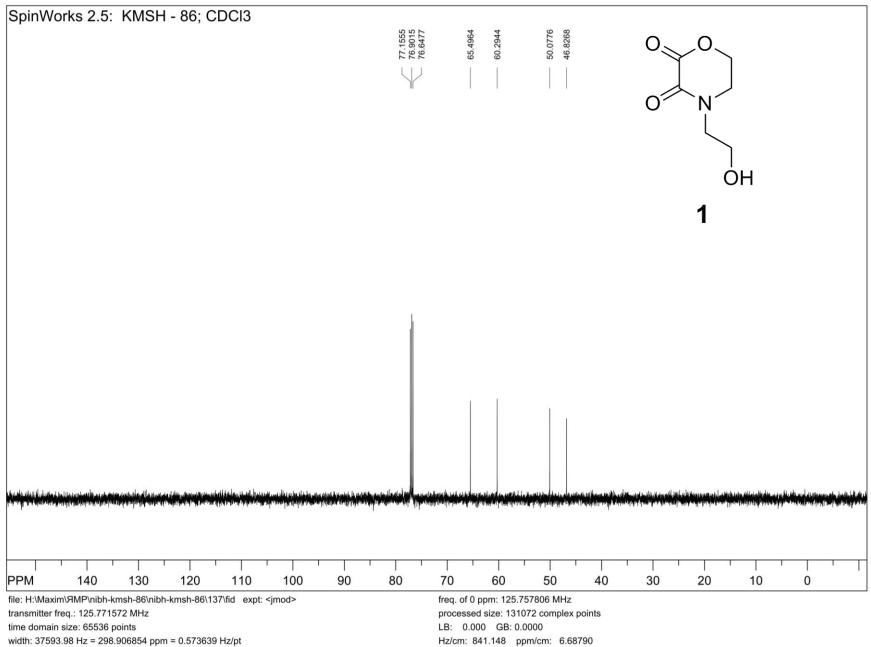
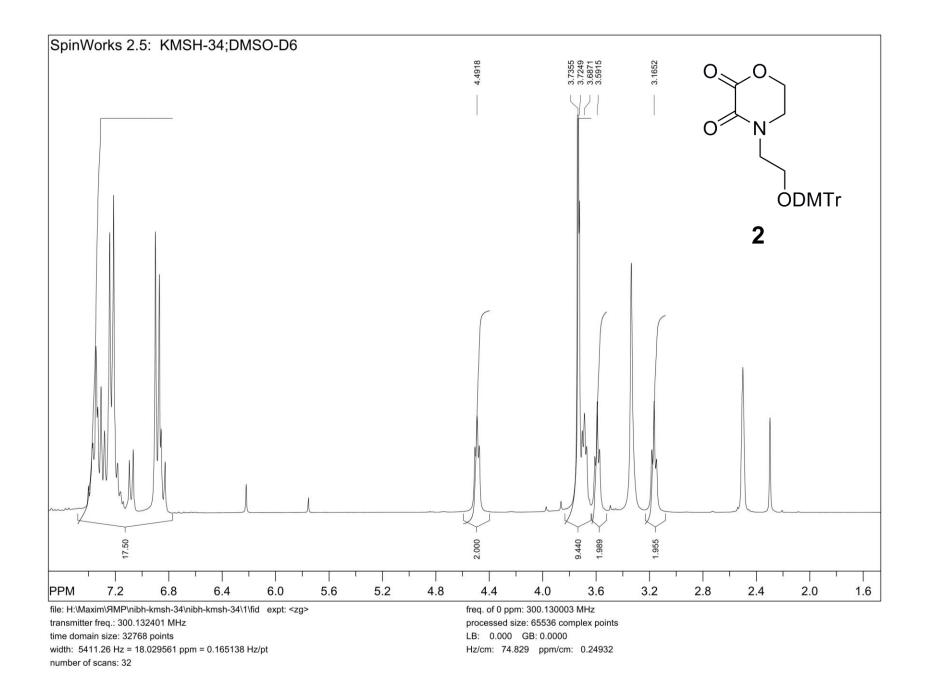


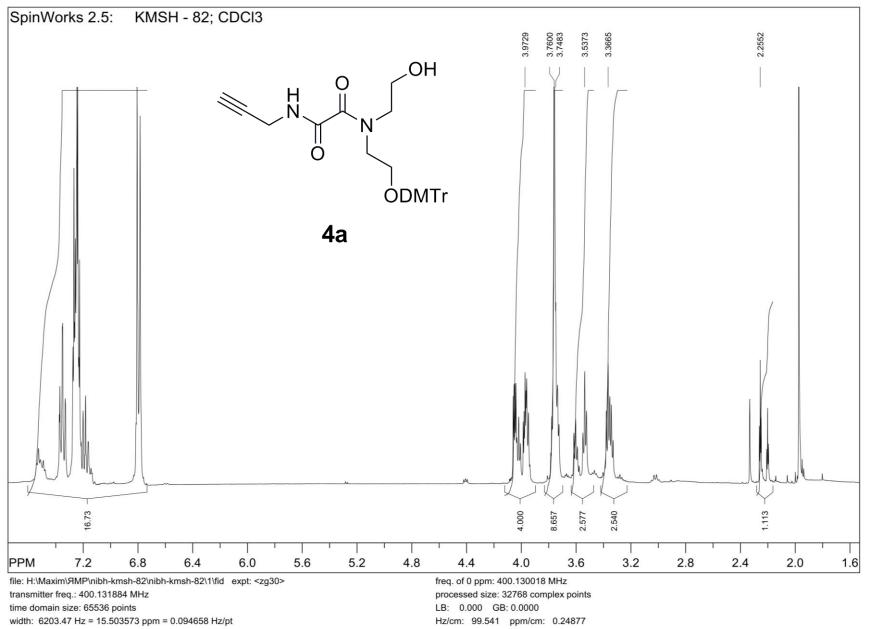
Fig. 2. ³¹P NMR spectrum of phosphoramidite **5b** purified by gel filtration on Sephadex LH-20.

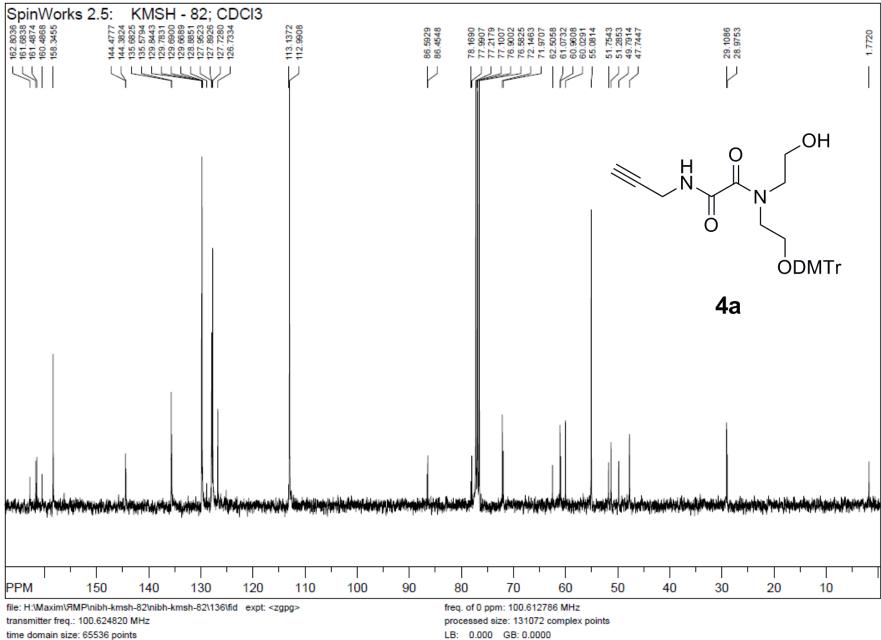
Analytically pure samples of phosphoramidites **5** could be obtained after gel filtration on a Sephadex LH-20 column eluted with dichloromethane (Fig. 2).





