

Continuous Flow, Highly Enantioselective Michael Additions Catalyzed by a PS-Supported Squaramide

Supporting Information

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General Methods	S2
General procedure for the Michael reaction	S2
General procedure for the recycling reactions in batch conditions	S2
Description of the experimental setup for the continuous-flow process	S2
Description of the experimental setup for the continuous-flow process with different substrates	S3
Physical and spectroscopical data of Michael adducts 3a-m	S4
¹ H and ¹³ C NMR spectra of 3a-m	S9
HPLC chromatograms of 3a-m	S22

General Methods

Unless otherwise stated, all commercial reagents were used as received and solvents were used from solvent drying system, all reactions were carried out directly under open air. Merrifield resin (1% DVB, $f = 0.53 \text{ mmol Cl g}^{-1} \text{ resin}$) was obtained from Novabiochem. All flash chromatography was carried out using 60 mesh silica gel and dry-packed columns. The ^1H and ^{13}C NMR spectra were recorded at 400 MHz and 500 MHz for ^1H or at 100 MHz and 125 MHz for ^{13}C , respectively. TMS was used as internal standard for ^1H NMR and CDCl_3 for ^{13}C NMR. Chemical shifts are reported in ppm referred to TMS. FT-IR measurements carried out on a FTIR spectrometer equipped with a DTGS detector, KBr beamsplitter at 4 cm^{-1} resolution. Elemental analyses were performed on a CHNS 932 micro-analyzer. Specific optical rotation measurement was carried out on a polarimeter equipped with a PMT detector using the Sodium line at 589 nm. High performance liquid chromatography (HPLC) was performed by using Chiralpak IA, and IC columns and guard columns. Racemic standard products were prepared using DABCO (20 mol%) as catalyst in order to establish HPLC conditions. Catalyst **PS-SQ** was synthesized according to the reported procedures.¹

General procedure for the Michael reaction

To a solution of squaramide organocatalyst **PS-SQ** (11.5 mg, 0.004 mmol, 2 mol%, $f = 0.38 \text{ mmol} \cdot \text{g}^{-1}$) in CH_2Cl_2 (0.5 mL) was added nitroolefin (0.2 mmol) and 2-hydroxy-1,4-naphthoquinone (0.2 mmol). Reactions were monitored by TLC until the consumption of starting compounds, and then the reaction mixture was directly purified by column chromatography on silica gel to afford the Michael product.

General procedure for the recycling reactions in batch conditions

To a solution of squaramide organocatalyst **PS-SQ** (23.0 mg, 0.008 mmol, 4 mol%, $f = 0.38 \text{ mmol} \cdot \text{g}^{-1}$) in CH_2Cl_2 (0.5 mL) was added nitroolefin (0.2 mmol) and 2-hydroxy-1,4-naphthoquinone (0.2 mmol). Reactions were monitored by TLC until the consumption of starting compounds. Then the reaction mixture was filtered, and washed with 15 mL of CH_2Cl_2 . After that the filtrate was concentrated at reduced pressure and purified by column chromatography on silica gel to afford the Michael product.

Description of the experimental setup for the continuous flow process

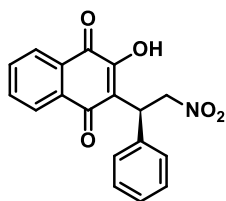
The packed bed reactor consisted of a vertically mounted and fritted low-pressure glass chromatography column (10 mm bore size and up to maximal 70 mm of adjustable bed height)

loaded with **PS-SQ** (0.25 g, 0.095 mmol, $f = 0.38 \text{ mmol} \cdot \text{g}^{-1}$). At the start, $\text{CH}_2\text{Cl}_2/\text{THF}$ (10:1) was flushed for half an hour at $0.2 \text{ ml} \cdot \text{min}^{-1}$ flow rate to swell the resin. After that, the solvent channel was switched to a solution of 2-hydroxy-1,4-naphthoquinone (27.0 mmol, 4.79 g) and *trans*- β -nitrostyrene (32.4 mmol, 4.83 g) in 270 ml $\text{CH}_2\text{Cl}_2/\text{THF}$ (10:1) of this solution were pumped to the reactor with a flow rate $0.2 \text{ ml} \cdot \text{min}^{-1}$. The reactor outlet was connected to a receiving flask where the product was collected. After 22 h flow was stopped and **PS-SQ** washed with $\text{CH}_2\text{Cl}_2/\text{THF}$ (10:1) solvent system for an hour. Conversion and enantioselectivity of the formed product were determined by ^1H NMR and HPLC analysis of periodically collected samples. In the end, collected solution was concentrated at reduced pressure and purified by flash chromatography on silica gel.

Description of the experimental setup for the continuous flow process with different substrates

The packed bed reactor was loaded with swollen resin **PS-SQ** (0.50 g, 0.19 mmol, $f = 0.38 \text{ mmol} \cdot \text{g}^{-1}$) in $\text{CH}_2\text{Cl}_2/\text{THF}$ (10:1) and flushed half an hour with the same solvent mixture at $0.2 \text{ ml} \cdot \text{min}^{-1}$ flow rate. After the resin was conditioned, the solvent channel was switched to a solution of 2-hydroxy-1,4-naphthoquinone (1.80 mmol) and the corresponding nitroalkene (1.80 mmol) in 18 ml $\text{CH}_2\text{Cl}_2/\text{THF}$ (10:1) and pumped through the reactor with a flow rate of $0.2 \text{ ml} \cdot \text{min}^{-1}$. The reactor outlet was connected to a receiving flask where the product was collected. After 1 h, the flow was switched to a washing solvent $\text{CH}_2\text{Cl}_2/\text{THF}$ (10:1) to clean the resin for half an hour. This process was repeated for each nitroalkene. Conversion and enantioselectivity of the product were determined by ^1H NMR and HPLC analysis of periodically collected samples. In the end, collected solutions were concentrated at reduced pressure and purified by flash chromatography on silica gel.

Physical and spectroscopical data of Michael adducts 3a-m

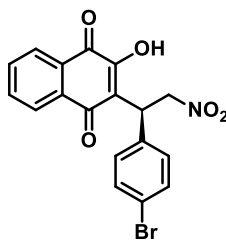


(S)-2-Hydroxy-3-(2-nitro-1-phenylethyl)naphthalene-1,4-dione (3a)²

¹H NMR (500 MHz, CDCl₃): δ 5.14 (dd, J = 6.9, 13.4 Hz, 1H), 5.31 (dd, J = 6.8, 9.1 Hz, 1H), 5.47 (dd, J = 9.1, 13.4 Hz, 1H), 7.24-7.32 (m, 3H), 7.46 (d, J = 7.2 Hz, 2H), 7.68 (dt, J = 1.3, 7.5 Hz, 1H), 7.76 (dt, J = 1.3, 7.5 Hz, 1H), 8.05 (d, J = 7.6 Hz, 1H), 8.11 (d, J = 7.6 Hz, 1H) ppm.

¹³C NMR (100 MHz, CDCl₃): 39.7, 76.4, 120.8, 126.3, 127.2, 127.8, 128.3 ($\times 2$), 129.0 ($\times 3$), 132.6, 133.3, 135.4, 137.5, 153.2, 181.1, 183.7.

HPLC analysis: Chiralpak IA Column, Hexane (0.1% TFA)/Dichloromethane/Ethanol (90:5:5), flow rate = 1.0 mL/min, wavelength = 254 nm, r_t = 17.8 (major), 23.9 min (minor).

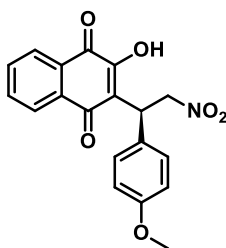


(S)-2-(1-(4-Fluorophenyl)-2-nitroethyl)-3-hydroxynaphthalene-1,4-dione (3b)²

¹H NMR (400 MHz, DMSO- d_6): δ 5.17 (t, J = 7.7 Hz, 1H), 5.33-5.28 (m, 1H), 5.45 (dd, J = 8.3, 13.7 Hz, 1H), 7.34 (d, J = 8.5 Hz, 2H), 7.48 (d, J = 8.5 Hz, 2H), 7.76 (t, J = 7.4 Hz, 1H), 7.82 (t, J = 7.4 Hz, 1H), 7.97-7.94 (m, 2H) ppm.

¹³C NMR (100 MHz, DMSO- d_6): 38.5, 76.9, 120.1, 120.6, 126.2, 126.4, 130.3, 130.5, 131.8, 132.5, 133.6, 135.2, 138.8, 158.6, 181.8, 183.6.

HPLC analysis: Chiralpak IA Column, Hexane (0.1% TFA)/Dichloromethane/Ethanol (85:14:1), flow rate = 1.0 mL/min, wavelength = 254 nm, r_t = 25.6 (major), 26.6 min (minor).

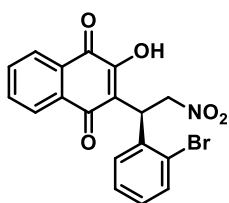


(S)-2-Hydroxy-3-(1-(4-methoxyphenyl)-2-nitroethyl)naphthalene-1,4-dione (3c)²

¹H NMR (500 MHz, CDCl₃): δ 3.76 (s, 3H), 5.11 (dd, J = 6.9, 13.3 Hz, 1H), 5.25 (dd, J = 7.0, 9.0 Hz, 1H), 5.43 (dd, J = 9.0, 13.3 Hz, 1H), 6.84 (d, J = 8.9 Hz, 2H), 7.39 (d, J = 8.8 Hz, 2H), 7.69 (dt, J = 1.3, 7.6 Hz, 1H), 7.77 (dt, J = 1.3, 7.6 Hz, 1H), 8.06 (d, J = 7.6 Hz, 1H), 8.11 (d, J = 7.6 Hz, 1H) ppm.

^{13}C NMR (100 MHz, CDCl_3): 39.0, 55.3, 76.6, 114.3, 121.1, 126.3, 127.3, 129.0, 129.4, 129.5, 132.7, 133.3, 135.4, 152.9, 159.2, 181.2, 183.3.

HPLC analysis: Chiralpak IA Column, Hexane (0.1% TFA)/Dichloromethane/Ethanol (92:4:4), flow rate = 0.6 mL/min, wavelength = 254 nm, r_t = 68.4 (major), 70.4 min (minor).

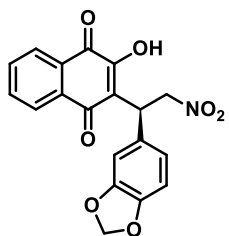


(R)-2-(1-(2-Bromophenyl)-2-nitroethyl)-3-hydroxynaphthalene-1,4-dione (3d)³

^1H NMR (400 MHz, CDCl_3): δ 4.91 (dd, J = 5.8, 13.7 Hz, 1H), 5.42 (dd, J = 10.4, 13.9 Hz, 1H), 5.70 (dd, J = 5.7, 10.4 Hz, 1H), 7.13 (dt, J = 1.5, 7.8 Hz, 1H), 7.27-7.23 (m, 1H), 7.41 (dd, J = 1.6, 7.8 Hz, 1H), 7.60 (dd, J = 1.2, 7.9 Hz, 1H), 7.80-7.69 (m, 2H), 8.13-8.09 (m, 2H) ppm.

^{13}C NMR (100 MHz, CDCl_3): 39.9, 74.9, 119.6, 124.6, 126.4, 127.3, 127.9, 129.0, 129.4, 129.6, 132.7, 133.3, 133.6, 135.6, 136.2, 154.0, 181.0, 183.8.

HPLC analysis: Chiralpak IA Column, Hexane (0.1% TFA)/Dichloromethane/Ethanol (90:9:1), flow rate = 1.0 mL/min, wavelength = 254 nm, r_t = 23.2 (major), 24.1 min (minor).



(S)-2-(1-(Benzo[d][1,3]dioxol-5-yl)-2-nitroethyl)-3-hydroxynaphthalene-1,4-dione (3e)

^1H NMR (400 MHz, CDCl_3): δ 5.13 (dd, J = 7.1, 13.1 Hz, 1H), 5.22 (t, J = 7.1, 15.6 Hz, 1H), 5.38 (dd, J = 8.4, 13.1 Hz, 1H), 5.90 (dd, J = 1.2, 4.0 Hz, 2H), 6.72 (d, J = 8.1 Hz, 1H), 6.93 (dd, J = 2.1, 7.8 Hz, 1H), 6.97 (d, J = 1.7 Hz, 1H), 7.67 (dt, J = 1.3, 7.5 Hz, 1H), 7.79-7.75 (m, 2H), 8.05 (dd, J = 1.4, 7.6 Hz, 1H), 8.10 (dd, J = 1.4, 7.8 Hz, 1H) ppm.

^{13}C NMR (100 MHz, CDCl_3): 39.5, 76.6, 101.2, 108.6, 108.7, 120.9, 121.8, 126.3, 127.2, 129.0, 131.1, 132.6, 133.3, 135.4, 147.2, 148.0, 153.0, 181.1, 183.7.

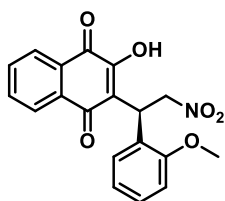
HRMS (ESI): m/z calcd. for $\text{C}_{19}\text{H}_{12}\text{NO}_7$ [$M - \text{H}$] $^-$ 366.0621, found 366.0619.

$[\alpha]_D^{26} = +2.29$ (c 7.40, CHCl_3)

m.p. 136-138 $^\circ\text{C}$.

IR (ATR): ν = 3229, 2891, 1673, 1635, 1546, 1271, 1235, 1038, 790.

HPLC analysis: Chiralpak IA Column, Hexane (0.1% TFA)/Dichloromethane/Ethanol (88:6:6), flow rate = 1.0 mL/min, wavelength = 254 nm, r_t = 29.9 (major), 49.8 min (minor).

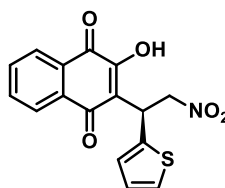


(S)-2-Hydroxy-3-(1-(2-methoxyphenyl)-2-nitroethyl)naphthalene-1,4-dione (3f)³

¹H NMR (400 MHz, CDCl₃): δ 3.87 (s, 3H), 4.96 (dd, J = 5.6, 13.6 Hz, 1H), 5.41 (dd, J = 10.2, 13.6 Hz, 1H), 5.65 (dd, J = 5.6, 10.2 Hz, 1H), 6.90-6.86 (m, 2H), 7.26-7.22 (m, 2H), 7.68 (dt, J = 1.3, 6.2 Hz, 1H), 7.71 (s, 1H), 7.76 (dt, J = 1.4, 6.2 Hz, 1H), 8.11- 8.07 (m, 2H) ppm.

¹³C NMR (100 MHz, CDCl₃): 34.2, 55.6, 75.4, 110.9, 120.5, 120.7, 124.9, 126.3, 127.2, 128.8, 129.0, 129.1, 132.9, 133.1, 135.3, 154.0, 156.9, 181.2, 183.8.

HPLC analysis: Chiralpak IC Column, Hexane (0.1% TFA)/Dichloromethane/Ethanol (75:23:2), flow rate = 0.7 mL/min, wavelength = 254 nm, r_t = 13.7 (major), 14.4 min (minor).

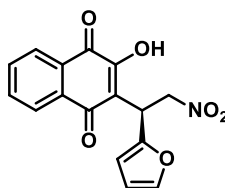


(S)-2-Hydroxy-3-(2-nitro-1-(thiophen-2-yl)ethyl)naphthalene-1,4-dione (3g)²

¹H NMR (400 MHz, CDCl₃): δ 5.13 (dd, J = 6.8, 13.5 Hz, 1H), 5.43 (dd, J = 8.9, 13.5 Hz, 1H), 5.61 (dd, J = 6.6, 9.0 Hz, 1H), 6.93 (dd, J = 3.5, 5.1 Hz, 1H), 7.10-7.11 (m, 1H), 7.20 (dd, J = 1.2, 5.1 Hz, 1H), 7.71 (dt, J = 1.3, 7.6 Hz, 1H), 7.81-7.77 (m, 2H), 8.08 (d, J = 7.5 Hz, 1H), 8.14 (d, J = 7.5 Hz, 1H) ppm.

¹³C NMR (100 MHz, CDCl₃): 34.8, 76.7, 119.9, 125.4, 126.4, 126.6, 127.0, 127.2, 129.0, 132.6, 133.4, 135.6, 139.0, 153.1, 181.0, 183.3.

HPLC analysis: Chiralpak IA Column, Hexane (0.1% TFA)/Dichloromethane/Ethanol (90:5:5), flow rate = 1.0 mL/min, wavelength = 254 nm, r_t = 21.2 (major), 22.9 min (minor).

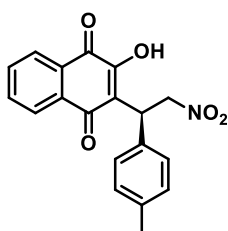


(S)-2-(1-(Furan-2-yl)-2-nitroethyl)-3-hydroxynaphthalene-1,4-dione (3h)²

¹H NMR (500 MHz, CDCl₃): δ 5.19 (dd, J = 6.9, 13.5 Hz, 1H), 5.27 (dd, J = 8.6, 13.5 Hz, 1H), 5.47 (t, J = 8.7 Hz, 1H), 6.23 (d, J = 3.3 Hz, 1H), 6.30 (dd, J = 1.8, 3.3 Hz, 1H), 7.32 (dd, J = 0.7, 1.8 Hz, 1H), 7.71 (dt, J = 1.2, 7.6 Hz, 1H), 7.81-7.78 (m, 2H), 8.09 (dd, J = 1.4, 7.7 Hz, 1H), 8.14 (dd, J = 1.4, 7.7 Hz, 1H) ppm.

^{13}C NMR (100 MHz, CDCl_3): 33.4, 74.7, 107.4, 110.7, 118.0, 126.4, 127.3, 129.0, 132.6, 133.4, 135.6, 142.3, 149.8, 153.9, 181.0, 183.2.

HPLC analysis: Chiralpak IA Column, Hexane (0.1% TFA)/Dichloromethane/Ethanol (92:4:4), flow rate = 1.0 mL/min, wavelength = 254 nm, r_t = 24.1 (major), 27.5 min (minor).

