## **Supplementary Information**

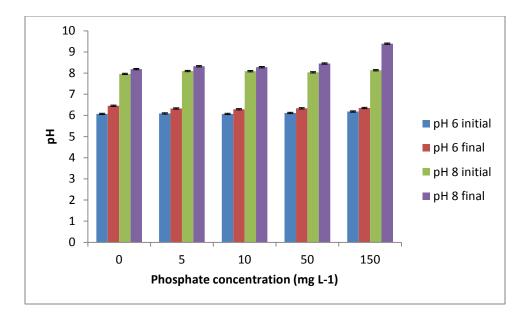
## A multi-technique investigation of the pH dependence of phosphate

## induced transformations of ZnO nanoparticles

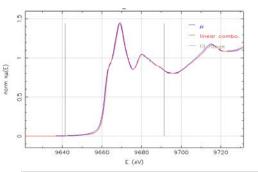
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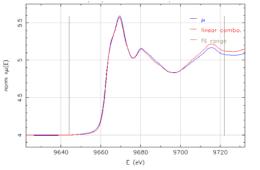


**Figure S1**. pH values at the beginning and end of aging as a function of initial pH and phosphate concentration. Results were similar regardless of aging duration so the different time points and replicates were averaged for each pH value and phosphate concentration.

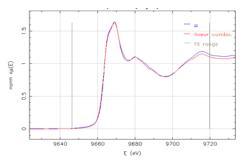


A – 90% ZnO, 10% Zn-phosphate

R	0.000304
Chi square	0.05186
Reduced chi square	0.0002284
ZnO	91.2
Zn-phosphate	8.8



R	0.000473
Chi square	0.09710
Reduced chi square	0.0004296
ZnO	73.9
Zn-phosphate	26.1

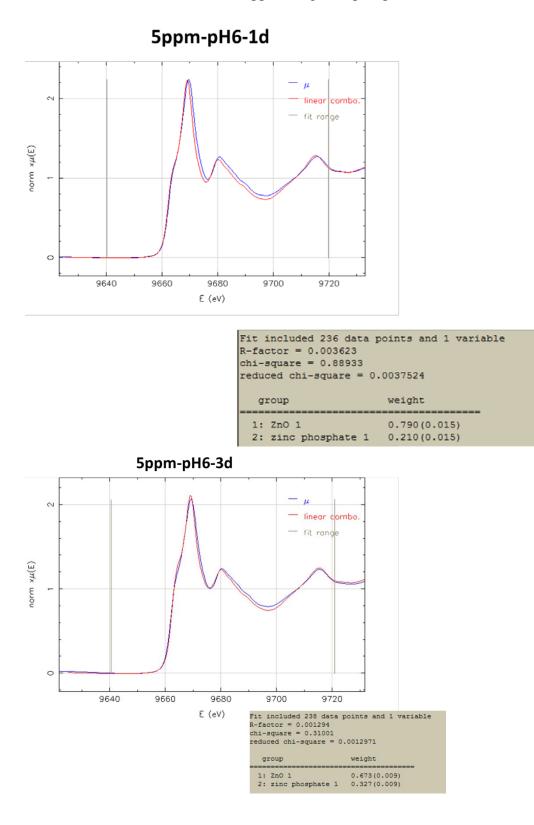


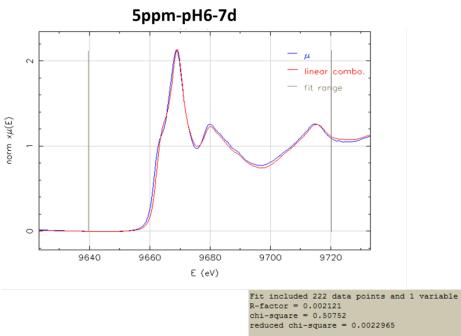
C – 50% ZnO, 50% Zn-phosphate

R	0.000320
Chi square	0.06283
Reduced chi square	0.0002869
ZnO	50.8
Zn-phosphate	49.2

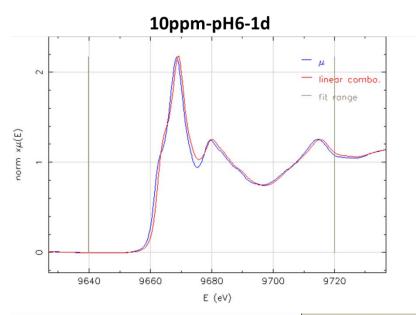
**Figure S2**. Linear combination fits for X-ray absorption near edge spectra (XANES) of standards containing known proportions of ZnO and  $Zn_3(PO_4)_2$ .

