

Enantioselective Synthesis of *trans*-Aryl- and Heteroaryl-Substituted Cyclopropylboronates by Copper(I)-Catalyzed Reactions of Allylic Phosphates with a Diboron Derivative

Chongmin Zhong,[†] Shun Kunii,[†] Yuki Kosaka,[†] Masaya Sawamura,[†] and Hajime Ito^{*,†,‡}

[†]*Department of Chemistry, Faculty of Science, Hokkaido University, Sapporo 060-0810, Japan and*

[‡]*PRESTO, Japan Science and Technology Agency (JST), Honcho, Kawaguchi, Saitama 332-0012, Japan*

E-mail: hajito@sci.hokudai.ac.jp

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1. General and Materials.

NMR spectra were recorded on a Varian Gemini 2000 (^1H : 300 MHz, ^{13}C :75.4 MHz) spectrometer. ^1H NMR chemical shifts (δ) are reported in parts per million (ppm) downfield of TMS and are referenced relative to TMS (δ 0.00 ppm). Chemical shifts for carbon atoms are reported in parts per million downfield of TMS and are referenced to the carbon resonance of the solvent (CDCl_3 : δ 77.2 ppm). GLC analyses were conducted on a Shimadzu GC-14B equipped with a flame ionization detector. HPLC analyses were carried out using Hitachi Elite LaChrome HPLC system with L-2400 UV detector. High-resolution mass spectra were recorded on a JEOL JMS-700TZ, JMS-T100L, JMS-T100GC, Thermo Fisher Scientific LTQ-Orbitrap XL, and Thermo Fisher Scientific Exactive mass spectrometers at the Center for Instrumental Analysis, Hokkaido University. Melting points were determined on a micro melting point apparatus (Yanaco: MP-500D) using micro cover glass. All reactions were performed under a nitrogen atmosphere, using oven-dried glassware unless otherwise indicated. Column chromatography was performed using silica gel (Kanto Chemical Co., 60Å, 40-100 μm) unless otherwise indicated.

Materials were obtained from commercial suppliers and purified by the standard procedures unless otherwise noted. Solvents for reactions were purchased from commercial suppliers, degassed via three freeze-pump-thaw cycles, and further dried on MS 4A. Bis(pinacolato)diboron was purchased from AllyChem, Co., Ltd, China. (*R,R*)-QuinoxP* was provided from Nippon Chemical Industrial Co. and used as received. (*R,R*)-*i*-Pr-DuPhos, copper(I) chloride (Reagent Plus grade) and THF solution of potassium *tert*-butoxide (1.0 M or 1.2 M) were purchased from Aldrich Chemical Co. and used as received. Other phosphine ligands were purchased from appropriate commercial suppliers.

3-Aryl substituted propargylic alcohols were prepared from the corresponding aryl iodide and propargyl alcohol via Sonogashira coupling reaction.¹ All (*Z*)-3-aryl allylic alcohols, except (*Z*)-3-(2-thiophenyl)prop-2-en-1-ol, were prepared from the corresponding propargylic alcohols according to the reported method for the preparation of (*Z*)-3-phenyl-2-propenol² through Ni-catalyzed stereoselective hydrogenation [$\text{Ni}(\text{OAc})_2 \cdot 4\text{H}_2\text{O}/\text{NaBH}_4/1,2\text{-diaminoethane}/\text{H}_2$]. The *E*-configured allylic alcohols in the crude products were removed by careful silica gel column chromatography (ethyl acetate/hexane). (*Z*)-3-(2-thiophenyl)prop-2-en-1-ol was prepared via hydrogenation of the corresponding propargylic alcohol in the presence of Lindlar catalyst³ and further purified by crystallization from CH_2Cl_2 /hexane.

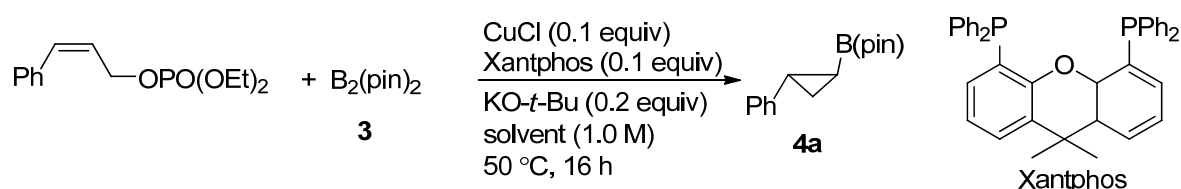
2. General Experimental Procedure.

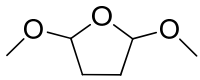
A typical procedure for the copper(I)-catalyzed enantioselective synthesis of cyclopropylboronates (Table 2) is shown as followings:

In a glove box filled with nitrogen, bis(pinacolato)diboron **3** (122 mg, 0.48 mmol), CuCl (2.0 mg, 0.02 mmol), (*R,R*)-*i*-Pr-DuPhos (10.0 mg, 0.024 mmol) and toluene (1.2 mL) were added into a vial tube with a magnetic stirrer bar. The tube was sealed with a screw cap equipped with a rubber septum and removed out of the glove box. The mixture was stirred at room temperature for ca. 30 min to form a clear yellow solution. After an allylic phosphate **2** (0.4 mmol) was added using a syringe, a THF solution of potassium *tert*-butoxide (1.2 M, 0.4 mmol) was then added dropwise with stirring at room temperature. After the reaction was complete, the reaction mixture was passed through a Florisil short column (hexane : ethyl acetate = 80:20). After evaporation under reduced pressure, the residue was subjected to silica gel column chromatography to obtain the product (typically, hexane : ethyl acetate = 95 : 5).

3. Detailed Investigation on Impact of Reaction Conditions and Catalysts.

Table S1. Solvent Effect on the Cyclopropylboronate Formation ^a



entry	solvent	GC conv. (%)	GC yield (%)
1	THF	85	63
2	Et ₂ O	62	46
3	dioxane	75	54
4	DME	68	47
5		76	50
6	CH ₂ ClCH ₂ Cl	21	9
7	CH ₂ Cl ₂	6	trace
8	CH ₃ CN	27	14
9	acetone	38	17
10	DMI	66	36

^a Phosphate (0.2 mmol), **3** (0.3 mmol), CuCl (0.02 mmol), Xantphos (0.02 mmol), KO-*t*-Bu (1.0 M THF solution, 0.04 mmol), solvent (0.2 mL).

Table S2. Effect of Base on the Cyclopropylboronate Formation ^a

$\text{Ph}-\text{CH}=\text{CH}-\text{CH}_2-\text{OP(O)(OEt)}_2 + \text{B}_2(\text{pin})_2 \xrightarrow[\text{base}]{\text{CuCl (10 mol\%)}, \text{Xantphos (12 mol\%)}, \text{THF, 50 } ^\circ\text{C, 12 h}} \text{Ph}-\text{CH}(\text{cyclopropyl})-\text{CH}_2-\text{B}(\text{pin})_2$ <div style="display: flex; justify-content: space-around; width: 100%;"> 3 4a </div>			
entry	additives	GC conv. (%)	GC yield (%)
1	LiCl	60	9
2	NaOAc	99	73
3	NaOMe	98	25
4	NaO- <i>t</i> -Bu	98	77
5	CsCO ₃	92	71
6	K ₃ PO ₄	60	41
7	Et ₃ N	73	57

Table S3. Effect of Copper(I) Salt and Base on the Cyclopropylboronate Formation ^a

$\text{Ph}-\text{CH}=\text{CH}-\text{CH}_2-\text{OPO(OEt)}_2 + \text{B}_2(\text{pin})_2 \xrightarrow[\text{THF, 50 } ^\circ\text{C, 16 h}]{\text{copper salt (10 mol\%)}, \text{Xantphos (x mmol\%)}, \text{KO-}t\text{-Bu (0.2 equiv)}, \text{additive (1.0 equiv)}} \text{Ph}-\text{CH}(\text{cyclopropyl})-\text{CH}_2-\text{B}(\text{pin})_2$ <div style="display: flex; justify-content: space-around; width: 100%;"> 3 4a </div>						
entry	copper salt	mol%	additive	x (mol%)	phosphate concentration (mol/L)	GC yield (%)
1	CuCl	10	NaOAc	10	0.5	75
2	CuCl	10	KO- <i>t</i> -Bu	10	0.5	77
3	CuCl	10	KO- <i>t</i> -Bu	12	0.5	78
4	CuCl	10	KO- <i>t</i> -Bu	12	0.25	78
5	CuCl	5	KO- <i>t</i> -Bu	10	0.5	64
6	CuCl	10	LiO- <i>t</i> -Bu	12	0.25	34
7	CuI	10	NaOAc	10	1.0	30
8	CuI	10	NaO- <i>t</i> -Bu	10	1.0	58
9	CuOAc	10	NaO- <i>t</i> -Bu	10	0.5	76
10	CuOAc	5	NaOAc	10	0.5	39

11	CuOAc	5	none	10	0.5	11
12	(CuOTf) ₂ ·toluene	5	KO- <i>t</i> -Bu	12	0.25	67
13	(CuOTf) ₂ ·toluene	5	LiO- <i>t</i> -Bu	12	0.25	30
14	Cu(CH ₃ CN) ₄ ·PF ₆	10	KO- <i>t</i> -Bu	12	0.25	57
15	Cu(CH ₃ CN) ₄ ·PF ₆	10	LiO- <i>t</i> -Bu	12	0.25	19
16 ^a	Cu(O- <i>t</i> -Bu)	5	none	5	0.25	3

^a Toluene was used as solvent, rt, 20 h.

Table S4. Effect of Leaving Group in the Reaction

entry	X	yield(%)	<i>trans/cis</i>
1	OP(O)(OEt) ₂	80	>99:1
2	OP(O)(O- <i>i</i> -Pr) ₂	89	>99:1
3	OCO ₂ Me	49	>99:1
4	OCOCF ₃	0	-
5	Cl	<1	-

Table S5. Effect of Achiral Phosphine Ligands in the Copper(I)-catalyzed Reaction ^a

entry	ligand	x (mol%)	GC yield (%)	<i>trans/cis</i> ^b
1	PPh ₃	24	14	93:7
2	PCy ₃	24	26	99:1
3	PBu ₃	24	40	96:4
4	P(<i>t</i> -Bu) ₃	24	20	97:3
5	dppe	12	34	93:7
6	dppp	12	52	94:6
7	dppb	12	11	94:6
8	dpph	12	2	nd

9	dppf	12	25	94:4
10	dtbpf	12	4	90:10
11	BDP	12	85	97:3

^a Phosphate (0.2 mmol), **3** (0.3 mmol), CuCl (0.02 mmol), THF (0.6 mL). ^b Determined by GC.

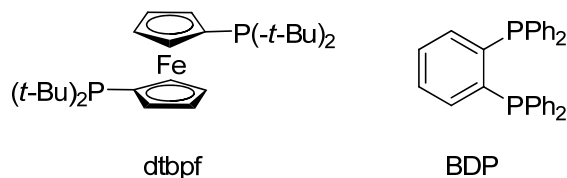
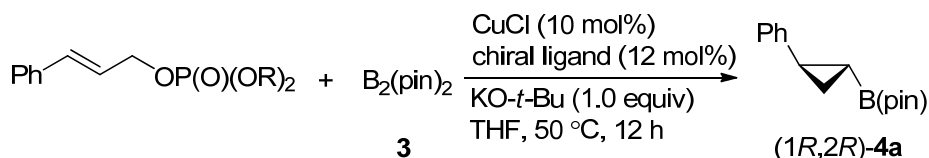
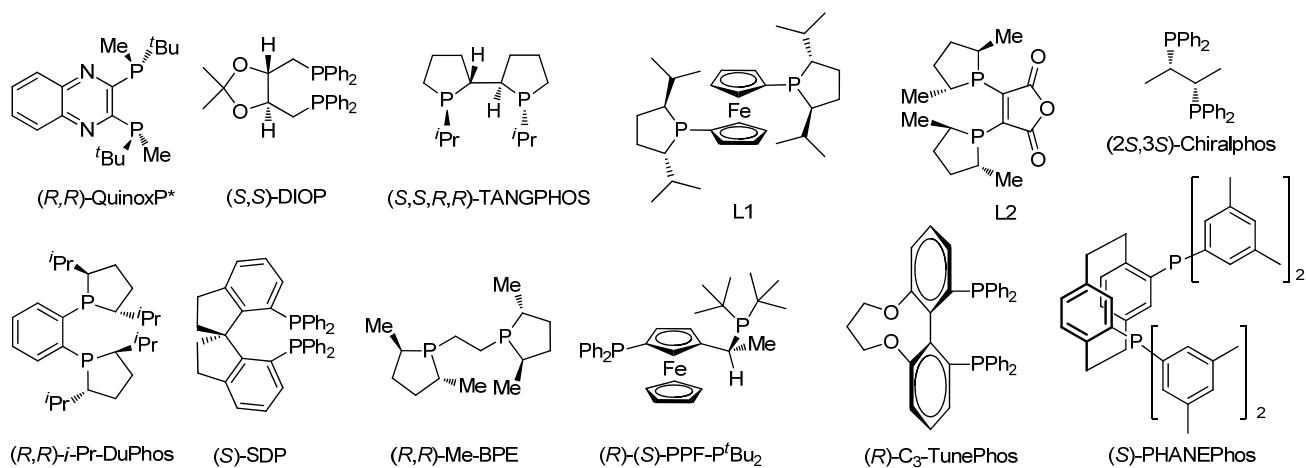


Table S6. Effect of Chiral Phosphine Ligands in the Reaction of (*E*)-Configured Substrates



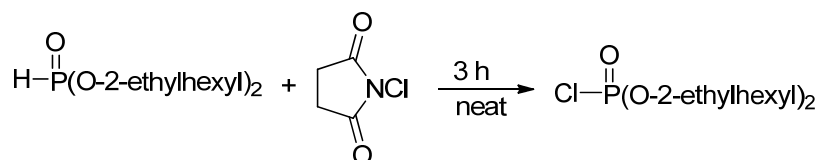
entry	R	ligand	yield (%)	<i>trans/cis</i> (ee)
1	Et	(<i>R,R</i>)-QuinoxP*	36	>99:1
2	Et	(<i>R</i>)-Segphos	20	97:3
3	<i>i</i> -Pr	(<i>S,S</i>)-DIOP	79	98:2 (−49 % ee for <i>trans</i>)
4	<i>i</i> -Pr	TANGPHOS	13	89:11
5	<i>i</i> -Pr	L1	trace	nd
6	<i>i</i> -Pr	(2 <i>S</i> ,3 <i>S</i>)-ChiralPhos	10	93:7
7	<i>i</i> -Pr	(<i>R</i>)-SDP	trace	nd
8	<i>i</i> -Pr	L2	10	88:22
9	<i>i</i> -Pr	(<i>R,R</i>)-Me-BPE	trace	nd
10	<i>i</i> -Pr	(<i>R</i>)-(<i>S</i>)-PPF-P- <i>t</i> -Bu ₂	16	49:51
11	<i>i</i> -Pr	(<i>S</i>)-PHANEPhos	trace	nd
12	<i>i</i> -Pr	(<i>R</i>)-C ₃ -TunePhos	23	91:9
13	<i>i</i> -Pr	(<i>R</i>)-BINAP	4	67:33
14	<i>i</i> -Pr	(<i>R</i>)-DTBM-Segphos	trace	nd
15	<i>i</i> -Pr	(<i>S,S</i>)-Reetz-Diphosphonite	2	nd
16	<i>i</i> -Pr	(<i>R,S</i>)-NMe ₂ -PPh ₂ -Mandyphos	trace	nd

17	<i>i</i> -Pr	(<i>R</i>)-Ph-MeOBIPHEP	7	71:29
18	<i>i</i> -Pr	(<i>R,R</i>)-BDPP	49	84:16 (−40% ee for trans)
19	<i>i</i> -Pr	(<i>S</i>)-BINAPHANE	27	81:19
20	<i>i</i> -Pr	Trost ligand	trace	nd
21	<i>i</i> -Pr	(<i>R,R</i>)-Me-DuPhos	33	73:27
22	<i>i</i> -Pr	(<i>R,R</i>)-Et-DuPhos	23	63:37
23	CH ₂ Ph	(<i>R,R</i>)-QuinoxP*	76	75:25 (73% ee for trans)
24	CH ₂ Ph	(<i>R,R</i>)- <i>i</i> -Pr-DuPhos	87	7:93 (25% ee for trans)



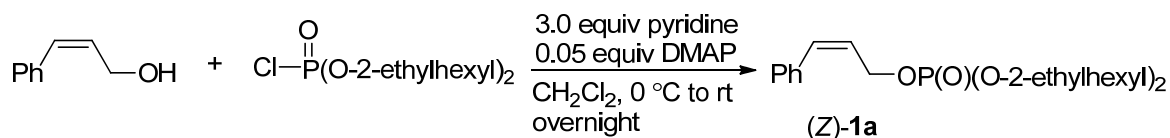
4. Substrate Synthesis and Characterization.

4.1. Preparation of bis(2-ethylhexyl) phosphorochloridate.



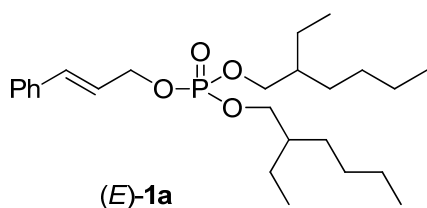
N-chlorosuccinimide (36 mmol, 4.807 g) was added to a dry 50 ml round bottom flask equipped with a magnetic stirrer bar and capped with a septum. The flask was evacuated and filled with argon using a vacuum line through a needle. The flask was cooled to $-78\text{ }^{\circ}\text{C}$ and bis(2-ethylhexyl) phosphonate (30 mmol, 9.193 g) was added dropwise. The flask was removed from the cooling bath and stirred at room temperature for 3 h. It should be careful that the reaction is strongly exothermic and that the flask should be connected to an argon line. The material was then diluted with dry hexane and filtrated through a cannula. The solid residue was washed twice with hexane. The combined solution was concentrated under reduced pressure to yield the product in 97% yield as a colorless oil. ^1H NMR (300 MHz, CDCl_3): δ 0.85–0.95 (m, 12H), 1.20–1.49 (m, 16H), 1.56–1.69 (m, 2H), 4.02–4.19 (m, 4H). ^{13}C NMR (75.4 MHz, CDCl_3): δ 10.9 (d, $^5J = 2.3\text{ Hz}$, CH_3), 14.0 (CH_3), 22.9 (CH_2), 23.2 (CH_2), 28.9 (d, $^4J = 2.3\text{ Hz}$, CH_2), 29.9 (CH_2), 39.9 (d, $^3J = 8.6\text{ Hz}$, CH), 71.8 (d, $^2J = 8.0\text{ Hz}$, CH_2). HRMS–ESI (m/z): $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{16}\text{H}_{35}\text{ClO}_3\text{P}$ 341.20123, found 341.20195.

4.2 Synthesis of (Z)-1a, (E)-1a, (E)-2a, (Z)-1a-o.

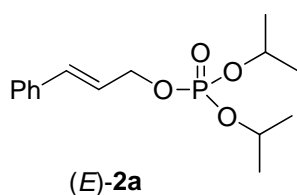


In a 50 ml dry two-neck flask equipped with a magnetic bar was added DMAP (0.042 mmol, 51 mg). The flask was then evacuated and back-filled with argon three times. Dry dichloromethane (10 ml), pyridine (24.9 mmol, 2.0 ml) and (Z)-3-phenylpropenol (8.3 mmol, 1.114 g) were added in turn to the flask. The reaction mixture was cooled to $0\text{ }^{\circ}\text{C}$ and then bis(2-ethylhexyl) phosphorochloridate (8.3 mmol, 2.286 g) was added dropwise. After the reaction was slowly warmed to room temperature through overnight with stirring, it was quenched with water (10 ml) at $0\text{ }^{\circ}\text{C}$. The organic layer was separated and the water layer was extracted with hexane ($3\times 15\text{ ml}$). The combined organic layer was then washed with water (50 ml). The solution was concentrated *in vacuo*, which afforded an oil that was

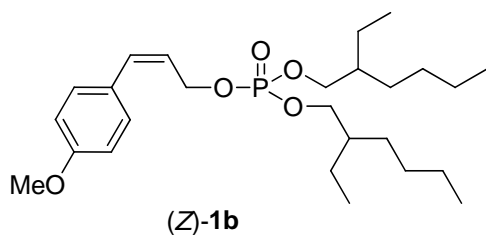
purified by flash chromatography (10:90 ethyl acetate : hexane) to give (*Z*)-**1a** (2.68 g, 74%) as a colorless oil. ^1H NMR (300 MHz, CDCl_3): δ 0.84–0.92 (m, 12H), 1.22–1.45 (m, 16H), 1.48–1.60 (m, 2H), 3.91–3.97 (m, 4H), 4.82 (ddd, $J = 8.3, 6.6, 1.7$ Hz, 2H), 5.88 (dt, $J = 11.8, 6.3$ Hz, 1H), 6.67 (d, $J = 11.8$ Hz, 1H), 7.18–7.22 (m, 2H), 7.25–7.39 (m, 3H). ^{13}C NMR (75.4 MHz, CDCl_3): δ 10.9 (CH_3), 14.1 (CH_3), 23.0 (CH_2), 23.3 (CH_2), 28.9 (d, $^4J = 1.1$ Hz, CH_2), 29.9 (CH_2), 40.2 (d, $^3J = 7.4$ Hz, CH), 64.2 (d, $^2J = 5.7$ Hz, CH_2), 69.9 (d, $^2J = 6.3$ Hz, CH_2), 126.8 (d, $^3J = 7.4$ Hz, CH), 127.9 (CH), 128.6 (CH), 129.0 (CH), 133.1 (d, $^4J = 1.1$ Hz, CH), 136.1 (C). HRMS–ESI (m/z): $[\text{M}+\text{Na}]^+$ calcd. for $\text{C}_{25}\text{H}_{43}\text{ClO}_4\text{NaP}$ 461.27967, found 461.27900.



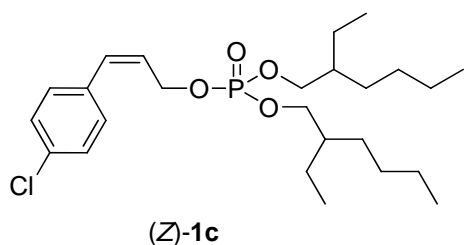
^1H NMR (300 MHz, CDCl_3): δ 0.84–0.92 (m, 12H), 1.18–1.50 (m, 16H), 1.51–1.62 (m, 2H), 3.88–4.05 (m, 4H), 4.70 (ddd, $J = 8.0, 6.3, 1.0$ Hz, 2H), 6.31 (dt, $J = 15.8, 6.3$ Hz, 1H), 6.68 (d, $J = 15.8$ Hz, 1H), 7.23–7.42 (m, 5H). ^{13}C NMR (75.4 MHz, CDCl_3): δ 10.9, 14.1, 23.0, 23.3, 28.9, 29.9, 40.2 (d, $J = 7.4$ Hz), 68.0 (d, $J = 5.3$ Hz), 69.9 (d, $J = 6.4$ Hz), 123.9 (d, $J = 6.7$ Hz), 126.9, 128.4, 128.9, 134.0, 136.3. HRMS–ESI (m/z): $[\text{M}+\text{Na}]^+$ calcd. for $\text{C}_{25}\text{H}_{43}\text{O}_4\text{NaP}$ 461.27967, found 461.27877.



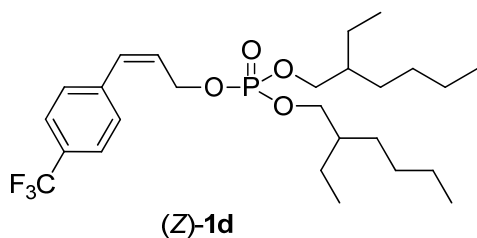
^1H NMR (300 MHz, CDCl_3): δ 1.35 (m, 12H), 4.61–4.73 (m, 4H), 6.31 (dt, $J = 15.9, 6.0$ Hz, 1H), 6.68 (d, $J = 16.0$ Hz, 1H), 7.23–7.42 (m, 5H). ^{13}C NMR (75.4 MHz, CDCl_3): δ 23.7 (d, $J = 5.0$ Hz, CH_3), 67.7 (d, $J = 5.6$ Hz), 72.5 (d, $J = 6.0$ Hz), 124.0 (d, $J = 7.4$ Hz), 126.8, 128.3, 128.8, 133.6, 136.3. HRMS–EI (m/z): $[\text{M}]^+$ calcd. for $\text{C}_{15}\text{H}_{23}\text{O}_4\text{P}$ 298.13340, found 298.13316.