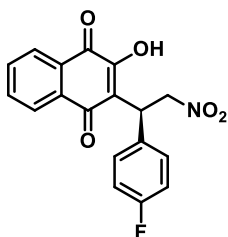


(S)-2-Hydroxy-3-(2-nitro-1-(p-tolyl)ethyl)naphthalene-1,4-dione (3i)²

^1H NMR (400 MHz, CDCl_3): δ 2.29 (s, 3H), 5.12 (dd, J = 6.8, 13.3 Hz, 1H), 5.28 (dd, J = 6.8, 10.4 Hz, 1H), 5.46 (dd, J = 9.0, 13.3 Hz, 1H), 7.12 (d, J = 8.0 Hz, 2H), 7.34 (d, J = 8.0 Hz, 2H), 7.66-7.78 (m, 3H), 8.06 (d, J = 7.6 Hz, 1H), 8.11 (d, J = 7.6 Hz, 1H) ppm.

^{13}C NMR (100 MHz, CDCl_3): 21.0, 39.3, 76.4, 121.0, 126.3, 127.2, 128.1, 129.0, 129.7, 132.7, 133.2, 134.5, 135.4, 137.7, 153.1, 181.2, 183.7.

HPLC analysis: Chiralpak IA Column, Hexane (0.1% TFA)/Dichloromethane/Ethanol (90:5:5), flow rate = 1.0 mL/min, wavelength = 254 nm, r_t = 20.9 (major), 25.8 min (minor).

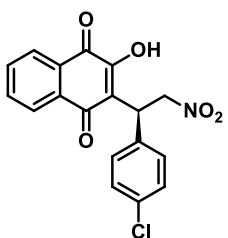


(S)-2-(1-(4-Fluorophenyl)-2-nitroethyl)-3-hydroxynaphthalene-1,4-dione (3j)²

^1H NMR (500 MHz, CDCl_3): δ 5.15 (dd, J = 6.8, 12.7 Hz, 1H), 5.29 (t, J = 15.2 Hz, 1H), 5.40 (dd, J = 8.4, 12.7 Hz, 1H), 6.99 (t, J = 8.7 Hz, 2H), 7.45 (dd, J = 5.2, 8.8 Hz, 2H), 7.70 (dt, J = 1.4, 7.6 Hz, 1H), 7.78 (dt, J = 1.4, 7.6 Hz, 1H), 8.07 (d, J = 7.6 Hz, 1H), 8.11 (d, J = 7.7 Hz, 1H) ppm.

^{13}C NMR (100 MHz, CDCl_3): 39.0, 76.4, 115.9 (d, J = 21.3 Hz), 120.6, 126.4, 127.2, 128.9, 130.0 (d, J = 8.3 Hz), 130.1, 132.6, 133.3, 133.4, 135.6, 153.3, 162.2 (d, J = 246.9 Hz), 181.1, 183.7.

HPLC analysis: Chiralpak IA Column, Hexane (0.1% TFA)/Dichloromethane/Ethanol (90:5:5), flow rate = 1.0 mL/min, wavelength = 254 nm, r_t = 20.5 (major), 23.4 min (minor).

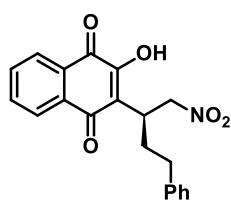


(S)-2-(1-(4-Chlorophenyl)-2-nitroethyl)-3-hydroxynaphthalene-1,4-dione (3k)²

^1H NMR (400 MHz, DMSO- d_6): δ 5.19 (t, J = 7.8 Hz, 1H), 5.32 (dd, J = 7.5, 13.9 Hz, 1H), 5.43 (dd, J = 8.2, 13.8 Hz, 1H), 7.41-7.35 (m, 4H), 7.79 (dt, J = 1.4, 7.4 Hz, 1H), 7.85 (dt, J = 1.4, 7.4 Hz, 1H), 8.00-7.97 (m, 2H) ppm.

^{13}C NMR (100 MHz, DMSO- d_6): 38.3, 76.9, 120.8, 126.3, 126.4, 129.0 ($\times 2$), 130.2 ($\times 3$), 130.4, 132.2, 133.9, 135.3, 138.0, 157.1, 181.3, 184.2.

HPLC analysis: Chiralpak IA Column, Hexane (0.1% TFA)/Dichloromethane/Ethanol (90:5:5), flow rate = 1.0 mL/min, wavelength = 254 nm, r_t = 20.4 (major), 21.8 min (minor).

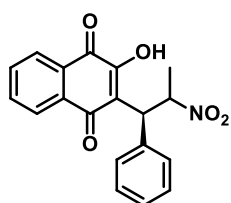


(S)-2-Hydroxy-3-(1-nitro-4-phenylbutan-2-yl)naphthalene-1,4-dione
(3l)²

^1H NMR (500 MHz, CDCl_3): δ 2.04-1.97 (m, 1H), 2.33-2.25 (m, 1H), 2.61-2.55 (m, 1H), 2.72-2.66 (m, 1H), 4.13-4.07 (m, 1H), 4.71 (dd, J = 12.6, 6.2 Hz, 1H), 4.96 (dd, J = 12.7, 9.1 Hz, 1H), 7.10-7.04 (m, 3H), 7.17-7.14 (m, 2H), 7.70 (t, J = 7.5 Hz, 1H), 7.78 (t, J = 7.5 Hz, 1H), 8.06 (d, J = 7.6 Hz, 1H), 8.11 (d, J = 7.6 Hz, 1H) ppm.

^{13}C NMR (100 MHz, CDCl_3): 31.7, 33.8, 34.8, 77.1, 120.4, 126.1, 126.3, 127.1, 128.2, 128.3, 129.1, 132.8, 133.2, 135.3, 140.9, 153.9, 180.7, 183.9.

HPLC analysis: Chiralpak IA Column, Hexane (0.1% TFA)/Dichloromethane/Ethanol (90:9:1), flow rate = 1.0 mL/min, wavelength = 254 nm, r_t = 23.8 (major), 25.4 min (minor).



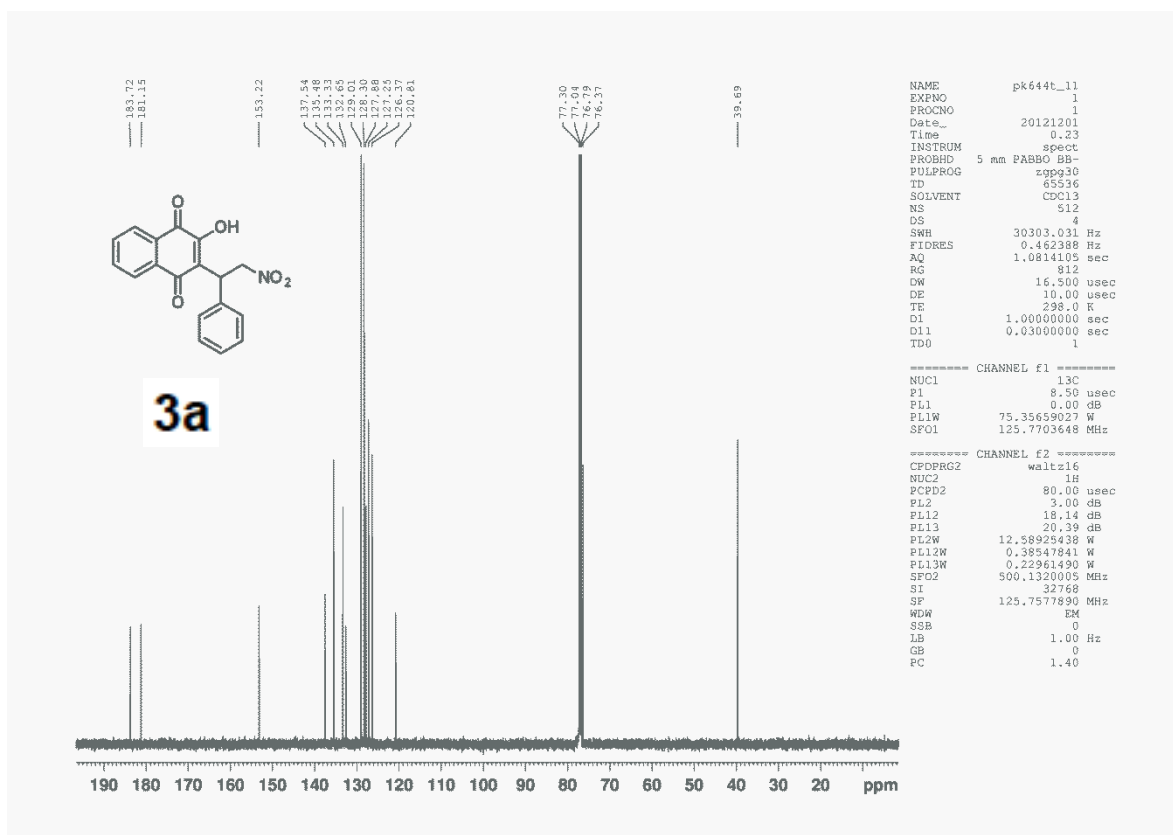
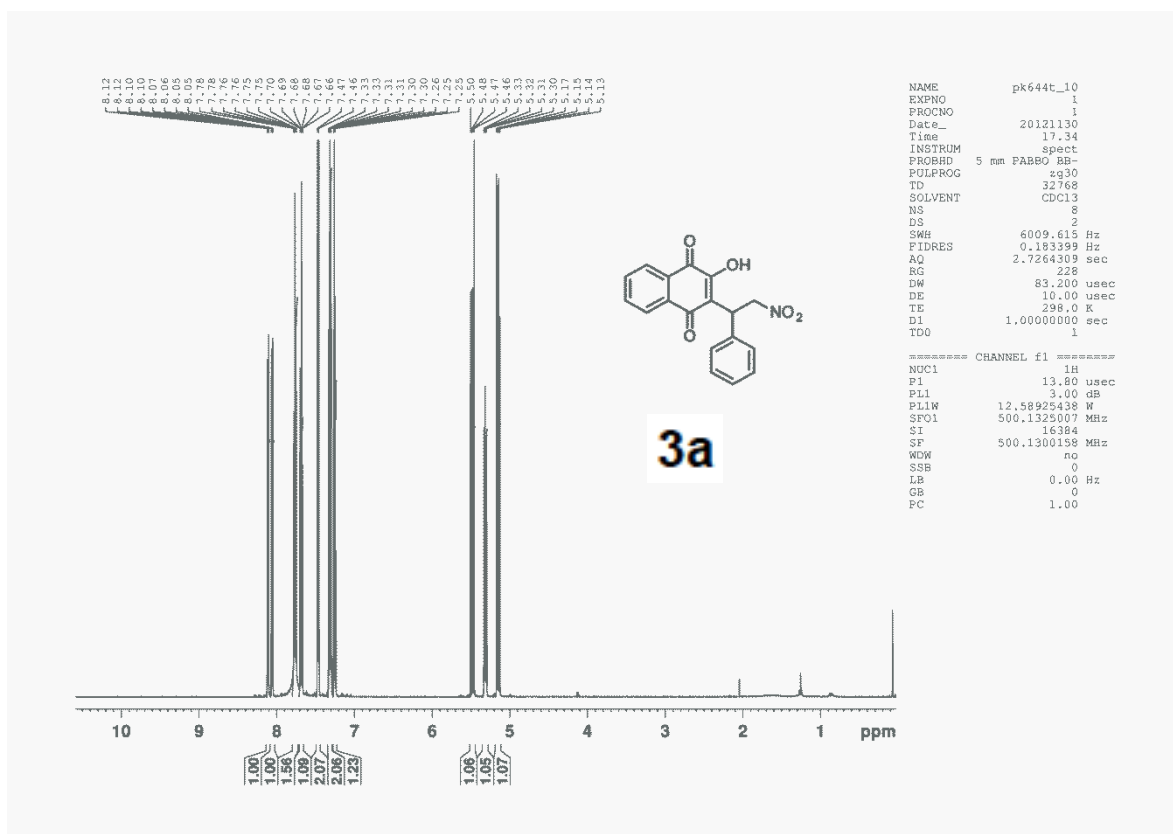
2-Hydroxy-3-((1S)-2-nitro-1-phenylpropyl)naphthalene-1,4-dione
(3m)³

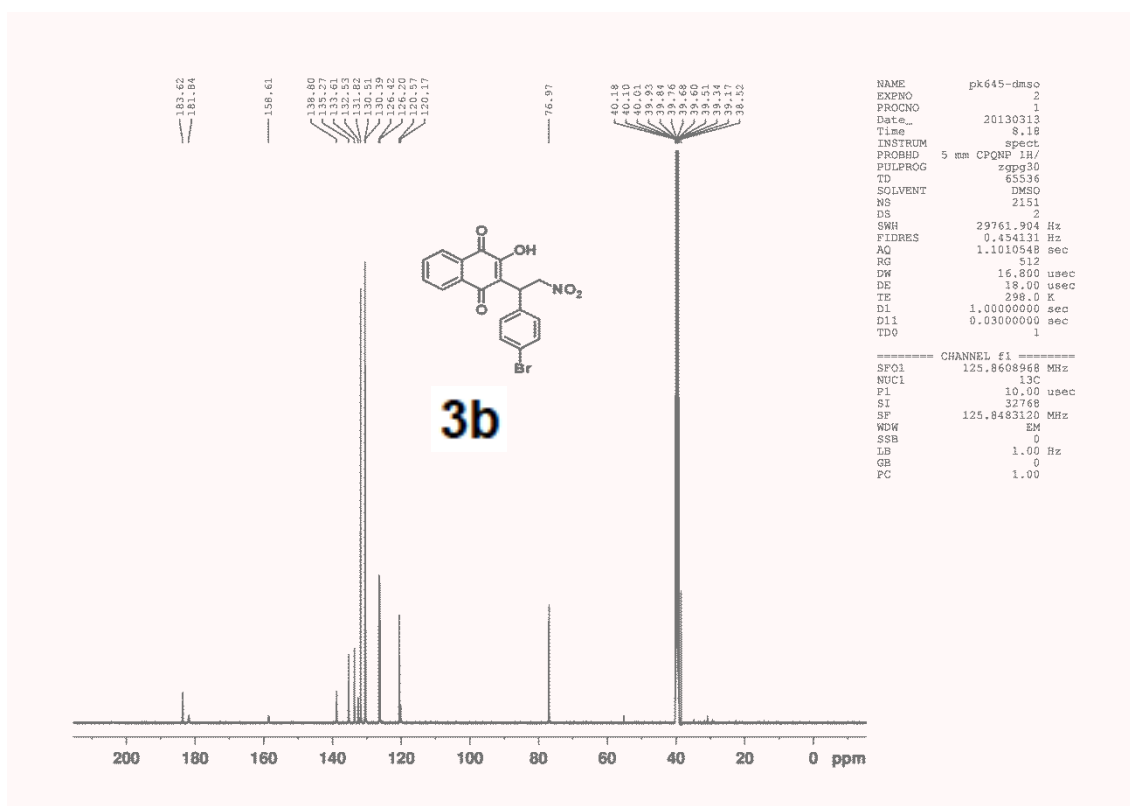
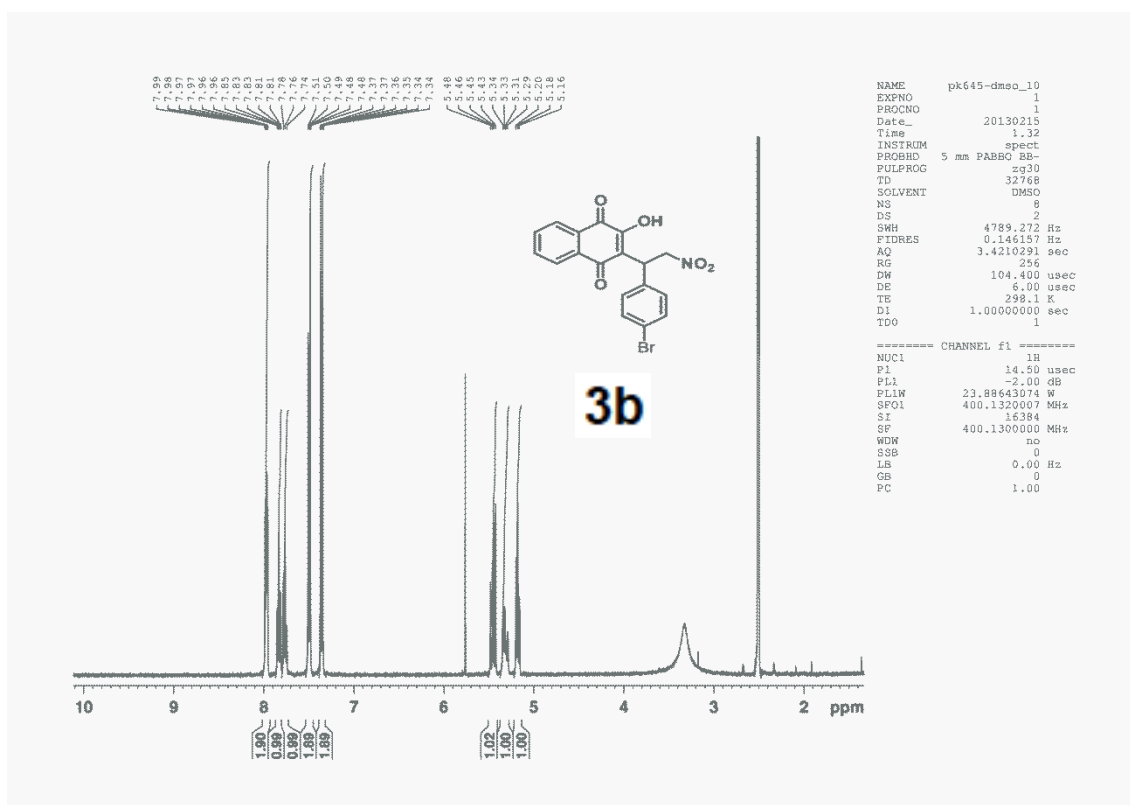
^1H NMR (500 MHz, CDCl_3): δ 1.53 (d, J = 6.7 Hz, 3H), 4.93 (d, J = 11.6 Hz, 1H), 6.05-6.12 (m, 1H), 7.24-7.33 (m, 3H), 7.52-7.54 (m, 2H), 7.64 (dt, J = 1.2, 7.5 Hz, 1H), 7.73 (dt, J = 1.3, 7.5 Hz, 1H), 7.78 (s, 1H), 8.01 (dd, J = 1.3, 7.6 Hz, 1H), 8.10 (dd, J = 1.0, 7.6 Hz, 1H) ppm.