Figure S3. Example linear combination fits for X-ray absorption near edge spectra (XANES) aged ZnO manufactured nanomaterials. The treatment conditions are given above each panel (ppm = mg/L of phosphate).

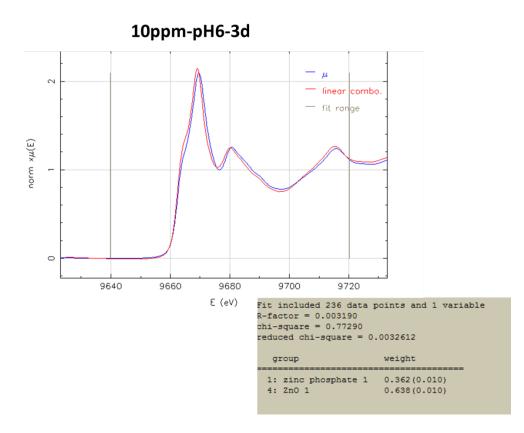


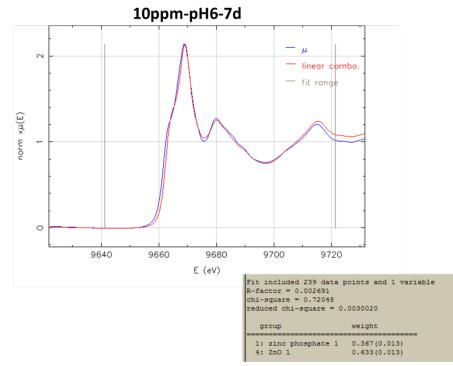


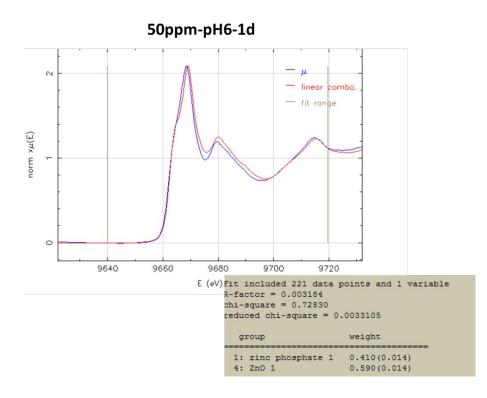
gı	roup		weight
2:	zinc phosphate	1	0.300(0.012)
1:	ZnO 1		0.700(0.012)



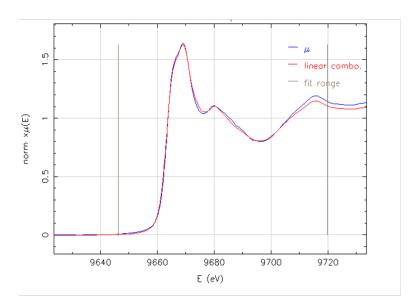
Fit included 221 data p R-factor = 0.004497 chi-square = 1.05227 reduced chi-square = 0.	
group	weight
1: ZnO 1 2: zinc phosphate 1	0.672(0.016) 0.328(0.016)