^1H NMR (300 MHz, CDCl_3): δ 0.84–0.93 (m, 12H), 1.20–1.48 (m, 16H), 1.48–1.61 (m, 2H), 3.82 (s, 3H), 3.83–4.01 (m, 4H), 4.82 (ddd, $J = 8.1, 6.6, 1.6$ Hz, 2H), 5.77 (dt, $J = 11.5, 6.5$ Hz, 1H), 6.60 (d, $J = 11.5$ Hz, 1H), 6.85–6.91 (m, 2H), 7.12–7.18 (m, 2H). ^{13}C NMR (75.4 MHz, CDCl_3): δ 10.9, 14.1, 23.0, 23.3, 28.9 (d, $J = 0.9$ Hz), 29.9, 40.1 (d, $J = 7.3$ Hz), 55.4, 64.3 (d, $J = 5.3$ Hz), 69.9 (d, $J = 6.6$ Hz), 114.0, 125.0 (d, $J = 7.3$ Hz), 128.8, 130.3, 132.7, 159.4. HRMS–ESI (m/z): $[\text{M}+\text{Na}]^+$ calcd. for $\text{C}_{26}\text{H}_{45}\text{O}_5\text{NaP}$ 491.29023, found 491.28999.

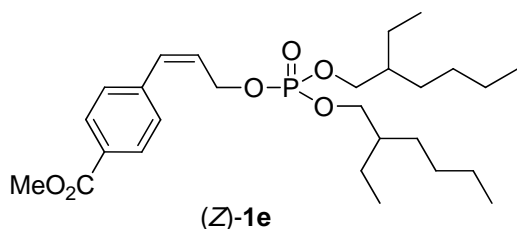


^1H NMR (300 MHz, CDCl_3): δ 0.79–0.97 (m, 12H), 1.18–1.47 (m, 16H), 1.48–1.61 (m, 2H), 3.88–4.01 (m, 4H), 4.77 (ddd, $J = 8.1, 6.6, 1.6$ Hz, 2H), 5.90 (dt, $J = 11.7, 6.6$ Hz, 1H), 6.61 (d, $J = 11.7$ Hz, 1H), 7.11–7.17 (m, 2H), 7.30–7.35 (m, 2H). ^{13}C NMR (75.4 MHz, CDCl_3): δ 10.9, 14.1, 23.0, 23.3, 28.9 (d, $J = 1.1$ Hz), 29.9, 40.1 (d, $J = 7.4$ Hz), 63.9 (d, $J = 5.1$ Hz), 70.0 (d, $J = 6.4$ Hz), 127.5 (d, $J = 7.2$ Hz), 128.8, 130.2, 132.0 (d, $J = 1.1$ Hz), 133.8, 134.5. HRMS–ESI (m/z): $[\text{M}+\text{Na}]^+$ calcd. for $\text{C}_{25}\text{H}_{42}\text{O}_4\text{ClNaP}$ 495.24069, found 495.23988.

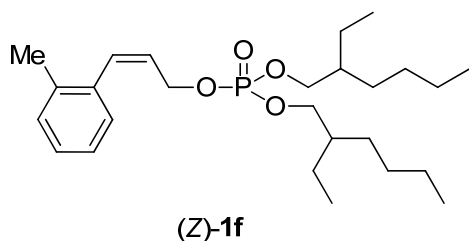


^1H NMR (300 MHz, CDCl_3): δ 0.80–0.97 (m, 12H), 1.18–1.47 (m, 16H), 1.48–1.61 (m, 2H), 3.88–4.01 (m, 4H), 4.78 (ddd, $J = 8.1, 6.7, 1.6$ Hz, 2H), 6.00 (dt, $J = 11.8, 6.7$ Hz, 1H), 6.69 (d, $J = 11.8$

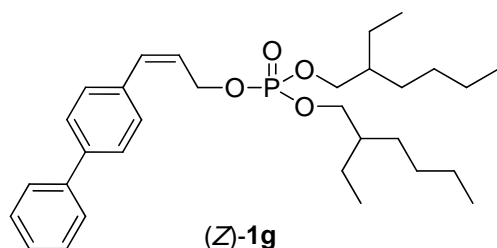
Hz, 1H), 7.32 (d, $J = 8.4$ Hz, 2H), 7.62 (d, $J = 8.4$ Hz, 2H). ^{13}C NMR (75.4 MHz, CDCl_3): δ 10.9, 14.1, 23.1, 23.3, 28.9 (d, $J = 1.1$ Hz), 29.9, 40.1 (d, $J = 7.4$ Hz), 63.8 (d, $J = 5.1$ Hz), 70.0 (d, $J = 6.4$ Hz), 124.3 (q, $J = 271.5$ Hz), 125.6 (q, $J = 11.3$ Hz), 128.9, 129.0, 129.2, 131.8, 139.6 (d, $J = 1.6$ Hz). HRMS–ESI (m/z): $[\text{M}+\text{Na}]^+$ calcd. for $\text{C}_{26}\text{H}_{42}\text{O}_4\text{F}_3\text{NaP}$ 529.26705, found 529.26610.



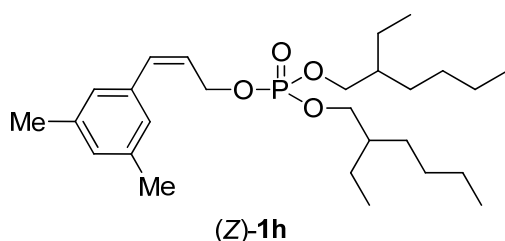
^1H NMR (300 MHz, CDCl_3): δ 0.84–0.92 (m, 12H), 1.20–1.47 (m, 16H), 1.46–1.60 (m, 2H), 3.93 (s, 3H), 3.91–3.99 (m, 4H), 4.80 (ddd, $J = 8.1, 6.4, 1.6$ Hz, 2H), 5.98 (dt, $J = 11.8, 6.4$ Hz, 1H), 6.69 (d, $J = 11.8$ Hz, 1H), 7.24–7.29 (m, 2H), 8.00–8.05 (m, 2H). ^{13}C NMR (75.4 MHz, CDCl_3): δ 10.9, 14.1, 23.0, 23.3, 28.9, 29.9, 40.1 (d, $J = 7.4$ Hz), 52.3, 63.9 (d, $J = 5.1$ Hz), 70.0 (d, $J = 6.3$ Hz), 128.9 (d, $J = 7.4$ Hz), 128.9, 129.4, 130.0, 132.1, 140.6, 167.0. HRMS–ESI (m/z): $[\text{M}+\text{Na}]^+$ calcd. for $\text{C}_{27}\text{H}_{45}\text{O}_6\text{NaP}$ 519.28515, found 519.28425.



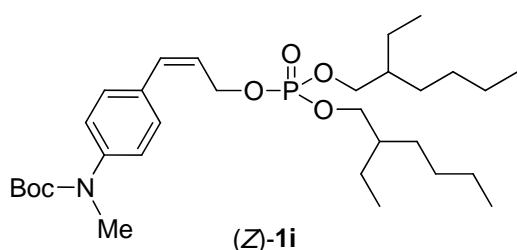
^1H NMR (300 MHz, CDCl_3): δ 0.84–0.92 (m, 12H), 1.14–1.47 (m, 16H), 1.47–1.60 (m, 2H), 2.27 (s, 3H), 3.86–3.99 (m, 4H), 4.67 (ddd, $J = 8.0, 6.6, 1.4$ Hz, 2H), 5.92 (dt, $J = 11.4, 6.6$ Hz, 1H), 6.73 (d, $J = 11.4$ Hz, 1H), 7.03–7.09 (m, 2H), 7.12–7.21 (m, 2H). ^{13}C NMR (75.4 MHz, CDCl_3): δ 10.9, 14.1, 19.9, 23.0, 23.3, 28.9, 29.9, 40.1 (d, $J = 7.4$ Hz), 64.2 (d, $J = 5.1$ Hz), 69.9 (d, $J = 6.4$ Hz), 125.9, 126.8 (d, $J = 7.2$ Hz), 128.1, 129.1, 130.2, 132.6, 135.1, 136.5. HRMS–ESI (m/z): $[\text{M}+\text{Na}]^+$ calcd. for $\text{C}_{26}\text{H}_{45}\text{O}_4\text{NaP}$ 475.29532, found 475.29449.



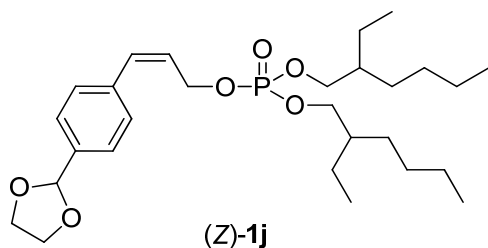
^1H NMR (300 MHz, CDCl_3 , δ): 0.88 (bt, $J = 7.4$ Hz, 12H), 1.15–1.46 (m, 16H), 1.48–1.64 (m, 2H), 3.96 (bt, $J = 5.5$ Hz, 4H), 4.87 (ddd, $J = 8.0, 6.6, 1.7$ Hz, 2H), 5.90 (dt, $J = 11.7, 6.4$ Hz, 1H), 6.70 (d, $J = 11.6$ Hz, 1H), 7.25–7.49 (m, 5H), 7.56–7.64 (m, 4H). ^{13}C NMR (75.4 MHz, CDCl_3 , δ): 10.7 (CH_3), 13.9 (CH_3), 22.8 (CH_2), 23.1 (CH_2), 28.7 (CH_2), 29.7 (CH_2), 39.9 (d, $^3J = 6.9$ Hz, CH), 64.1 (d, $^2J = 5.7$ Hz, CH_2), 69.8 (d, $^2J = 6.3$ Hz, CH_2), 126.7 (d, $^3J = 7.4$ Hz, CH), 127.0 (CH), 127.1 (CH), 127.5 (CH), 128.9 (CH), 129.3 (CH), 132.4 (CH), 134.9 (C), 140.5 (C), 140.6 (C). HRMS–ESI (m/z): $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{31}\text{H}_{47}\text{O}_4\text{PNa}$, 537.31042; found, 537.31001.



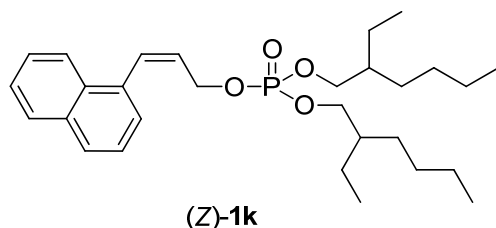
^1H NMR (300 MHz, CDCl_3 , δ): 0.88 (bt, $J = 7.4$ Hz, 12H), 1.18–1.46 (m, 16H), 1.49–1.64 (m, 2H), 2.31 (s, 6H), 3.94 (bt, $J = 5.5$ Hz, 4H), 4.83 (ddd, $J = 8.0, 6.3, 1.7$ Hz, 2H), 5.82 (dt, $J = 11.8, 6.3$ Hz, 1H), 6.60 (d, $J = 11.8$ Hz, 1H), 6.80 (s, 2H), 6.93 (s, 1H). ^{13}C NMR (75.4 MHz, CDCl_3 , δ): 10.7 (CH_3), 13.9 (CH_3), 21.1 (CH_3), 22.8 (CH_2), 23.1 (CH_2), 28.7 (d, $^4J = 1.1$ Hz, CH_2), 29.7 (CH_2), 39.9 (d, $^3J = 7.4$ Hz, CH), 64.1 (d, $^2J = 5.2$ Hz, CH_2), 69.6 (d, $^2J = 6.3$ Hz, CH_2), 126.3 (d, $^3J = 7.4$ Hz, CH), 126.6 (CH), 129.3 (CH), 132.92 (d, $^4J = 1.1$ Hz, CH), 135.8 (C), 137.9 (C). HRMS–ESI (m/z): $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{27}\text{H}_{47}\text{O}_4\text{PNa}$, 489.31042; found, 489.31000.



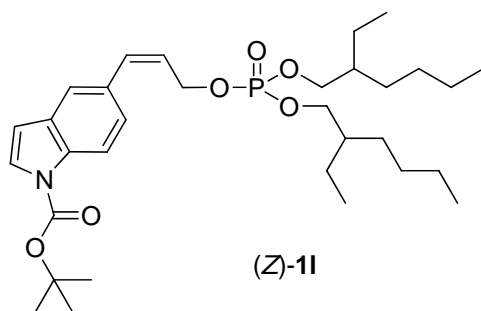
^1H NMR (300 MHz, CDCl_3 , δ): 0.88 (bt, $J = 7.1$ Hz, 12H), 1.18–1.44 (m, 16H), 1.46 (s, 9H), 1.49–1.64 (m, 2H), 3.26 (s, 3H), 3.95 (bt, 4H), 4.82 (ddd, $J = 8.0, 6.5, 1.6$ Hz, 2H), 5.85 (dt, $J = 11.8, 6.5$ Hz, 1H), 6.62 (d, $J = 11.8$ Hz, 1H), 7.13–7.26 (m, 4H). ^{13}C NMR (75.4 MHz, CDCl_3 , δ): 10.7 (CH_3), 13.9 (CH_3), 22.8 (CH_2), 23.1 (CH_2), 28.2 (CH_3), 28.7 (CH_2), 29.7 (CH_2), 37.0 (CH_3), 39.9 (d, $^3J = 7.4$ Hz, CH), 64.0 (d, $^2J = 5.2$ Hz, CH_2), 69.7 (d, $^2J = 6.9$ Hz, CH_2), 80.4 (C), 125.2 (CH), 126.4 (d, $^3J = 6.9$ Hz, CH), 129.0 (CH), 132.2 (CH), 132.8 (C), 143.2 (C), 154.7 (C). HRMS–ESI (m/z): $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{31}\text{H}_{54}\text{NO}_6\text{PNa}$, 590.35810; found, 590.35747.



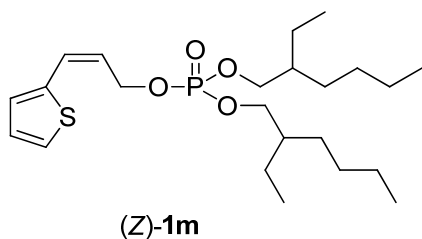
^1H NMR (300 MHz, CDCl_3 , δ): 0.83–0.94 (m, 12H), 1.17–1.46 (m, 16H), 1.49–1.61 (m, 2H), 3.86–4.01 (m, 4 H), 4.01–4.17 (m, 4H), 4.80 (ddd, $J = 8.0, 6.6, 1.7$ Hz, 2H), 5.82 (s, 1H), 5.89 (dt, $J = 11.8, 6.3$ Hz, 1H), 6.66 (d, $J = 11.8$ Hz, 1H), 7.22 (d, $J = 8.3$ Hz, 2H), 7.47 (d, $J = 8.3$ Hz, 2H). ^{13}C NMR (75.4 MHz, CDCl_3 , δ): 10.7 (CH_3), 13.8 (CH_3), 22.8 (CH_2), 23.0 (CH_2), 28.7 (d, $^4J = 1.1$ Hz, CH_2), 29.7 (CH_2), 39.9 (d, $^3J = 7.4$ Hz, CH), 63.9 (d, $^2J = 5.2$ Hz, CH_2), 65.2 (CH_2), 69.7 (d, $^2J = 6.3$ Hz, CH_2), 103.3 (CH), 126.5 (CH), 127.1 (d, $^3J = 6.9$ Hz, CH), 128.7 (CH), 132.4 (CH), 136.7 (C), 137.4 (C). HRMS–ESI (m/z): $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{28}\text{H}_{47}\text{O}_6\text{PNa}$, 533.30025; found, 533.29975.



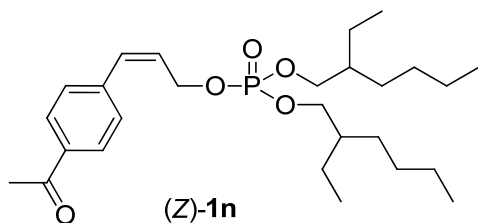
^1H NMR (300 MHz, CDCl_3): δ 0.76–0.91 (m, 12H), 1.12–1.42 (m, 16H), 1.42–1.56 (m, 2H), 3.83–3.96 (m, 4H), 4.69 (ddd, $J = 8.0, 6.6, 1.4$ Hz, 2H), 6.15 (dt, $J = 11.4, 6.6$ Hz, 1H), 7.20 (d, $J = 11.4$ Hz, 1H), 7.24–7.29 (m, 1H), 7.41–7.45 (m, 1H), 7.48–7.55 (m, 2H), 7.81 (d, $J = 8.3$ Hz, 1H), 7.83–7.90 (m, 1H), 7.92–7.98 (m, 1H). ^{13}C NMR (75.4 MHz, CDCl_3): δ 10.9, 14.1, 23.0, 23.2, 28.9 (d, $J = 1.1$ Hz), 29.9, 40.1 (d, $J = 7.4$ Hz), 64.3 (d, $J = 5.4$ Hz), 69.9 (d, $J = 6.4$ Hz), 124.9, 125.4, 126.3, 126.5, 126.9, 128.4 (d, $J = 7.2$ Hz), 128.6, 128.7, 131.6, 131.8, 133.0, 133.8. HRMS–ESI (m/z): $[\text{M}+\text{Na}]^+$ calcd. for $\text{C}_{29}\text{H}_{45}\text{O}_4\text{NaP}$ 511.29532, found 511.29445.



^1H NMR (300 MHz, CDCl_3): δ 0.87 (t, $J = 7.3$ Hz, 12H), 1.20–1.47 (m, 16H), 1.48–1.60 (m, 2H), 1.68 (s, 9H), 3.88–4.00 (m, 4H), 4.88 (ddd, $J = 8.0, 6.5, 1.6$ Hz, 2H), 5.86 (dt, $J = 11.8, 6.6$ Hz, 1H), 6.56 (dd, $J = 3.8, 0.5$ Hz, 1H), 6.77 (d, $J = 11.8$ Hz, 1H), 7.15 (dd, $J = 8.6, 1.6$ Hz, 1H), 7.37–7.39 (m, 1H), 7.61 (d, $J = 3.6$ Hz, 1H), 8.10 (d, $J = 8.6$ Hz, 1H). ^{13}C NMR (75.4 MHz, CDCl_3): δ 10.9, 14.1, 23.0, 23.3, 28.2, 28.9, 29.9, 40.1 (d, $J = 7.4$ Hz), 64.4 (d, $J = 5.6$ Hz), 69.9 (d, $J = 6.3$ Hz), 84.0, 107.5, 115.2, 121.3, 125.5, 125.8 (d, $J = 7.3$ Hz), 126.8, 130.7, 130.9, 133.5, 134.7, 149.9. HRMS–ESI (m/z): $[\text{M}+\text{Na}]^+$ calcd. for $\text{C}_{32}\text{H}_{52}\text{O}_6\text{NNaP}$ 600.34299, found 600.34252.

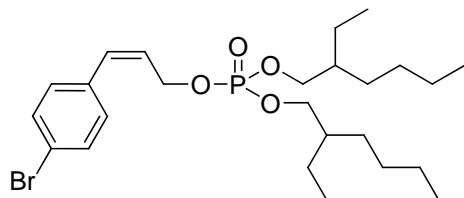


^1H NMR (300 MHz, CDCl_3): δ 0.88 (t, $J = 8.2$ Hz, 12H), 1.20–1.48 (m, 16H), 1.48–1.66 (m, 2H), 3.96 (t, $J = 6.0$ Hz, 4H), 4.92 (ddd, $J = 8.8, 6.7, 1.4$ Hz, 2H), 5.77 (dt, $J = 13.0, 6.7$ Hz, 1H), 6.67 (d, $J = 13.0$ Hz, 1H), 6.96–7.00 (m, 1H), 7.00–7.05 (m, 1H), 7.32 (d, $J = 5.5$ Hz, 1H). ^{13}C NMR (75.4 MHz, CDCl_3): δ 10.9, 14.1, 23.0, 23.3, 28.9, 29.9, 40.2 (d, $J = 7.3$ Hz), 64.5 (d, $J = 5.6$ Hz), 70.0 (d, $J = 6.6$ Hz), 124.4, 125.2 (d, $J = 7.7$ Hz), 126.8, 127.6, 128.5, 139.1. HRMS–ESI (m/z): $[\text{M}+\text{Na}]^+$ calcd. for $\text{C}_{23}\text{H}_{41}\text{O}_4\text{NaPS}$ 467.23609, found 467.23512.



^1H NMR (300 MHz, CDCl_3): δ 0.80–0.98 (m, 12H), 1.20–1.46 (m, 16H), 1.48–1.62 (m, 2H), 2.61 (s, 3H), 3.88–4.20 (m, 4H), 4.81 (ddd, $J = 7.8, 6.4, 1.7$ Hz, 2H), 5.99 (dt, $J = 11.8, 6.6$ Hz, 1H), 6.69 (d, $J = 11.8$ Hz, 1H), 7.30 (d, $J = 8.4$ Hz, 2H), 7.95 (d, $J = 8.4$ Hz, 2H). ^{13}C NMR (75.4 MHz, CDCl_3): δ 10.9,

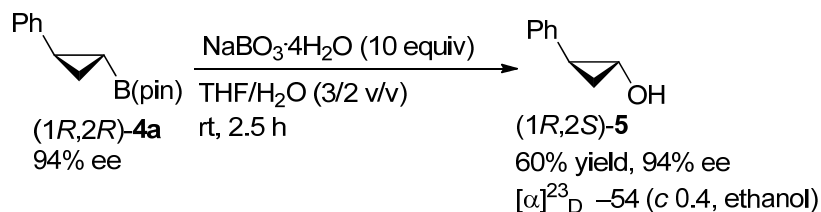
14.1, 23.0, 23.3, 26.7, 28.9 (d, $J = 1.1$ Hz), 29.9, 40.1 (d, $J = 7.3$ Hz), 63.9 (d, $J = 5.2$ Hz), 70.0 (d, $J = 6.3$ Hz), 128.7, 129.0 (d, $J = 7.4$ Hz), 129.1, 132.0, 136.3, 140.8, 197.8. HRMS–ESI (m/z): $[M+Na]^+$ calcd. for $C_{27}H_{45}O_5NaP$ 503.29023, found 503.28950.



(Z)-1o

1H NMR (300 MHz, $CDCl_3$): δ 0.79–0.96 (m, 12H), 1.20–1.47 (m, 16H), 1.48–1.61 (m, 2H), 3.90–3.97 (m, 4H), 4.76 (ddd, $J = 8.1, 6.6, 1.6$ Hz, 2H), 5.91 (dt, $J = 11.7, 6.6$ Hz, 1H), 6.59 (d, $J = 11.7$ Hz, 1H), 7.05–7.10 (m, 2H), 7.45–7.51 (m, 2H). ^{13}C NMR (75.4 MHz, $CDCl_3$): δ 10.2, 14.1, 23.0, 23.3, 28.9 (d, $J = 1.1$ Hz), 29.9, 40.1 (d, $J = 7.3$ Hz), 63.9 (d, $J = 5.6$ Hz), 70.0 (d, $J = 6.3$ Hz), 122.0, 127.6 (d, $J = 7.2$ Hz), 130.5, 131.8, 132.0, 134.9. HRMS–ESI (m/z): $[M+Na]^+$ calcd. for $C_{25}H_{42}O_4BrNaP$ 539.19018, found 539.18929.

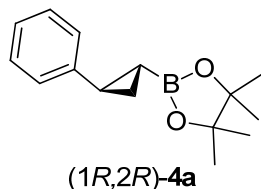
5. Determination of Absolute Configuration of *trans*-**4a**.