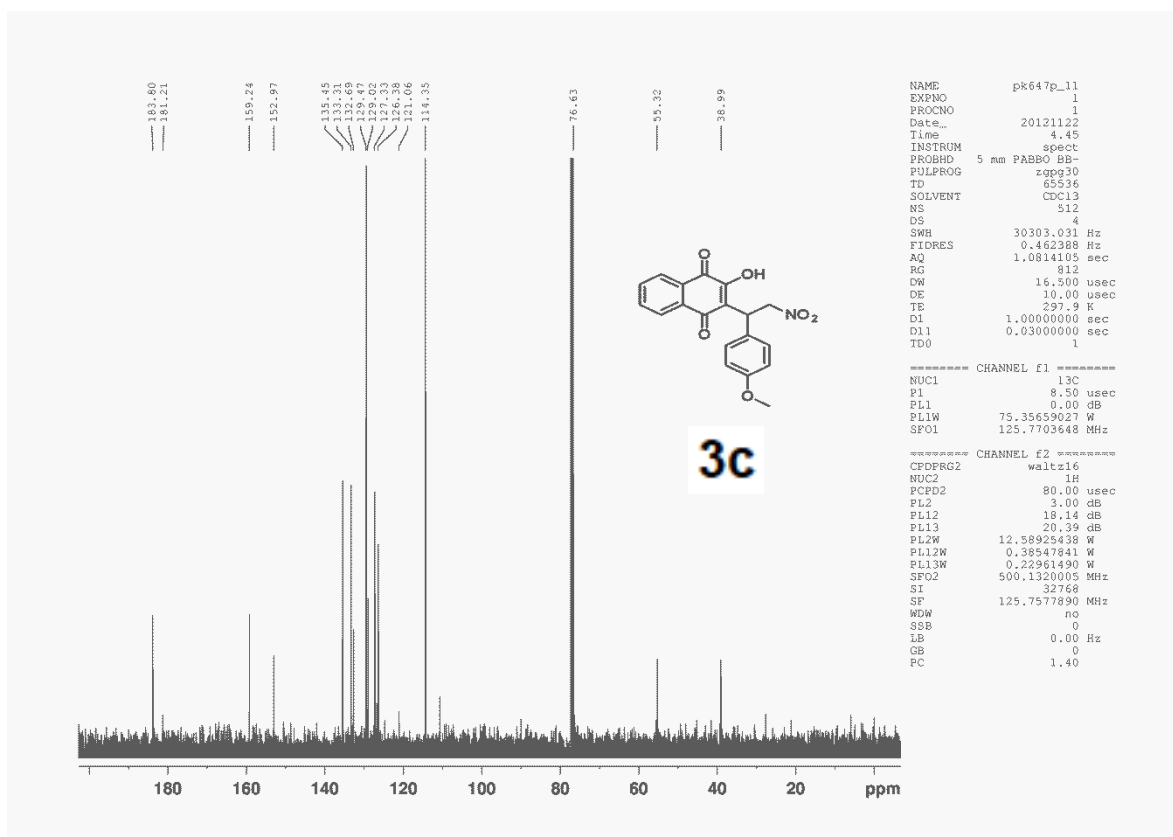
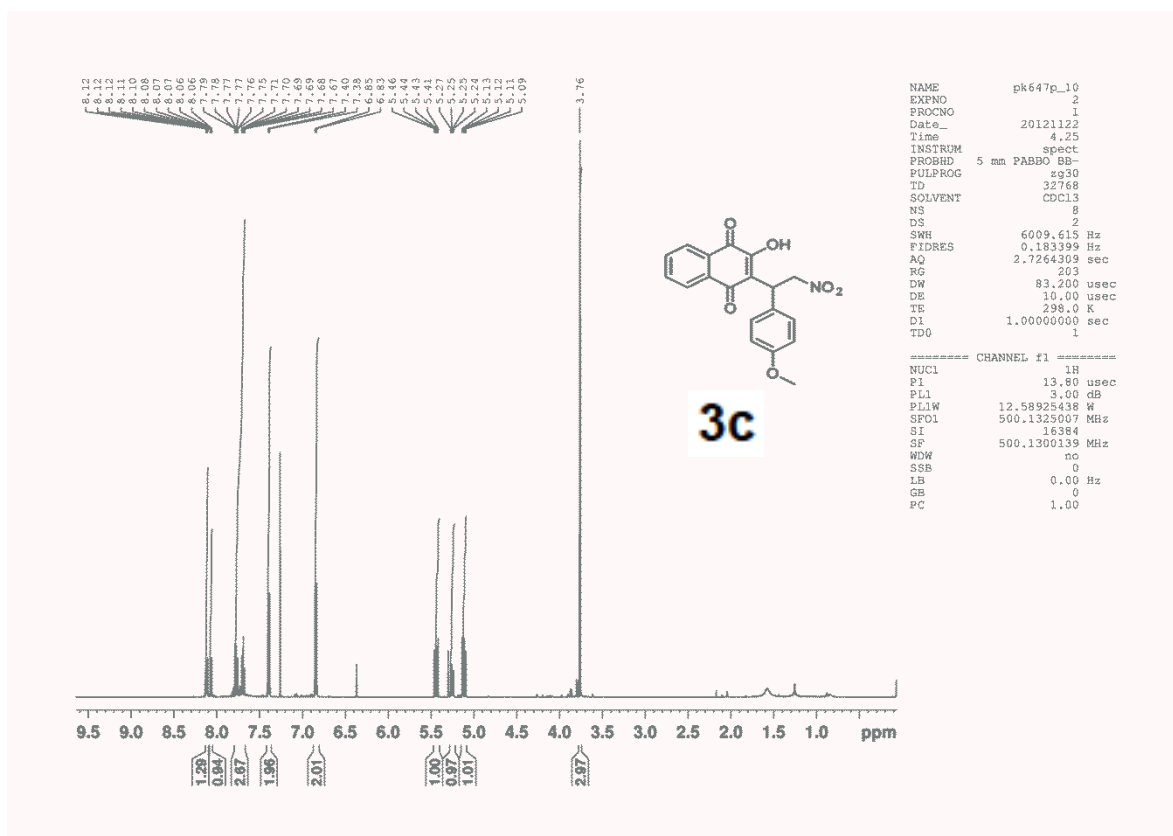
^{13}C NMR (100 MHz, CDCl_3): 19.7, 46.6, 83.4, 121.2, 126.2, 127.2, 127.9, 129.1, 129.2, 129.9, 132.7, 133.1, 135.3, 137.3, 152.8, 181.1, 183.6.

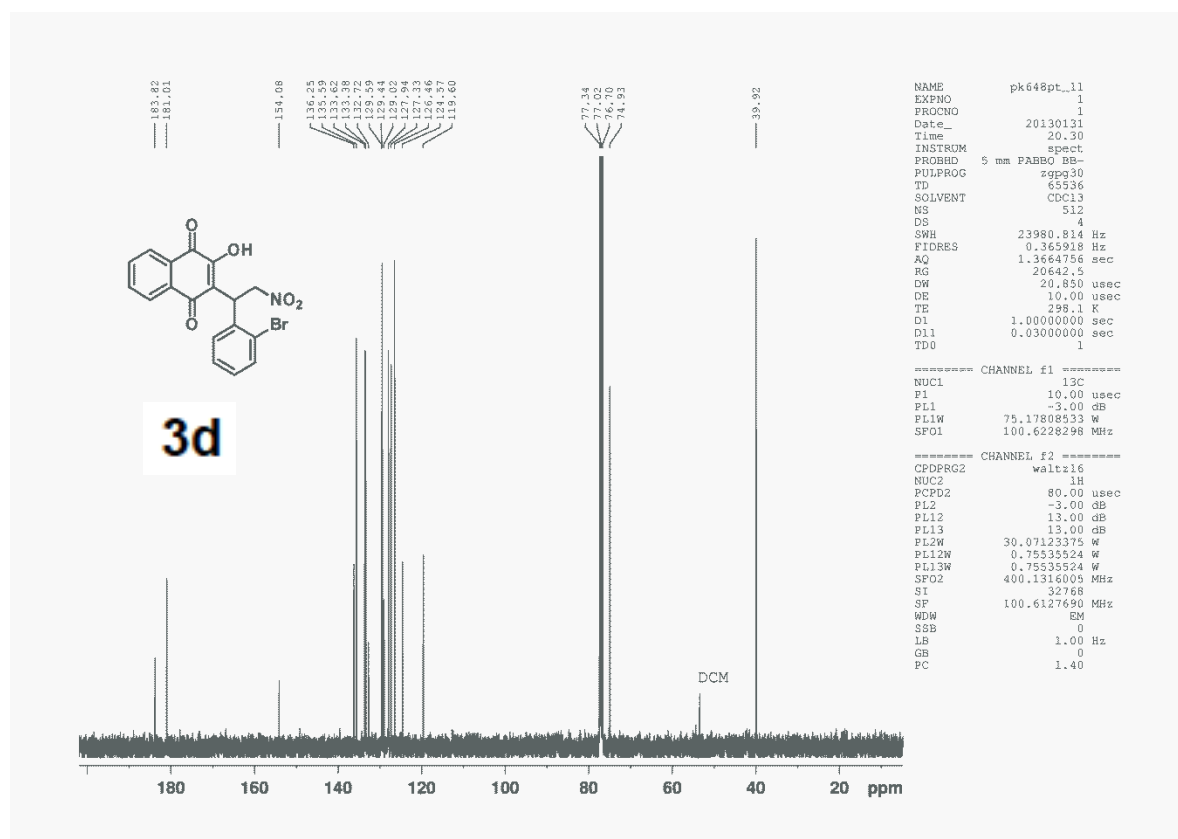
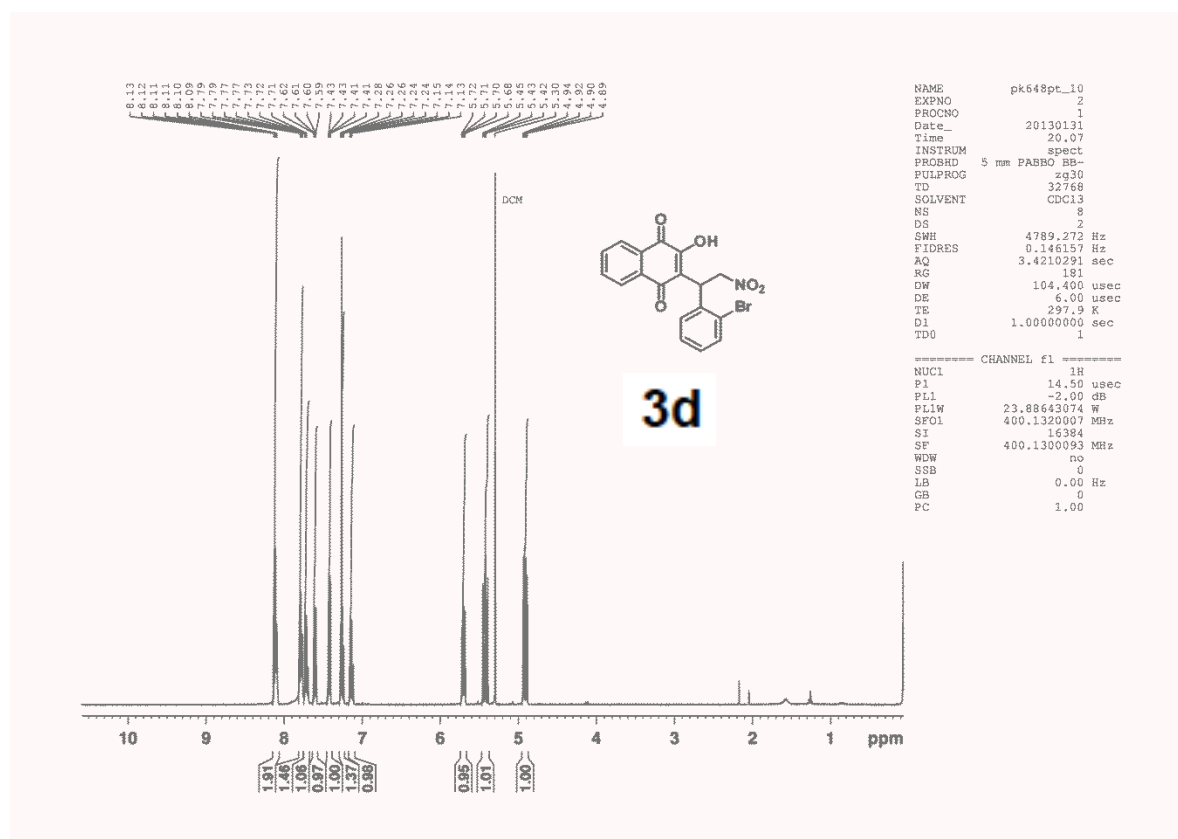
HPLC analysis: Chiralpak IA Column, Hexane (0.1% TFA)/Dichloromethane/Ethanol (90:5:5), flow rate = 1.0 mL/min, wavelength = 254 nm, r_t = 15.2 (major), 23.9 min (minor).

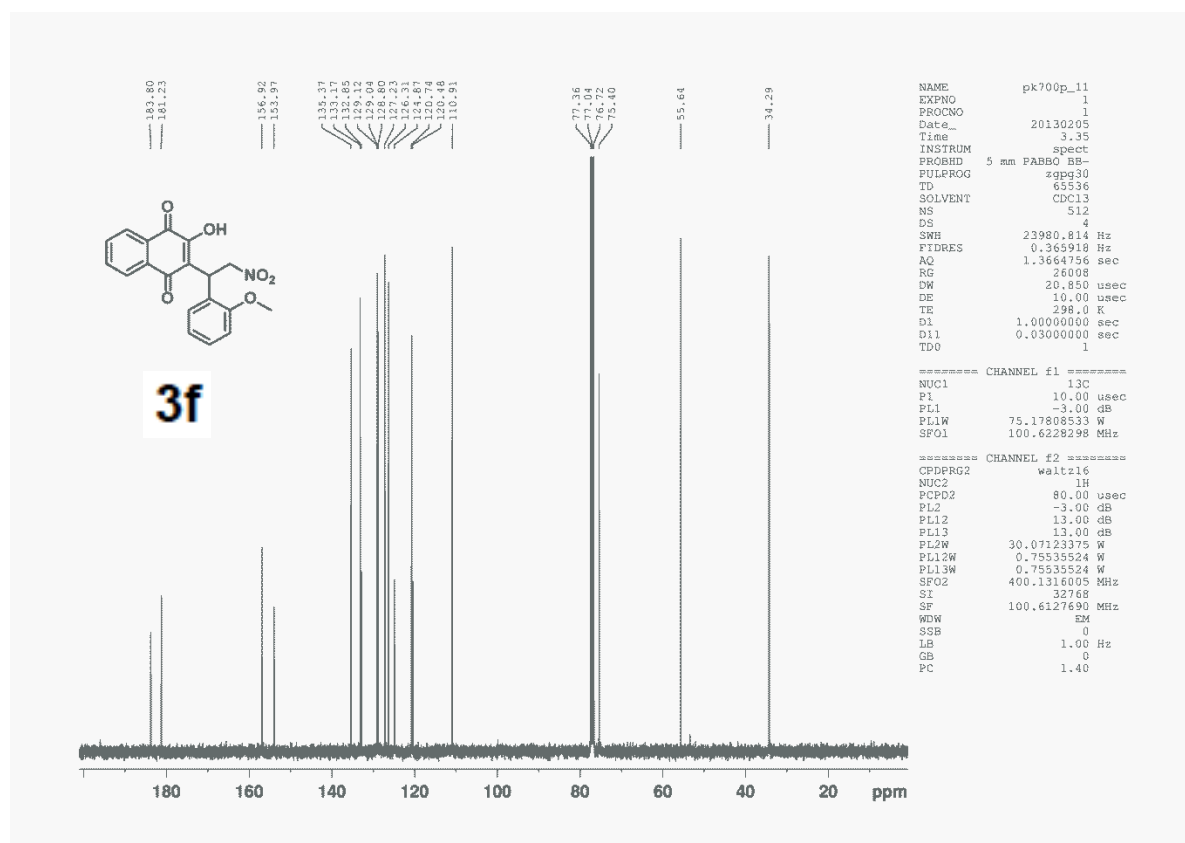
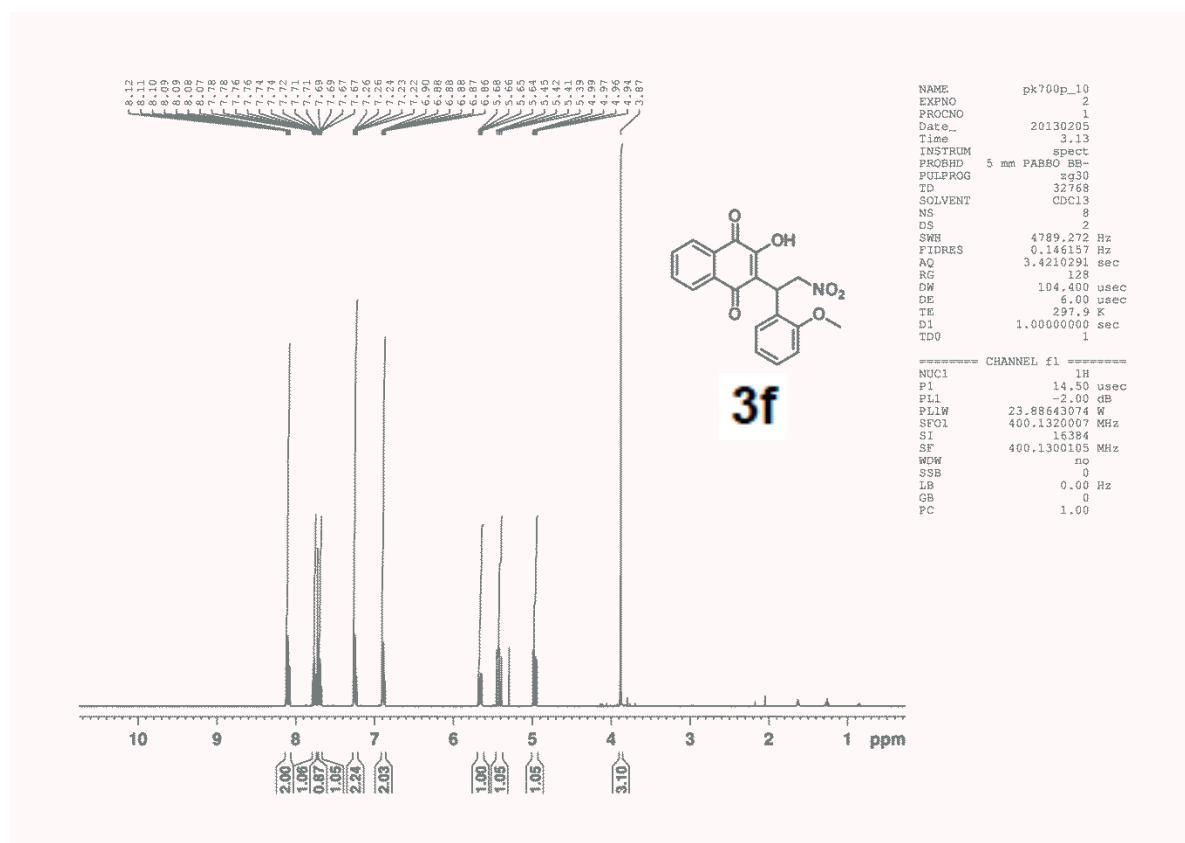
¹H and ¹³C NMR spectra of 3a-m