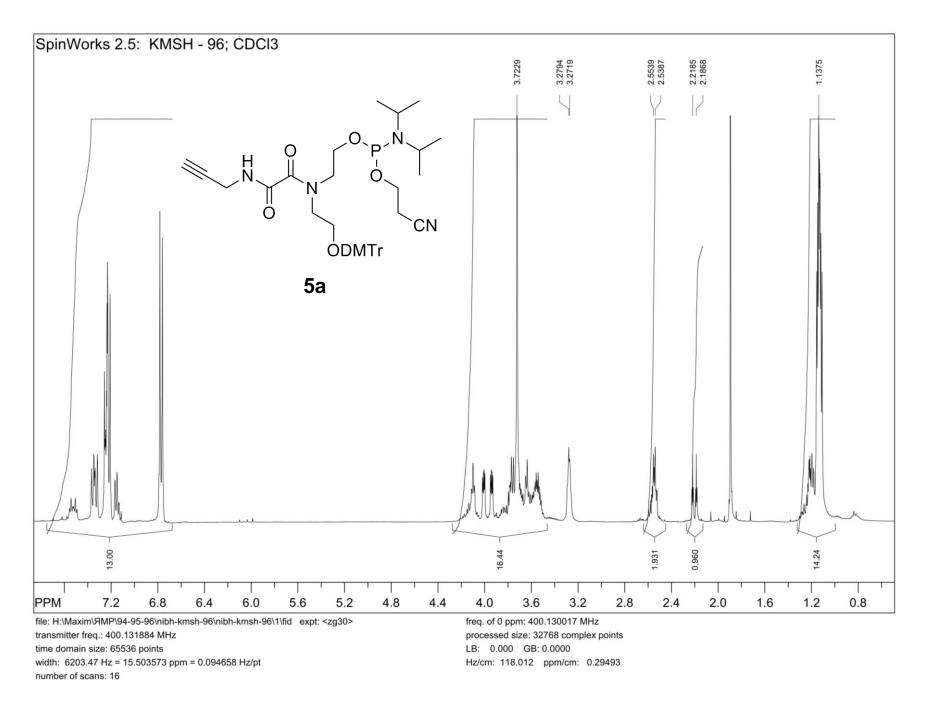


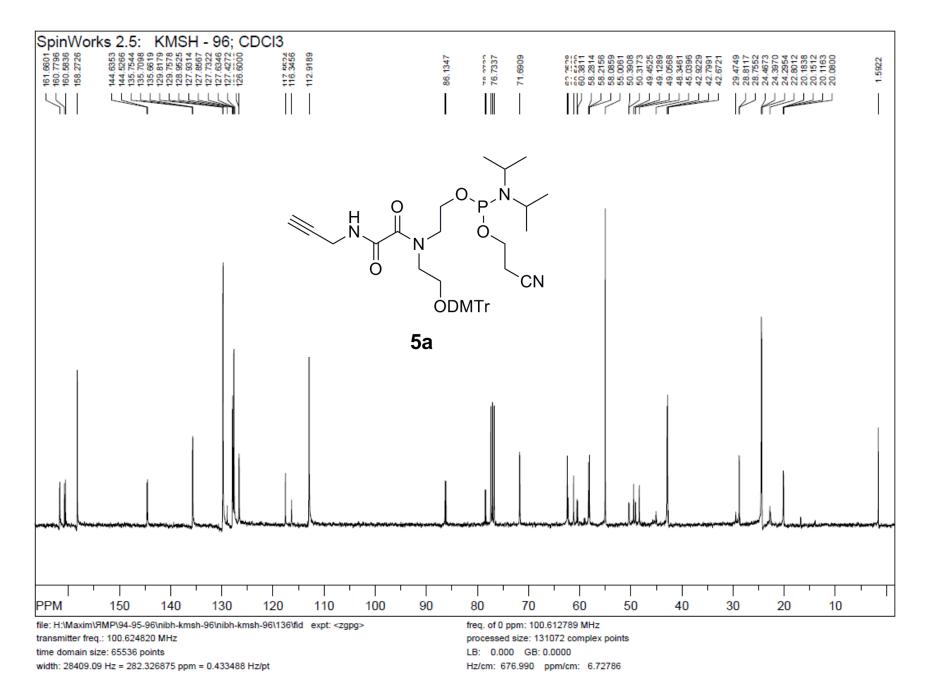


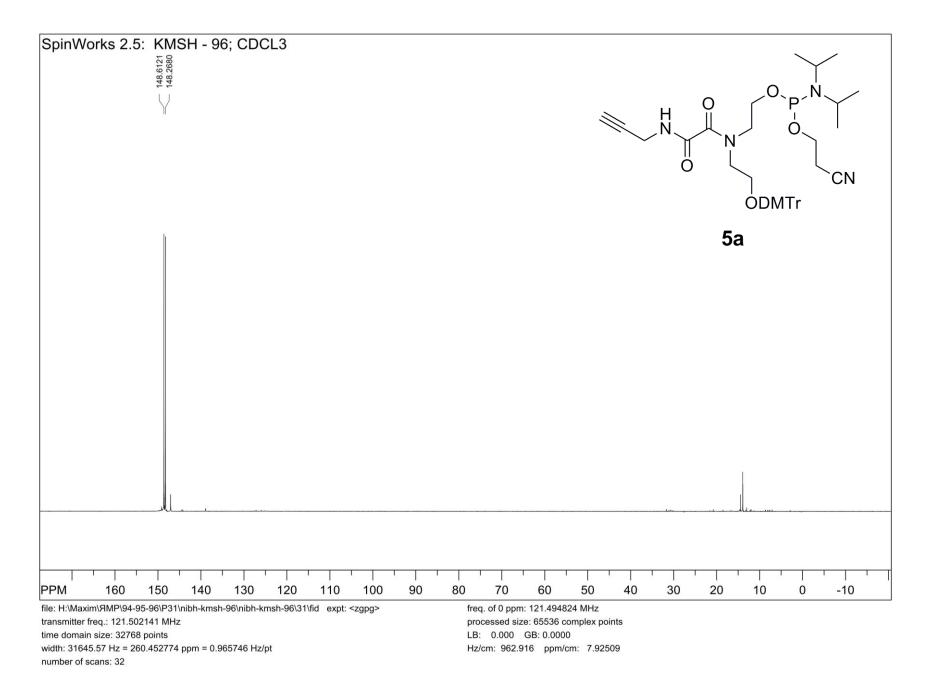


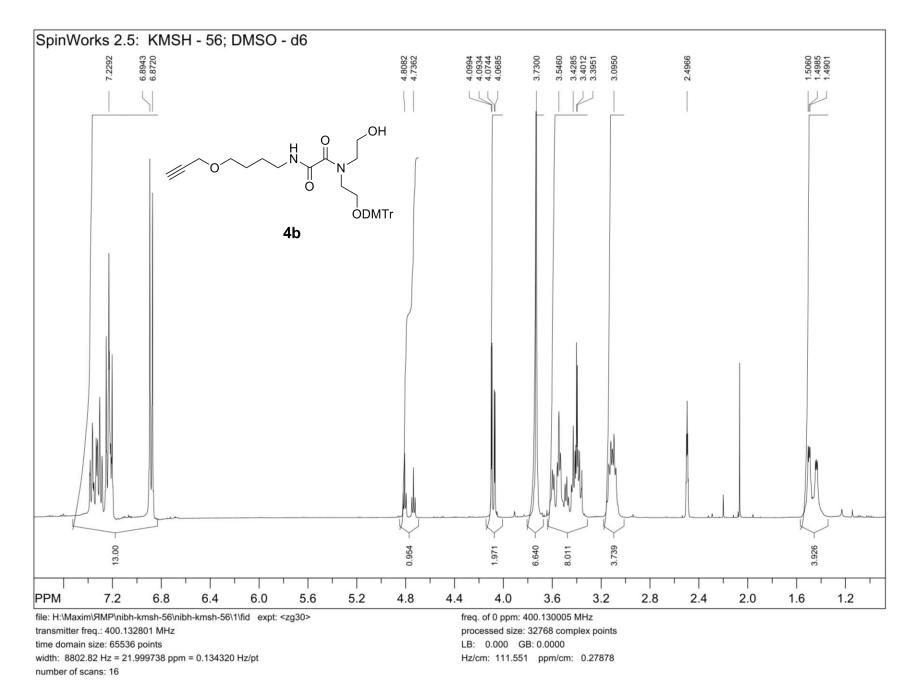
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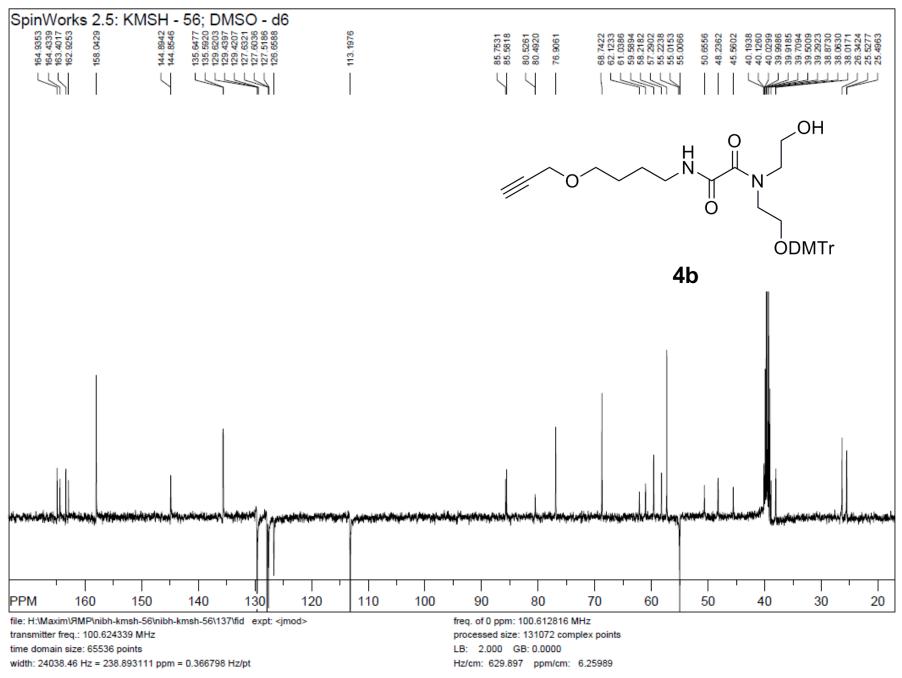
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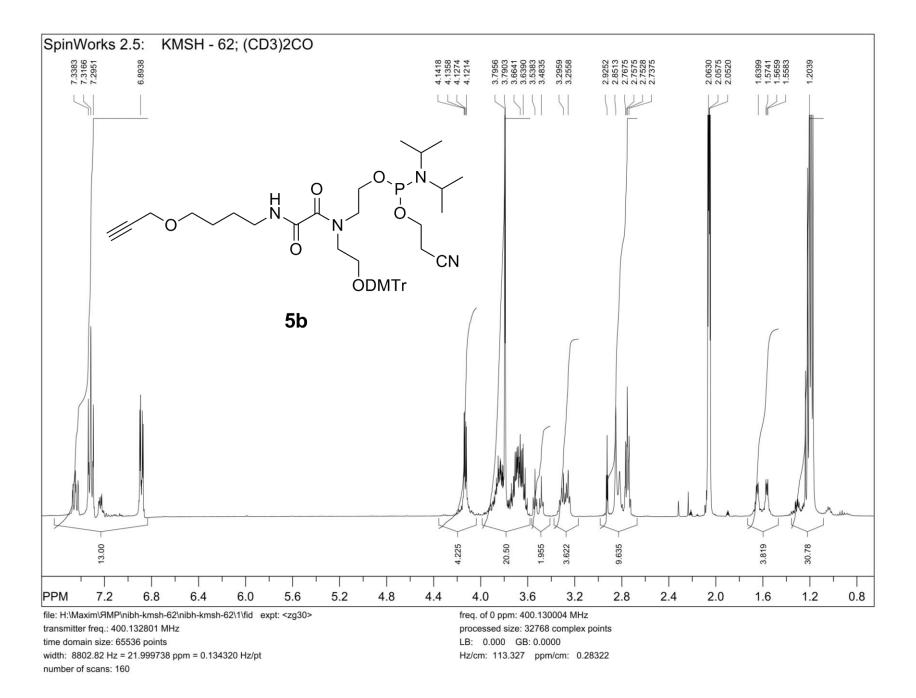