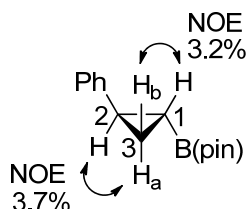
The absolute configuration of *trans*-**4a** was determined by comparing the optical rotation of the cyclopropyl alcohol obtained by oxidation. The spectral data were identical with those reported.⁴ Yield 60%. ¹H NMR (300 MHz, CDCl₃): δ 1.05 (q, J = 6.3 Hz, 1H), 1.28 (ddd, J = 9.6, 6.3, 2.5 Hz, 1H), 2.11 (ddd, J = 9.6, 6.3, 2.5 Hz, 1H), 2.23 (br, 1H), 3.62 (ddd, J = 6.5, 3.5, 2.6 Hz, 1H), 6.98–7.04 (m, 2H), 7.13–7.21 (m, 1H), 7.23–7.29 (m, 2H). $[\alpha]_{\text{D}}^{23.0} -54$ (c 0.4, ethanol, 94% ee). The ee value was determined by HPLC analysis (CHIRALCEL OD-3, 2-PrOH/Hexane = 5/95, 0.5 mL/min, UV detector at 220 nm, 40 °C, (1*R*,2*S*)-**5** t_{R} = 30.5 min, (1*S*,2*R*)-**5** t_{R} = 26.6 min).

6. Characterization Data for Cyclopropylboronates.

4,4,5,5-Tetramethyl-2-[(1*R*,2*R*)-2-phenylcyclopropyl]1,3,2-dioxaborolane.

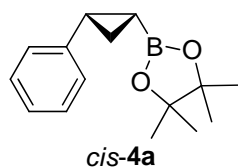


^1H NMR (300 MHz, CDCl_3): δ 0.30 (ddd, $J = 9.8, 6.8, 5.4$ Hz, 1H), 1.00 (ddd, $J = 9.8, 5.4, 3.7$ Hz, 1H), 1.16 (ddd, $J = 8.1, 6.8, 3.6$ Hz, 1H), 1.24 (s, 6H), 1.25 (s, 6H), 2.10 (dt, $J = 8.1, 5.4$ Hz, 1H), 7.05–7.17 (m, 3H), 7.21–7.28 (m, 2H). NOEs were observed between resonances of H-1 and H-3b, H-2 and H-3a.

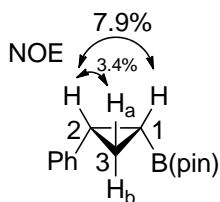


^{13}C NMR (75.4 MHz, CDCl_3): δ 15.1, 22.0, 24.8, 24.8, 83.3, 125.8, 125.9, 128.5, 143.7. The carbon directly attached to the boron atom was not detected, likely due to quadropolar relaxation.^{4,5,6} HRMS–EI (m/z): $[\text{M}]^+$ calcd. for $\text{C}_{15}\text{H}_{21}\text{BO}_2$ 244.16346, found 244.16310. $[\alpha]_D^{26} -175.8$ (c 1.085, CHCl_3 , 94% ee). The ee value was determined by HPLC analysis (CHIRALCEL OD-3, hexane, 0.5 mL/min, UV detector at 220 nm, 40 °C, (1*R*,2*R*)-**4a** t_R = 18.2 min, (1*S*,2*S*)-**4a** t_R = 16.3 min).

4,4,5,5-Tetramethyl-2-(2-phenylcyclopropyl)-1,3,2-dioxaborolane.

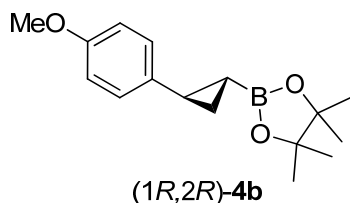


^1H NMR (300 MHz, CDCl_3): δ 0.44 (ddd, $J = 10.1, 9.3, 7.3$ Hz, 1H), 0.88 (s, 6H), 1.01 (s, 6H), 1.10 (ddd, $J = 9.3, 8.0, 4.2$ Hz, 1H), 1.28 (ddd, $J = 7.1, 6.1, 4.2$ Hz, 1H), 2.35 (ddd, $J = 10.1, 7.8, 6.1$ Hz, 1H), 7.08–7.16 (m, 1H), 7.18–7.31 (m, 4H). NOEs were observed between resonances of H-2 and H-1, H-2 and H-3a.



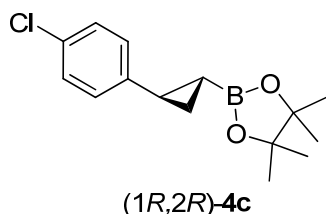
^{13}C NMR (75.4 MHz, CDCl_3): δ 3.4 (br), 8.9, 21.8, 24.5, 24.8, 83.1, 126.0, 127.9, 129.1, 141.0. HRMS–EI (m/z): $[\text{M}]^+$ calcd. for $\text{C}_{15}\text{H}_{21}\text{BO}_2$ 244.16346, found 244.16352. $[\alpha]^{22.6}_{\text{D}} +17.6$ (c 0.98, CHCl_3 , 27% ee). The ee value was determined by HPLC analysis (CHIRALPAK IC, 2-PrOH/Hexane = 0.1/99.9, 0.5 mL/min, UV detector at 220 nm, 40 °C, $t_{\text{minor}} = 11.3$ min, $t_{\text{major}} = 12.3$ min). The absolute configuration of this compound could not be determined because the C–B bond oxidation resulted in fast decomposition of the alcohol derivative.

2-[(1*R*,2*R*)-2-(4-Methoxyphenyl)cyclopropyl]-4,4,5,5-tetramethyl-1,3,2-dioxaborolane.



^1H NMR (300 MHz, CDCl_3): δ 0.22 (ddd, $J = 9.8, 6.6, 5.6$ Hz, 1H), 0.93 (ddd, $J = 9.6, 5.4, 3.7$ Hz, 1H), 1.11 (ddd, $J = 8.1, 6.8, 3.6$ Hz, 1H), 1.24 (s, 6H), 1.25 (s, 6H), 2.07 (dt, $J = 8.1, 5.4$ Hz, 1H), 3.77 (s, 3H), 6.76–6.82 (m, 2H), 6.99–7.05 (m, 2H). ^{13}C NMR (75.4 MHz, CDCl_3): δ 5.5 (br), 14.6, 21.3, 24.8, 24.8, 55.4, 83.3, 114.0, 127.0, 135.6, 158.0. HRMS–EI (m/z): $[\text{M}]^+$ calcd. for $\text{C}_{16}\text{H}_{23}\text{BO}_3$ 274.17402, found 274.17399. $[\alpha]^{26.0}_{\text{D}} -152.5$ (c 1.055, CHCl_3 , 94% ee). The ee value was determined by HPLC analysis (CHIRALPAK IC, 2-PrOH/Hexane = 0.25/99.75, 0.5 mL/min, UV detector at 220 nm, 40 °C, (1*R*,2*R*)-**4b** $t_{\text{R}} = 16.8$ min, (1*S*,2*S*)-**4b** $t_{\text{R}} = 24.6$ min).

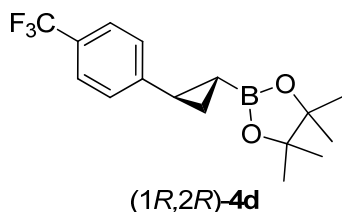
2-[(1*R*,2*R*)-2-(4-Chlorophenyl)cyclopropyl]-4,4,5,5-tetramethyl-1,3,2-dioxaborolane.



^1H NMR (300 MHz, CDCl_3): δ 0.25 (ddd, $J = 9.8, 6.8, 5.6$ Hz, 1H), 0.96 (ddd, $J = 9.8, 5.2, 3.8$ Hz, 1H), 1.16 (ddd, $J = 8.1, 6.9, 3.8$ Hz, 1H), 1.24 (s, 6H), 1.25 (s, 6H), 2.07 (dt, $J = 8.1, 5.4$ Hz, 1H), 6.97–7.03 (m, 2H), 7.17–7.23 (m, 2H). ^{13}C NMR (75.4 MHz, CDCl_3): δ 6.2 (br), 15.1, 21.4, 24.77, 24.80, 83.4, 127.3, 128.5, 131.3, 142.2. HRMS–EI (m/z): $[\text{M}]^+$ calcd. for $\text{C}_{15}\text{H}_{20}\text{BClO}_2$ 278.12449,

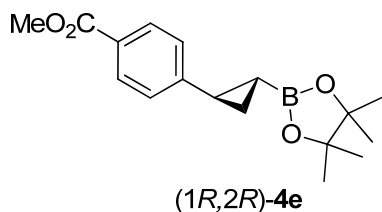
found 278.12453. $[\alpha]^{26.0}_{\text{D}} -155.7$ (c 0.87, CHCl_3 , 84% ee). The ee value was determined by HPLC analysis (CHIRALCEL OD-3, hexane, 0.5 mL/min, UV detector at 220 nm, 40 °C, (1*R*,2*R*)-**4c** t_{R} = 13.0 min, (1*S*,2*S*)-**4c** t_{R} = 14.5 min).

4,4,5,5-Tetramethyl-2-[(1*R*,2*R*)-2-[4-(trifluoromethyl)phenyl]cyclopropyl-1,3,2-dioxaborolane.



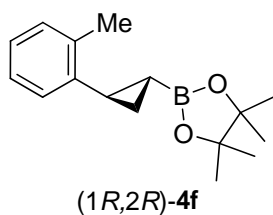
^1H NMR (300 MHz, CDCl_3): δ 0.35 (ddd, J = 9.8, 7.0, 5.6 Hz, 1H), 1.04 (ddd, J = 9.8, 5.2, 3.9 Hz, 1H), 1.19–1.32 (m, 1H), 1.25 (s, 6H), 1.26 (s, 6H), 2.14 (dt, J = 8.0, 5.2 Hz, 1H), 7.16 (d, J = 8.2 Hz, 2H), 7.48 (d, J = 8.2 Hz, 2H). ^{13}C NMR (75.4 MHz, CDCl_3): δ 6.7 (br), 15.6, 21.8, 24.78, 24.82, 83.6, 124.6 (q, J = 272.0 Hz), 125.4 (q, J = 3.8 Hz), 128.0 (q, J = 32.5 Hz), 126.1, 148.1 (q, J = 1.1 Hz). HRMS–EI (m/z): $[\text{M}]^+$ calcd. for $\text{C}_{16}\text{H}_{20}\text{BF}_3\text{O}_2$ 312.15084, found 312.15085. mp 41.8–44.0 °C. $[\alpha]^{26.0}_{\text{D}} -122.4$ (c 0.63, CHCl_3 , 82% ee). The ee value was determined by HPLC analysis (CHIRALCEL OD-3, hexane, 0.5 mL/min, UV detector at 220 nm, 40 °C, (1*R*,2*R*)-**4d** t_{R} = 11.0 min, (1*S*,2*S*)-**4d** t_{R} = 12.2 min).

Methyl 4-[(1*R*,2*R*)-2-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)cyclopropyl]benzoate.



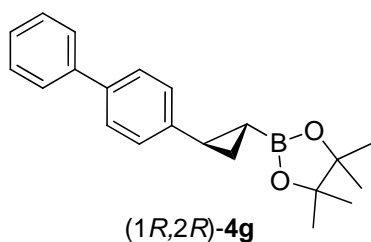
^1H NMR (300 MHz, CDCl_3): δ 0.37 (ddd, J = 9.9, 6.9, 5.5 Hz, 1H), 1.07 (ddd, J = 9.9, 5.2, 3.8 Hz, 1H), 1.24 (s, 6H), 1.26 (s, 6H), 1.19–1.30 (m, 1H), 2.13 (dt, J = 8.0, 5.3 Hz, 1H), 3.89 (s, 3H), 7.09–7.14 (m, 2H), 7.88–7.93 (m, 2H). ^{13}C NMR (75.4 MHz, CDCl_3): δ 15.8, 22.1, 24.8, 52.1, 83.5, 125.7, 127.6, 129.9, 149.7, 167.5. The carbon directly attached to the boron atom was not detected, likely due to quadrupolar relaxation.^{4,5,6} HRMS–EI (m/z): $[\text{M}]^+$ calcd. for $\text{C}_{17}\text{H}_{23}\text{BO}_4$ 302.16894, found 302.16861. mp 55.5–69.2 °C. $[\alpha]^{26.0}_{\text{D}} -132.9$ (c 0.91, CHCl_3 , 64% ee). The ee value was determined by HPLC analysis (CHIRALCEL OD-3, 2-PrOH/Hexane = 1:99, 0.5 mL/min, UV detector at 220 nm, 40 °C, (1*R*,2*R*)-**4e** t_{R} = 11.6 min, (1*S*,2*S*)-**4e** t_{R} = 12.7 min).

4,4,5,5-Tetramethyl-2-[(1*R*,2*R*)-2-(*o*-tolyl)cyclopropyl]-1,3,2-dioxaborolane.



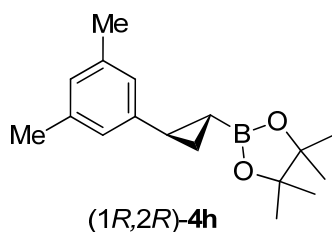
^1H NMR (300 MHz, CDCl_3): δ 0.16 (dt, $J = 9.6, 6.2$ Hz, 1H), 1.02 (ddd, $J = 9.7, 5.7, 3.4$ Hz, 1H), 1.13 (ddd, $J = 8.0, 6.5, 3.5$ Hz, 1H), 1.26 (s, 12H), 2.09 (dt, $J = 8.0, 5.8$ Hz, 1H), 2.40 (s, 3H), 6.98–7.02 (m, 1H), 7.07–7.16 (m, 3H). ^{13}C NMR (75.4 MHz, CDCl_3): δ 12.3, 19.7, 20.4, 24.7, 24.8, 83.3, 125.7, 126.0, 126.1, 129.8, 138.2, 141.2. The carbon directly attached to the boron atom was not detected, likely due to quadrupolar relaxation.^{4,5,6} HRMS–EI (m/z): $[\text{M}]^+$ calcd. for $\text{C}_{16}\text{H}_{23}\text{BO}_2$ 258.17911, found 258.17899. mp 60.8–68.8 °C. $[\alpha]_D^{26.0} -103.1$ (c 0.99, CHCl_3 , 94% ee). The ee value was determined by HPLC analysis (CHIRALCEL OD-3, Hexane, 0.5 mL/min, UV detector at 220 nm, 40 °C, (1*R*,2*R*)-**4f** t_R = 18.1 min, (1*S*,2*S*)-**4f** t_R = 20.2 min).

4,4,5,5-Tetramethyl-2-[(1*R*,2*R*)-2-(4-phenylphenyl)cyclopropyl]-1,3,2-dioxaborolane.



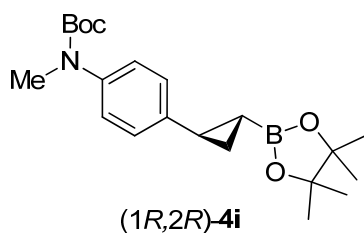
^1H NMR (300 MHz, CDCl_3 , δ): 0.35 (ddd, $J = 9.6, 6.6, 5.6$ Hz, 1H), 1.01–1.09 (m, 1H), 1.05 (ddd, $J = 9.9, 5.5, 3.9$ Hz, 1H), 1.25 (s, 6H), 1.26 (s, 6H), 2.15 (dt, $J = 8.0, 5.5$ Hz, 1H), 7.15 (d, $J = 8.3$ Hz, 2H), 7.23–7.64 (m, 7H). ^{13}C NMR (75.4 MHz, CDCl_3 , δ): 6.1 (br, CH), 15.0 (CH_2), 21.5 (CH), 24.6 (CH_3), 24.7 (CH_3), 83.2 (C), 126.1 (CH), 127.0 (CH), 127.01 (CH), 127.06 (CH), 128.8 (CH), 138.6 (C), 141.2 (C), 147.3 (C). $[\alpha]_D^{21} -149.8$ (c 0.995, CHCl_3 , 86% ee), mp 86°C. HRMS–ESI (m/z): $[\text{M}]^+$ calcd for $\text{C}_{21}\text{H}_{25}\text{BO}_2$, 320.19476; found, 320.19479. The ee value of 86% was determined by chiral HPLC analysis (CHIRALCEL OD-3, hexane/2-propanol 99.75/0.25, UV detector at 220 nm, 40 °C, (1*R*,2*R*)-**4g**: t_R = 23.5 min. (1*S*,2*S*)-**4g**: t_R = 32.4 min.).

4,4,5,5-Tetramethyl-2-[(1*R*,2*R*)-2-(3,5-dimethylphenyl)cyclopropyl]-1,3,2-dioxaborolane.



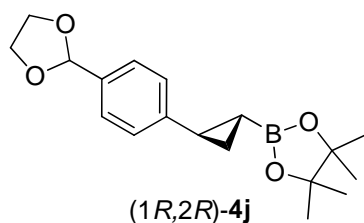
¹H NMR (300 MHz, CDCl₃, δ): 0.27 (ddd, *J* = 9.6, 6.9, 5.5 Hz, 1H), 0.98 (ddd, *J* = 9.7, 5.2, 3.7 Hz, 1H), 1.08–1.20 (m, 1H), 1.23 (s, 6H), 1.25 (s, 6H), 1.99–2.08 (m, 1H), 2.26 (s, 6H), 6.71 (s, 2H), 6.78 (s, 1H). ¹³C NMR (75.4 MHz, CDCl₃, δ): 5.3 (br, CH), 14.6 (CH₂), 21.1 (CH₃), 21.6 (CH), 24.5 (CH₃), 24.6 (CH₃), 83.1 (C), 123.6 (CH), 127.3 (CH), 137.8 (C), 143.4 (C). [α]_D²³ –147.6 (*c* 0.975, CHCl₃, 94% ee). HRMS–ESI (*m/z*): [M+H]⁺ calcd for C₁₇H₂₆BO₂, 273.20258; found, 273.20205. The ee value of 94% was determined by chiral HPLC analysis (CHIRALCEL OD-3, hexane, UV detector at 220 nm, 40 °C, (1*R*,2*R*)-4h: *t*_R = 13.4 min. (1*S*,2*S*)-4h: *t*_R = 16.2 min.).

4,4,5,5-Tetramethyl-2-[(1*R*,2*R*)-2-[4-(*N*-Boc-*N*-methylamino)phenyl]cyclopropyl-1,3,2-dioxaborolane.



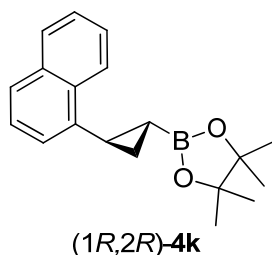
¹H NMR (300 MHz, CDCl₃, δ): 0.27 (ddd, *J* = 9.7, 6.8, 5.6 Hz, 1H), 0.98 (ddd, *J* = 9.7, 5.3, 3.8 Hz, 1H), 1.11–1.20 (m, 1H), 1.24 (s, 6H), 1.25 (s, 6H), 1.43 (s, 9H), 2.08 (dt, *J* = 8.3, 5.4 Hz, 1H), 3.22 (s, 3H), 6.98–7.14 (m, 4H). ¹³C NMR (75.4 MHz, CDCl₃, δ): 5.6 (br, CH), 14.8 (CH₂), 21.3 (CH), 24.57 (CH₃), 24.61 (CH₃), 28.23 (CH₃), 37.3 (CH₃), 80.1 (C), 83.2 (C), 125.5 (CH), 125.8 (CH), 140.6 (C), 141.5 (C), 155.0 (C). [α]_D²³ –90.5 (*c* 0.935, CHCl₃, 89% ee.). MS–ESI (*m/z*): [M+H]⁺ calcd for C₂₁H₃₂BNO₄, 374.25026; found, 374.25006. The ee value of 89% was determined by chiral HPLC analysis (CHIRALCEL IC, hexane/2-propanol 99/1, 40 °C, (1*R*,2*R*)-4i: *t*_R = 29.4 min. (1*S*,2*S*)-4i: *t*_R = 33.7 min.).

2-[(1*R*,2*R*)-2-[4-(1,3-Dioxolan-2-yl)phenyl]cyclopropyl]-4,4,5,5-tetramethyl-1,3,2-dioxaborolane



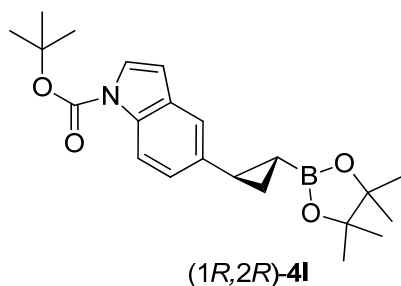
¹H NMR (300 MHz, CDCl₃, δ): 0.30 (ddd, *J* = 9.7, 6.8, 5.5 Hz, 1H), 0.96–1.04 (m, 1H), 1.12–1.21 (m, 1H), 1.24 (s, 6H), 1.25 (s, 6H), 2.11 (dt, *J* = 8.1, 5.4 Hz, 1H), 3.91–4.18 (m, 4H), 5.78 (s, 1H), 7.09 (d, *J* = 8.3 Hz, 2H), 7.35 (d, *J* = 8.3 Hz, 2H). ¹³C NMR (75.4 MHz, CDCl₃, δ): 5.8 (br, CH), 15.0 (CH₂), 21.6 (CH), 24.5 (CH₃), 24.6 (CH₃), 65.1 (CH₂), 83.1 (C), 103.7 (CH), 125.7 (CH), 126.4 (CH), 135.2 (C), 144.7 (C). [α]_D²² −127.9° (*c* 1.03, CHCl₃, 86% ee). HRMS–ESI (*m/z*): [M+H]⁺ calcd for C₁₈H₂₆BO₄, 317.19241; found, 317.19150. Anal. Calcd for C₁₈H₂₅BO₄: C, 68.37; H, 7.97. Found: C, 68.11; H, 8.13. The ee value of 86% was determined by chiral HPLC analysis (CHIRALCEL OD-3, hexane/2-propanol 99.5/0.5, 40 °C, (1*R*,2*R*)-**4j**: *t*_R = 22.7 min. (1*S*,2*S*)-**4j**: *t*_R = 27.3 min.).

4,4,5,5-Tetramethyl-2-[(1*R*,2*R*)-2-(naphthalen-1-yl)cyclopropyl]-1,3,2-dioxaborolane.



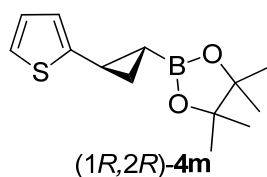
¹H NMR (300 MHz, CDCl₃): δ 0.37 (dt, *J* = 9.6, 6.2 Hz, 1H), 1.09 (ddd, *J* = 9.6, 5.6, 3.4 Hz, 1H), 1.28 (m, 1H), 1.31 (s, 12H), 2.59 (dt, *J* = 8.1, 5.9 Hz, 1H), 7.24–7.29 (m, 1H), 7.34–7.40 (m, 1H), 7.45–7.57 (m, 2H), 7.70 (d, *J* = 8.0 Hz, 1H), 7.82–7.87 (m, 1H), 8.33–8.38 (m, 1H). ¹³C NMR (75.4 MHz, CDCl₃): δ 12.3, 19.8, 24.63, 24.64, 83.2, 123.5, 124.4, 125.5, 125.6, 125.8, 126.7, 128.5, 133.4, 133.5, 138.9. The carbon directly attached to the boron atom was not detected, likely due to quadrupolar relaxation.^{4,5,6} HRMS–EI (*m/z*): [M]⁺ calcd. for C₁₉H₂₃BO₂ 294.17911, found 294.17897. [α]_D^{26.0} −58.2 (*c* 1.085, CHCl₃, 91% ee). The ee value was determined by HPLC analysis (CHIRALCEL OD-3, 2-PrOH/Hexane = 0.5/99.5, 0.5 mL/min, UV detector at 220 nm, 40 °C, (1*R*,2*R*)-**4k** *t*_R = 17.4 min, (1*S*,2*S*)-**4k** *t*_R = 15.7 min).