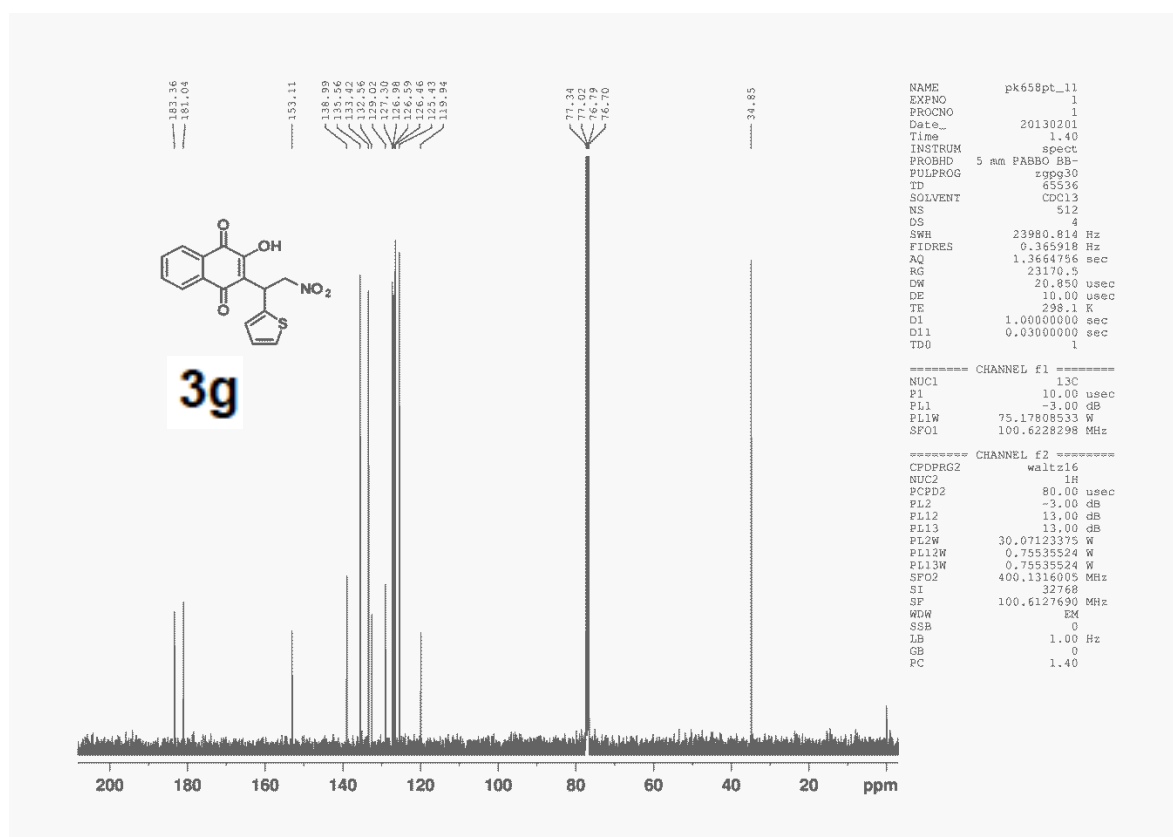
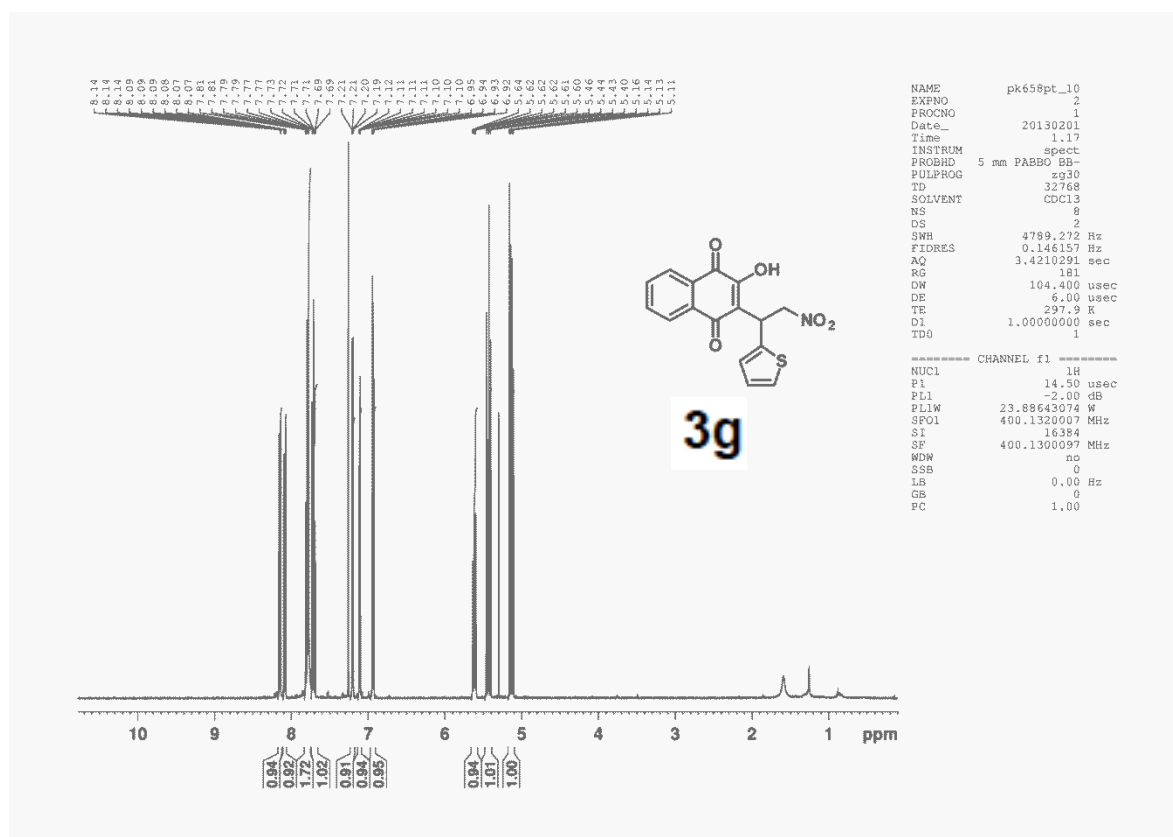


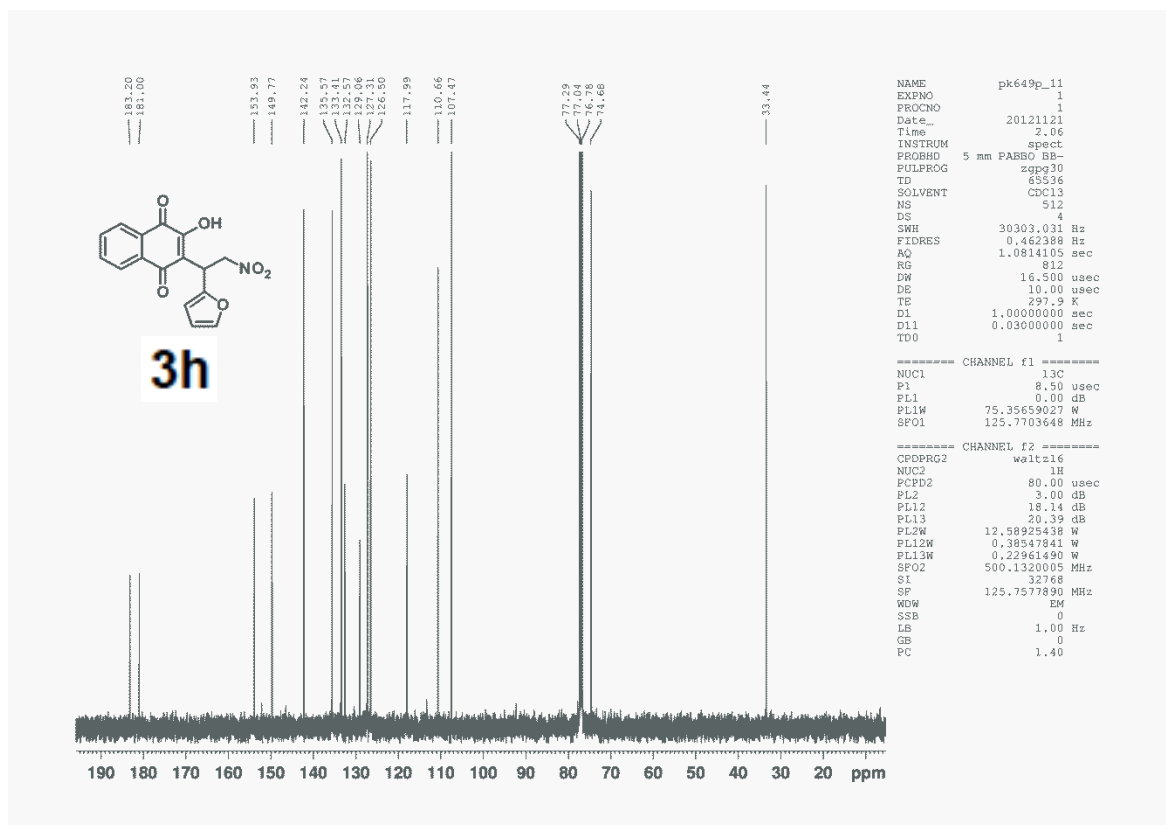
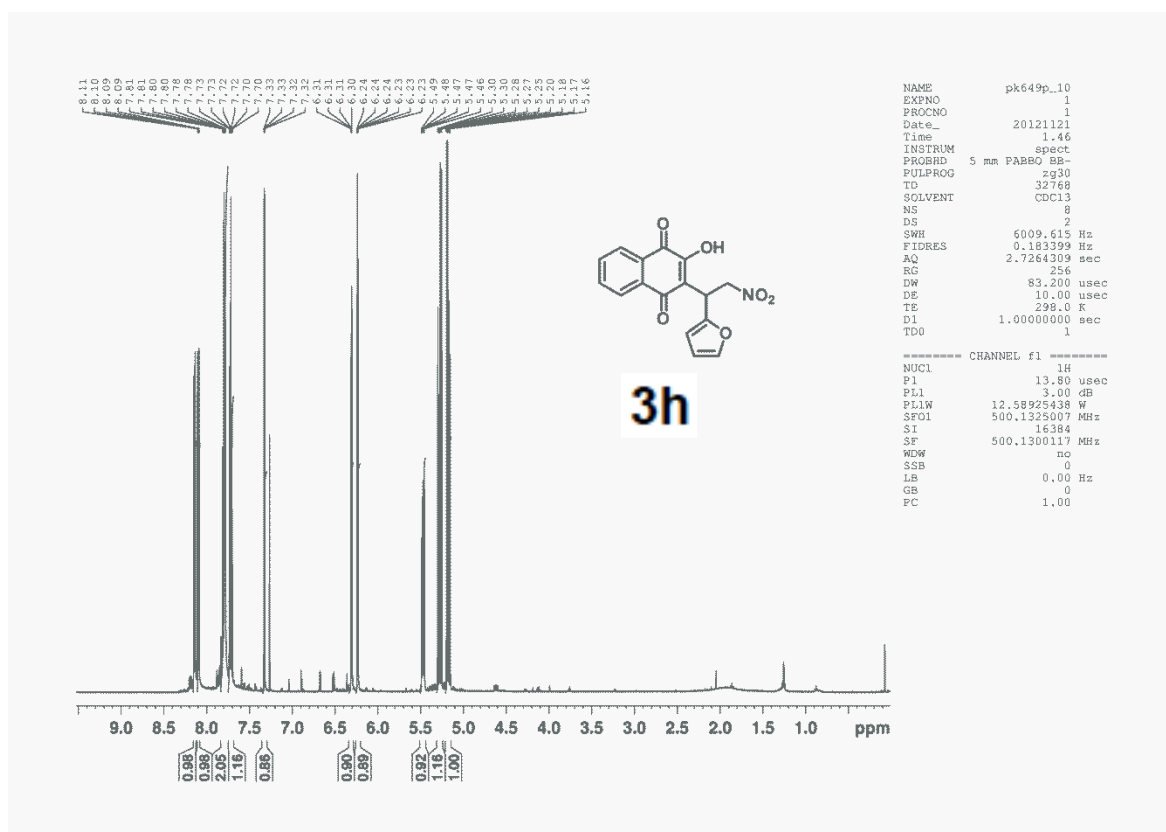


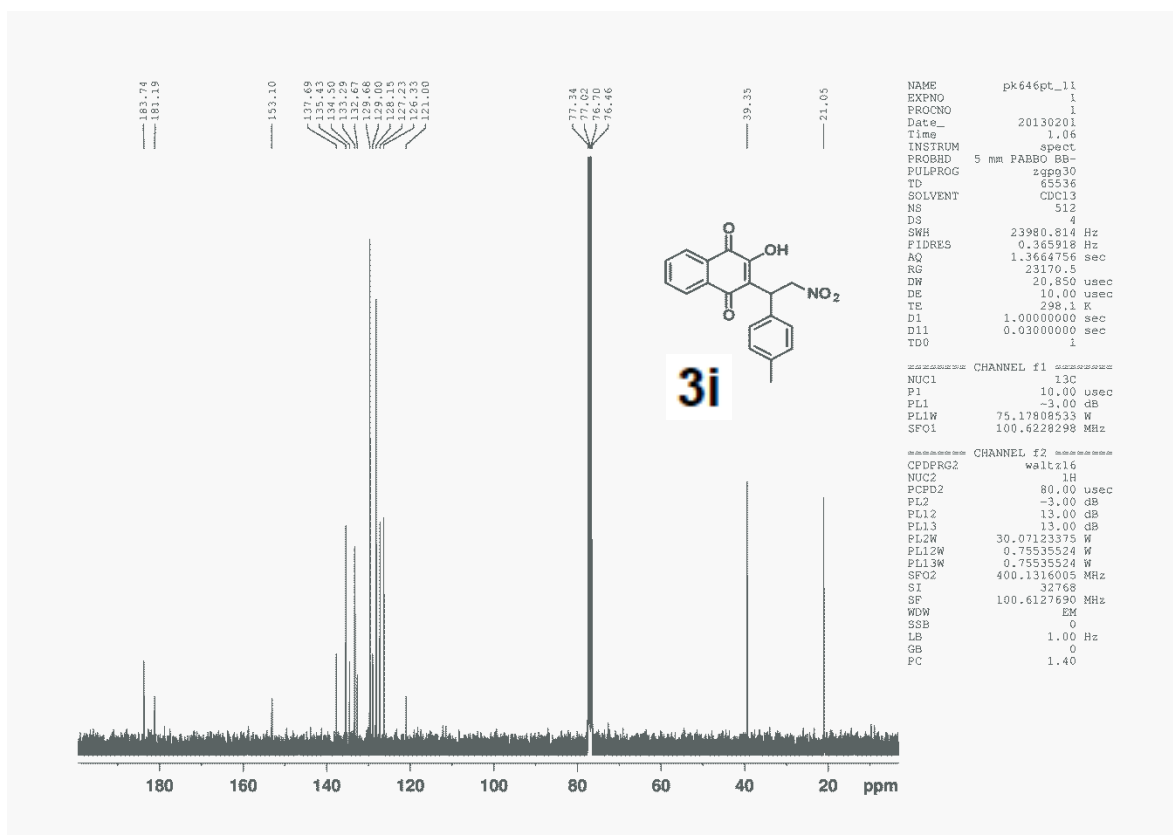
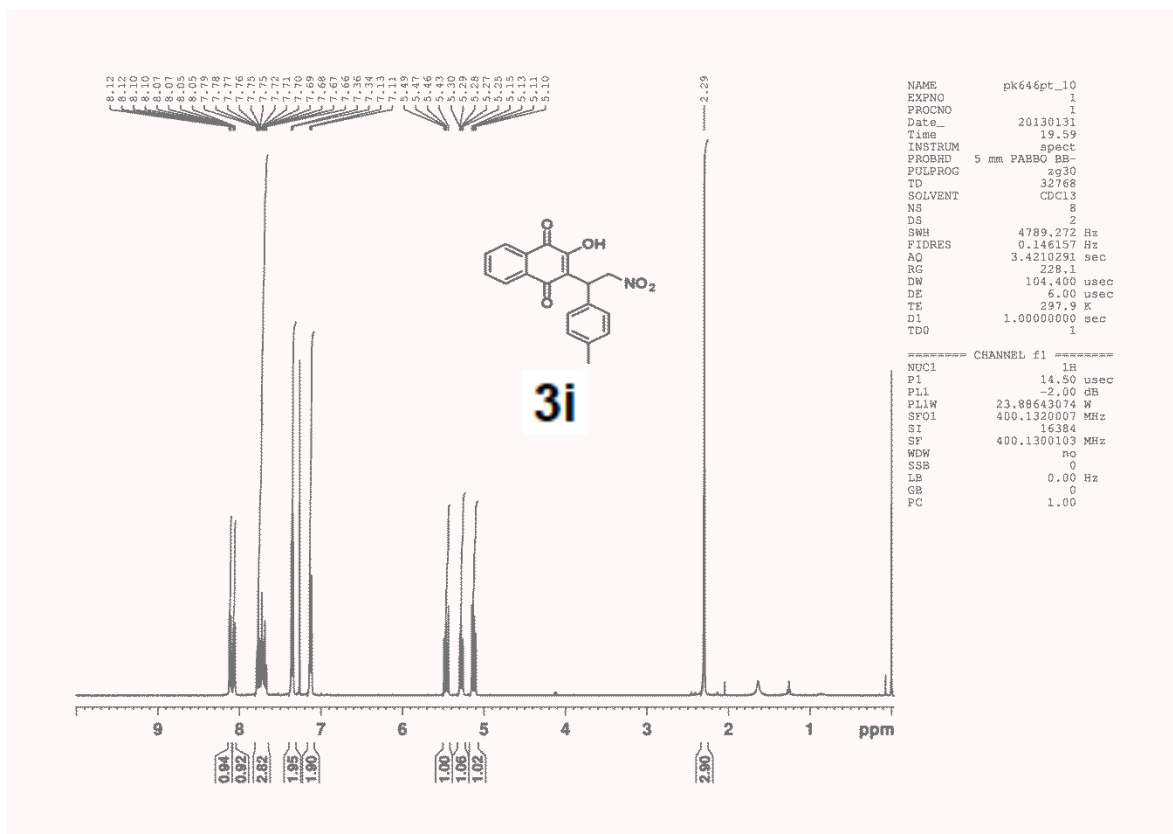