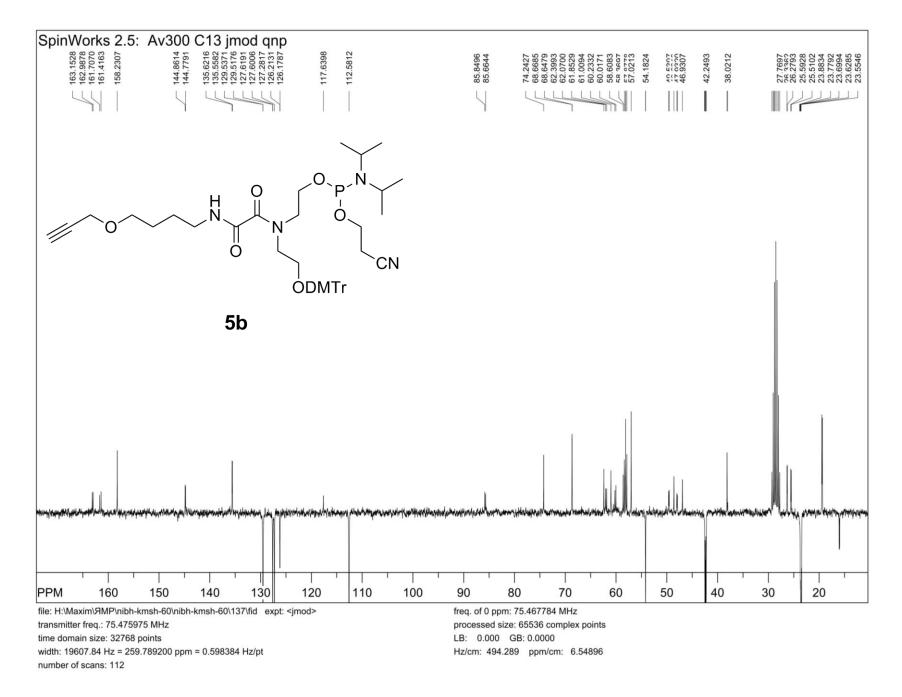


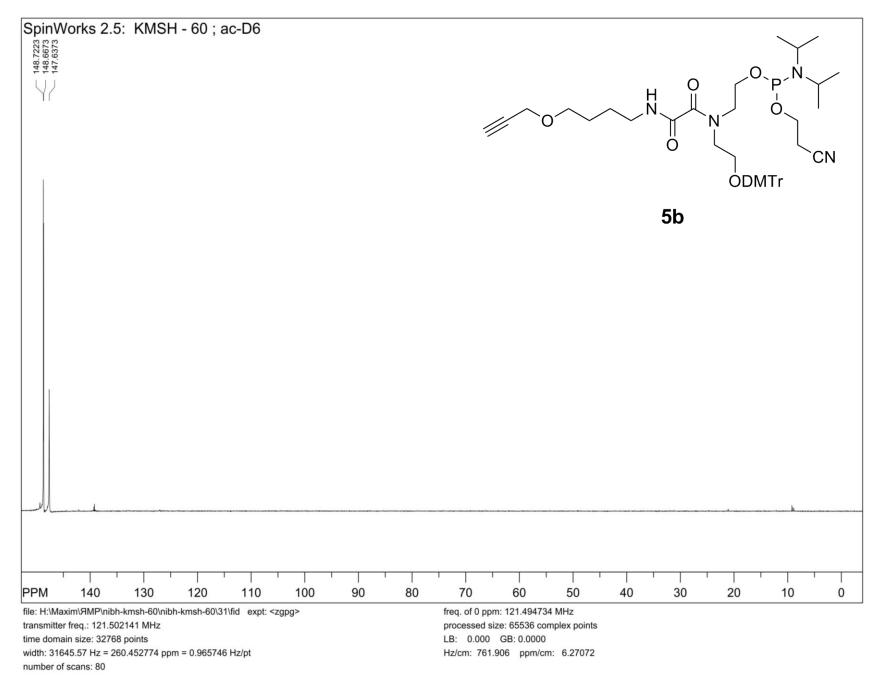


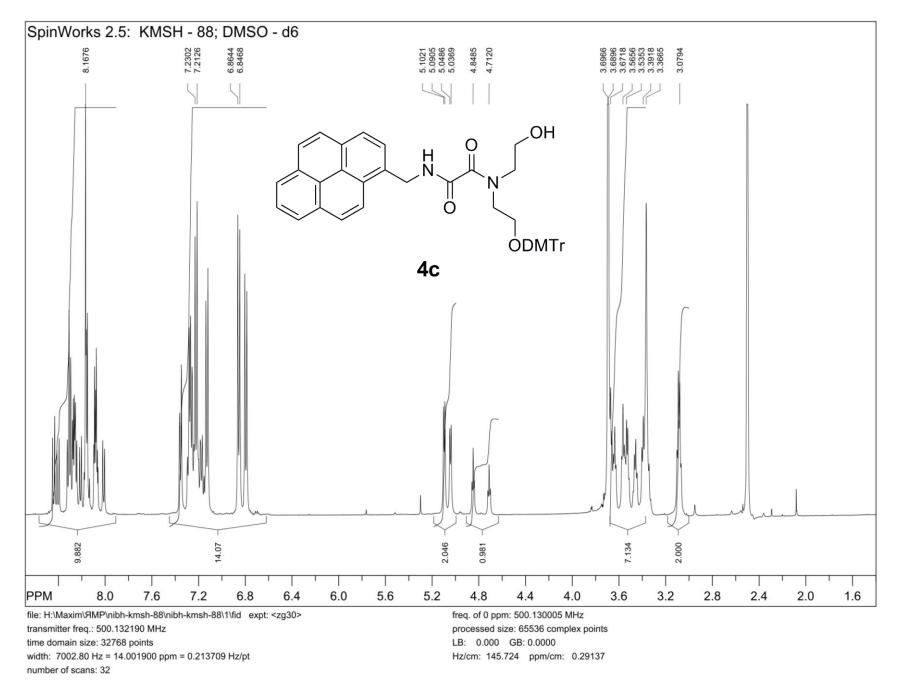


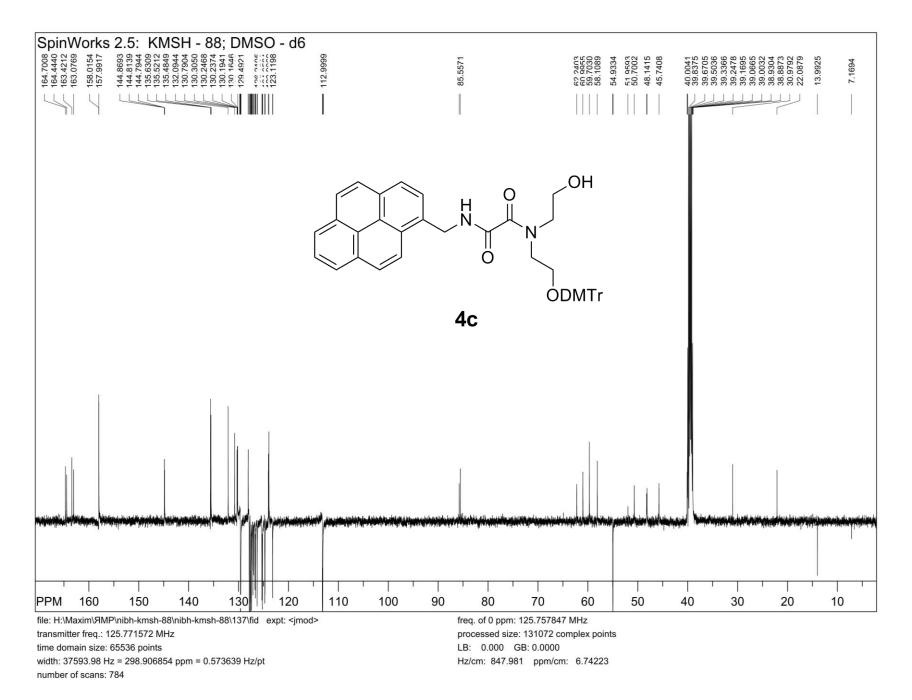