***tert*-Butyl 5-[(1*R*,2*R*)-2-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)cyclopropyl]-1*H*-indole-1-carboxylate.**



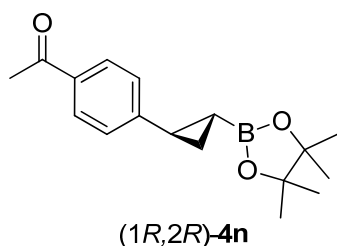
^1H NMR (300 MHz, CDCl_3): δ 0.32 (ddd, $J = 9.7, 6.8, 5.7$ Hz, 1H), 1.03 (ddd, $J = 9.7, 5.3, 3.7$ Hz, 1H), 1.16 (ddd, $J = 7.9, 6.9, 3.7$ Hz, 1H), 1.25 (s, 6H), 1.26 (s, 6H), 1.66 (s, 9H), 2.21 (dt, $J = 8.1, 5.5$ Hz, 1H), 6.48 (dd, $J = 3.8, 0.5$ Hz, 1H), 7.04 (dd, $J = 8.7, 1.8$ Hz, 1H), 7.28 (d, $J = 1.7$ Hz, 1H), 7.54 (d, $J = 3.6$ Hz, 1H), 7.98 (d, $J = 8.5$ Hz, 1H). ^{13}C NMR (75.4 MHz, CDCl_3): δ 14.9, 22.0, 24.80, 24.83, 28.3, 83.3, 83.6, 107.3, 115.1, 118.2, 122.8, 126.3, 131.0, 133.8, 137.9, 150.1. The carbon directly attached to the boron atom was not detected, likely due to quadropolar relaxation.^{4,5,6} HRMS–ESI (m/z): $[\text{M}+\text{Na}]^+$ calcd. for $\text{C}_{22}\text{H}_{30}\text{BNNaO}_4$ 406.21656, found 406.21662. $[\alpha]^{26.0}_{\text{D}} -117.6$ (c 0.95, CHCl_3 , 92% ee). The ee value was determined by HPLC analysis (CHIRALCEL OD-3, 2-PrOH/Hexane = 0.5/99.5, 0.5 mL/min, UV detector at 220 nm, 40 °C, (1*R*,2*R*)-4l t_{R} = 11.7 min, (1*S*,2*S*)-4l t_{R} = 13.2 min).

4,4,5,5-Tetramethyl-2-[(1*R*,2*R*)-2-(thiophen-2-yl)cyclopropyl]-1,3,2-dioxaborolane.



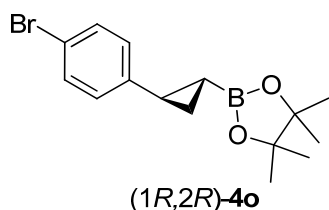
^1H NMR (300 MHz, CDCl_3): δ 0.34 (ddd, $J = 9.9, 6.9, 5.5$ Hz, 1H), 1.02 (ddd, $J = 9.9, 5.2, 3.6$ Hz, 1H), 1.17 (ddd, $J = 8.0, 7.0, 3.6$ Hz, 1H), 1.24 (s, 6H), 1.25 (s, 6H), 2.31 (dt, $J = 8.0, 5.4$ Hz, 1H), 6.78 (dt, $J = 3.5, 1.0$ Hz, 1H), 6.87 (dd, $J = 5.0, 3.5$ Hz, 1H), 7.03 (dd, $J = 5.0, 1.0$ Hz, 1H). ^{13}C NMR (75.4 MHz, CDCl_3): δ 15.8, 17.5, 24.8, 83.4, 122.2, 123.0, 126.9, 148.5. The carbon directly attached to the boron atom was not detected, likely due to quadropolar relaxation.^{4,5,6} HRMS–EI (m/z): $[\text{M}]^+$ calcd. for $\text{C}_{13}\text{H}_{19}\text{BO}_2\text{S}$ 250.11988, found 250.11976. $[\alpha]^{26.0}_{\text{D}} -160.4$ (c 1.06, CHCl_3 , 92% ee). The ee value was determined by HPLC analysis (CHIRALCEL OD-3, Hexane, 0.5 mL/min, UV detector at 220 nm, 40 °C, (1*R*,2*R*)-4m t_{R} = 16.1 min, (1*S*,2*S*)-4m t_{R} = 17.5 min).

1-[4-[(1*R*,2*R*)-2-(4,4,5,5-Tetramethyl-1,3,2-dioxaborolan-2-yl)cyclopropyl]]2phenylethanone.



^1H NMR (300 MHz, CDCl_3): δ 0.38 (ddd, $J = 9.8, 7.0, 5.5$ Hz, 1H), 1.08 (ddd, $J = 9.8, 5.1, 3.8$ Hz, 1H), 1.20–1.27 (m, 1H), 1.25 (s, 6H), 1.26 (s, 6H), 2.15 (dt, $J = 8.0, 5.3$ Hz, 1H), 2.57 (s, 3H), 7.11–7.16 (m, 2H), 7.82–7.87 (m, 2H). ^{13}C NMR (75.4 MHz, CDCl_3): δ 15.9, 22.1, 24.80, 24.82, 26.6, 83.6, 125.8, 128.8, 135.0, 150.0, 198.1. The carbon directly attached to the boron atom was not detected, likely due to quadropolar relaxation.^{4,5,6} HRMS–EI (m/z): $[\text{M}]^+$ calcd. for $\text{C}_{17}\text{H}_{23}\text{BO}_3$ 286.17402, found 286.17406. $[\alpha]^{25.3}_{\text{D}} -163.6$ (c 0.365, CHCl_3 , 85% ee). The ee value was determined by HPLC analysis (CHIRALCEL OD-3, 2-PrOH/Hexane = 1.5/98.5, 0.5 mL/min, UV detector at 220 nm, 40 °C, (1*R*,2*R*)-**4n** t_{R} = 14.6 min, (1*S*,2*S*)-**4n** t_{R} = 13.9 min).

2-[(1*R*,2*R*)-2-(4-Bromophenyl)cyclopropyl]-4,4,5,5-tetramethyl-1,3,2-dioxaborolane.



^1H NMR (300 MHz, CDCl_3): δ 0.26 (ddd, $J = 9.9, 6.9, 5.4$ Hz, 1H), 0.96 (ddd, $J = 9.8, 5.4, 3.7$ Hz, 1H), 1.16 (ddd, $J = 8.2, 7.0, 3.7$ Hz, 1H), 1.24 (s, 6H), 1.25 (s, 6H), 2.06 (dt, $J = 8.2, 5.4$ Hz, 1H), 6.91–6.97 (m, 2H), 7.32–7.37 (m, 2H). ^{13}C NMR (75.4 MHz, CDCl_3): δ 15.1, 21.5, 24.78, 24.81, 83.5, 119.2, 127.7, 131.5, 142.8. The carbon directly attached to the boron atom was not detected, likely due to quadropolar relaxation.^{4,5,6} HRMS–EI (m/z): $[\text{M}]^+$ calcd. for $\text{C}_{15}\text{H}_{20}\text{BBrO}_2$ 322.07397, found 322.07311. $[\alpha]^{26.0}_{\text{D}} -74.6$ (c 0.155, CHCl_3 , >50% ee). The ee value was determined by HPLC analysis (CHIRALCEL OD-3, Hexane, 0.5 mL/min, UV detector at 220 nm, 40 °C, (1*R*,2*R*)-**4o** t_{R} = 14.1 min, (1*S*,2*S*)-**4o** t_{R} = 16.1 min (overlapped with a peak from a side product)).

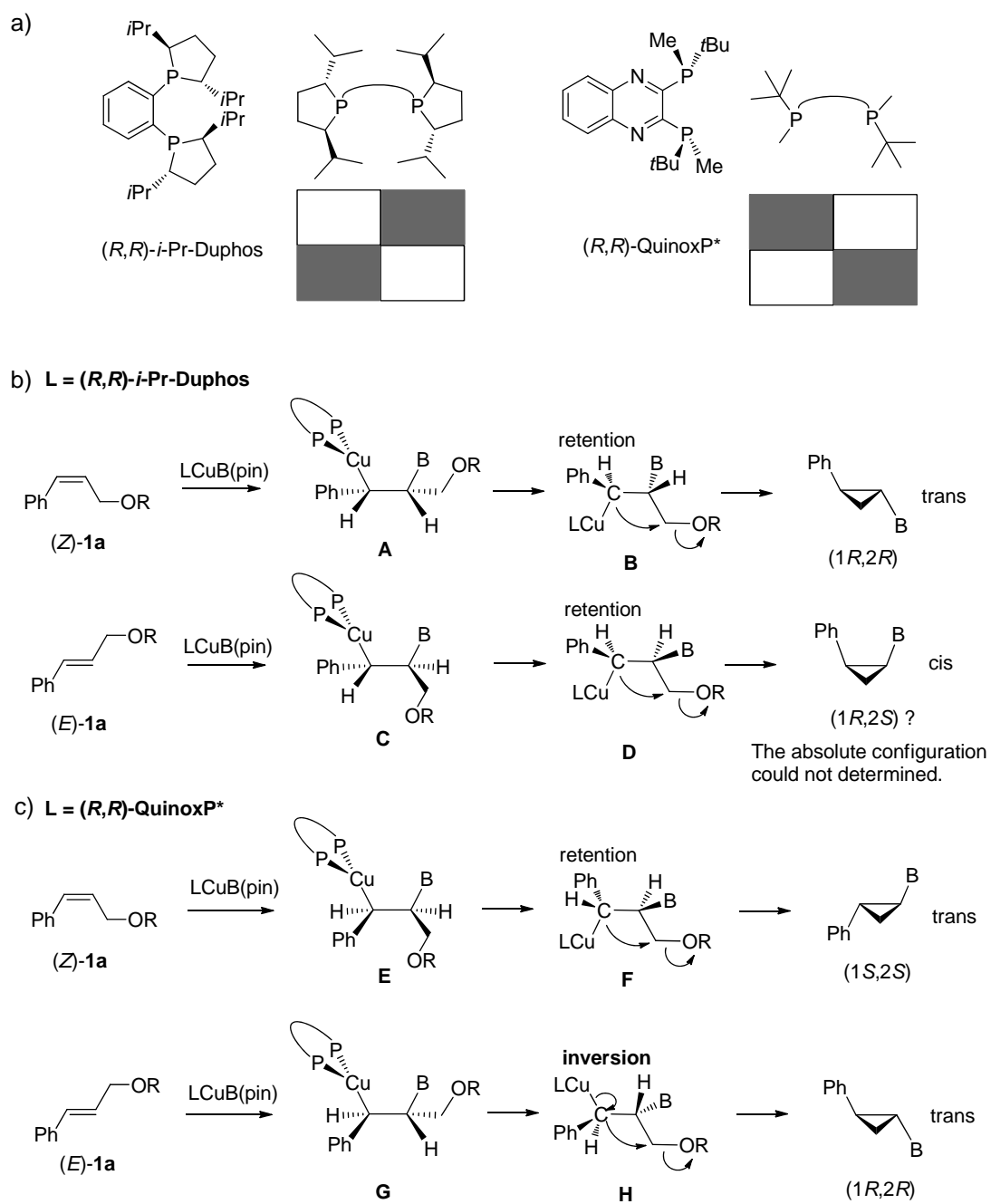
7. Possible Explanation on the Stereoselectivity in the reaction of (Z)- and (E)-1a.

The chiral environments created by (*R,R*)-*i*-Pr-DuPhos in the copper(I) catalyst intermediate is opposite to those with (*R,R*)-QuinoxP* or (*R,R*)-Me-DuPhos (Figure S1a). This explains the absolute configuration of the product **4a** in Table 1 (main text): the product with (*R,R*)-*i*-Pr-DuPhos [(1*R*,2*R*)-**4a**, Table 1, entries 1–3] shows the opposite configuration those with (*R,R*)-QuinoxP* or (*R,R*)-Me-DuPhos [(1*S*,2*S*)-**4a**, Table 1, entries 4, 5, 9].

The further explanation on the stereochemical outcomes in the main text was illustrated in Figure S1b and S1c. In the reaction of (Z)-**1a**, the borylcopper(I) species with (*R,R*)-*i*-Pr-DuPhos ligand first approaches to the (Z)-**1a** substrate such that the steric interaction between the ligand and phenyl group in (Z)-**1a** is minimized, affording the alkylcopper(I) intermediate **A** (Figure S1b). This intermediate **A** next undergoes intramolecular nucleophilic substitution (**B**) with retention of the configuration in terms of the carbon atom on the copper center to produce (1*R*,2*R*)-**4a** (94% ee). In the case of the reaction of (*E*)-**1a**, the borylcopper(I) intermediate reacts with (*E*)-**1a** to afford **C** with the favored steric interaction with the phenyl group. It is reasonable to suppose that the subsequent intramolecular nucleophilic substitution (**D**) proceeds with the retention of the configuration at the α -carbon center. This explains the *cis* product formation (Scheme 1, *cis*-**4a** (27% ee), the absolute configuration could not be determined).

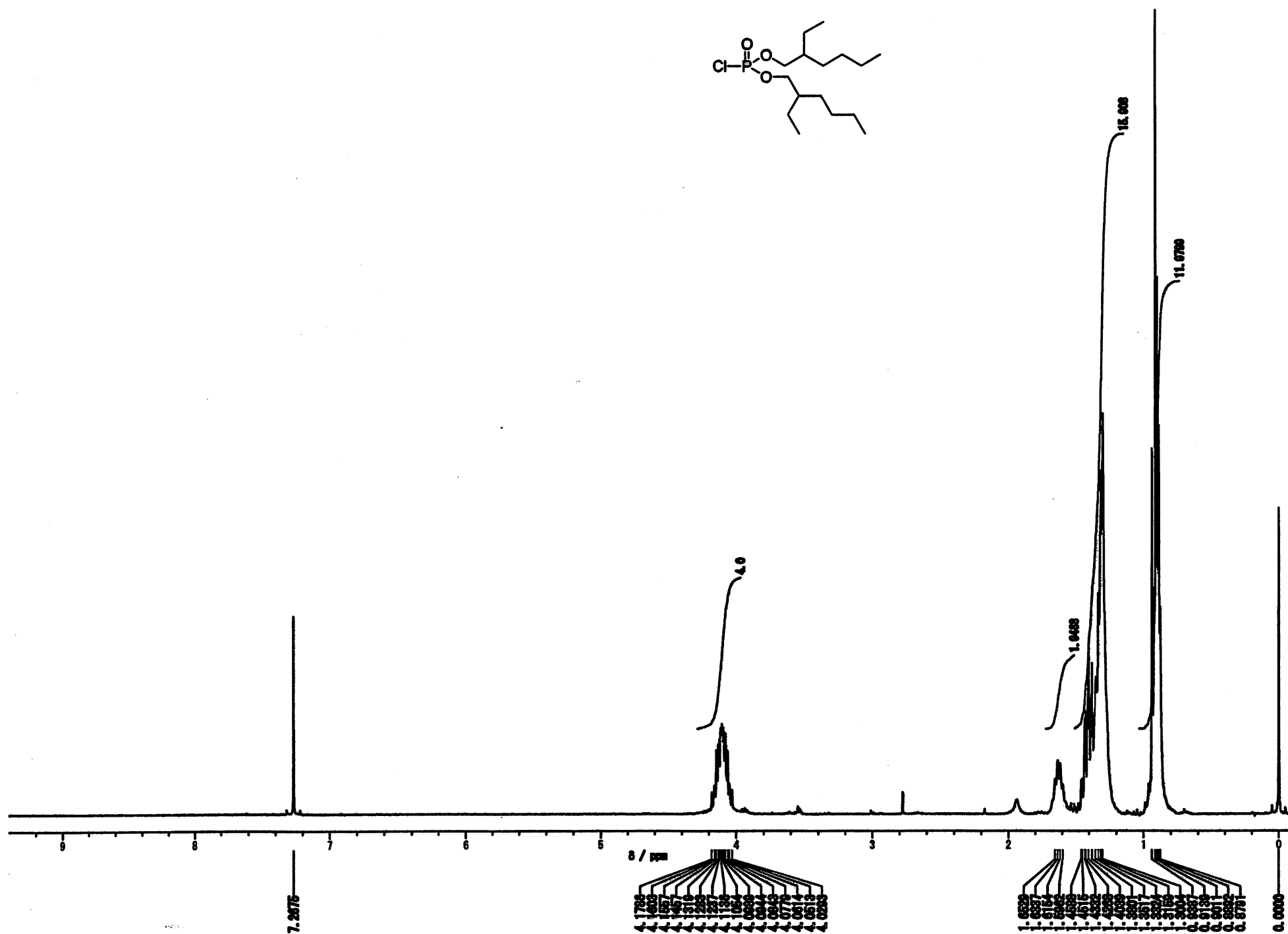
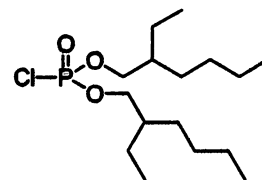
The reaction between the catalyst with (*R,R*)-QuinoxP* and (Z)-**1a** proceeds similarly: the steric interaction between the ligand and phenyl group is minimized. The resultant intermediate **E** undergoes intramolecular nucleophilic substitution (**F**) with retention of the configuration to afford (1*S*,2*S*)-**4a**. In the case of (*E*)-**1a**, the inversion of the stereochemistry of the α -carbon atom in the alkylcopper product **H** should take place during the cyclization process. Despite the rationale for the difference between (*R,R*)-*i*-Pr-DuPhos and (*R,R*)-QuinoxP* in their stereoselectivity at the cyclization step is not clear at this stage, this assumption can explain the stereochemical outcome observed in our results. The similar stereoselectivity switching of the copper(I) catalyst containing (*R,R*)-QuinoxP* ligand was observed in the reaction of silyl-substituted allylic carbonates.^{1b}

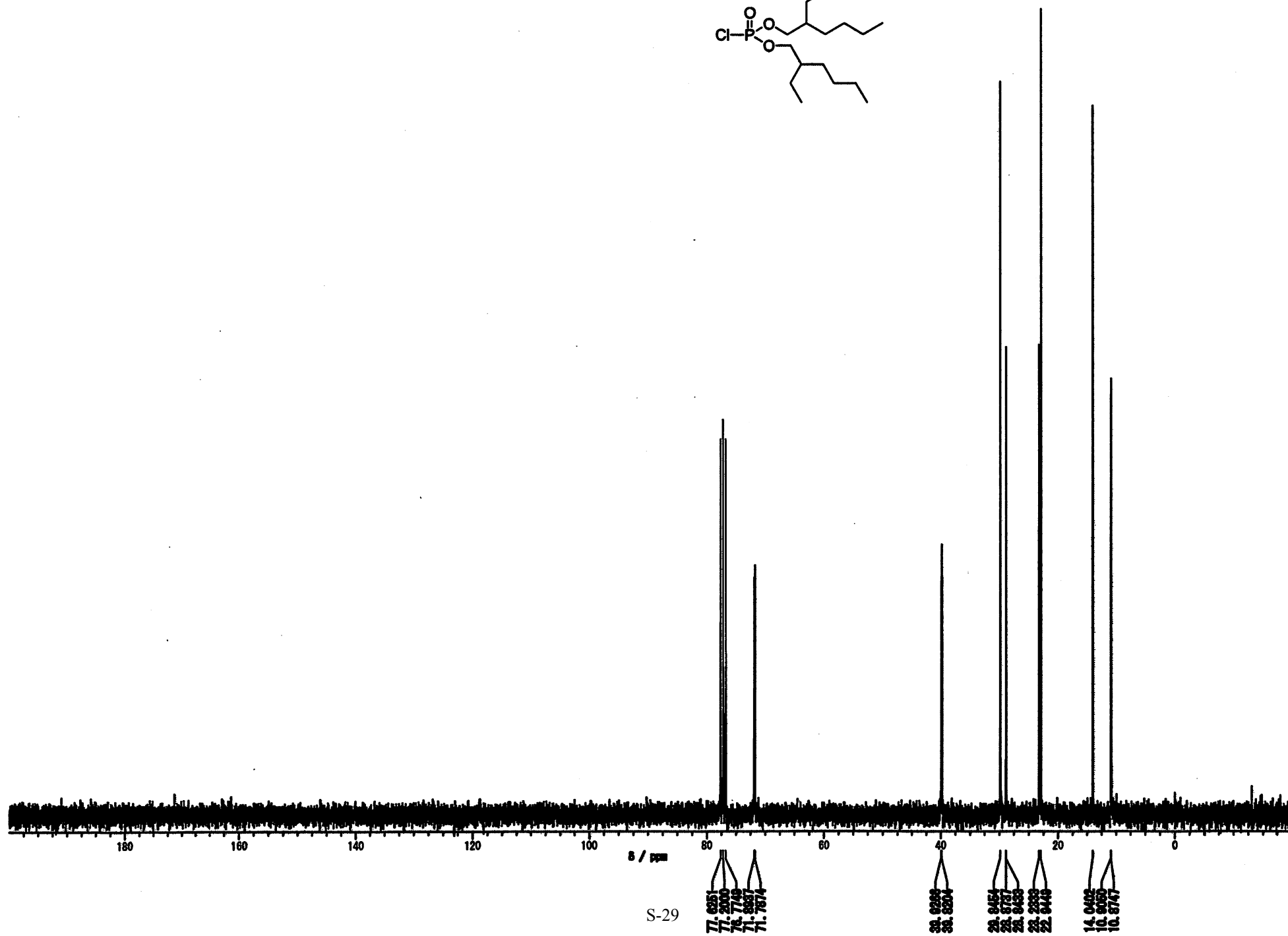
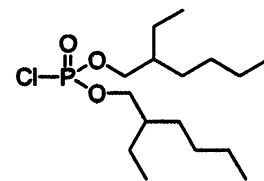
Figure S1.