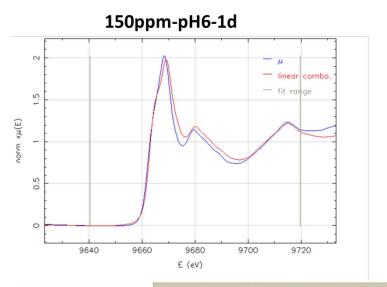




50ppm-pH6-7d

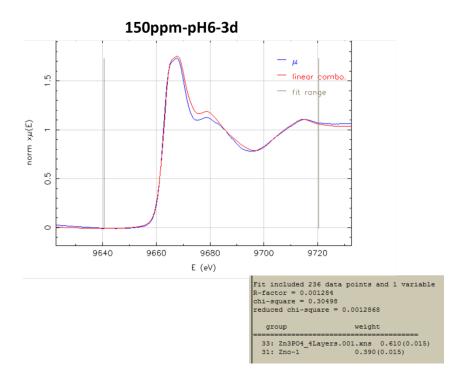


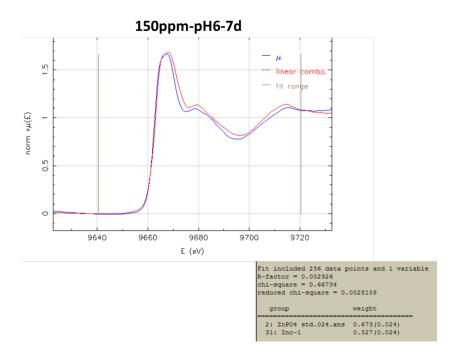
R	0.000320
Chi square	0.06283
Reduced chi square	0.0002869
ZnO	50.8
Zn-phosphate	49.2



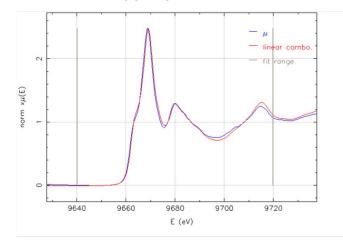
Fit included 220 data points and 1 variable R-factor = 0.005601 chi-square = 1.23505 reduced chi-square = 0.0056138

group			weight
	ZnPO4 ZnO 1	std.024.xns	0.600(0.019) 0.400(0.019)



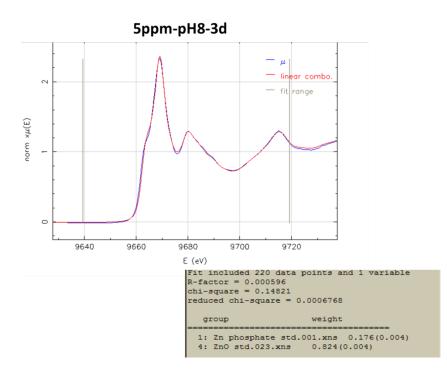


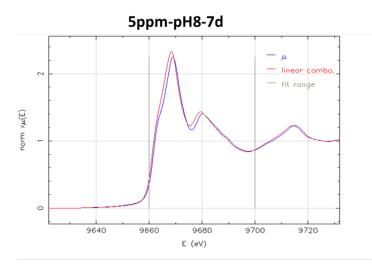
5ppm-pH8-1d



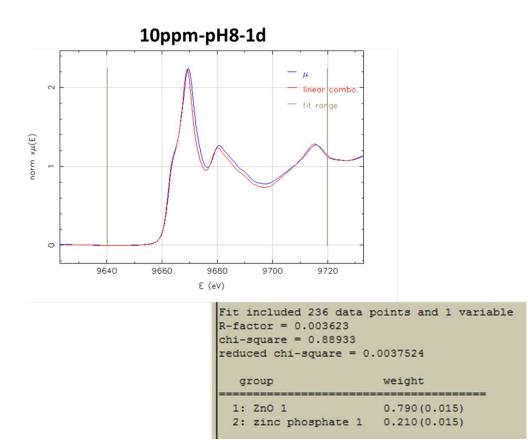
Fit included 220 data points and 1 variable R-factor = 0.001816 chi-square = 0.44799 reduced chi-square = 0.0020363

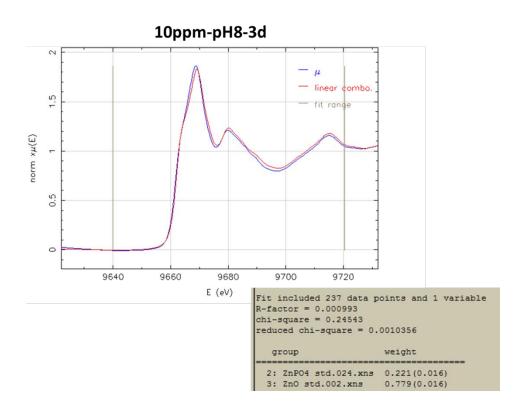
g1	roup	weight	
1:	Zn phosphate st	d.001.xns	0.095(0.008)
3:	ZnO std.023.xns	0.905	(0.008)