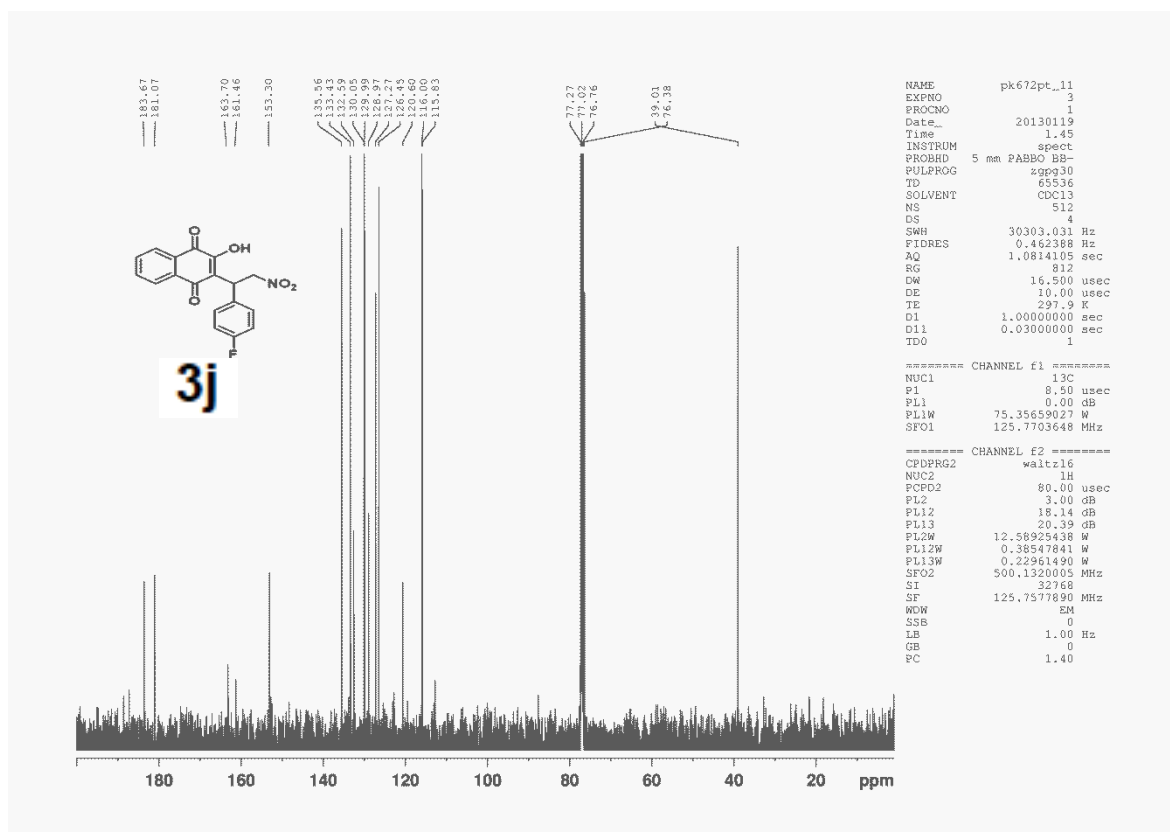
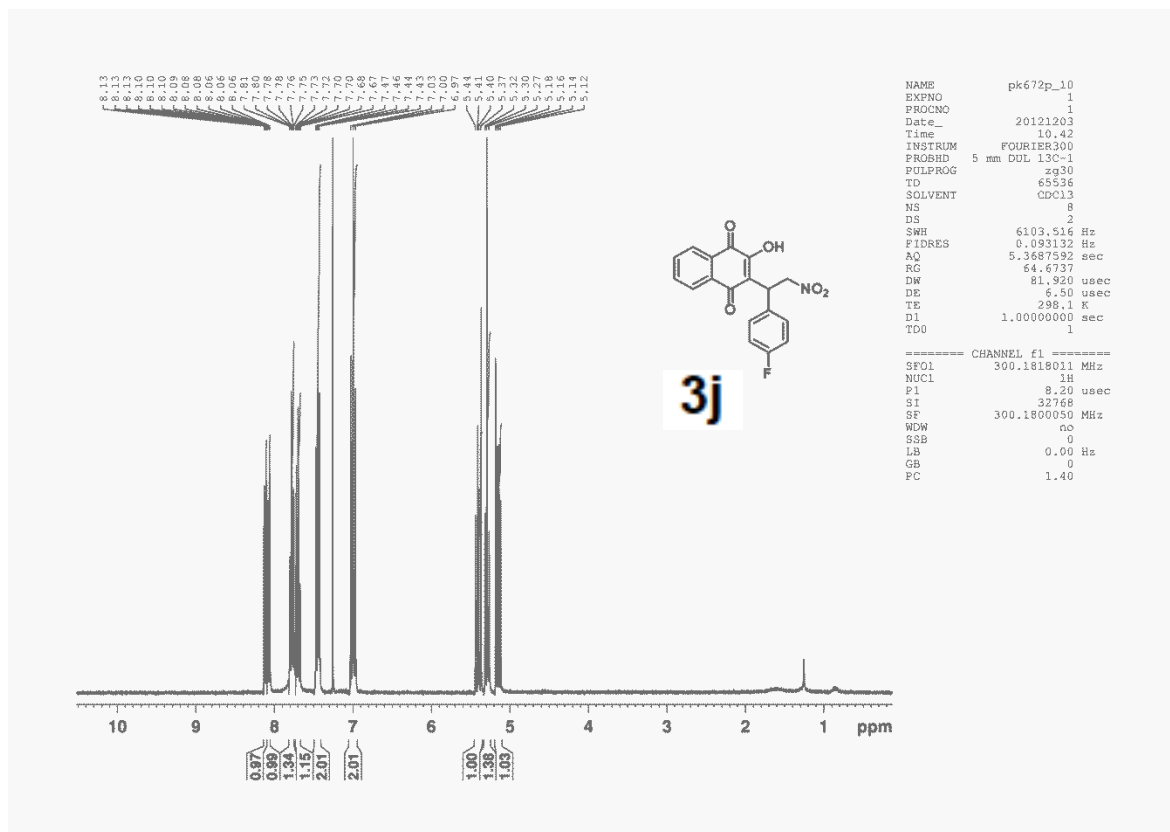


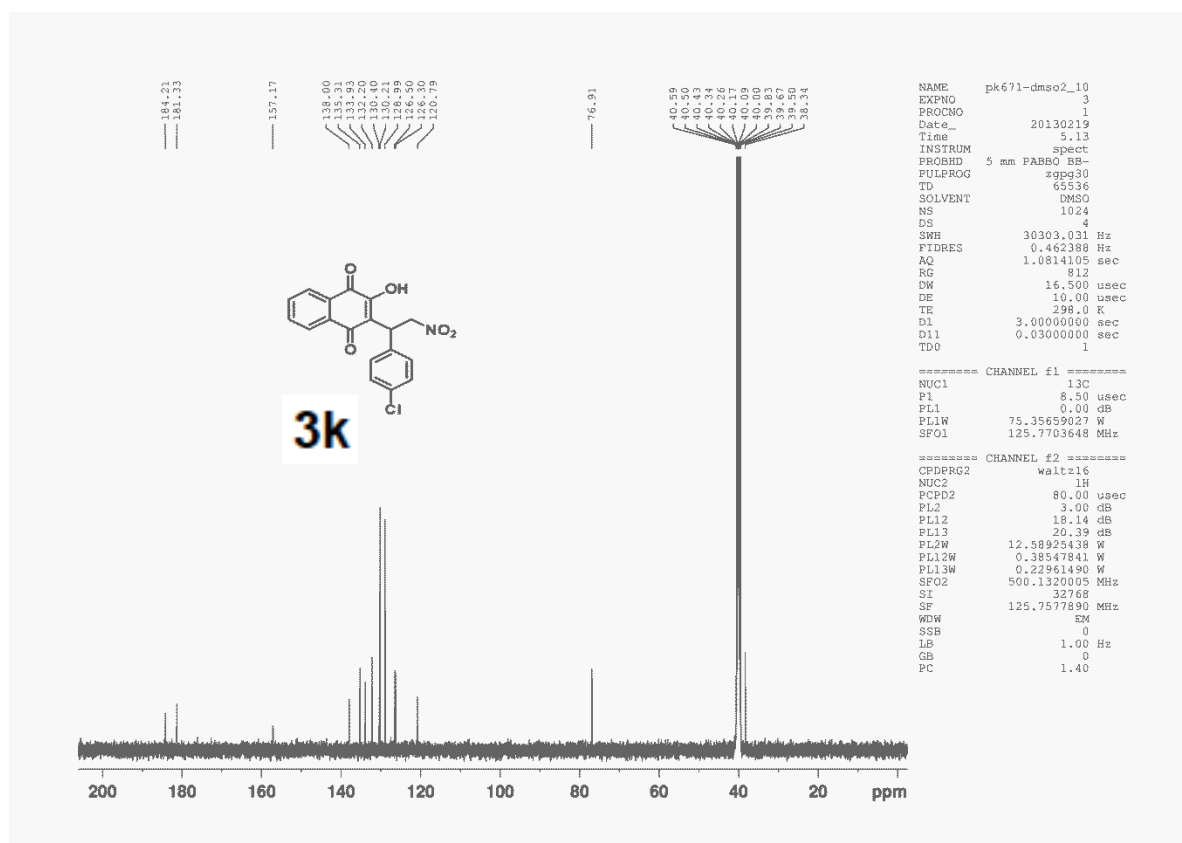
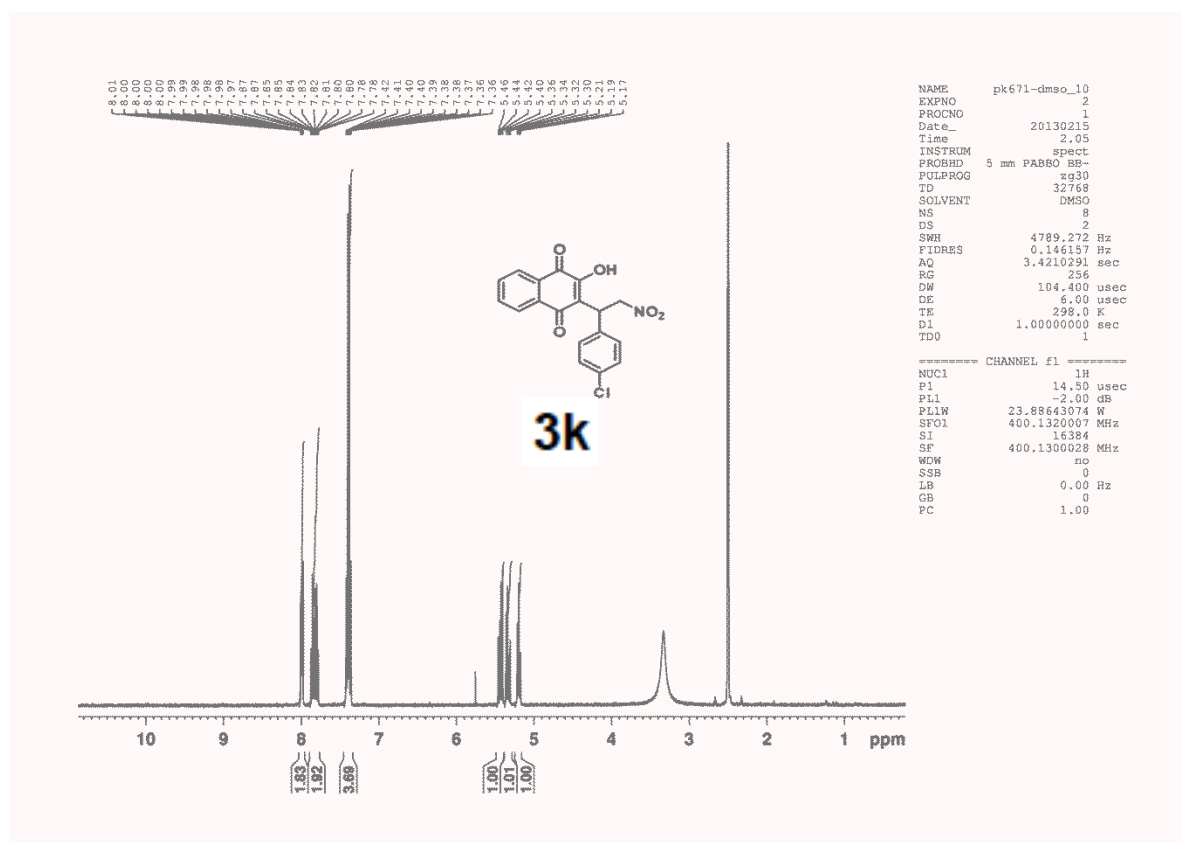


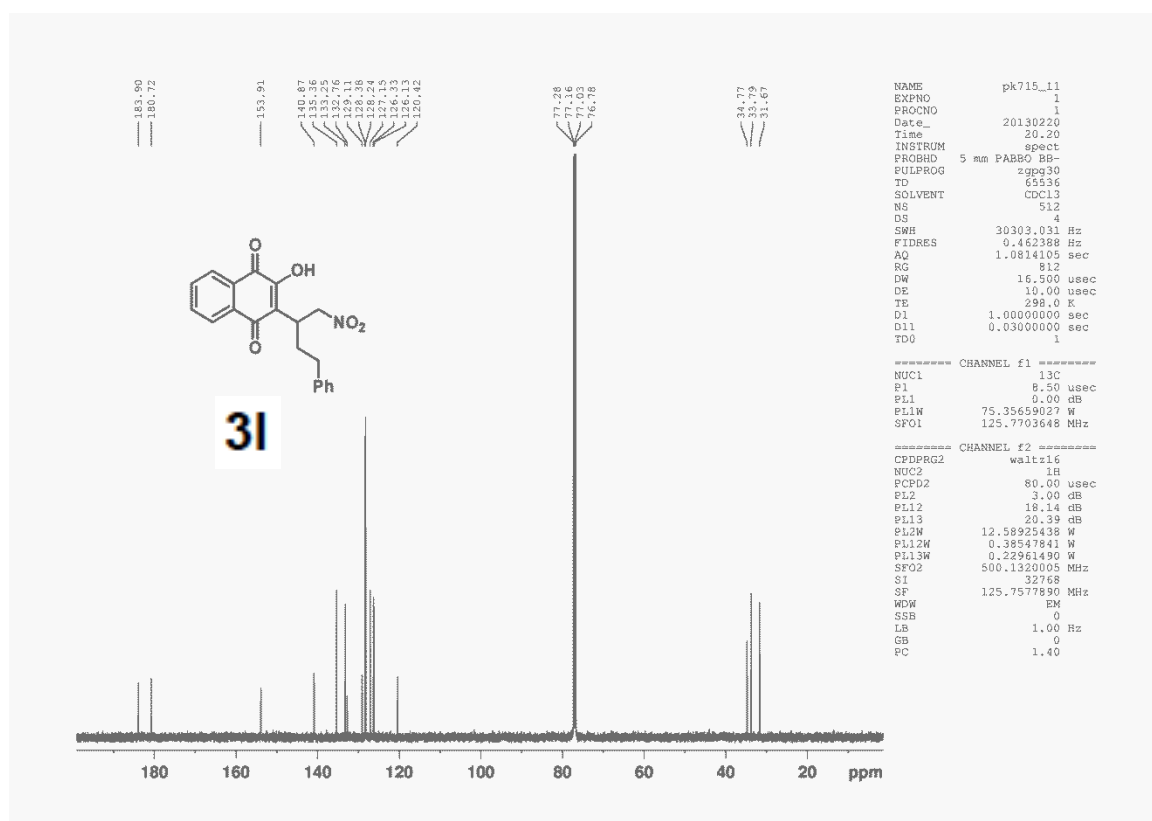
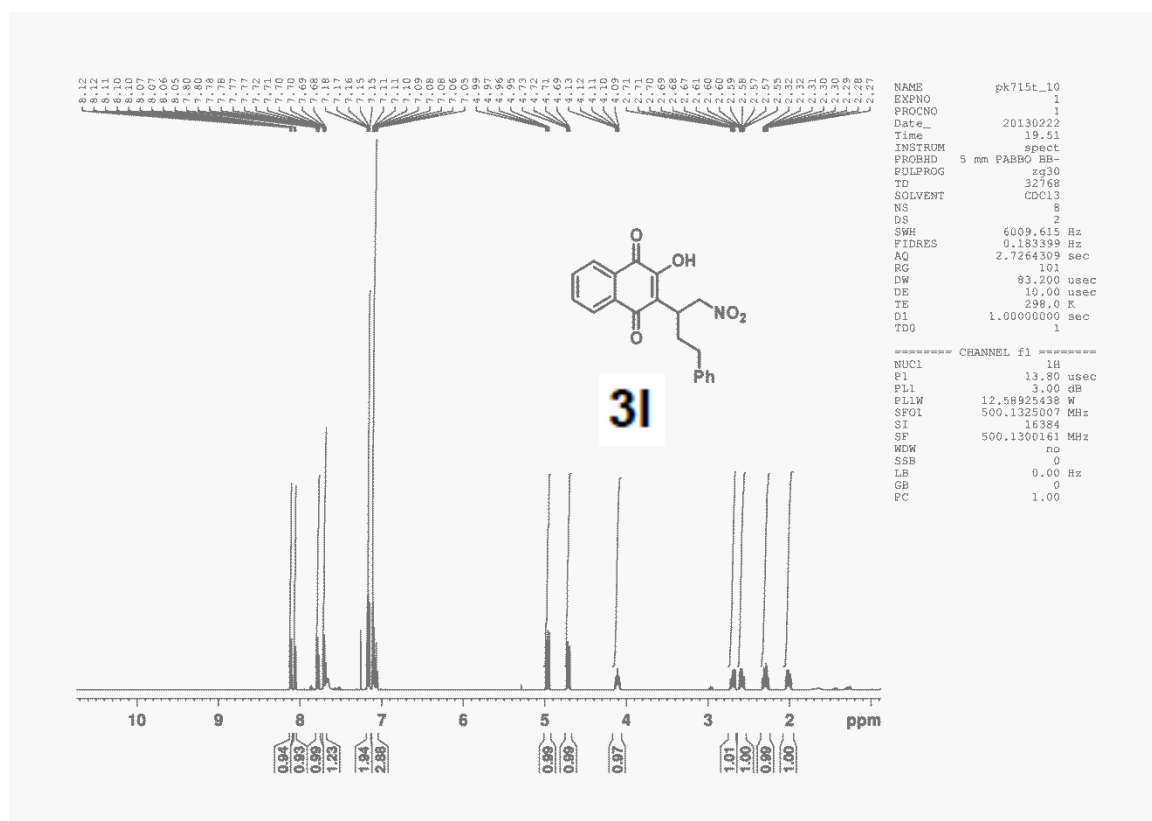