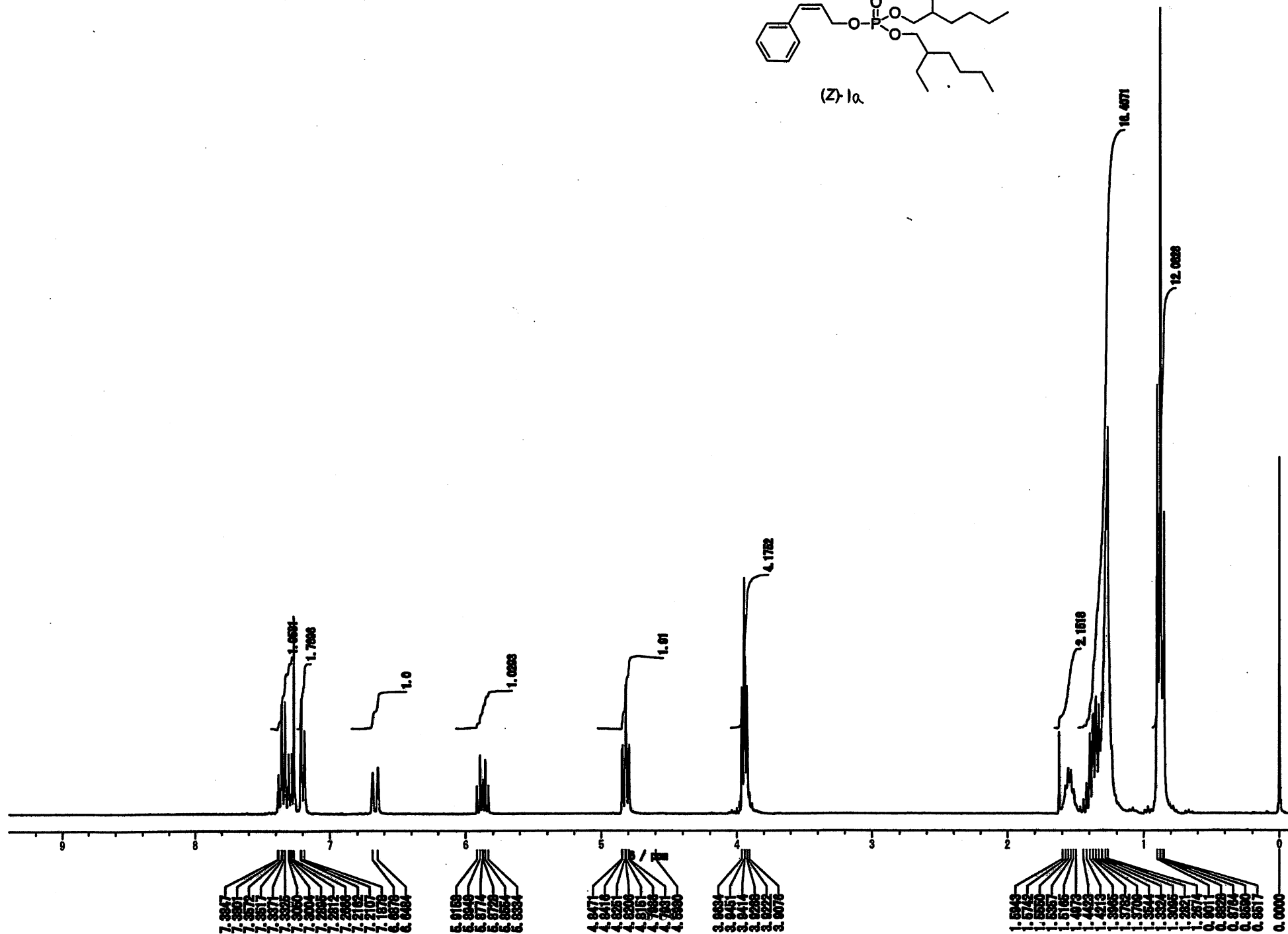
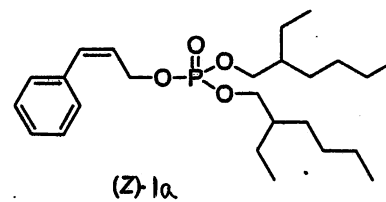


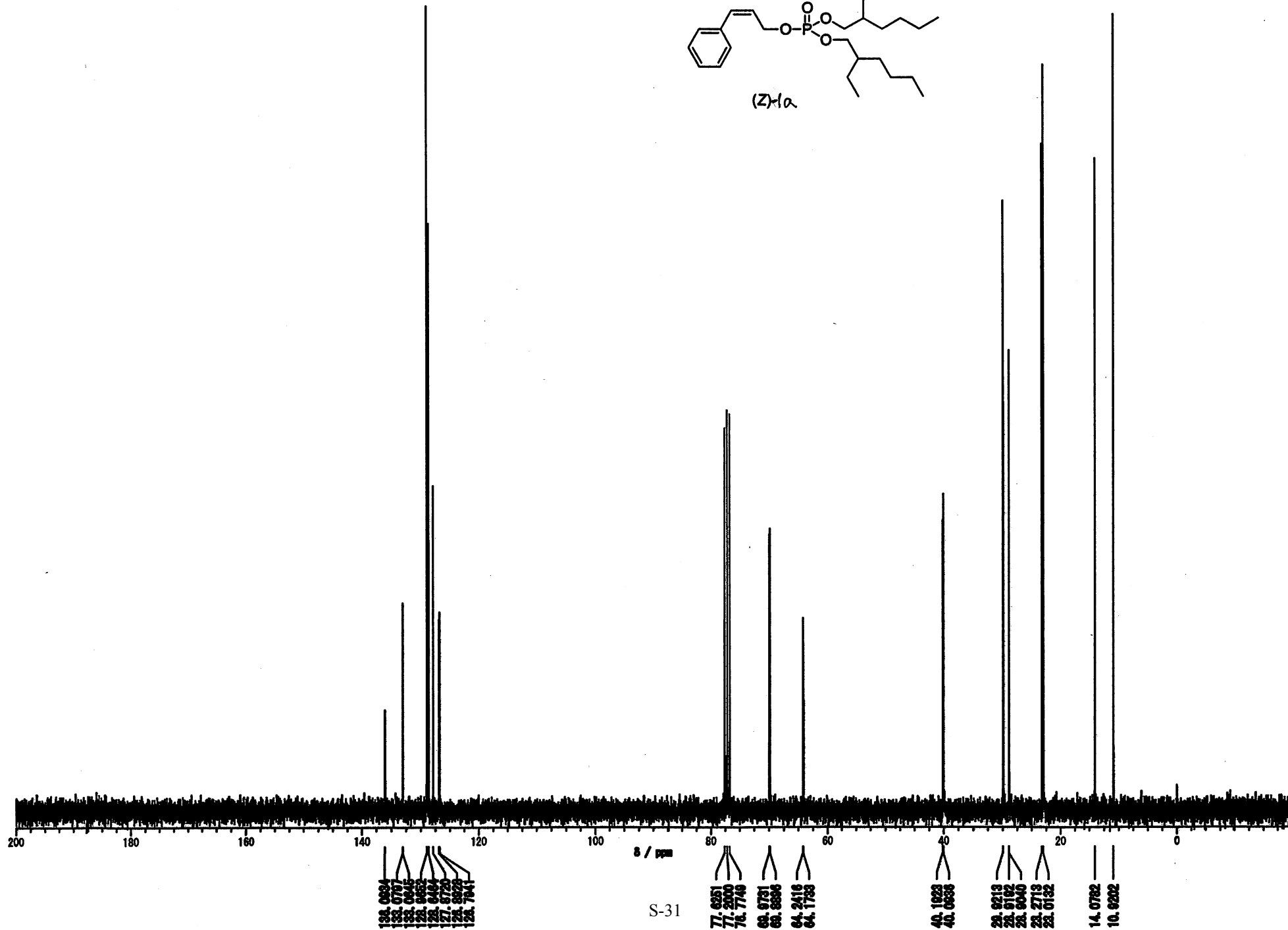
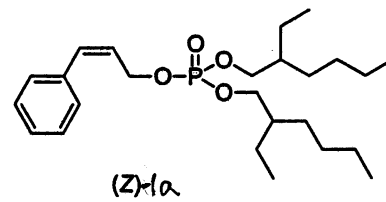
8. References.

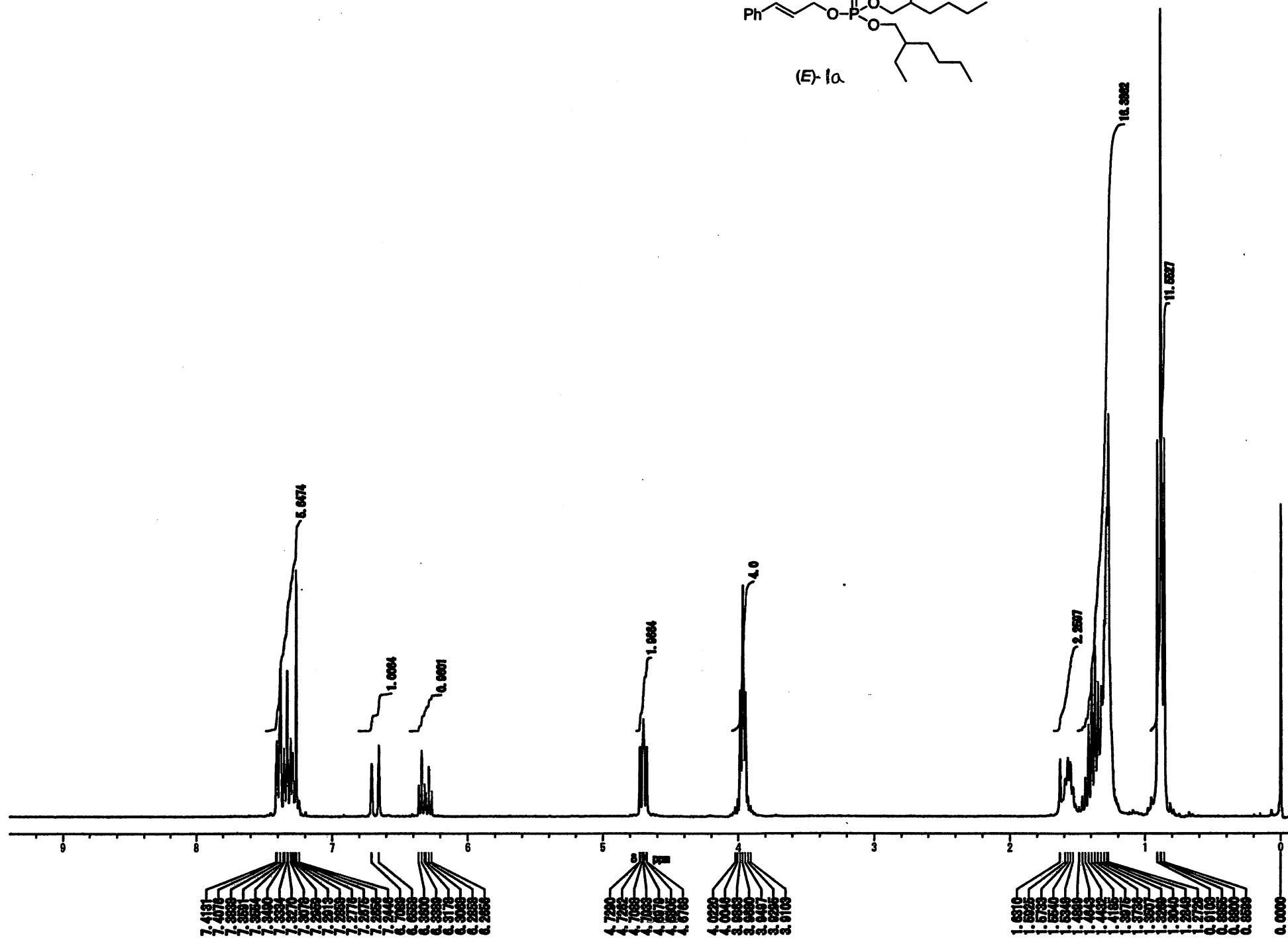
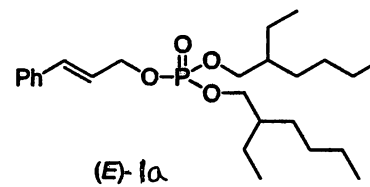
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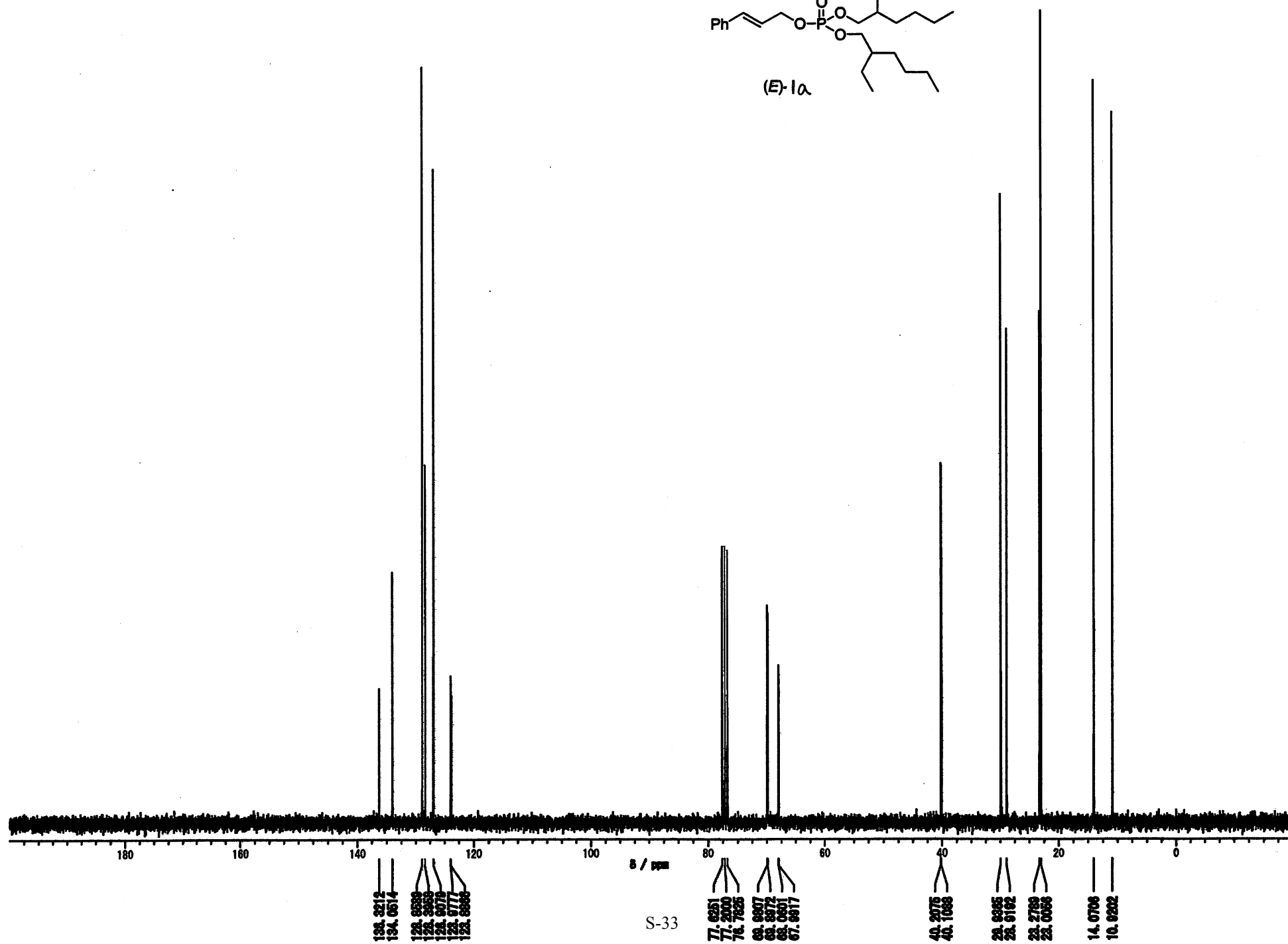
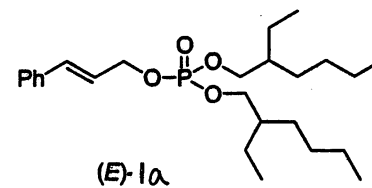


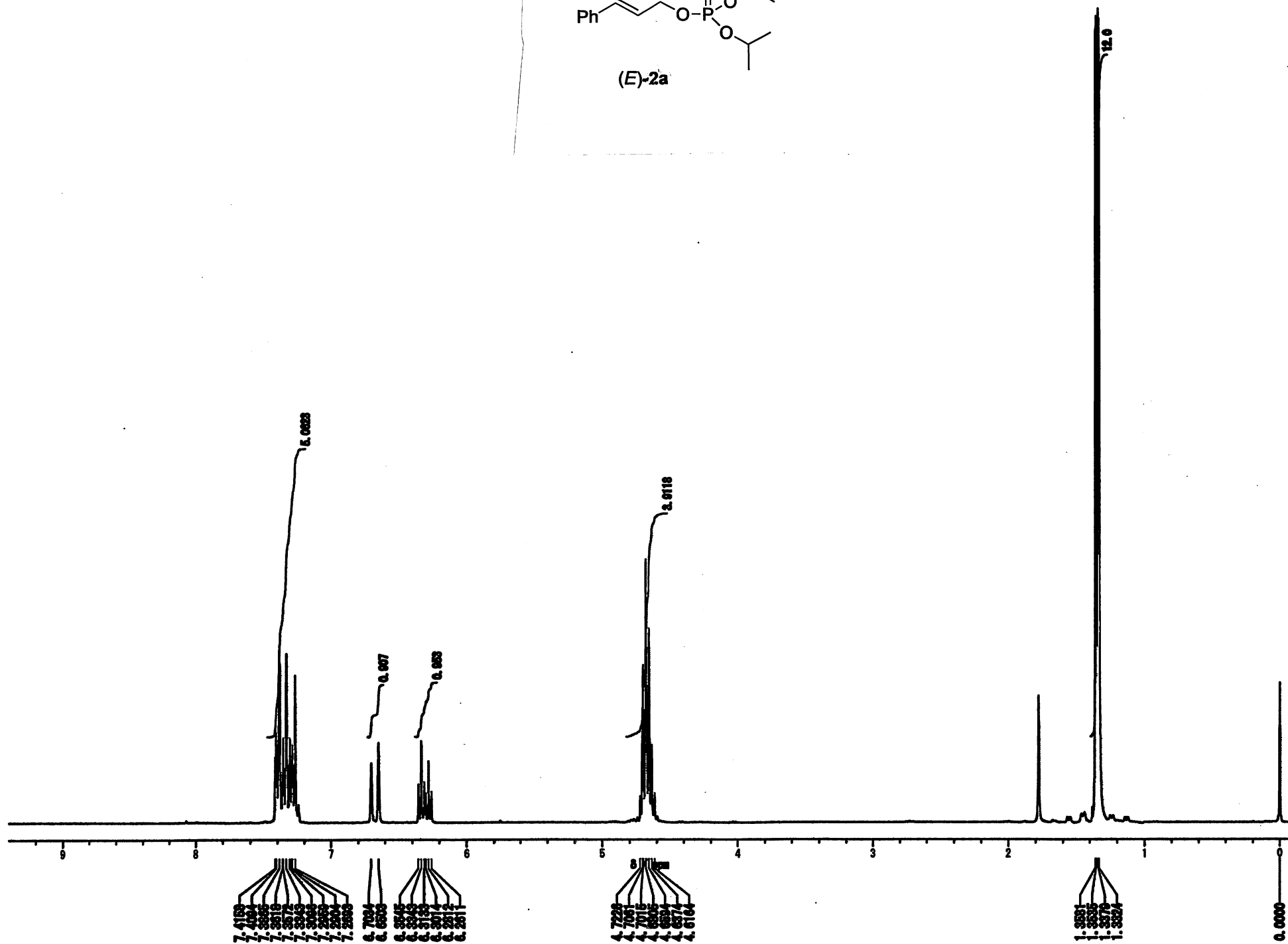
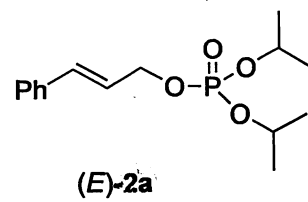


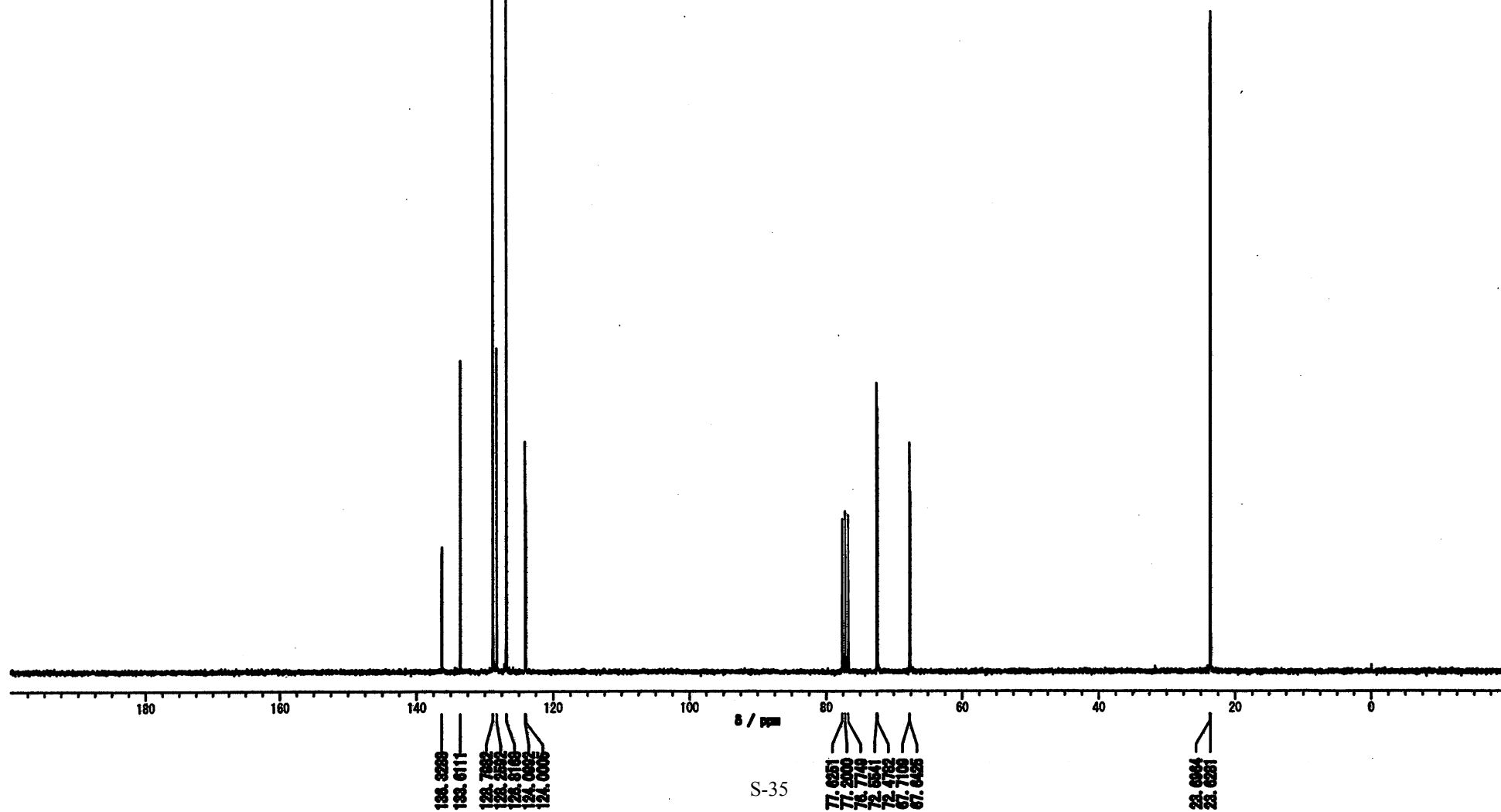
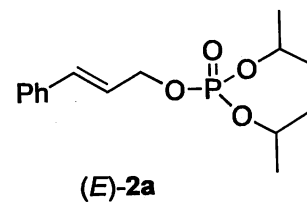


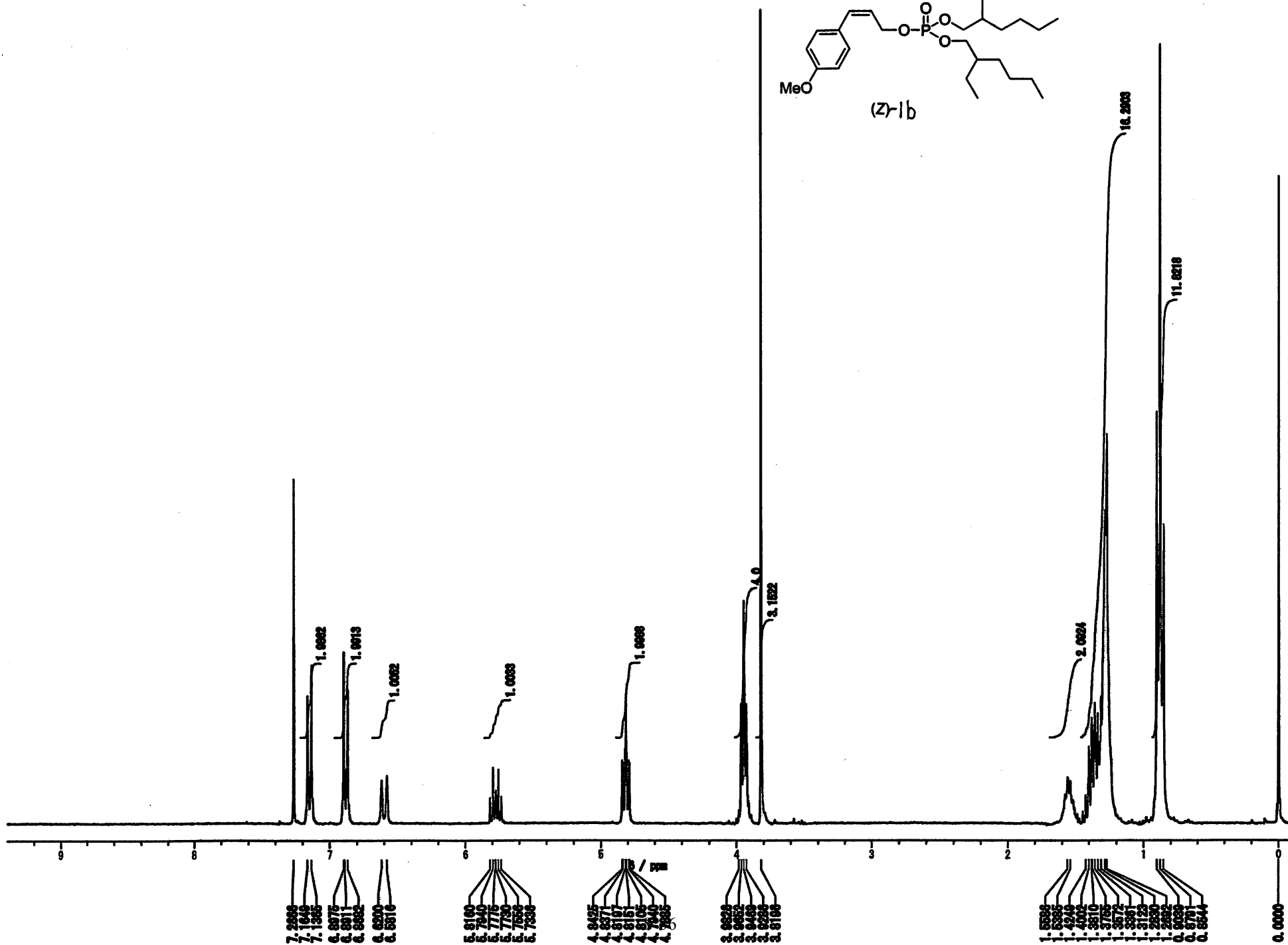
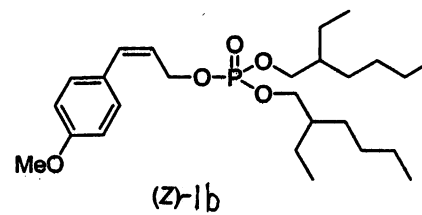