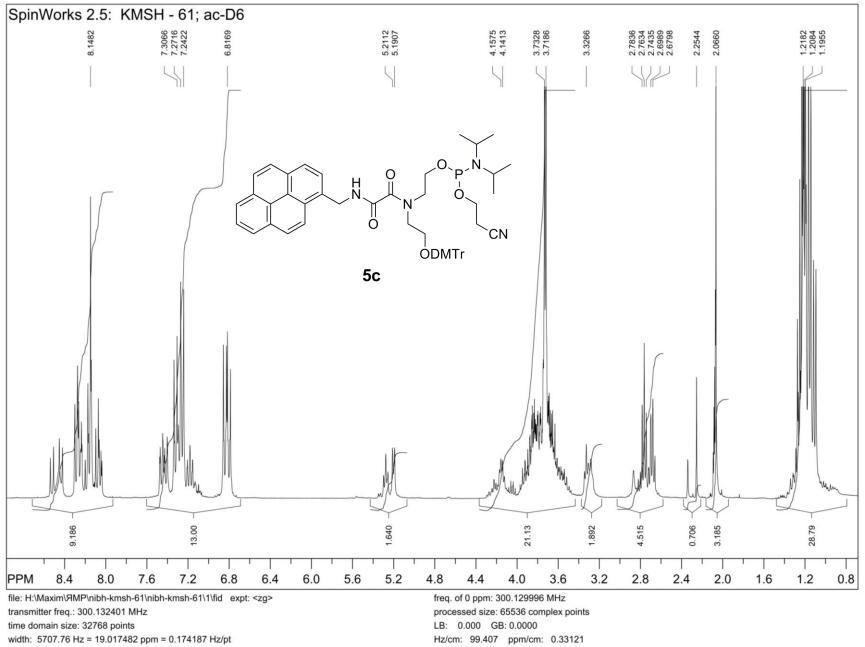


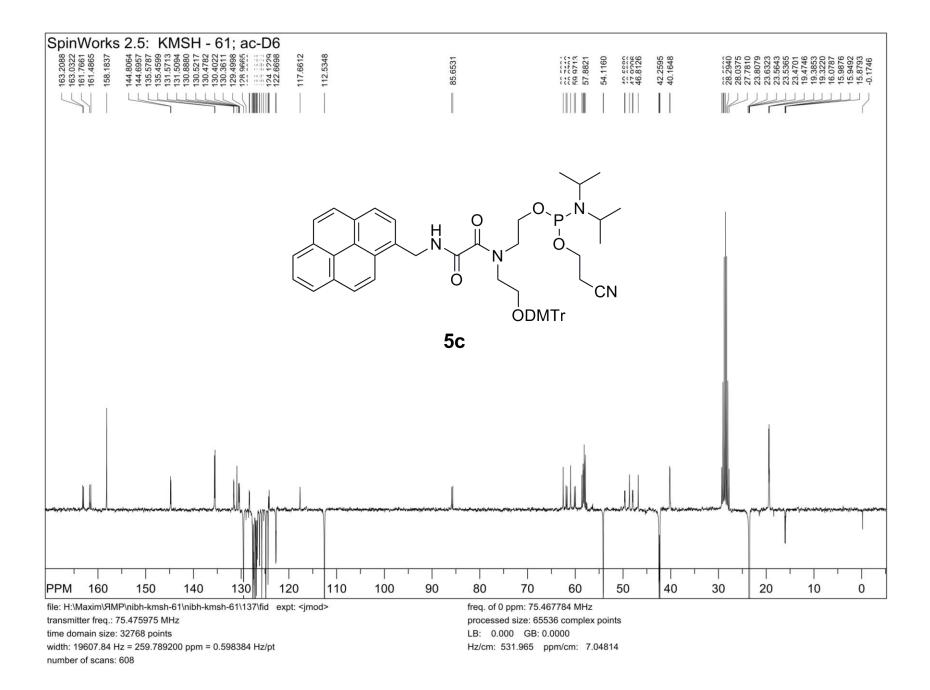


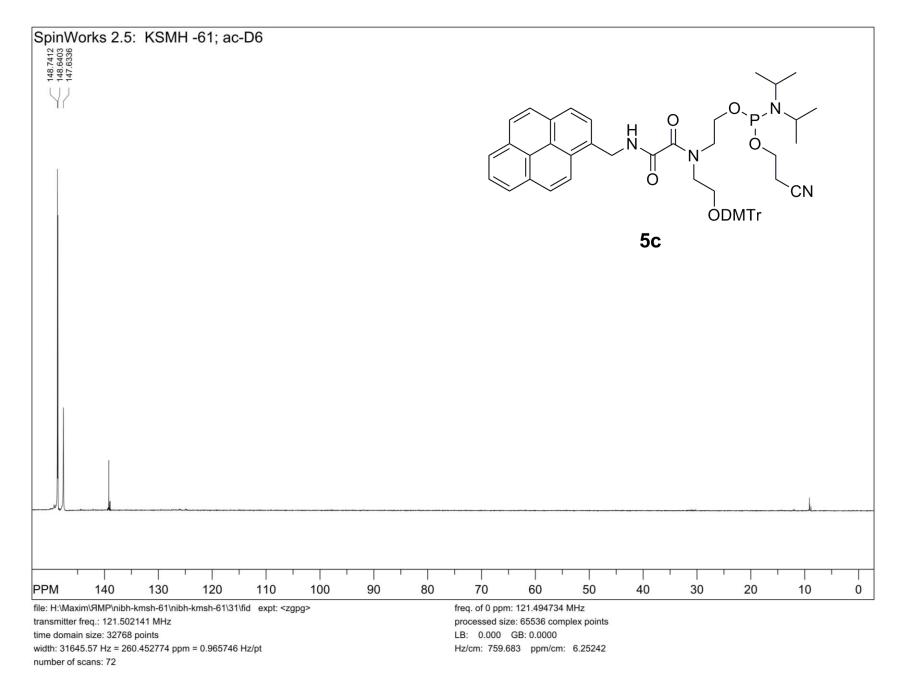


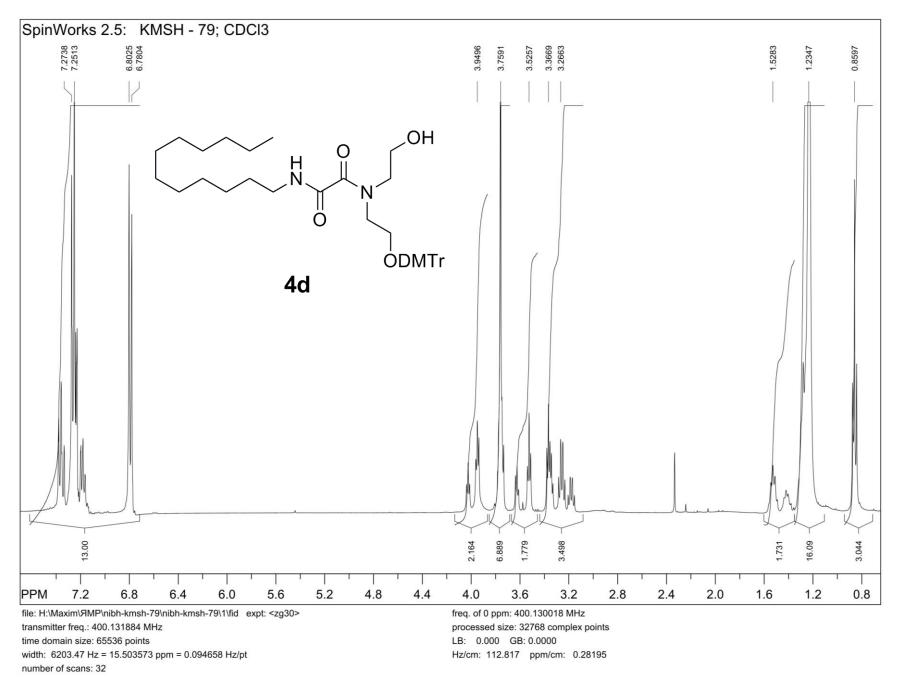


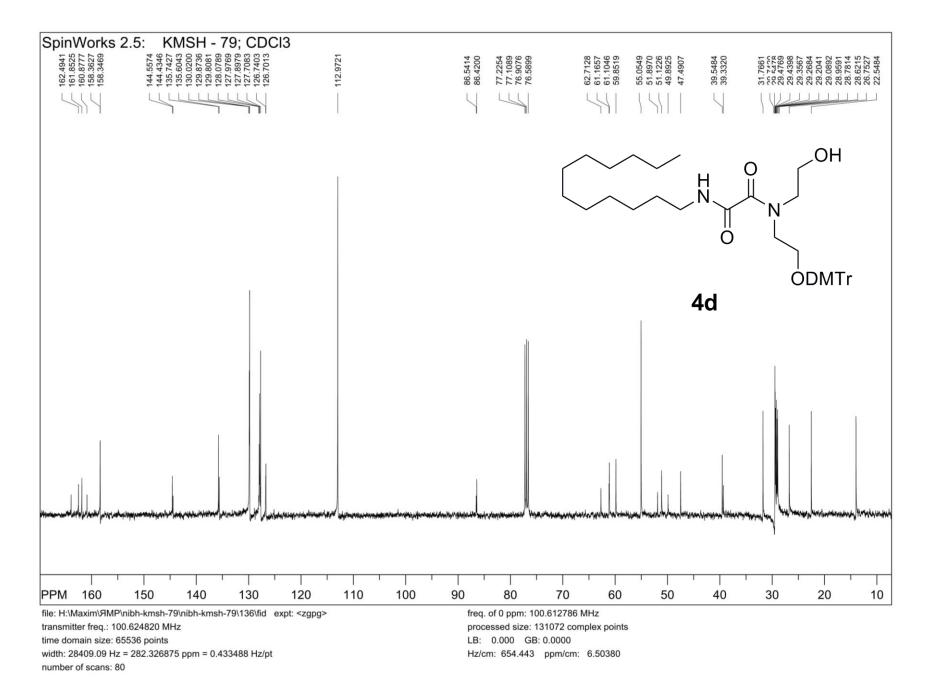