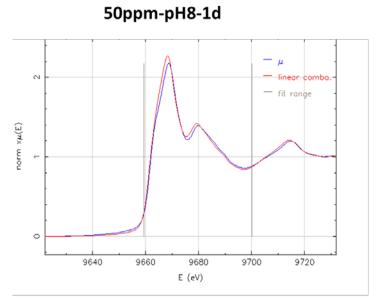




Chi square	1.0414
Reduced schi square	0.0078301
ZnO	82.5
Zn3(PO4)2	17.5

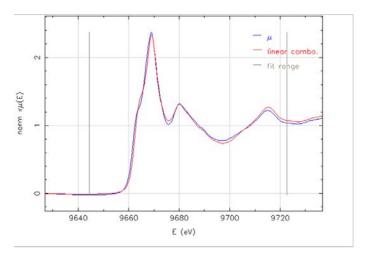






Chi square	0.46576
Reduced schi square	0.0034246
ZnO	73.1
Zn3(PO4)2	26.9





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Fit included 226 data points and 1 variable

R-factor = 0.001964

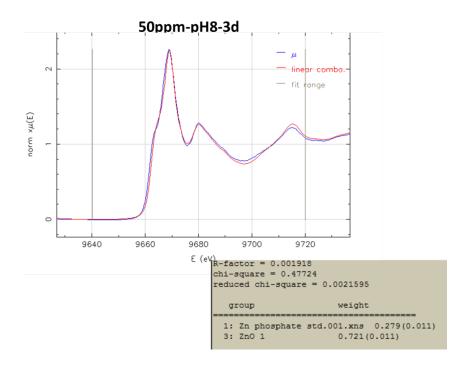
chi-square = 0.52810

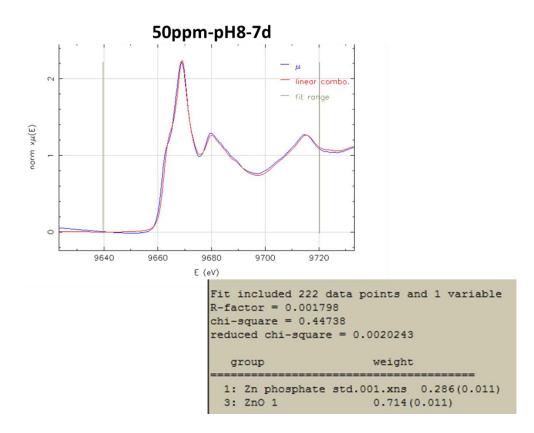
reduced chi-square = 0.0023471

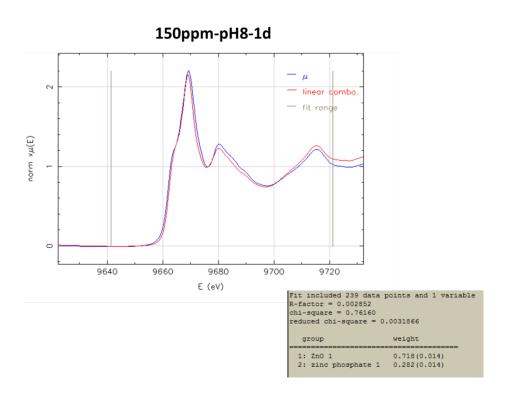
group weight

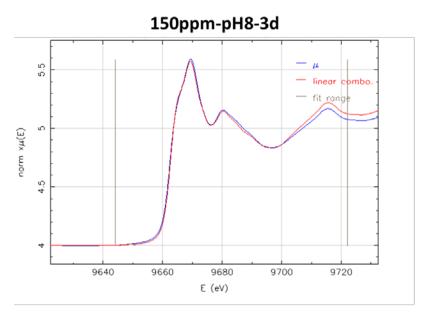
1: Zn phosphate std.001.xns 0.249(0.008)

4: ZnO std.023.xns 0.751(0.008)
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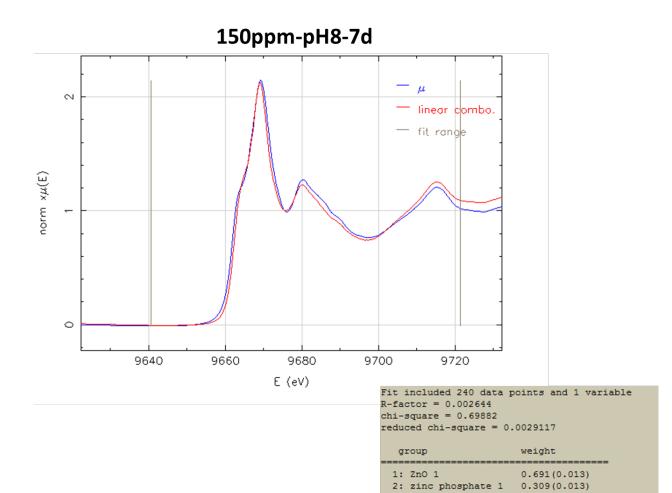