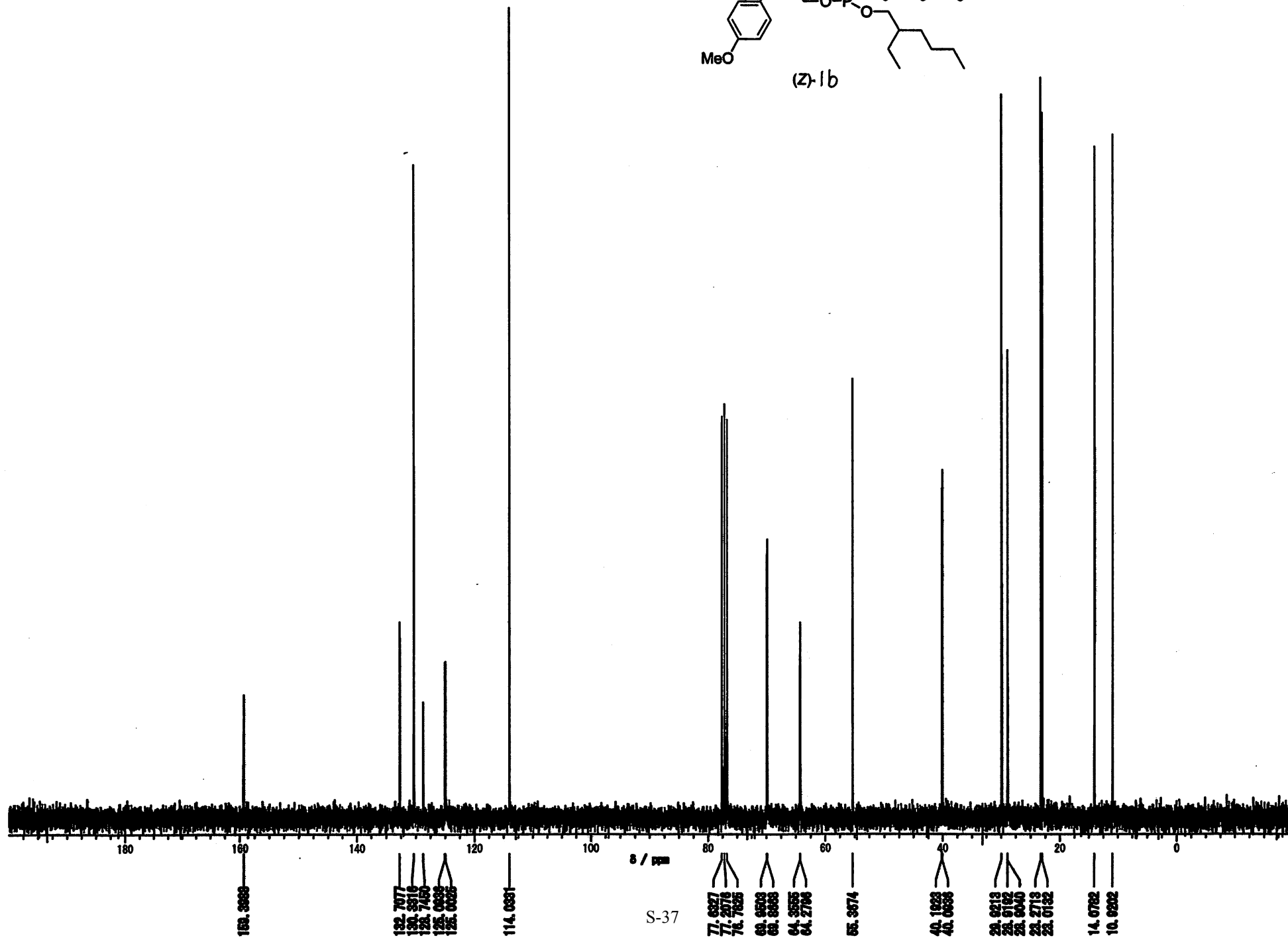
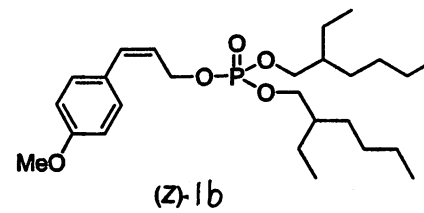


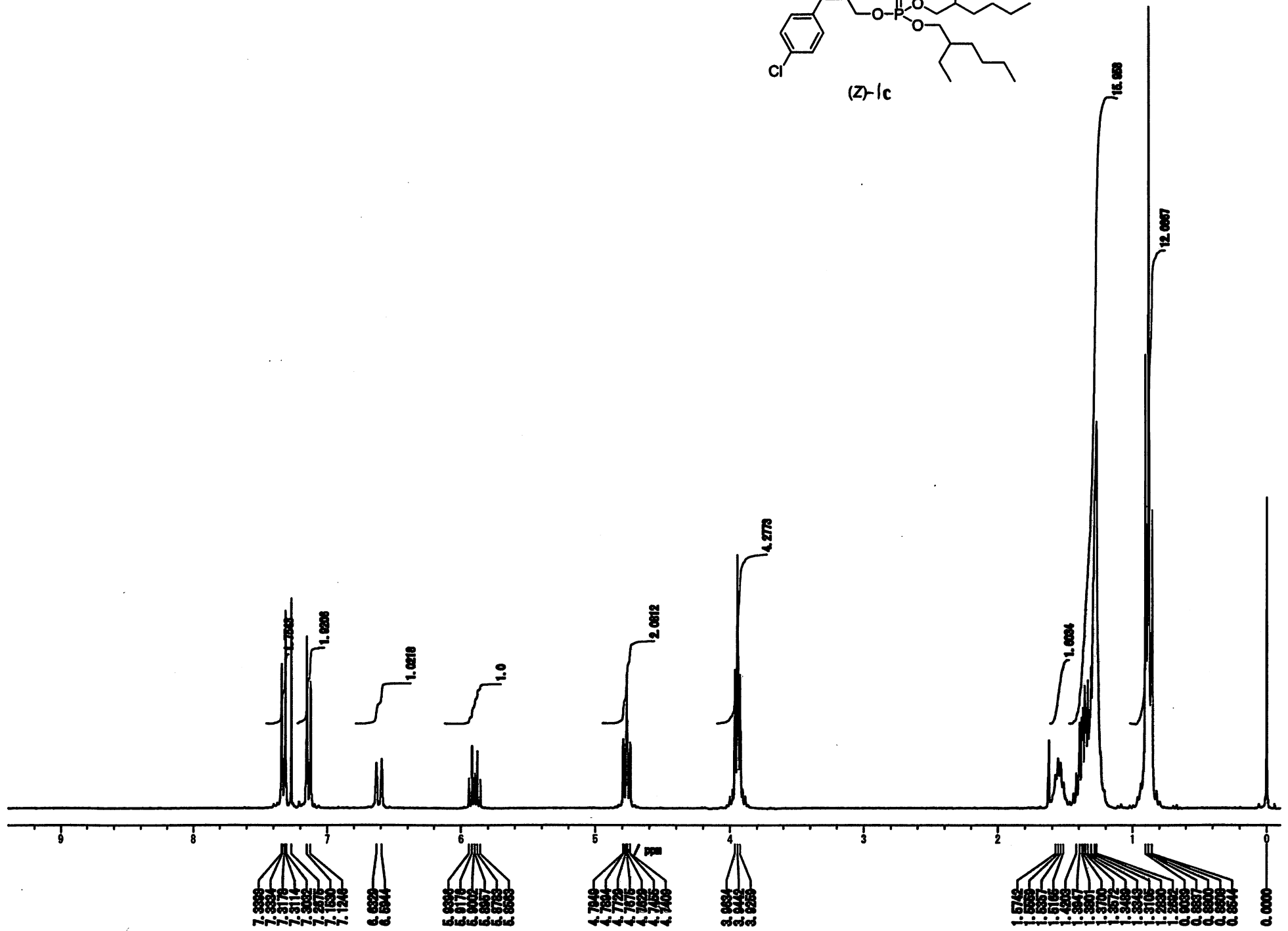
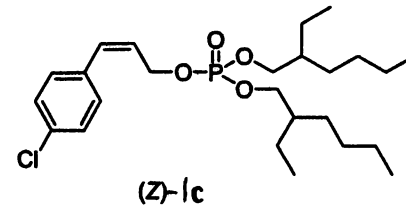


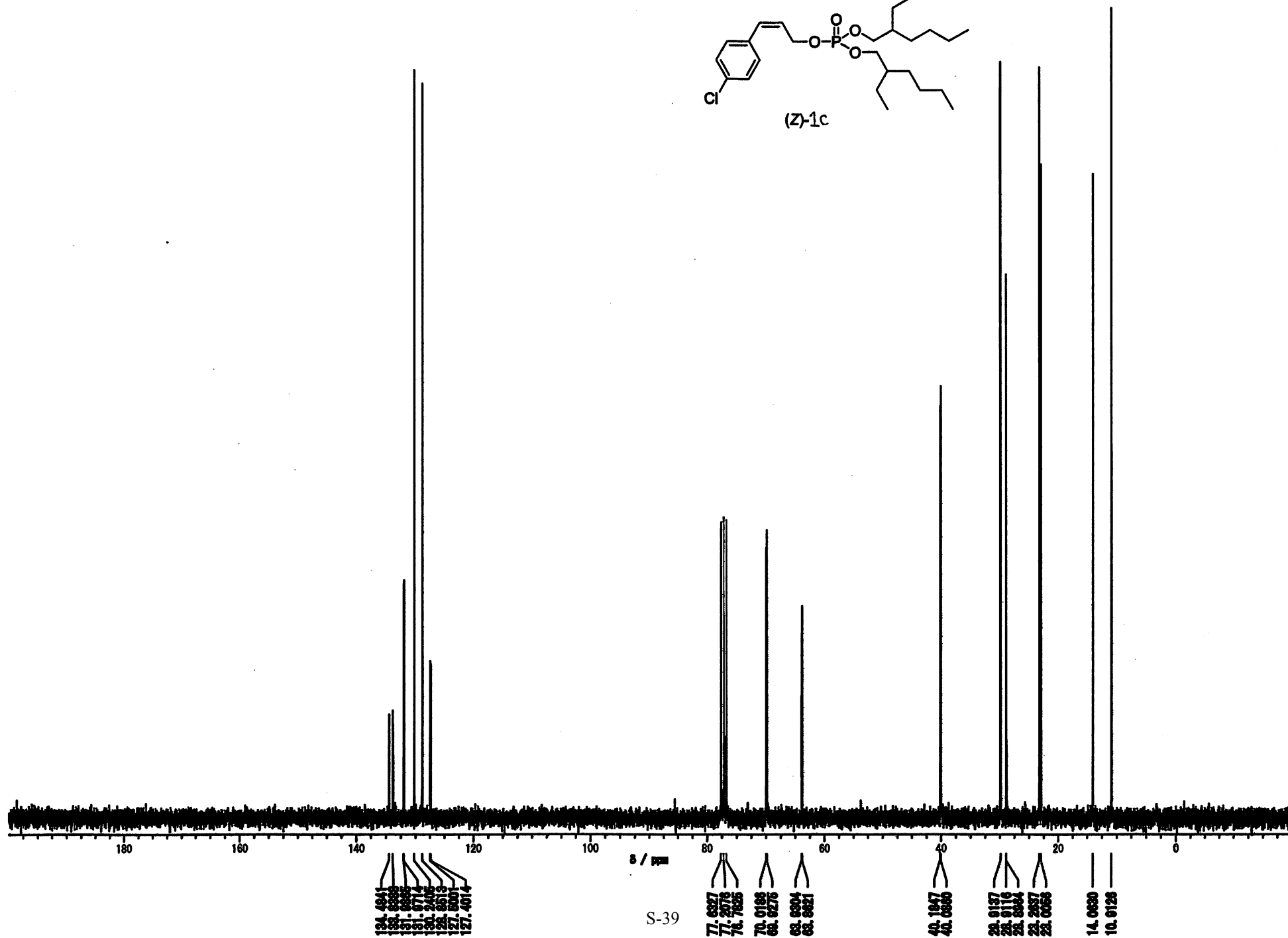
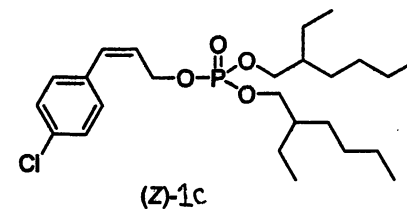


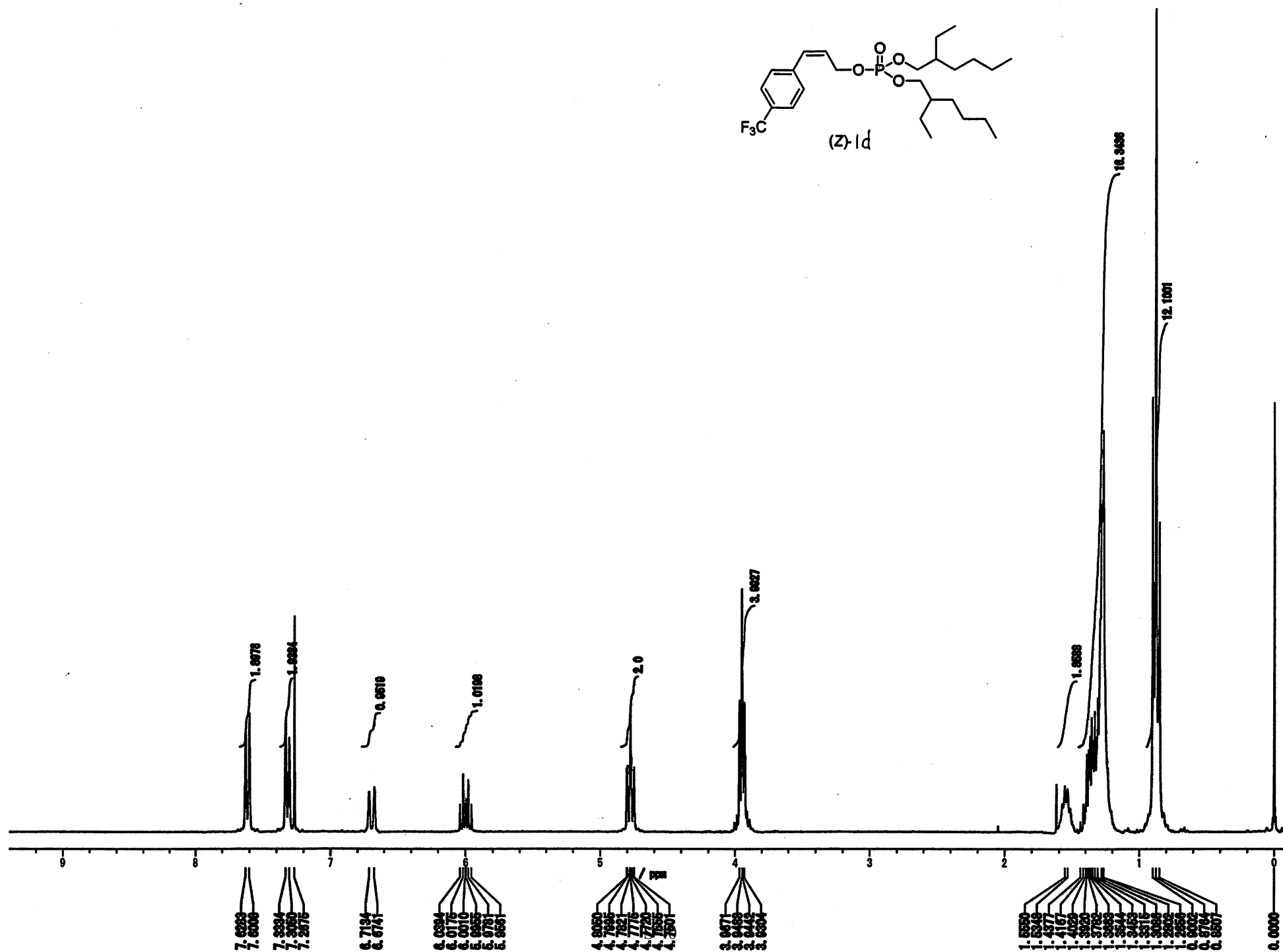
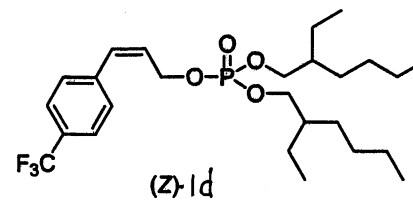


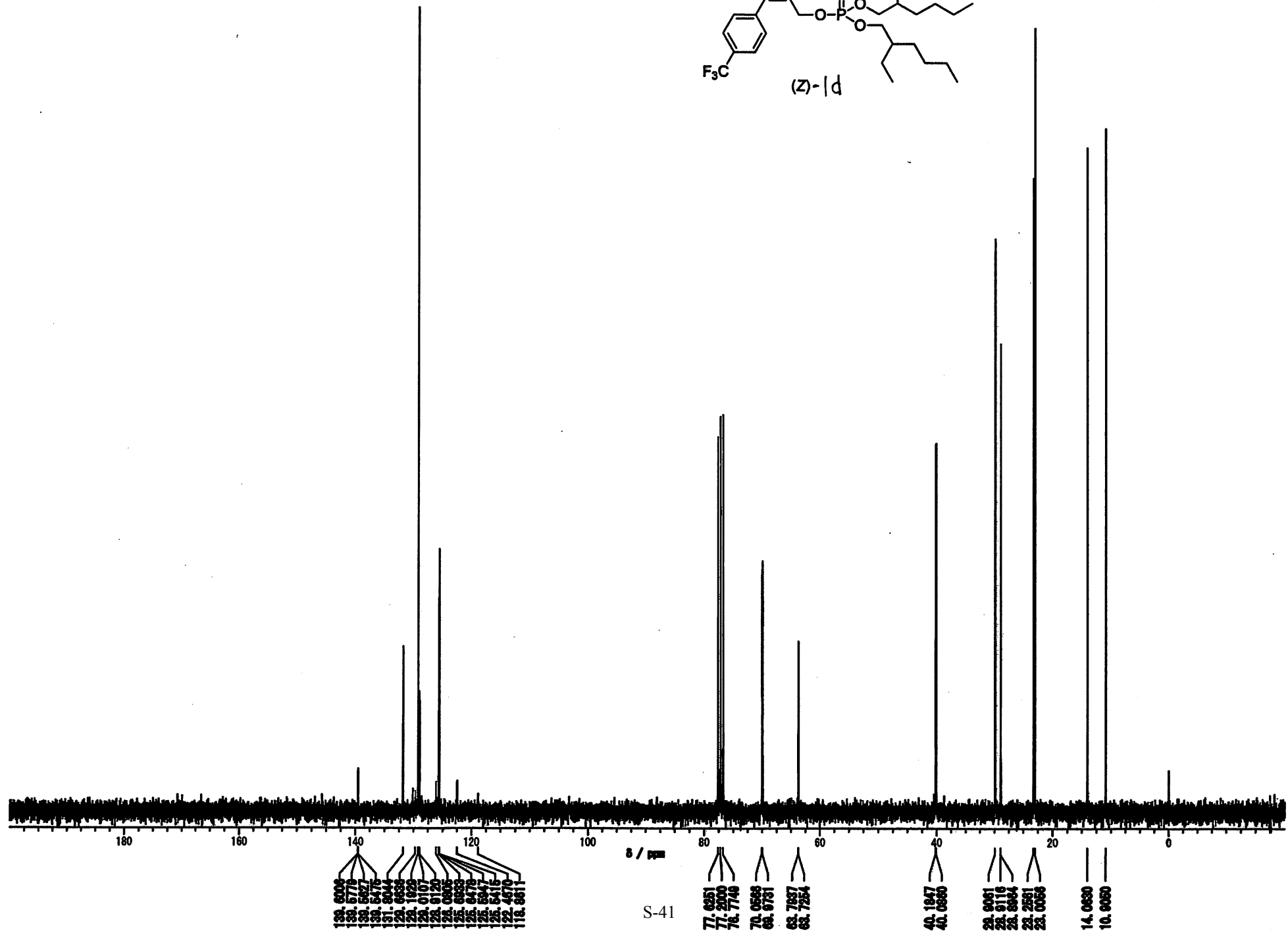
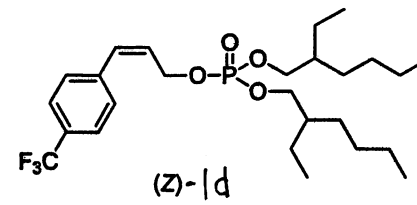


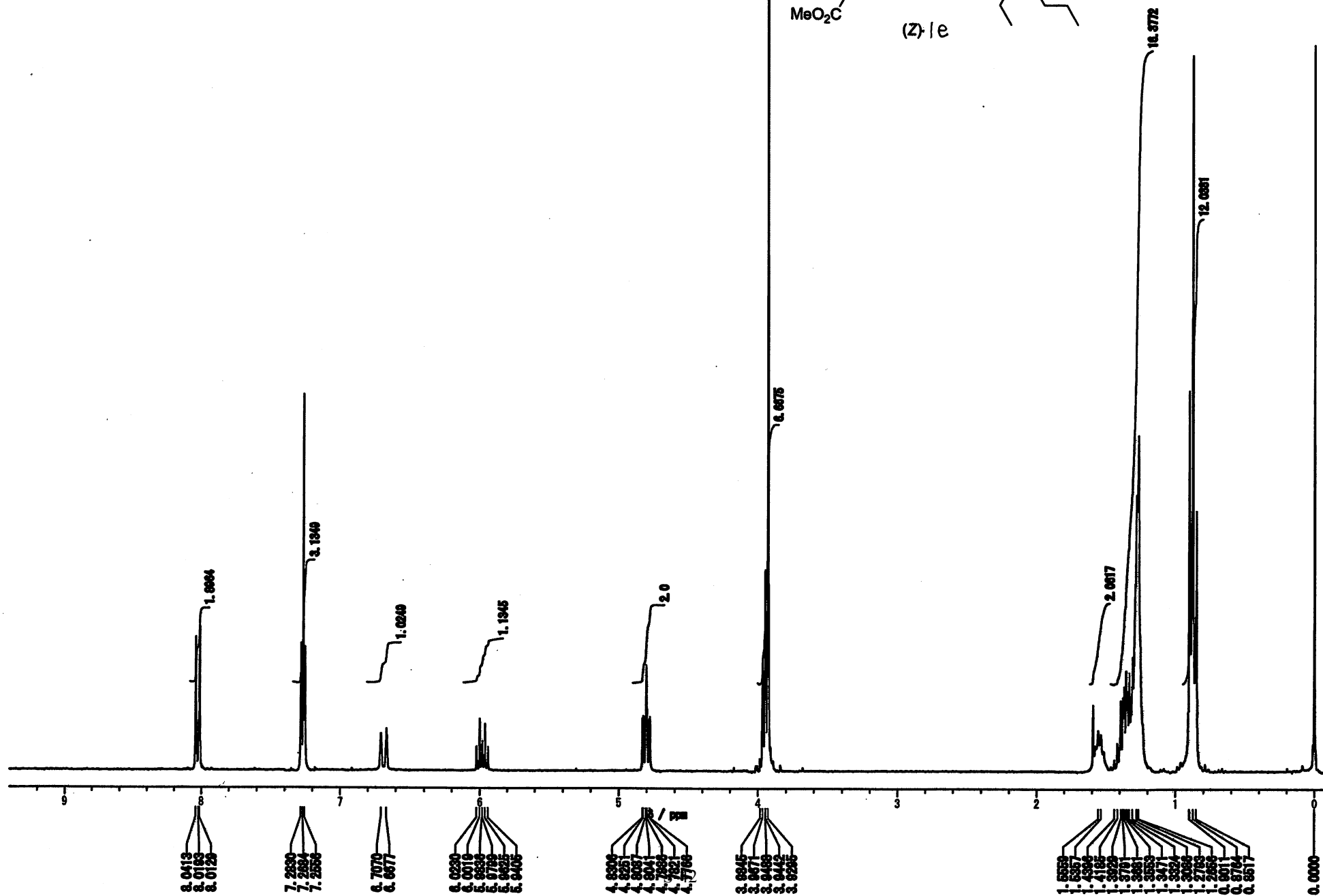
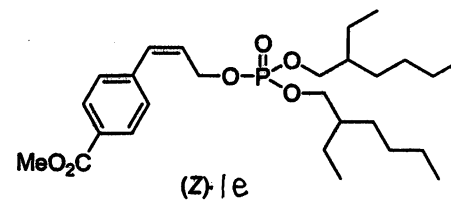