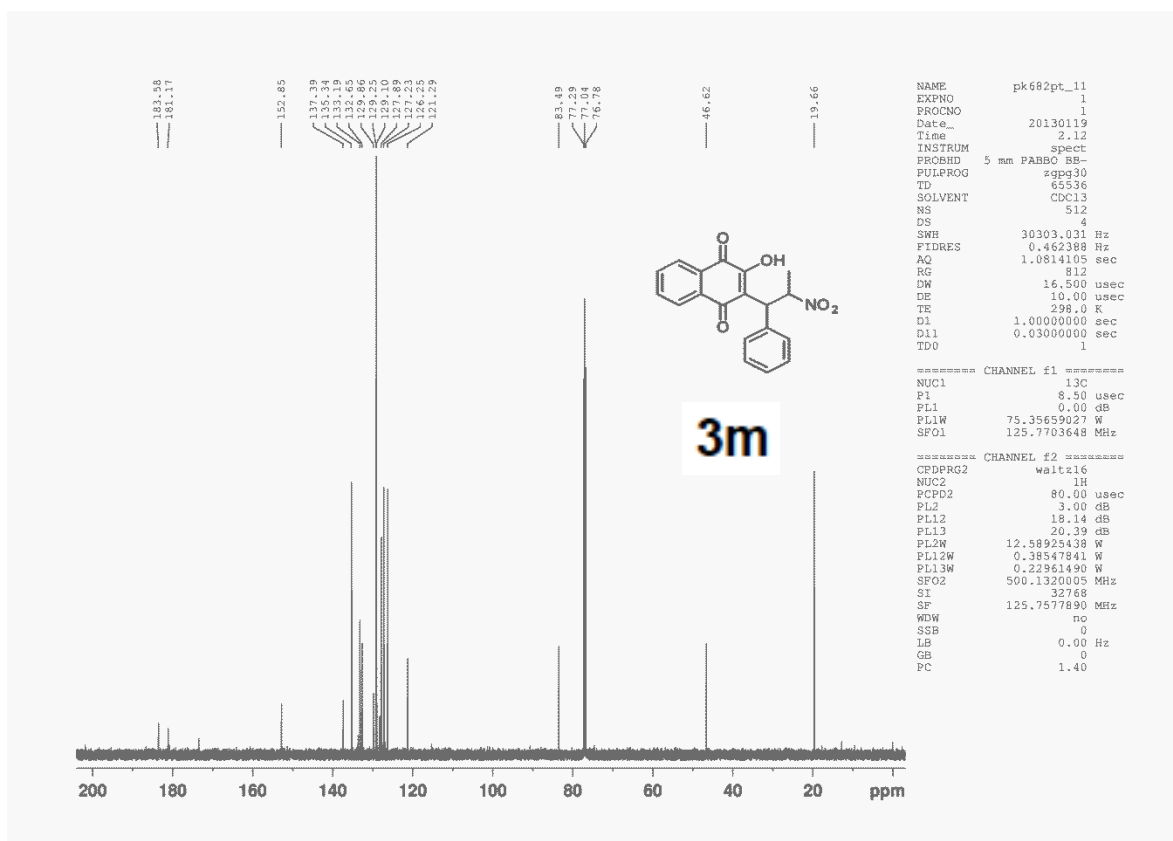
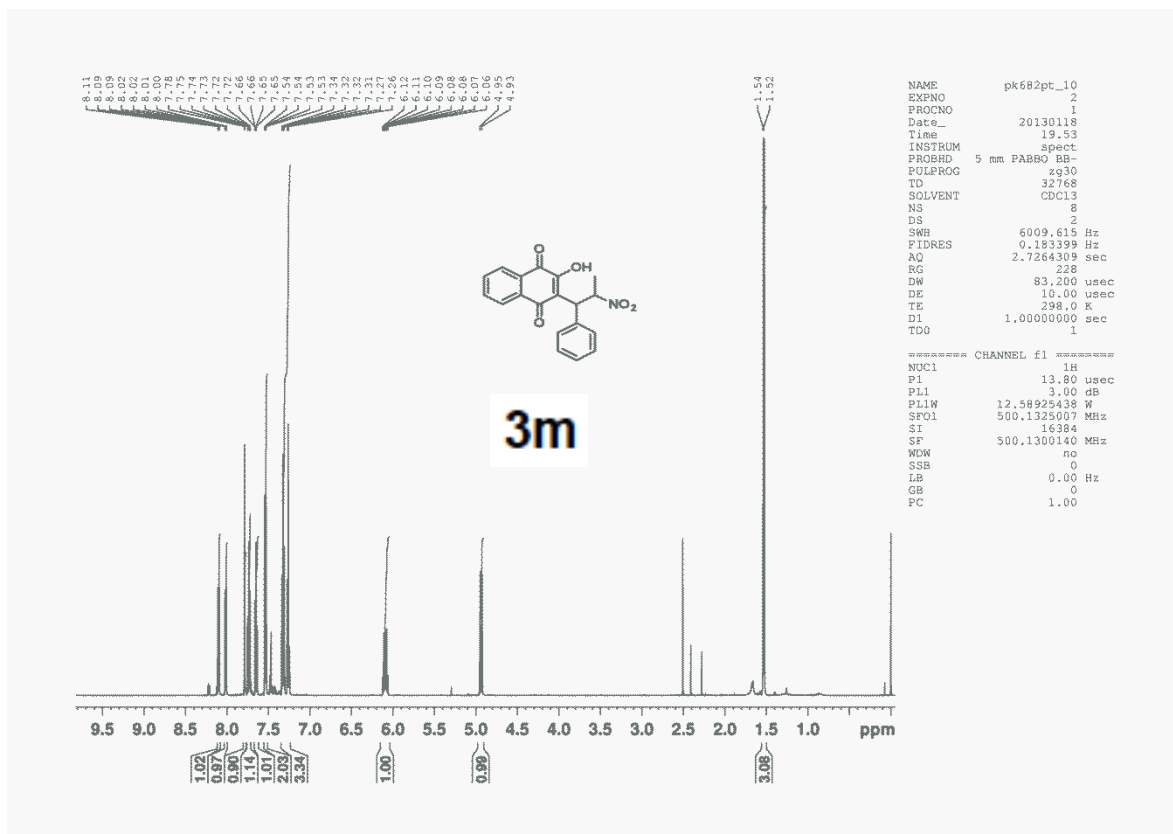






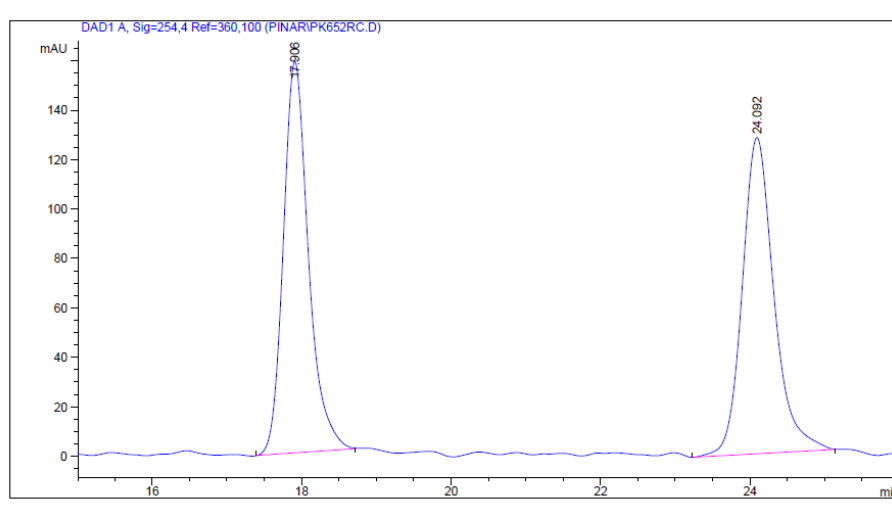






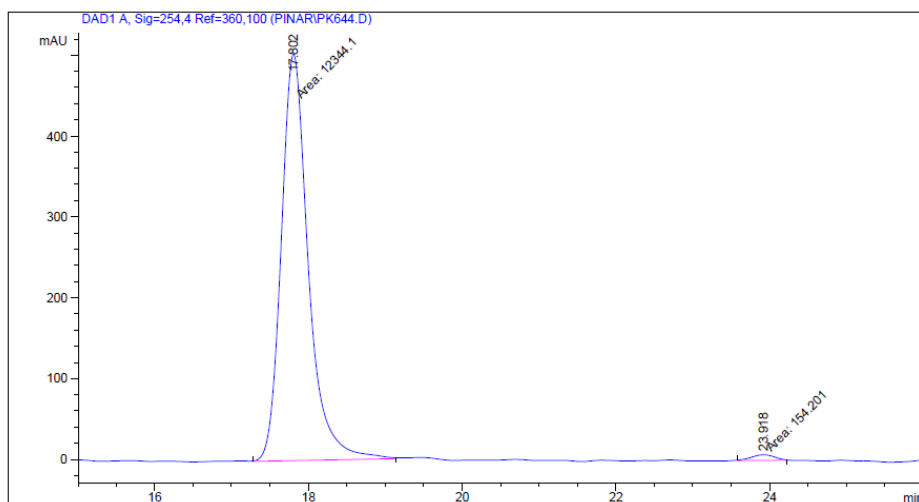
HPLC chromatograms of 3a-m

Racemic 3a



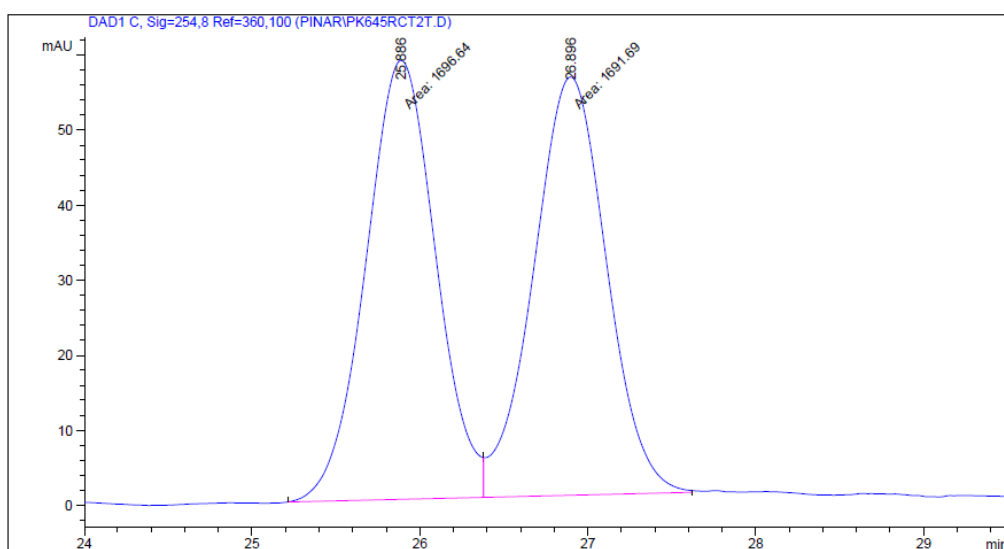
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	17.906	BB	0.3546	3704.31226	158.51859	49.4876
2	24.092	VB	0.4464	3781.01489	128.08470	50.5124

Enantioenriched 3a



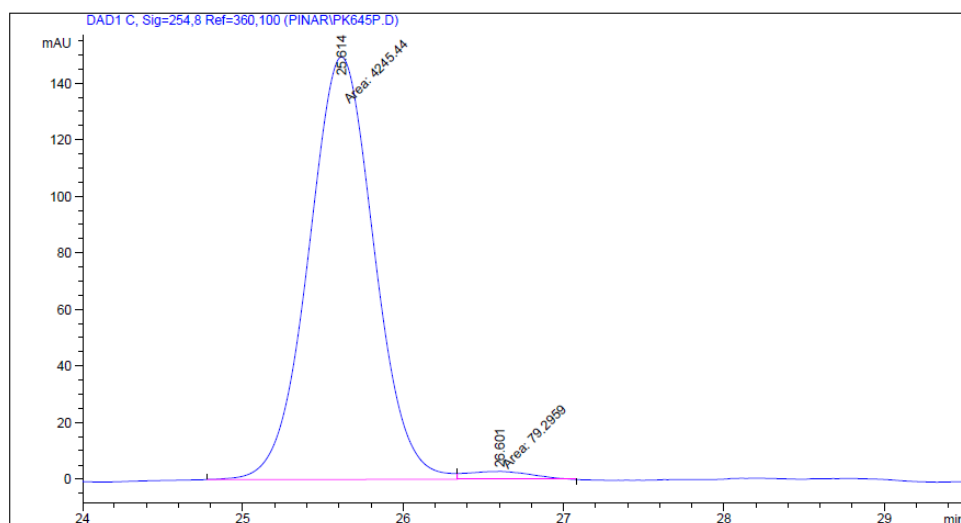
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	17.802	MM	0.4089	1.23441e4	503.11310	98.7662
2	23.918	MM	0.3580	154.20116	7.17821	1.2338

Racemic **3b**



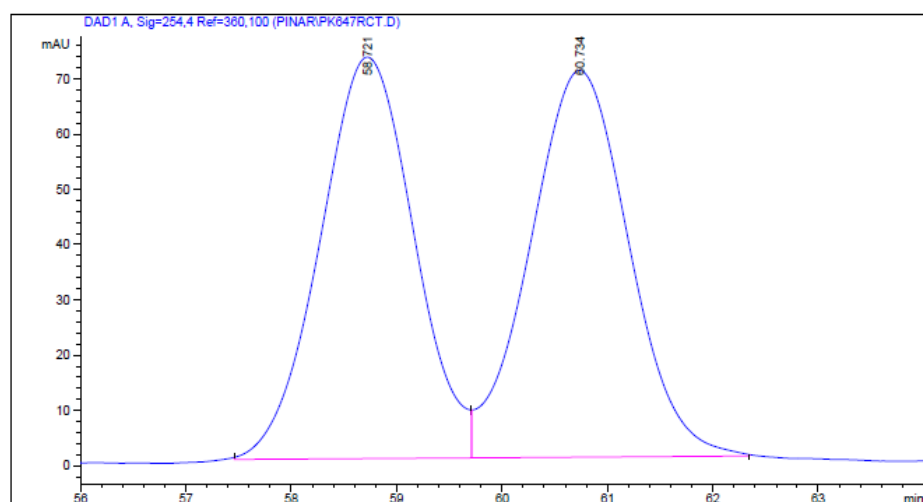
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	25.886	MF	0.4832	1696.63599	58.51677	50.0730
2	26.896	FM	0.5055	1691.68848	55.77486	49.9270

Enantioenriched **3b**



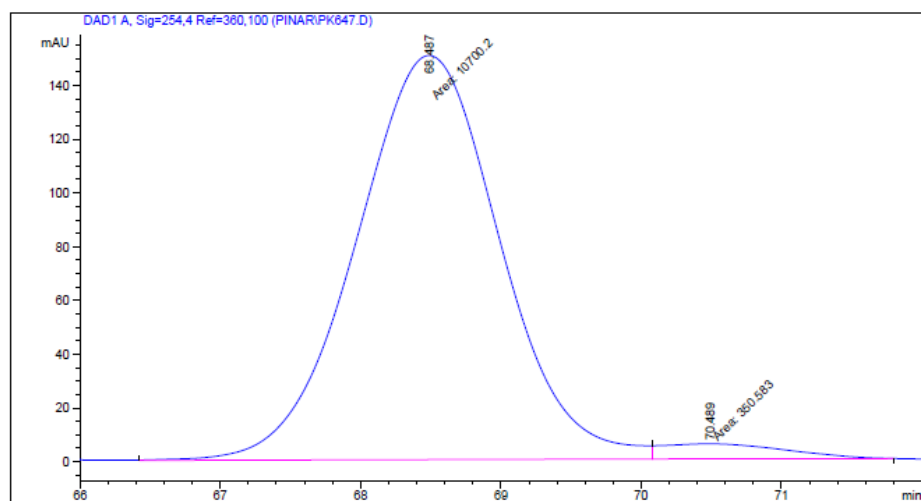
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	25.614	MF	0.4735	4245.44092	149.44862	98.1665
2	26.601	FM	0.4632	79.29591	2.85333	1.8335