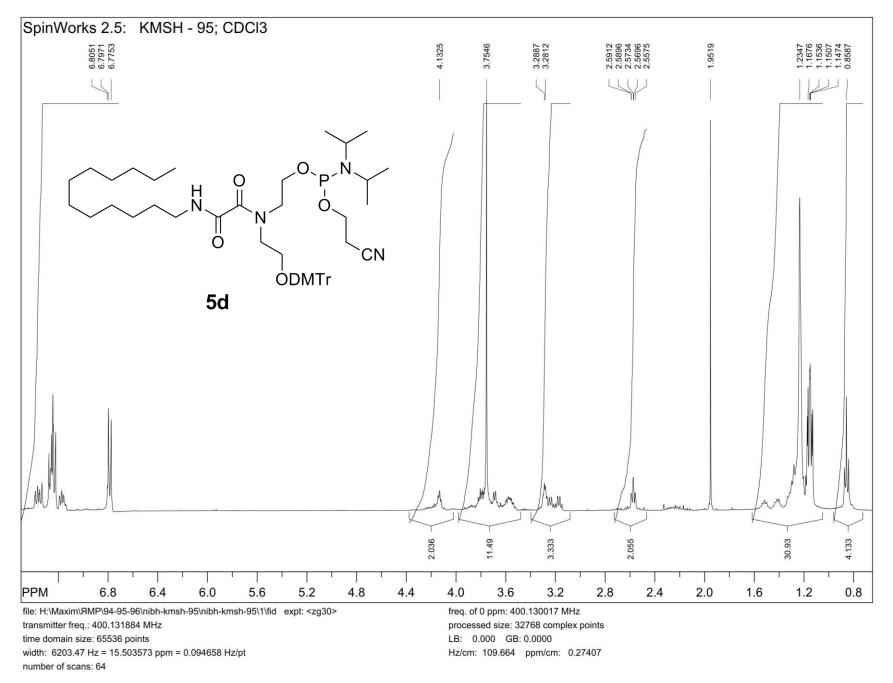


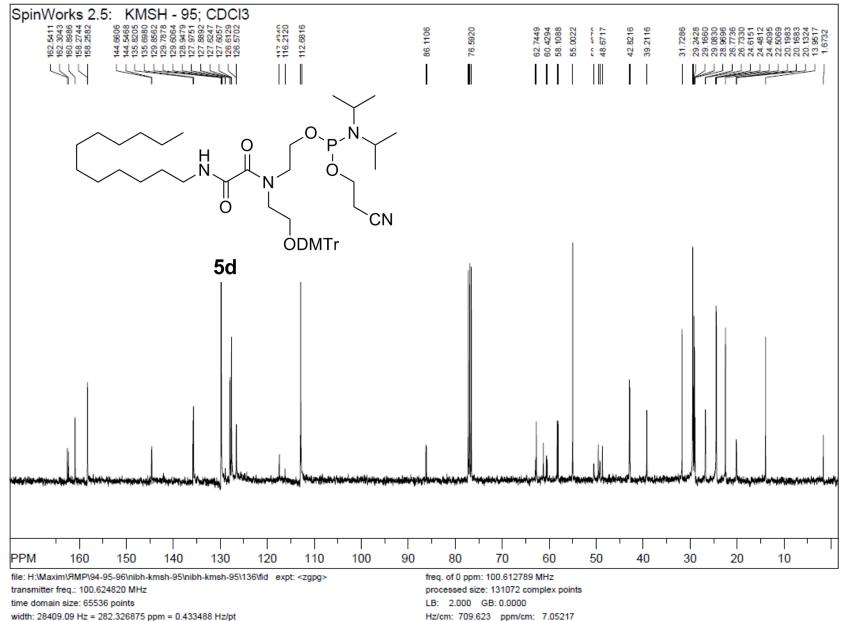




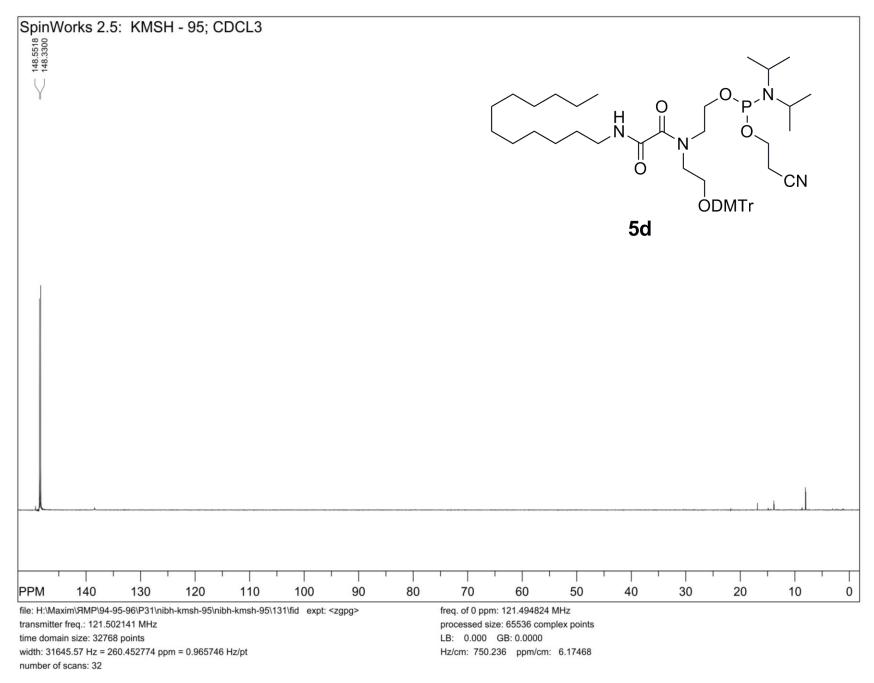


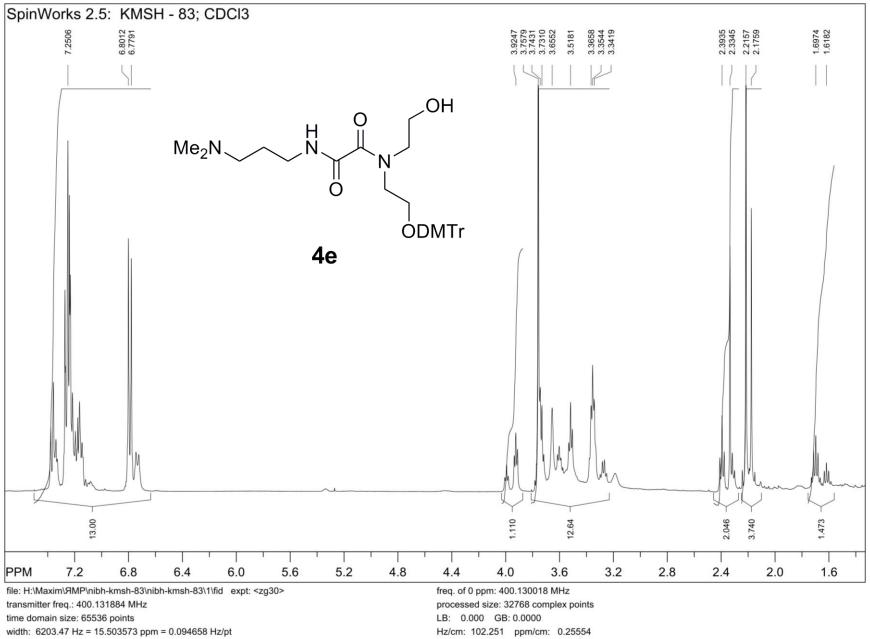