R	0.000473
Chi square	0.09710
Reduced chi square	0.0004296
ZnO	73.9
Zn-phosphate	26.1



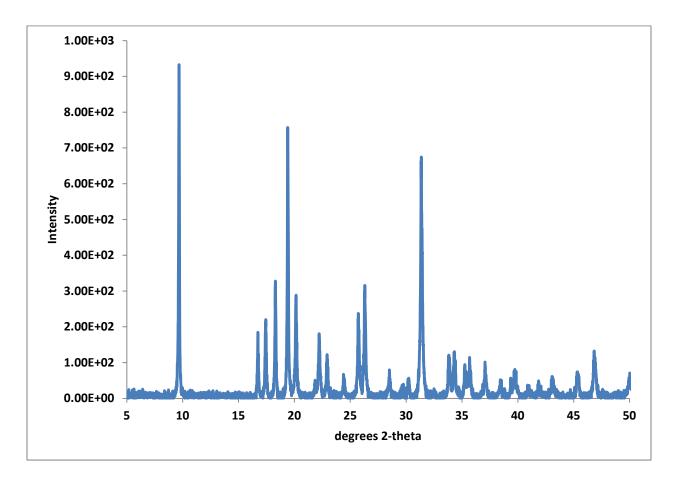
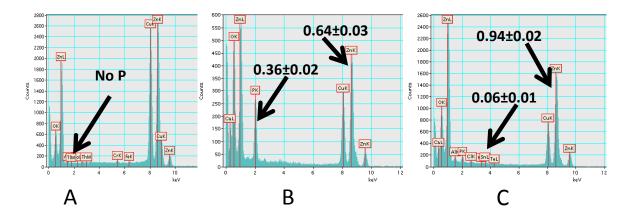


Figure S4. X-ray diffraction pattern for hopeite reported on the RUFF database (<u>http://rruff.info/</u>, RUFF ID R050254).



**Figure S5** X-ray energy dispersive spectra for pristine material (A) micron sized fraction of aged material (B) and nano sized fraction of aged material (C). The ratios of P and Zn are indicated on the figure.

## Additional interpretation of NMR spectra.

The NMR spectra provide additional detail regarding particles aged at pH 8, showing that one reason for the low hopeite formation is that a greater number of other species were formed in competition. Whereas the species with the chemical shift of 8.0 ppm (**Fig 6**) was the only byproduct at pH 6, only 47% of the pH 8 material being accounted for by the analogous species (7.8 ppm) with 27% of the pH 8 aged material was accounted for by a third component (5.3 ppm) not evident in particles aged at pH 6. The signals we observe near 4.2 ppm bear qualitative similarlities with that of  $\alpha$ -Zn<sub>3</sub>(PO<sub>4</sub>)<sub>2</sub> with respect to both isotropic chemical shift (3.9 ppm) and individual principal values (See **Table S1**). Our T<sub>1</sub>s were all considerably shorter than those reported by Roming et al. <sup>36</sup> for  $\alpha$ -Zn<sub>3</sub>(PO<sub>4</sub>)<sub>2</sub>, likely because our materials include water molecules which place <sup>1</sup>H near the phosphate <sup>31</sup>P. Indeed Roming et al. <sup>36</sup> report a T<sub>1</sub> of 48 s for 'as-prepared' zinc phosphate, in the range of our values ranging from 33 to 67 s for the signal near 4.2 ppm we assign to hopeite.

**Table S1**: Isotropic chemical shifts ( $\delta_{iso}$ ) and tensors ( $\delta_{11}$ ,  $\delta_{22}$ , and  $\delta_{33}$ ), T<sub>1</sub> relaxation times, cross-polarization buildup times ( $\tau_{cp}$ ) and T<sub>1H,p</sub> relaxation times for ZnO MNMs aged at pH 6 and pH 8 and standards.