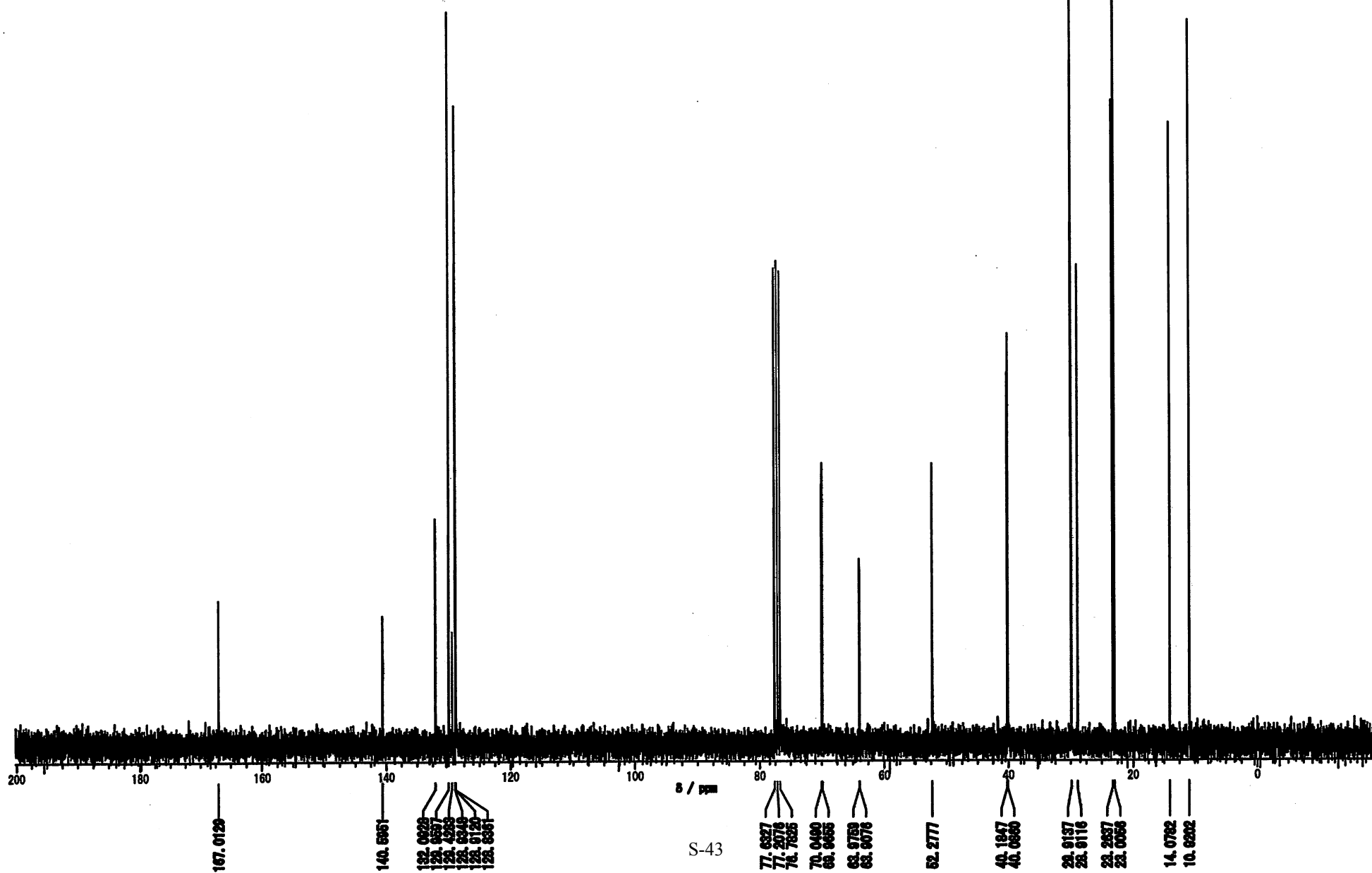
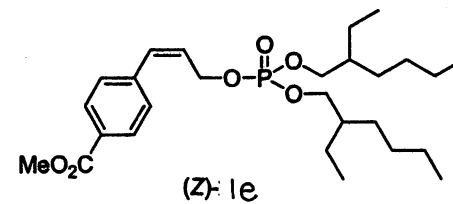


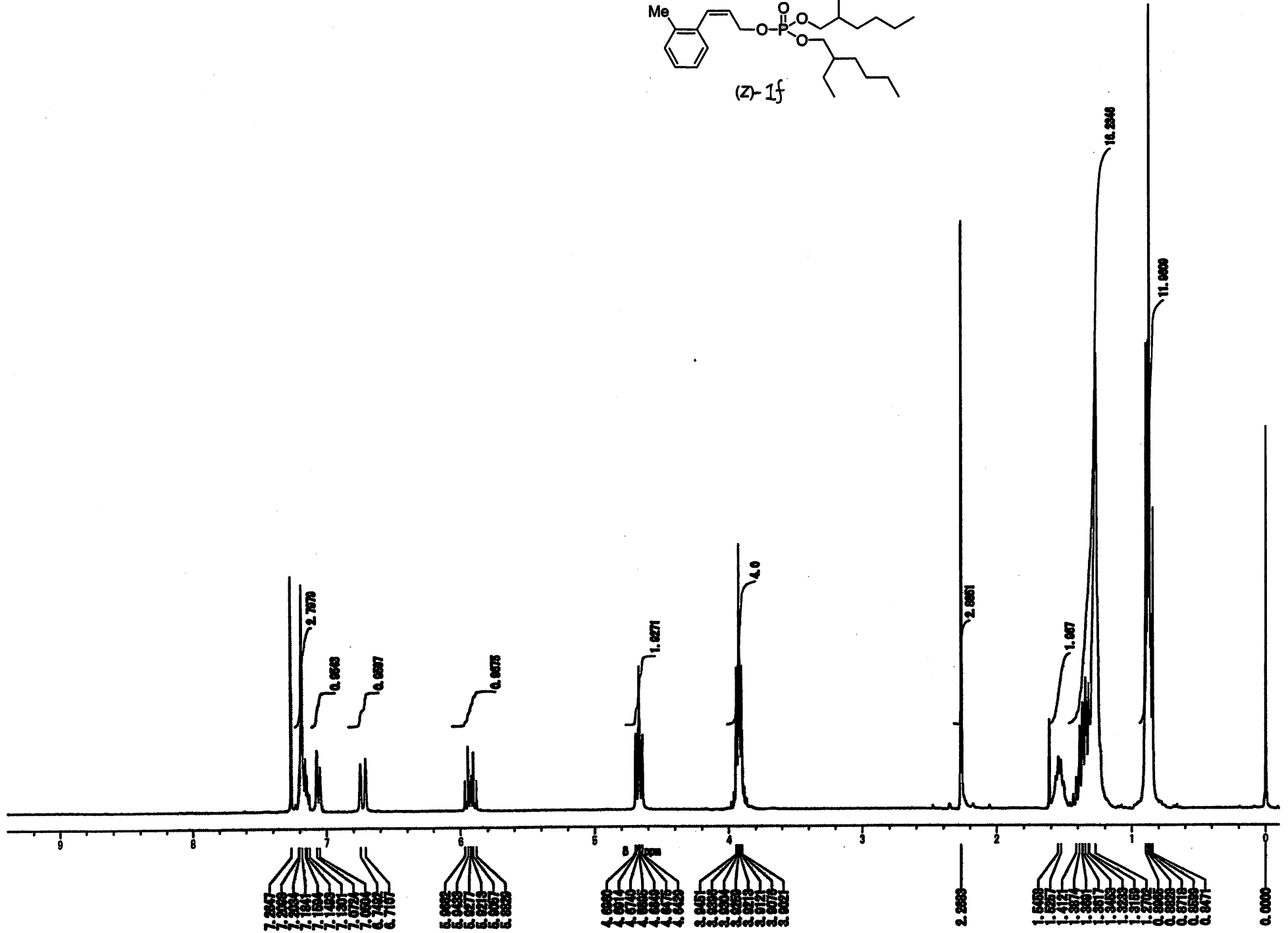
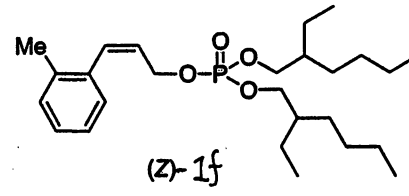


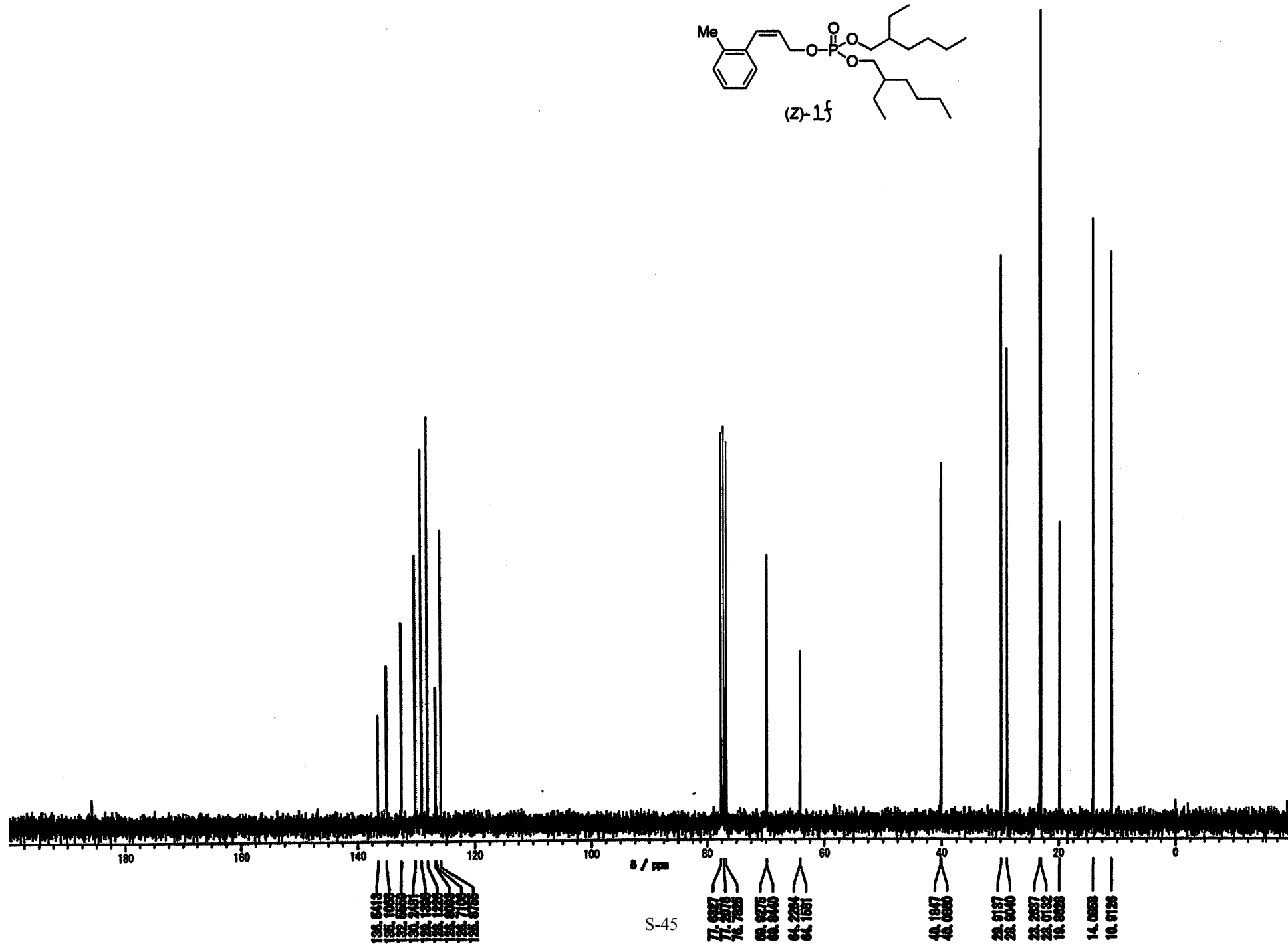
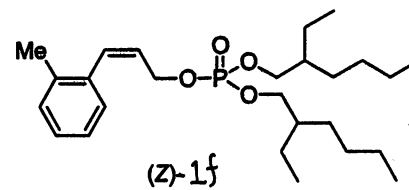


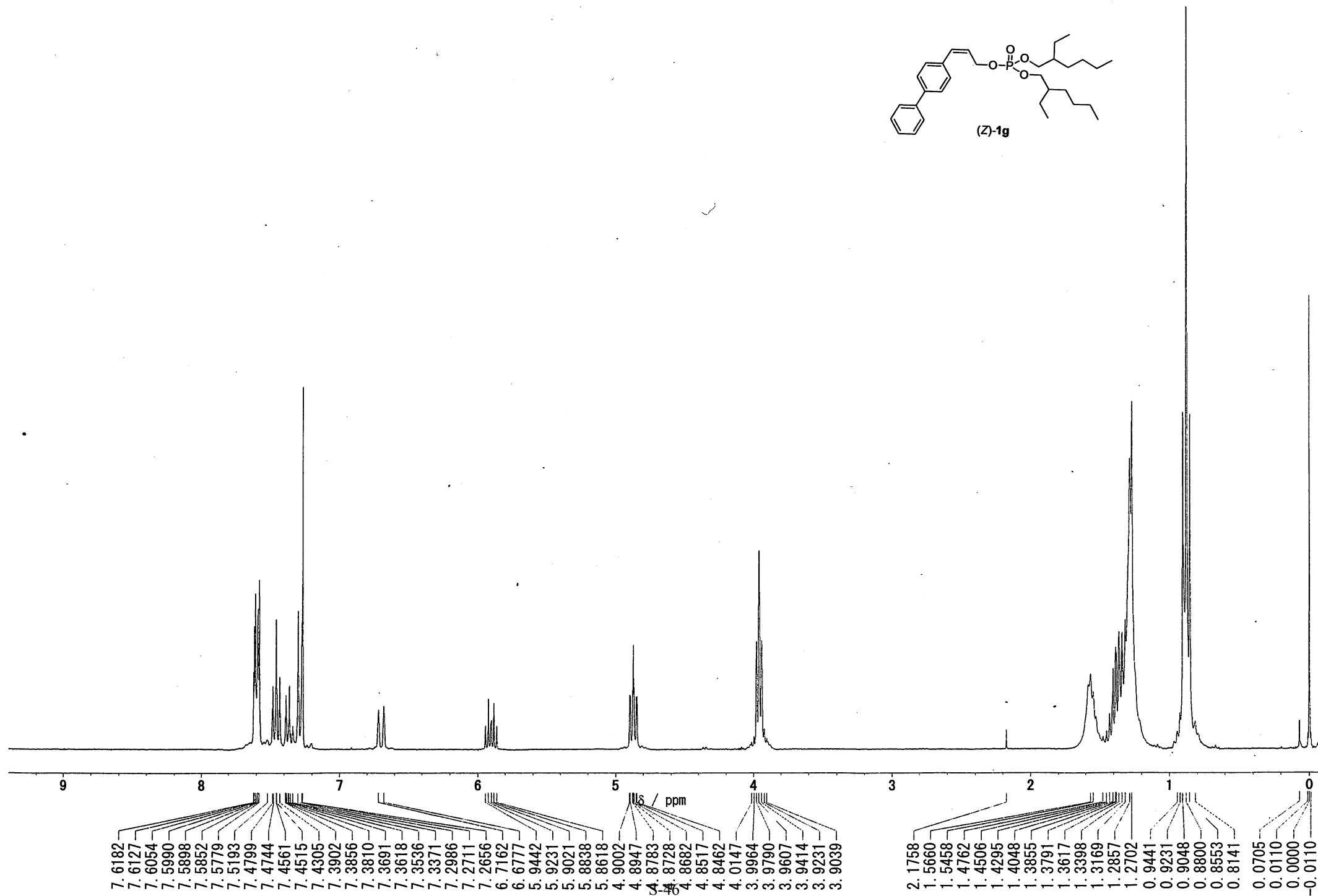
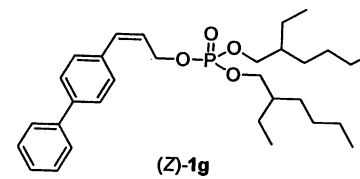


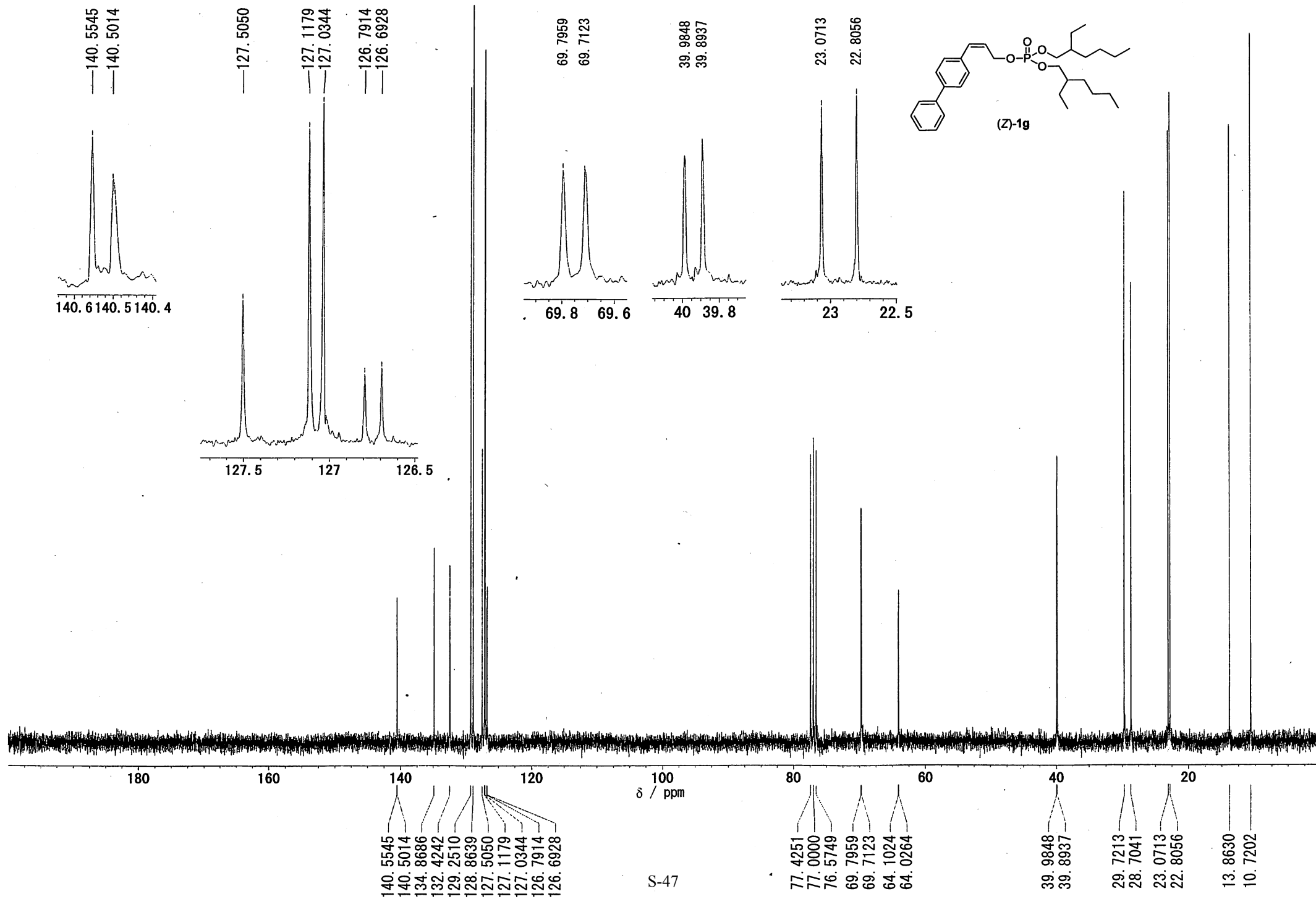


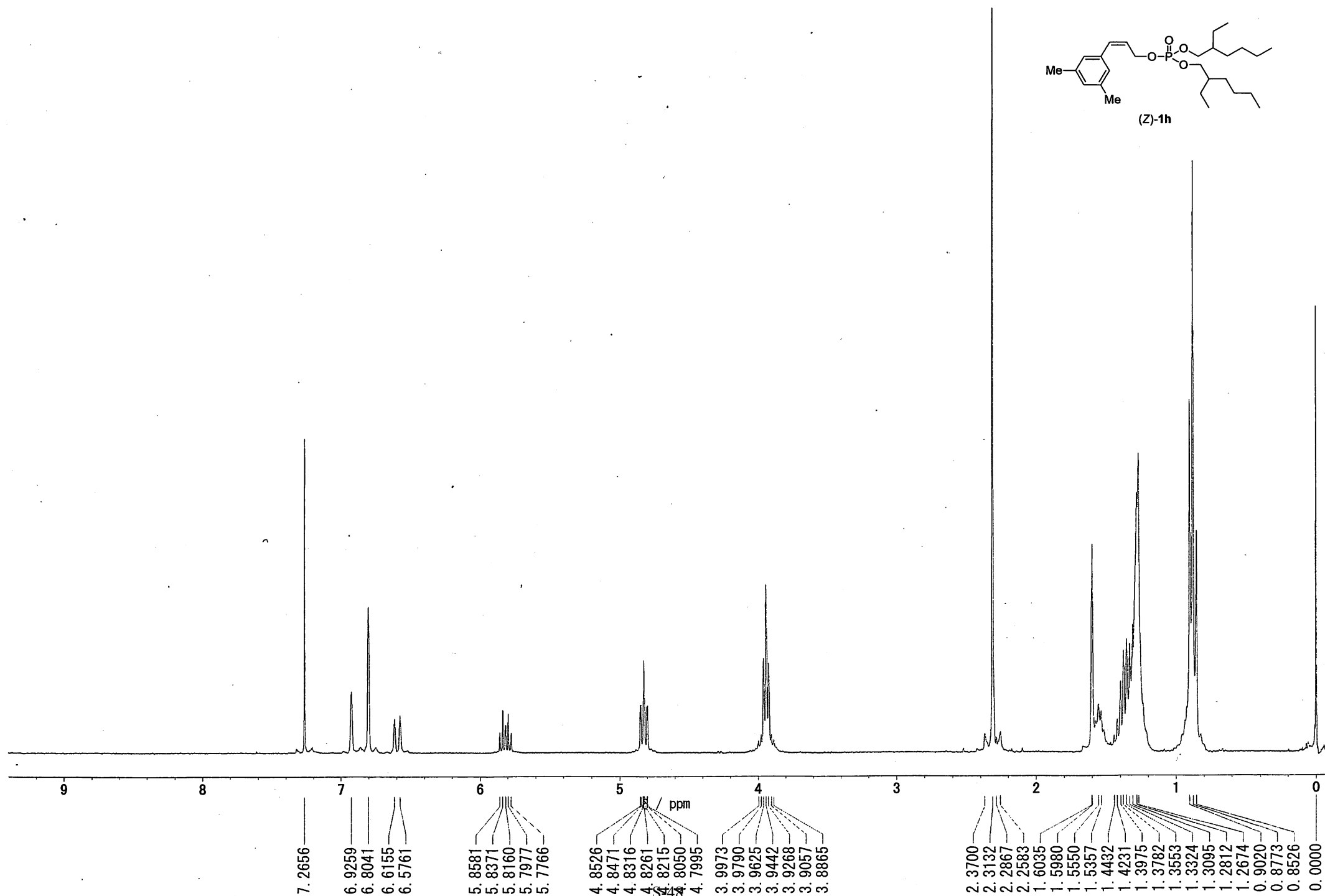
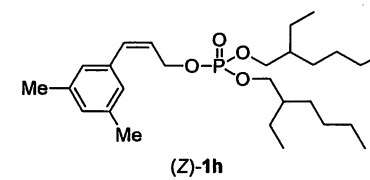