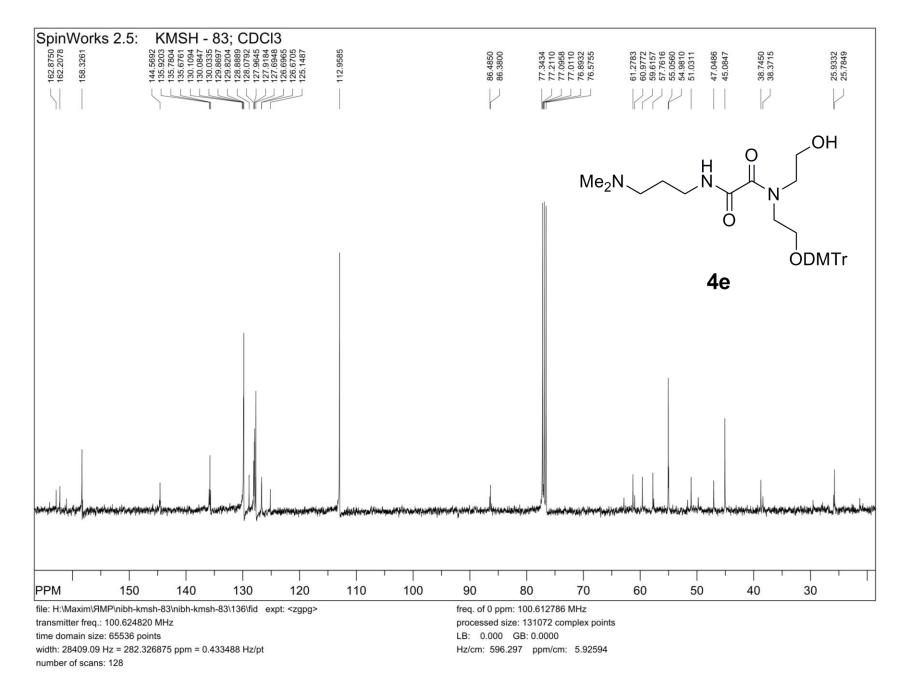


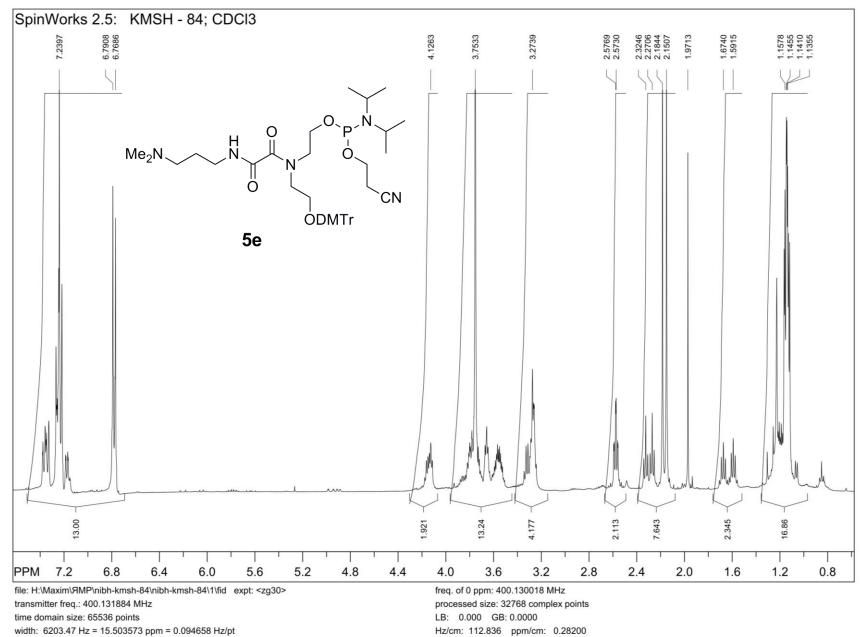
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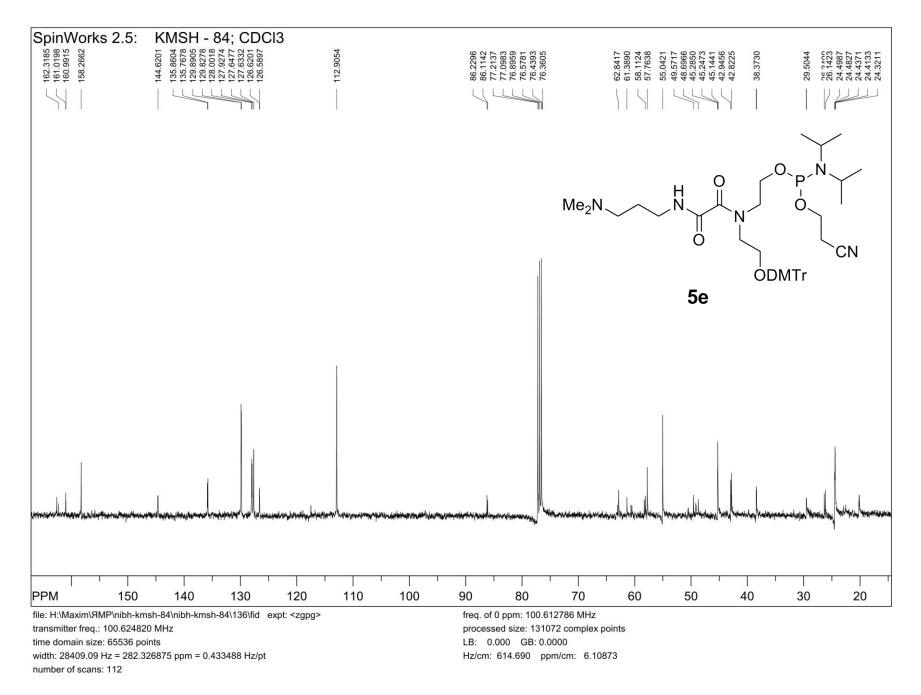