Species	$\delta_{iso}$	$\delta_{11}$	δ <sub>22</sub>	δ <sub>33</sub>	span	<b>T</b> <sub>1</sub>	$\tau_{cp}$	T <sub>1H,p</sub>
	(ppm)	(ppm)	(ppm)	(ppm)	(ppm)	(s)	(ms)	(ms)
рН 6 В	8.0	46.0 ±.5	-7 ± 4	-15 ± 4	61 ±4	102	1.2	26
рН 6 А	4.3	38.6 ±.3	1.2 ±.4	-27.0 ± .1	66 ± .3	67	.65	5.7
hopeite	4.2	38.8 ± .4	2.5 ± .3	-28.6 ± .1	67.4 ± .6	33	.67	24
pH 8 A	4.5	37.65±.0 7	1.3 ± .3	-25.5 ± .4	63.2 ± .4	67		
рН 8 В	5.3	33 ± 2	-9 ± 1	-9 ± 2	42 ± 3			
pH 8 D <sup>b</sup>	7.8	39 ± 2	6 ± 2	-21.6 ± .6	60.1 ± .9	71		
Zn <sub>3</sub> (PO <sub>4</sub> ) <sub>2</sub>	3.9	39.8 ±.2	6.7 ± .7	-34.9 ± .5	74.7 ± .3			
$\alpha$ - Zn <sub>3</sub> (PO <sub>4</sub> ) <sub>2</sub> <sup>c</sup>	3.9	37.0	6.4	-31.7	68.7	1948		
β- Zn <sub>3</sub> (PO <sub>4</sub> ) <sub>2</sub> <sup>c</sup>	7.6	27.7	3.8	-8.8	36.5	946		

The T<sub>1</sub>s of each of the <sup>31</sup>P signals were evaluated by varying the delay between scans from 900 s to 4 s in 9 steps and fitting the resulting signal amplitudes to the function  $A=M_o(1-e^{-t/T_1})$  where 'A' is the signal amplitude obtained using an interscan delay of 't', M<sub>o</sub> is the maximum amplitude expected after infinite delay and T<sub>1</sub> is the longitudinal relaxation time.

<sup>a</sup> Data for the particles aged at pH 6 and hopeite were obtained using 100 s delays between direct polarization <sup>31</sup>P scans or 20 s between cross-polarized scans and acquisition times of 80 ms. Data for particles aged at pH 8 were obtained similarly except that they employed 200 s delays between direct polarization scans.

<sup>b</sup> pH 8 C: a fourth component is needed to describe the spectrum of material aged at pH 8 in 150 mg  $L^{-1}$  phosphate. However due to extensive overlap we do not have a unique description for it at present. A shoulder is seen at 7.1 ppm (**Figure 6**) but simulation of the pH 8 150 mg  $L^{-1}$  spectrum with four components yields best agreement when the fourth component included is broad and centered at 10.6 ppm (data not shown).

<sup>c</sup> Our isotropic chemical shifts ranging from 4 -  $\approx 10$  ppm and spans smaller than 70 ppm demonstrate that the phosphate is orthophosphate not a polyphosphate, as Roming et al. <sup>30</sup> report isotropic shifts of 3.9 ppm and 7.6 ppm associated with spans of 68.7 ppm and 36.4 ppm for  $\alpha$ - and  $\beta$ - Zn<sub>3</sub>(PO<sub>4</sub>)<sub>2</sub>, respectively but spans greater than 80 ppm for most of the components of their Zn<sub>2</sub>P<sub>2</sub>O<sub>7</sub> samples 80.7, 84.4, 84.6, 81.2, 99.1 and 69.3 ppm. Similarly, our relatively modest signal spans are most consistent with un-protonated PO<sub>4</sub><sup>3-</sup> groups, as Rothwell et al. <sup>31</sup> found that for Ca<sup>2+</sup> salts, HPO<sub>4</sub><sup>2-</sup> and H<sub>2</sub>PO<sub>4</sub><sup>-1</sup> have broader spans of 123 ppm and 127 ppm for HPO<sub>4</sub><sup>2-</sup> species and 97 ppm, 125 ppm for H<sub>2</sub>PO<sub>4</sub><sup>-</sup> species vs. 34 ppm and 33 ppm for PO<sub>4</sub><sup>3-</sup> species.