Racemic **3c**⁴



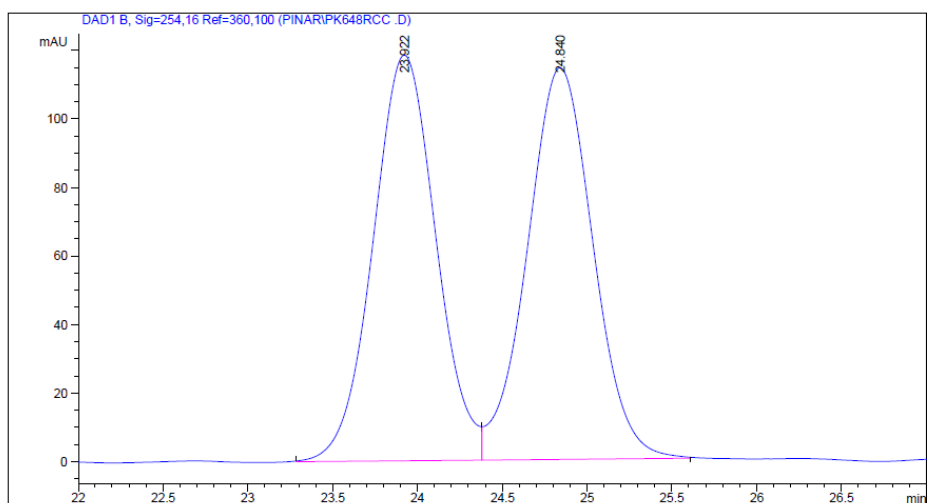
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	58.721	BV	0.9519	4486.43896	72.70438	49.5231
2	60.734	VB	1.0038	4572.84912	70.03064	50.4769

Enantioenriched **3c**



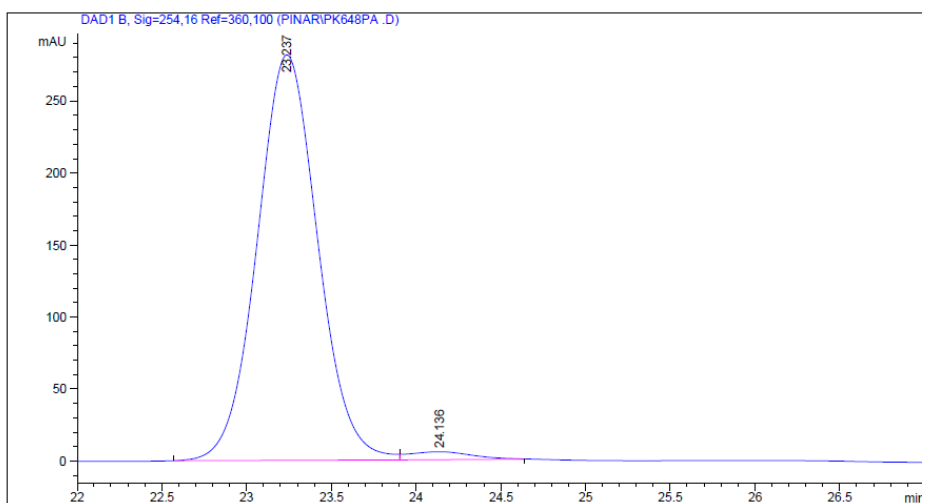
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	68.487	MF	1.1854	1.07002e4	150.44484	96.8275
2	70.489	FM	1.0188	350.58313	5.73535	3.1725

Racemic **3d**



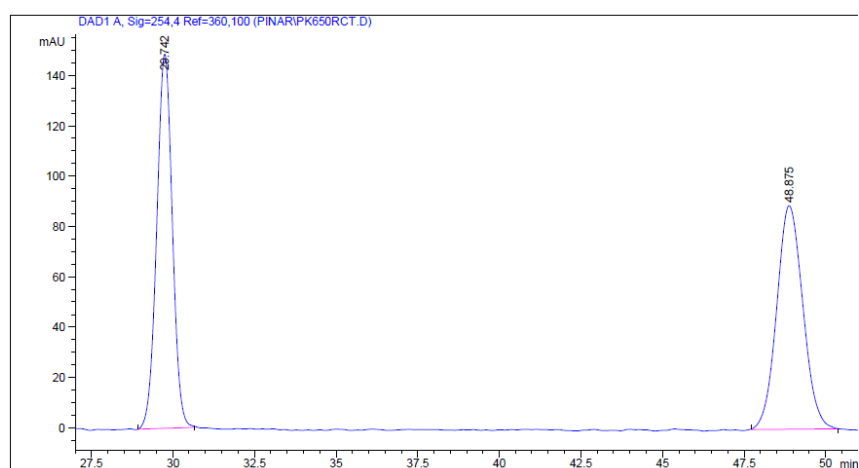
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	23.922	BV	0.3987	3051.84570	118.41208	49.7101
2	24.840	VB	0.4191	3087.43848	114.32726	50.2899

Enantioenriched **3d**



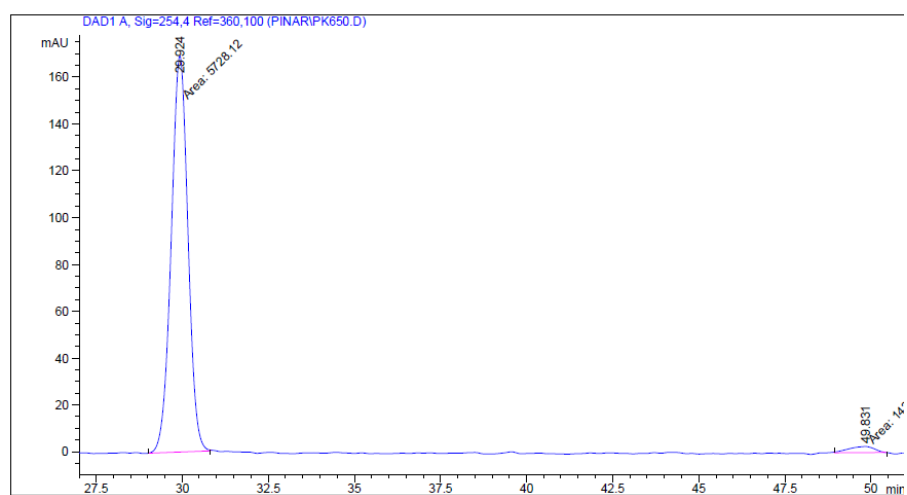
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	23.237	BB	0.3844	7013.01660	281.79114	97.9397
2	24.136	BB	0.3789	147.52817	5.64431	2.0603

Racemic **3e**



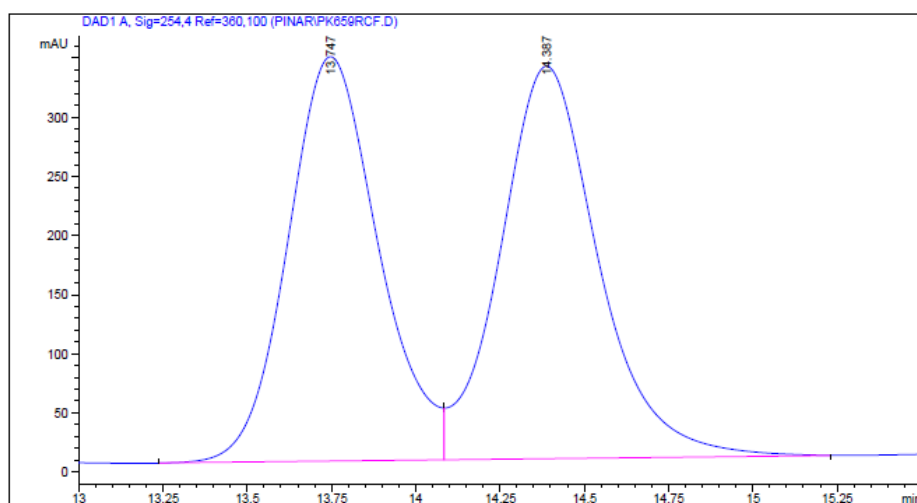
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	29.742	BB	0.5158	4947.71191	148.83781	49.8746
2	48.875	BB	0.8779	4972.58984	88.98877	50.1254

Enantioenriched **3e**



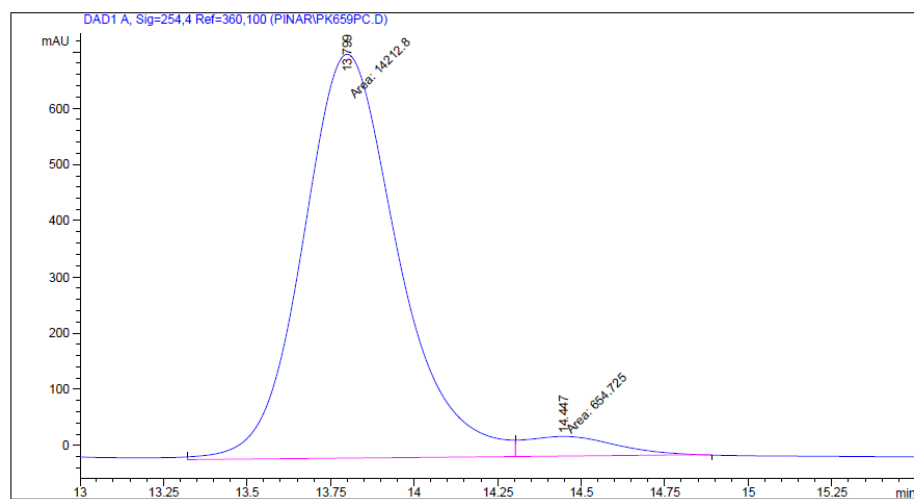
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	29.924	MM	0.5640	5728.11670	169.28139	97.5633
2	49.831	MM	0.8819	143.06218	2.70373	2.4367

Racemic **3f**



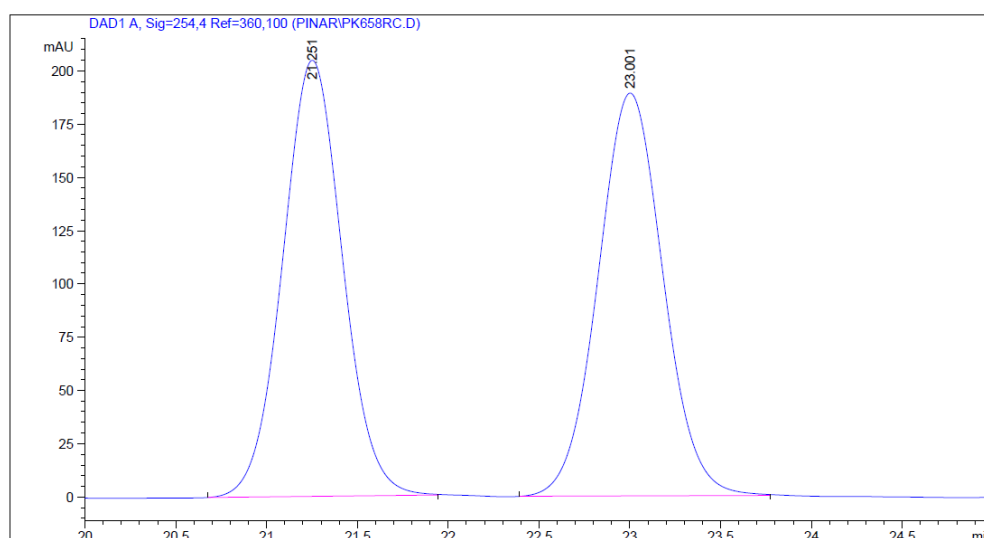
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	13.747	BV	0.2839	6386.39697	341.98383	48.6776
2	14.387	VB	0.3047	6733.39502	331.94006	51.3224

Enantioenriched **3f**



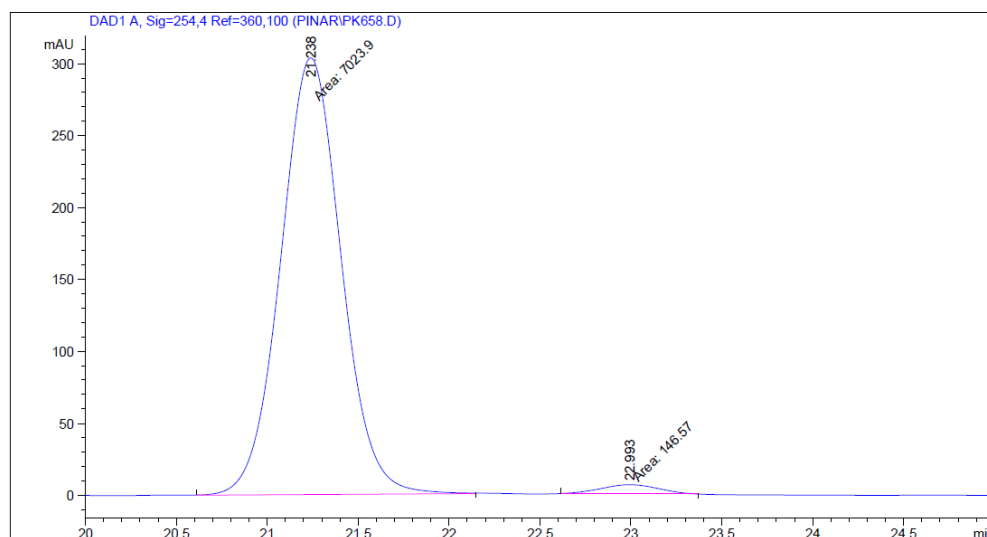
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	13.799	MF	0.3295	1.42128e4	718.94635	95.5963
2	14.447	FM	0.3105	654.72528	35.14039	4.4037

Racemic **3g**



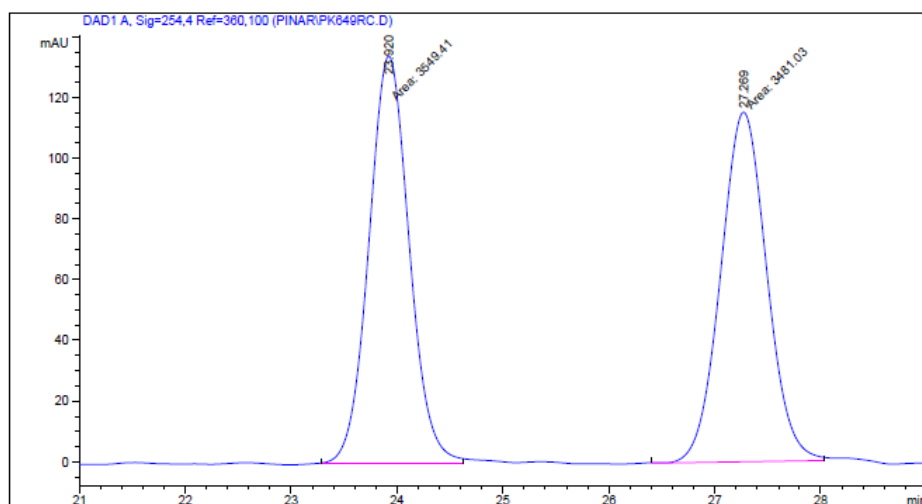
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	21.251	BB	0.3556	4697.72656	204.80247	49.8956
2	23.001	BB	0.3870	4717.39258	189.12001	50.1044

Enantioenriched **3g**



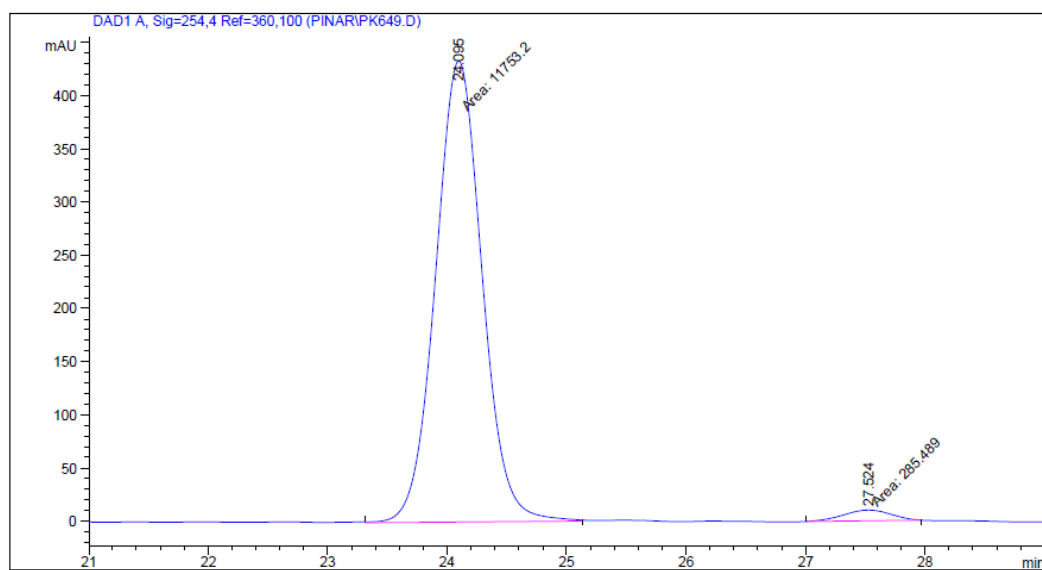
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	21.238	MM	0.3854	7023.89551	303.71033	97.9559
2	22.993	MM	0.3748	146.57024	6.51727	2.0441

Racemic **3h**



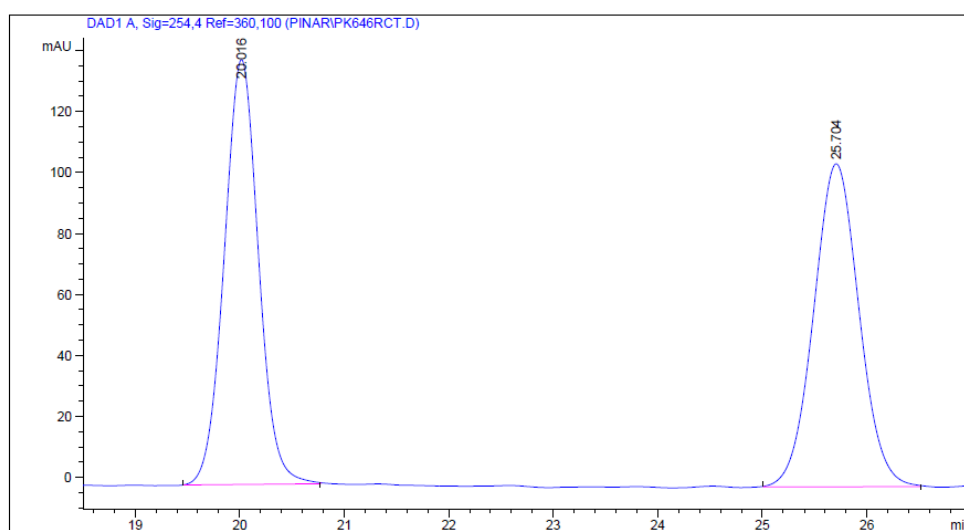
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	23.920	MM	0.4411	3549.41138	134.11685	50.4863
2	27.269	MM	0.5037	3481.03223	115.18871	49.5137