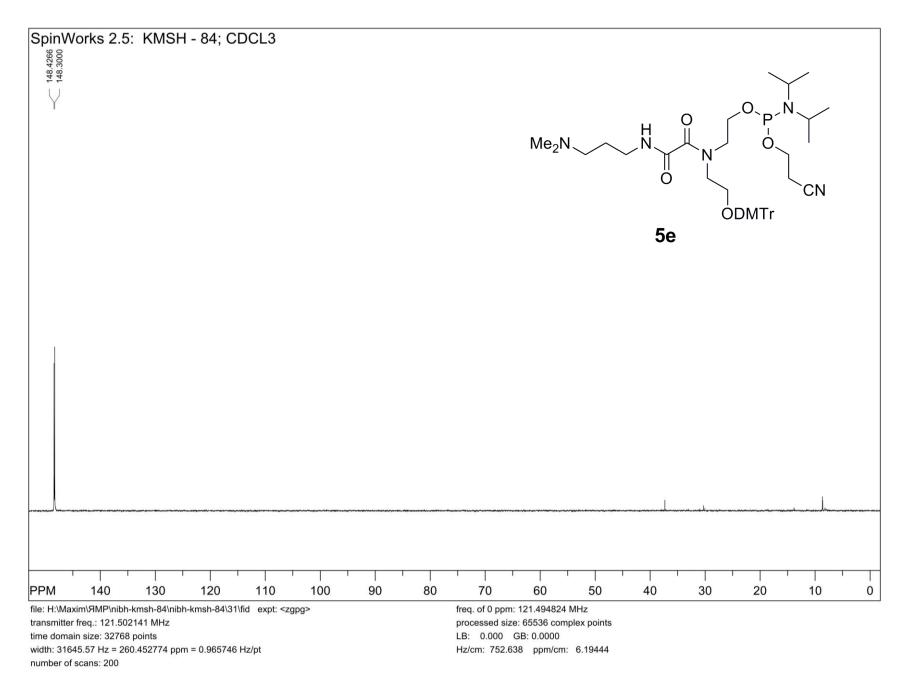


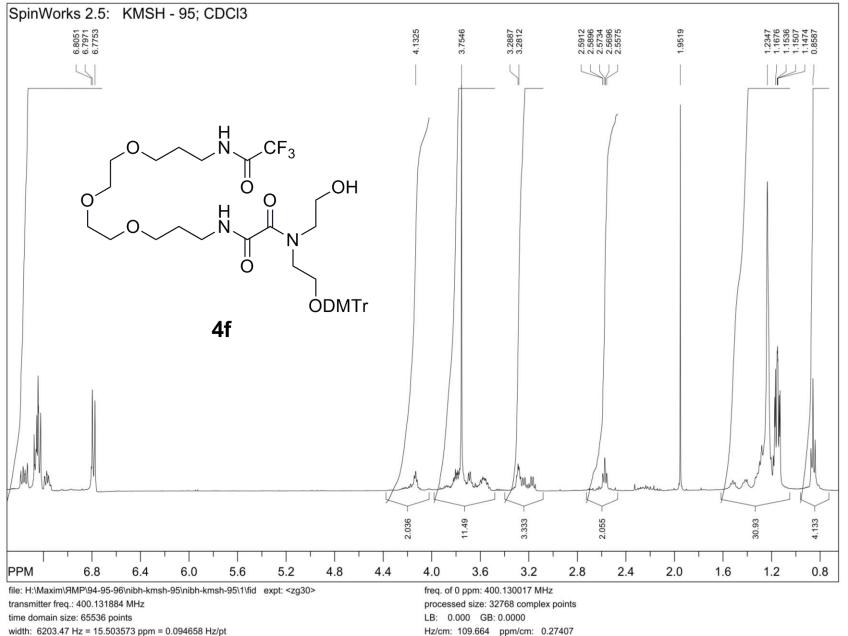




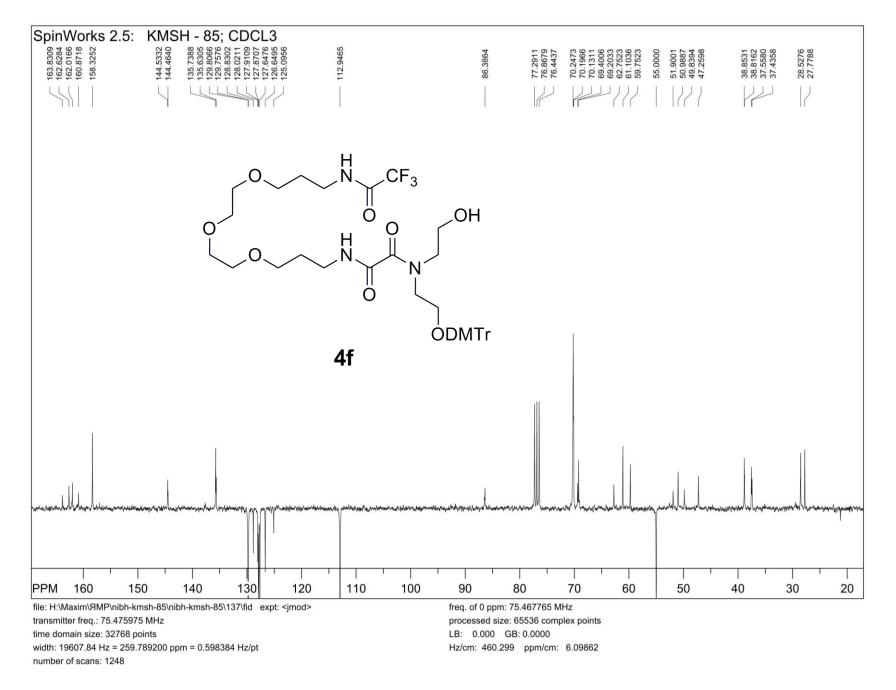


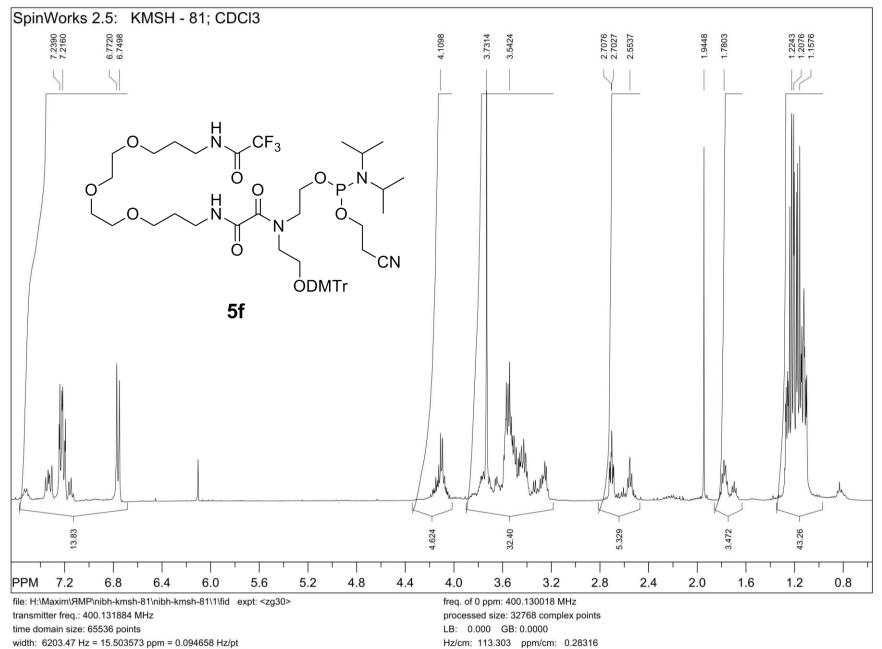


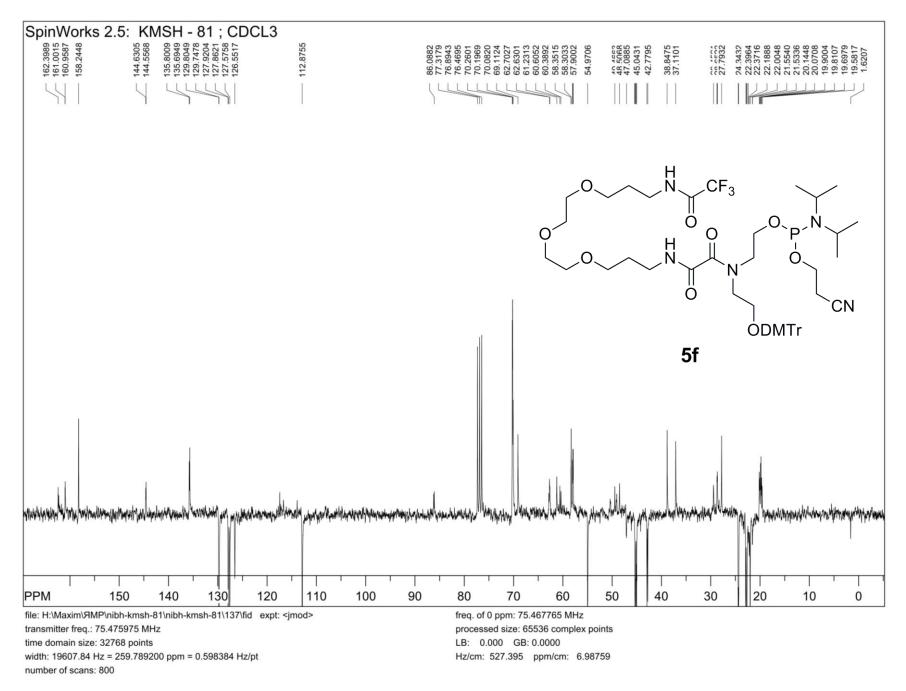


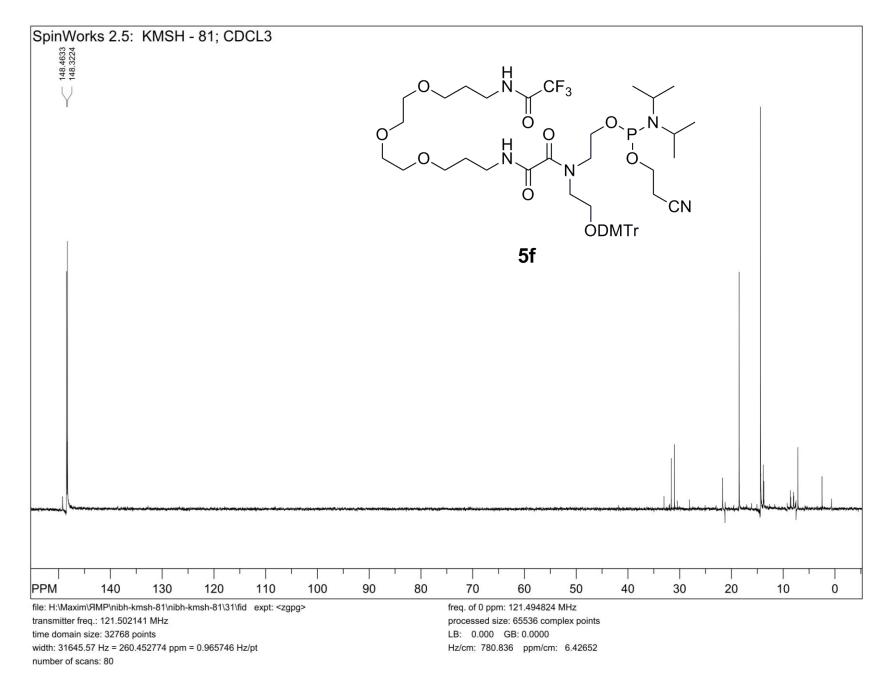