Enantioenriched **3h**



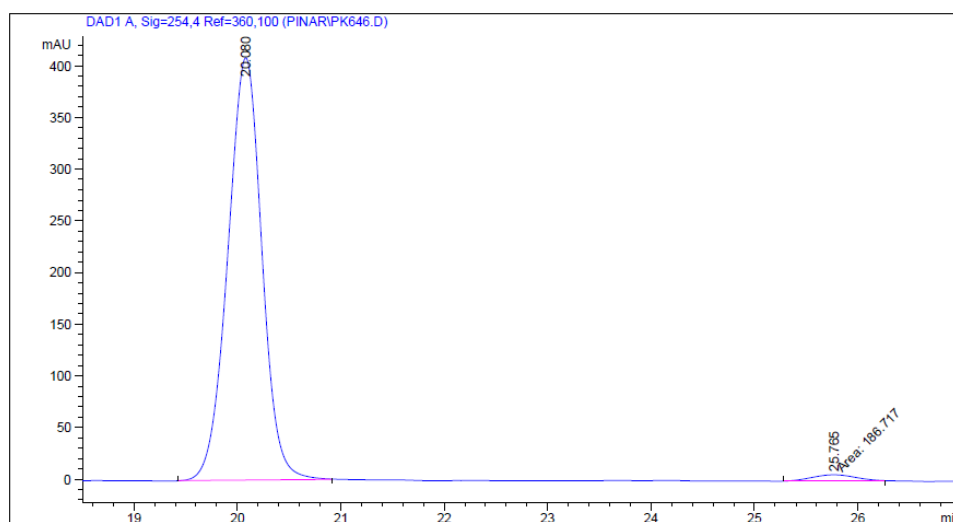
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	24.095	MM	0.4520	1.17532e4	433.41428	97.6286
2	27.524	MM	0.4689	285.48889	10.14751	2.3714

Racemic **3i**



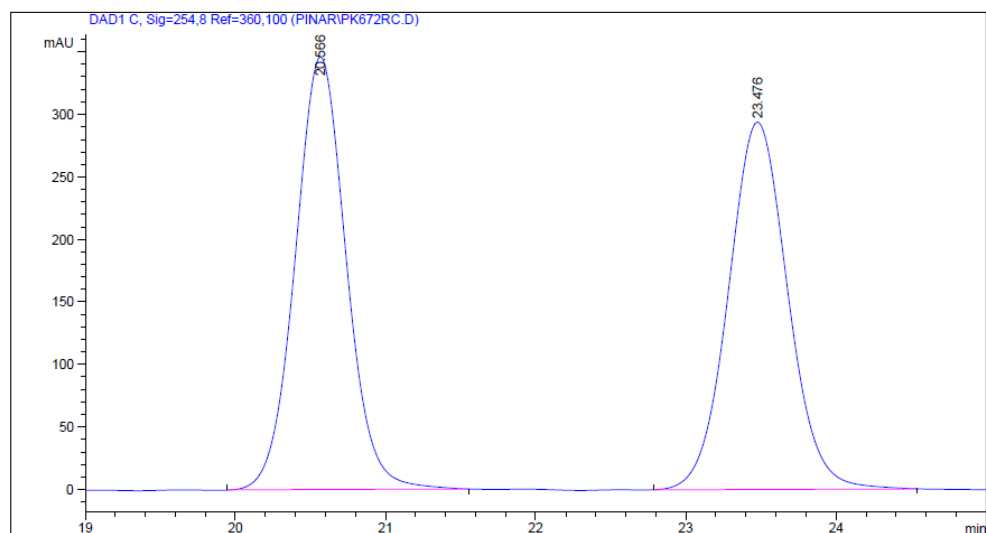
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	20.016	BB	0.3520	3181.97534	139.53908	49.8061
2	25.704	BB	0.4666	3206.75098	106.10830	50.1939

Enantioenriched **3i**



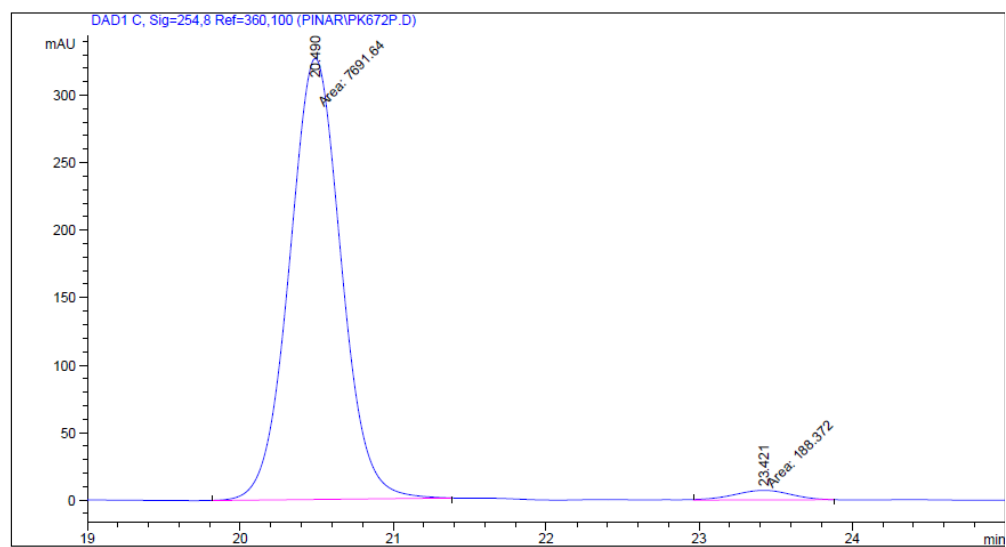
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	20.080	BB	0.3533	9366.82715	408.79440	98.0456
2	25.765	MM	0.5020	186.71684	6.19931	1.9544

Racemic **3j**



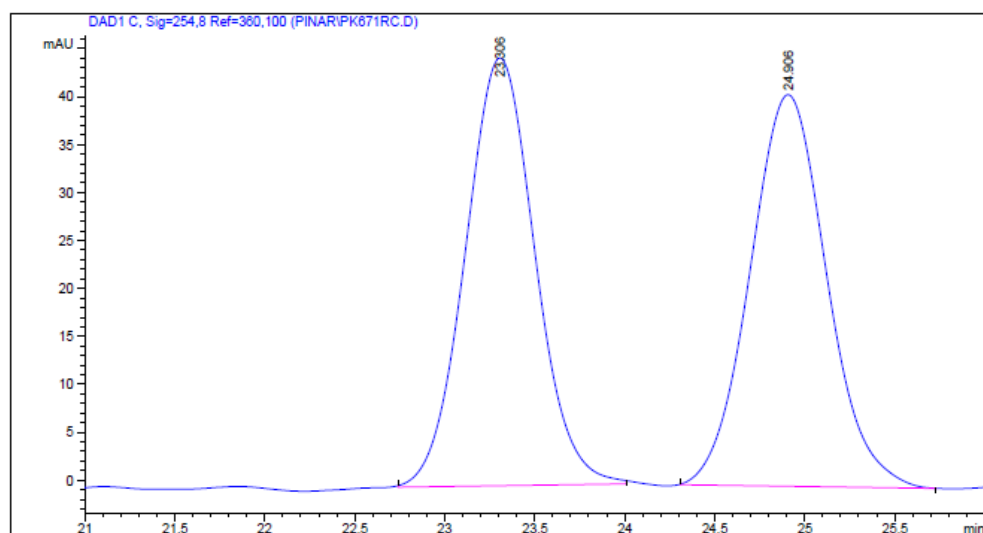
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	20.566	BB	0.3638	8186.64111	346.32211	50.3370
2	23.476	BB	0.4246	8077.02148	293.88864	49.6630

Enantioenriched **3j**



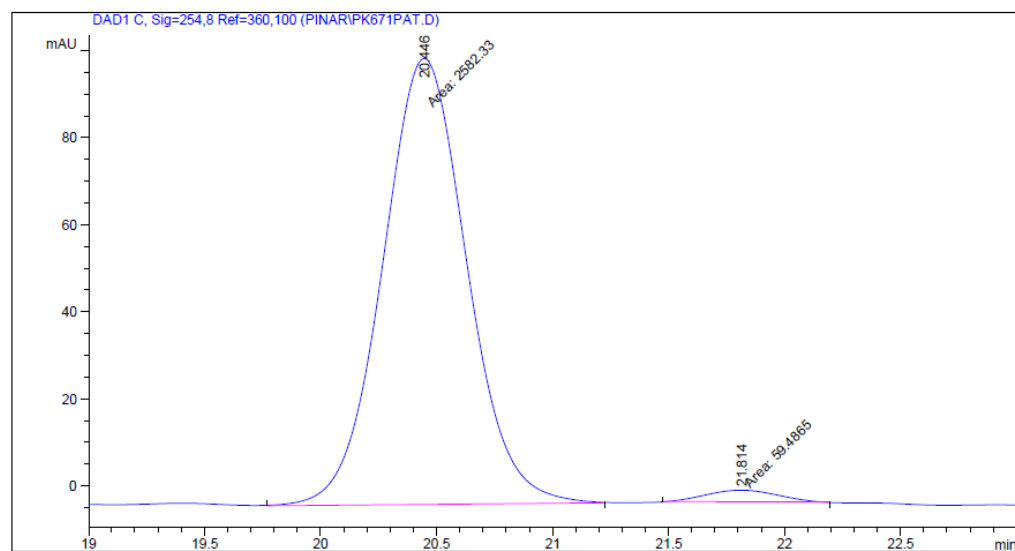
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	20.490	MM	0.3923	7691.63916	326.79651	97.6095
2	23.421	MM	0.4365	188.37187	7.19243	2.3905

Racemic **3k**



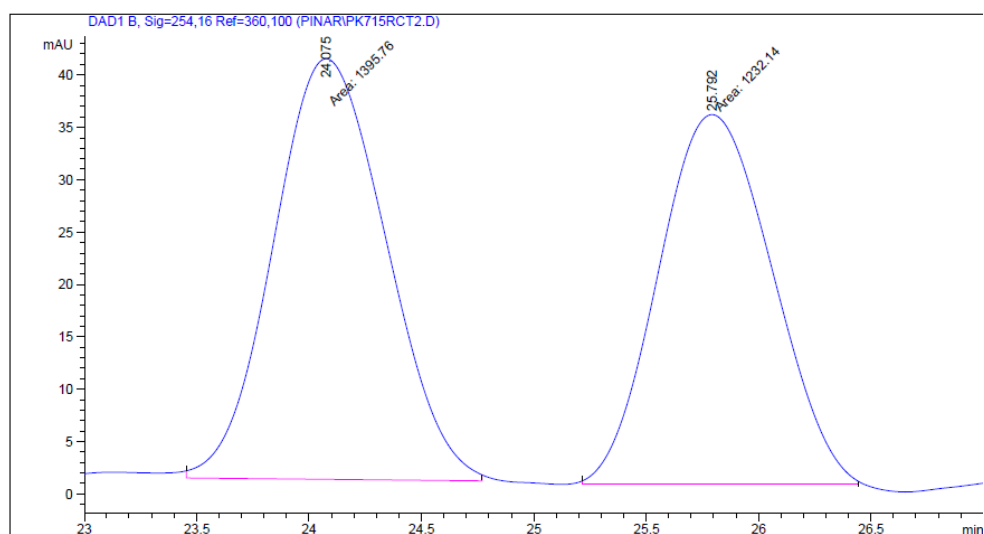
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	23.306	BB	0.4180	1201.36890	44.63844	49.8550
2	24.906	BB	0.4611	1208.35754	40.84908	50.1450

Enantioenriched **3k**



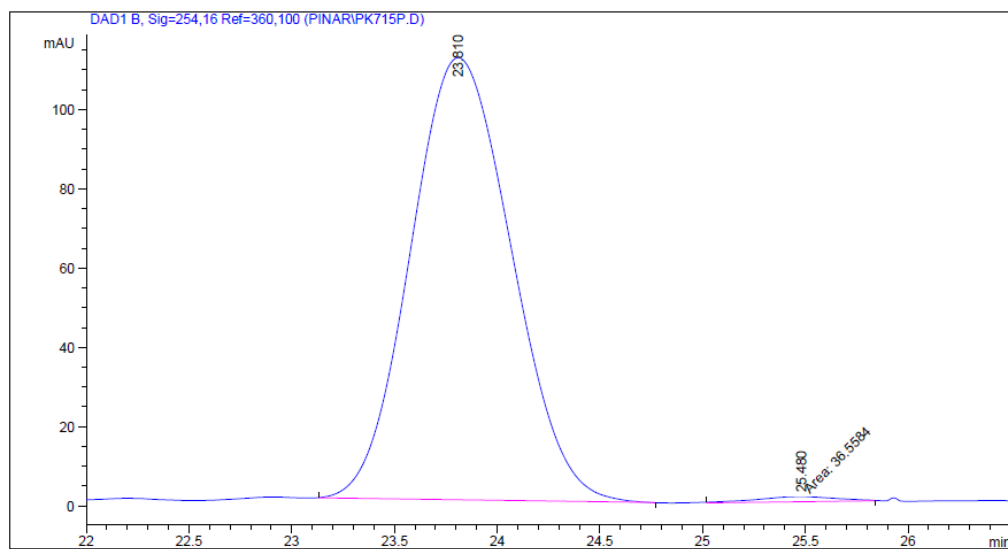
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	20.446	MM	0.4199	2582.33276	102.49802	97.7483
2	21.814	MM	0.3618	59.48645	2.74052	2.2517

Racemic **31**



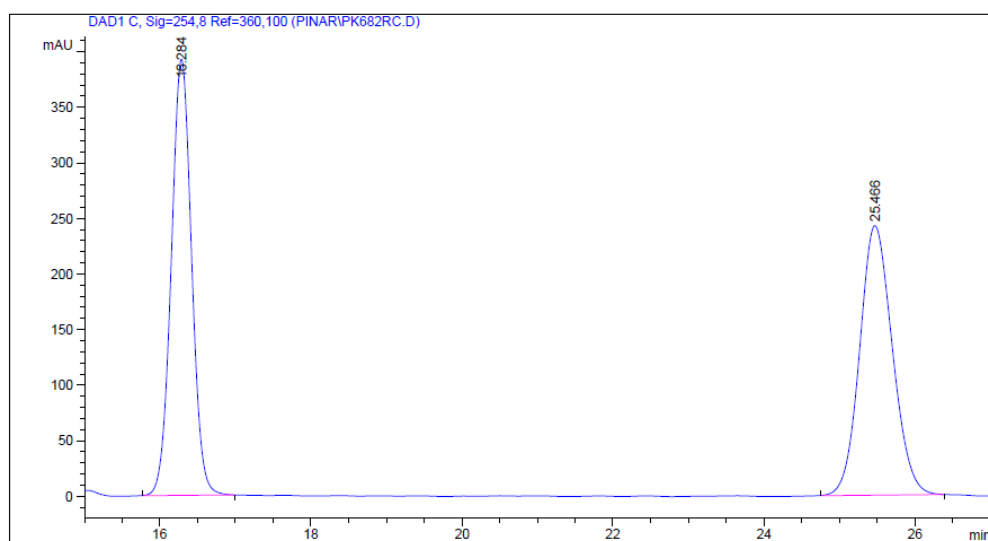
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	24.075	MM	0.5789	1395.75574	40.18335	53.1130
2	25.792	MM	0.5826	1232.14429	35.25049	46.8870

Enantioenriched **31**



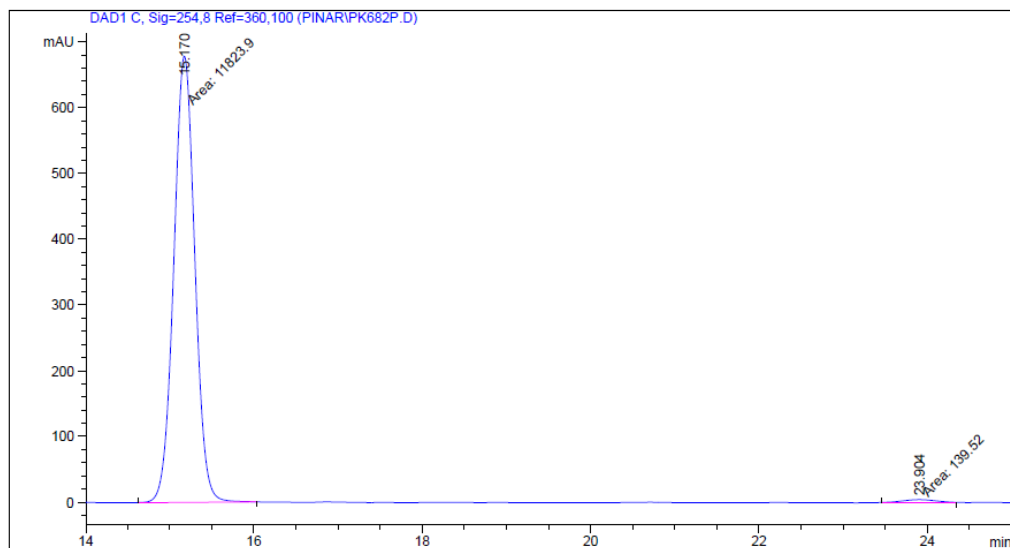
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	23.810	BB	0.5423	3808.82153	111.57813	99.0493
2	25.480	MM	0.4555	36.55839	1.33774	0.9507

Racemic **3m**



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	16.284	BB	0.2902	7346.96924	392.85043	50.1074
2	25.466	BB	0.4654	7315.48438	242.84973	49.8926

Enantioenriched **3m**



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	15.170	MM	0.2901	1.18239e4	679.20148	98.8338
2	23.904	MM	0.4884	139.51962	4.76158	1.1662

References

1. Kasaplar, P.; Riente, P.; Hartmann, C.; Pericàs, M. A., *Adv. Synth. Catal.* **2012**, 354, 2905.
2. Zhou, W.-M.; Liu, H.; Du, D.-M., *Org. Lett.* **2008**, 10, 2817.
3. Wu, R.; Chang, X.; Lu, A.; Wang, Y.; Wu, G.; Song, H.; Zhou, Z.; Tang, C., *Chem. Commun.* **2011**, 47, 5034.
4. For racemic samples, HPLC flow was 0.7 ml/min.