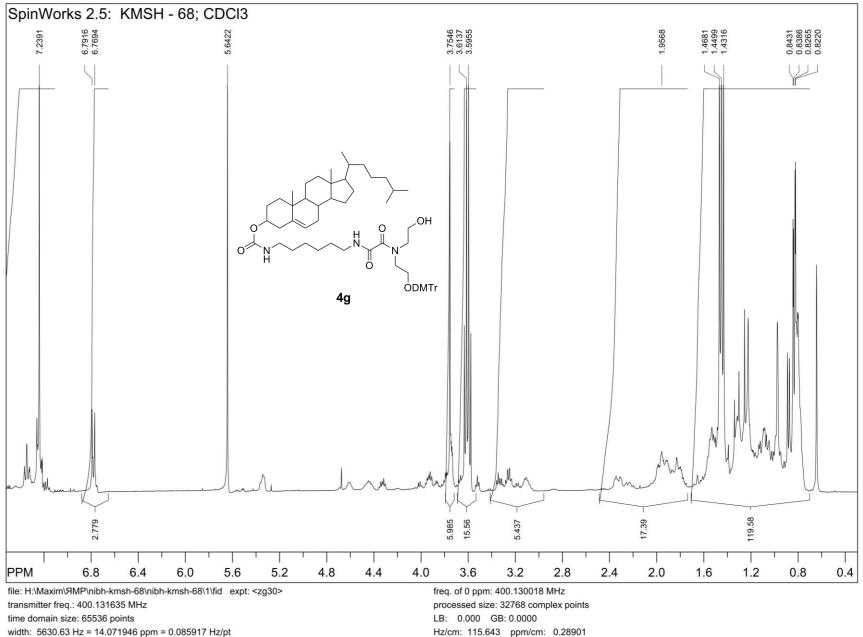
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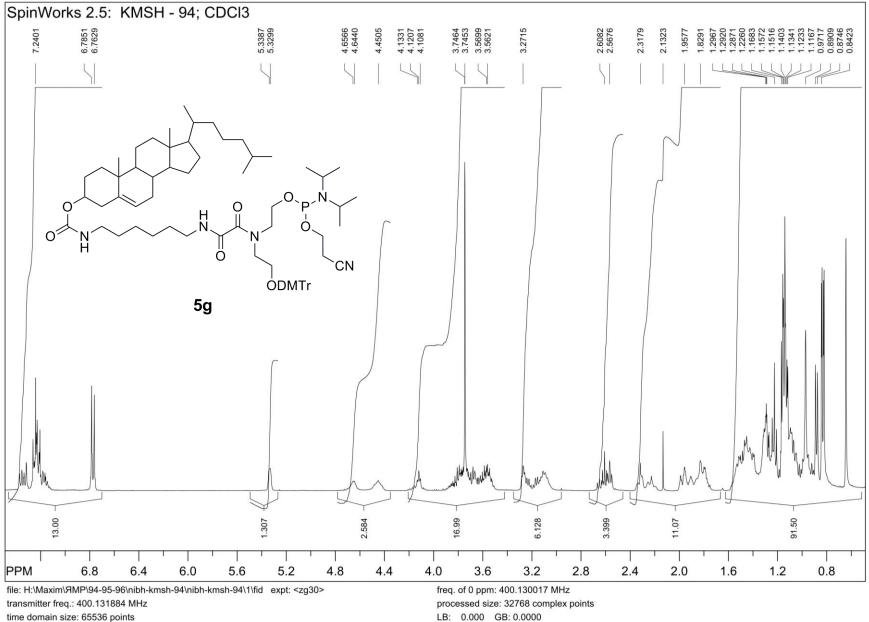








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94658 Hz/pt

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