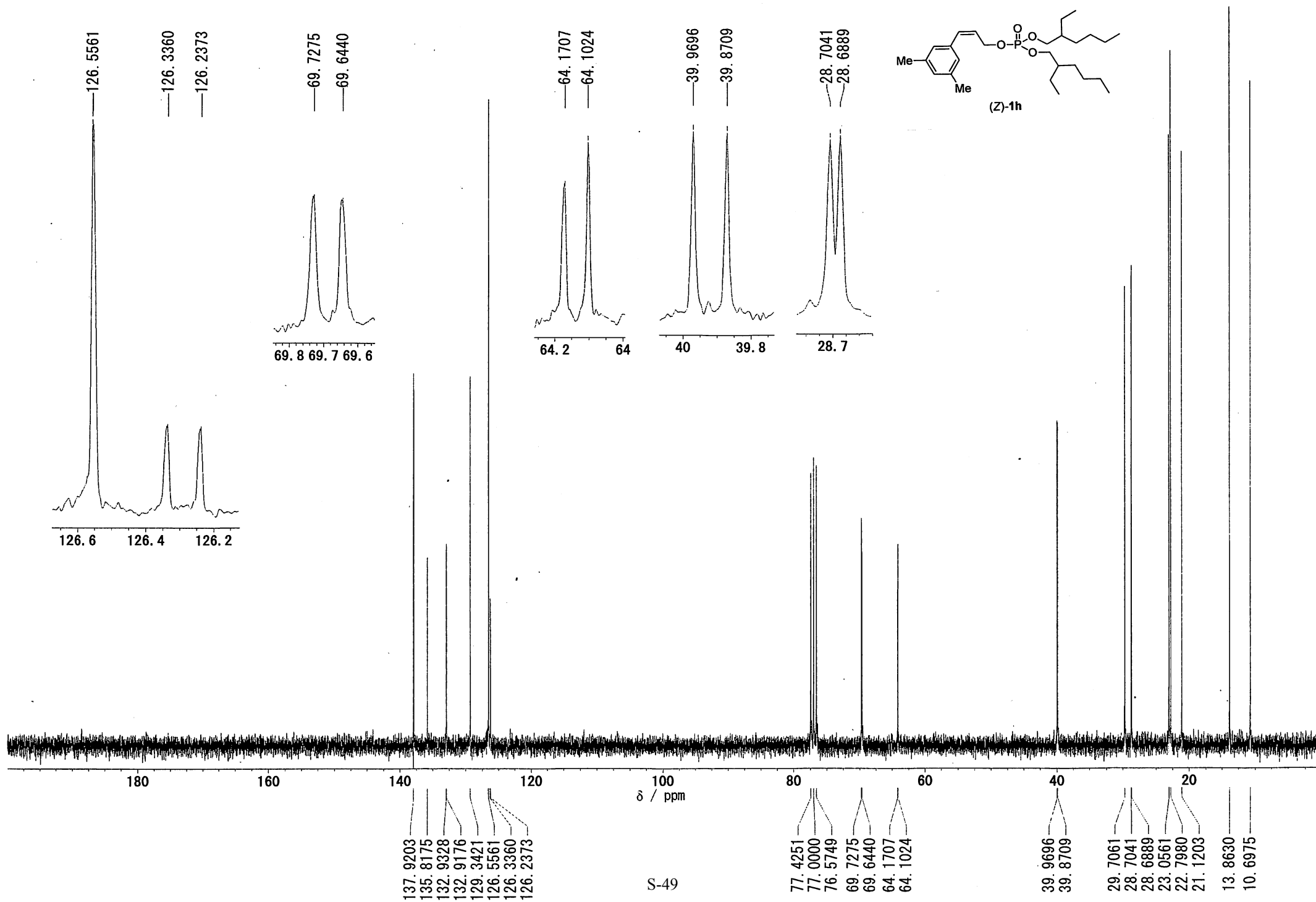


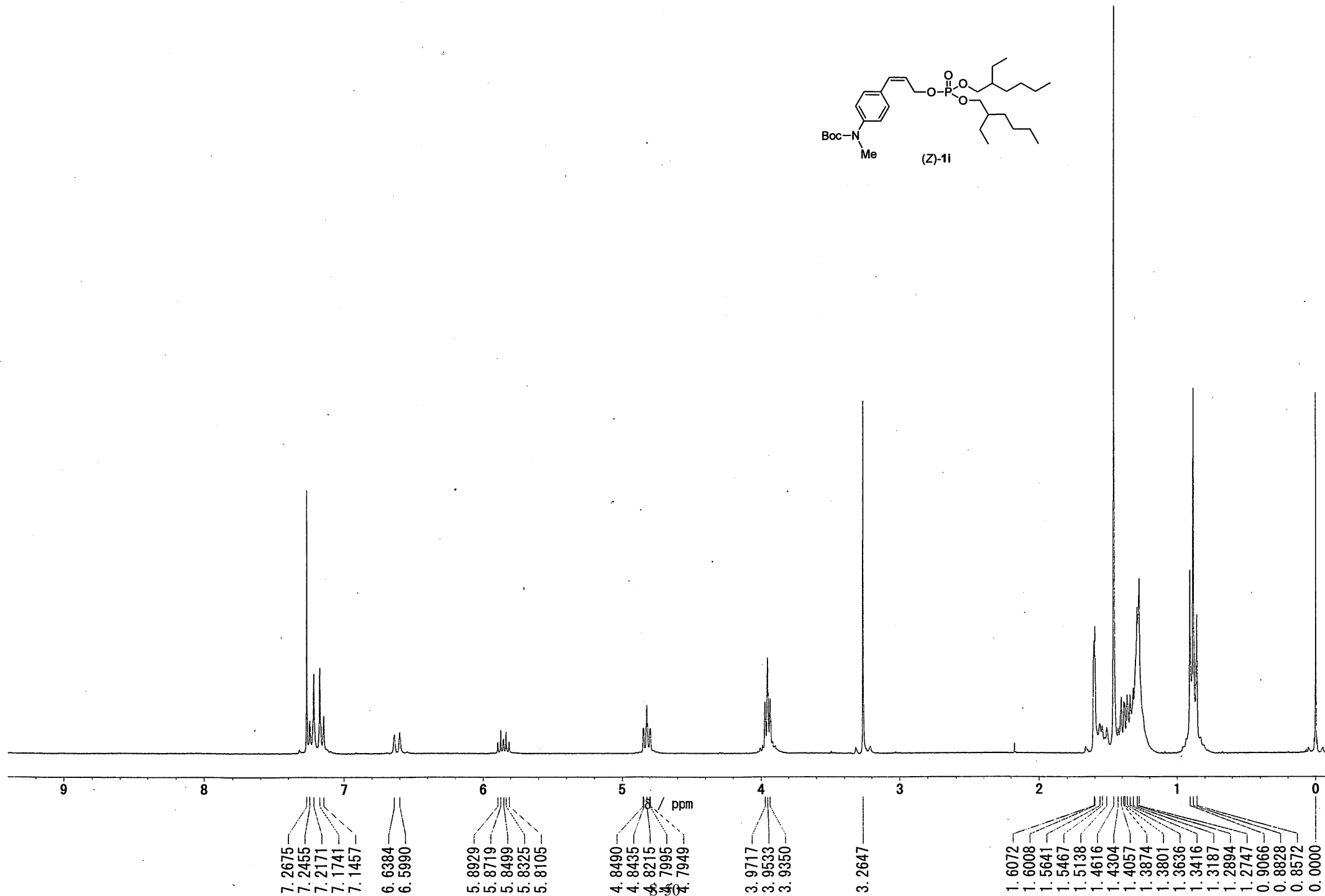
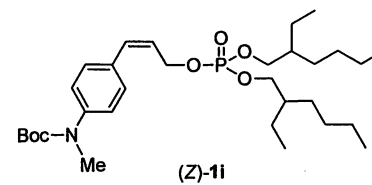


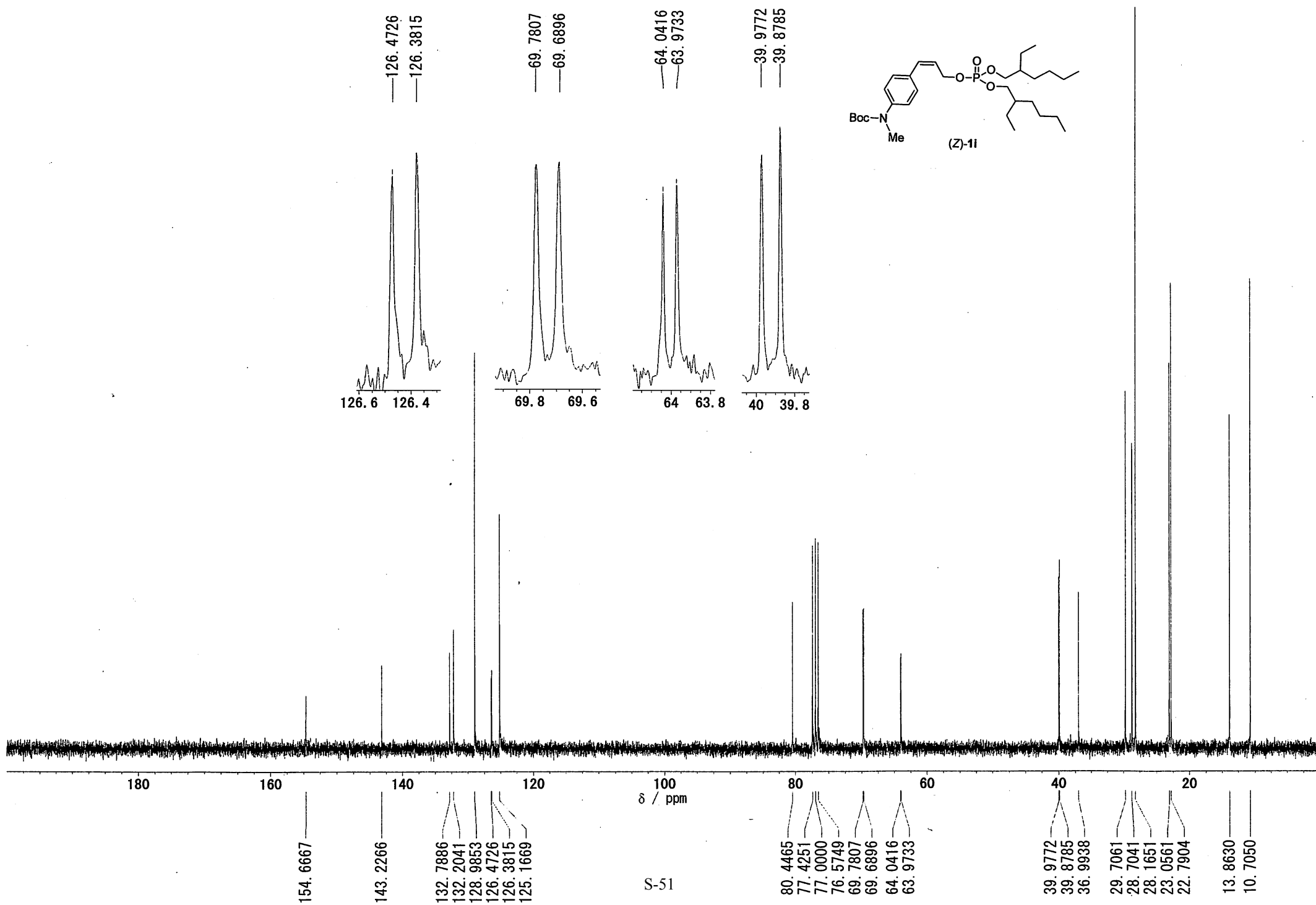


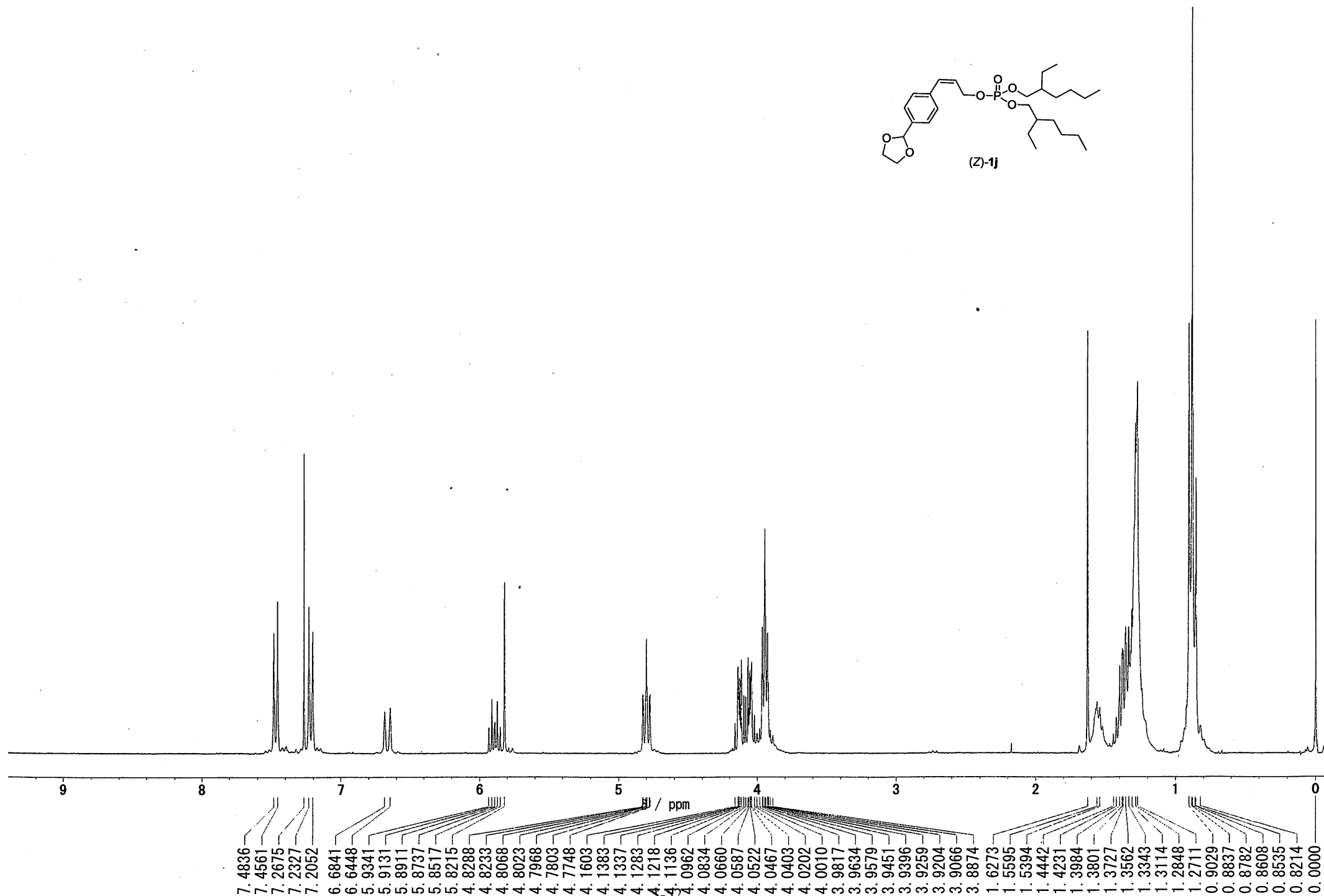
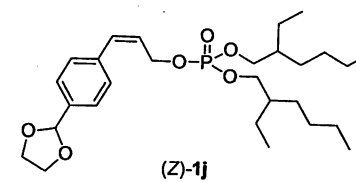


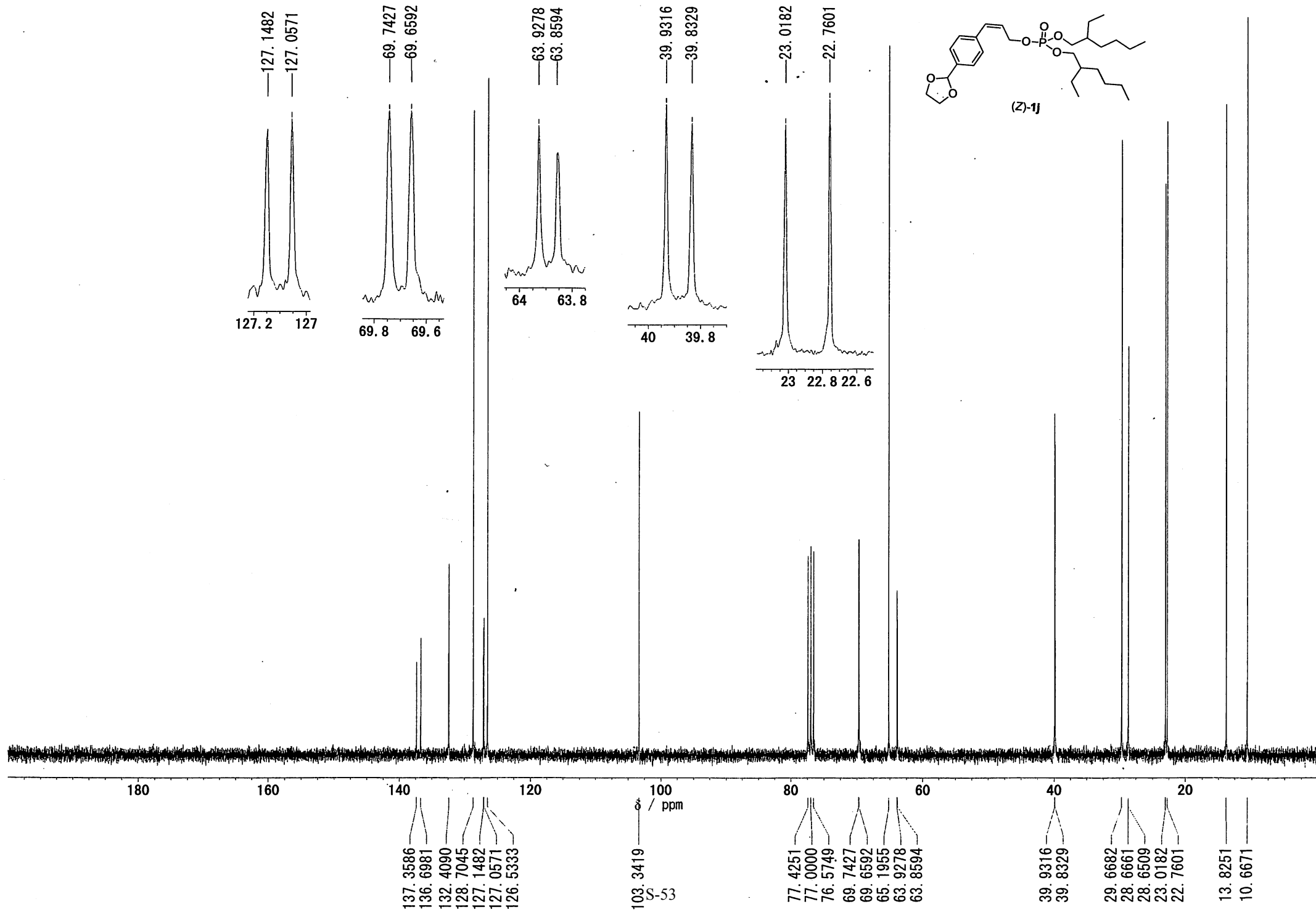


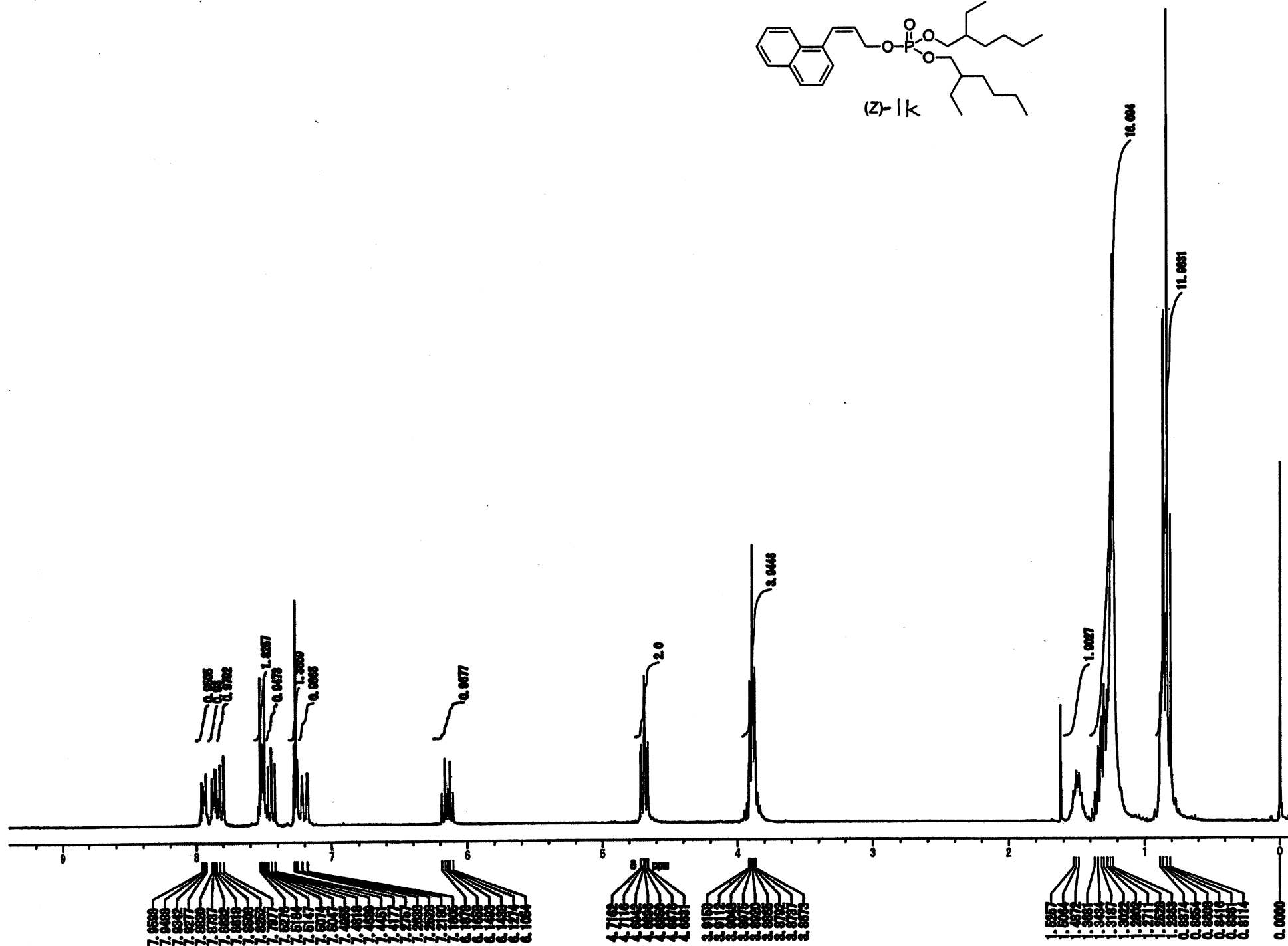
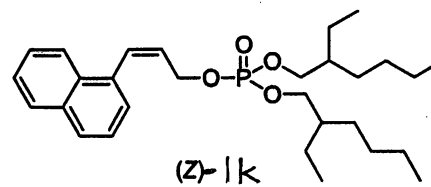


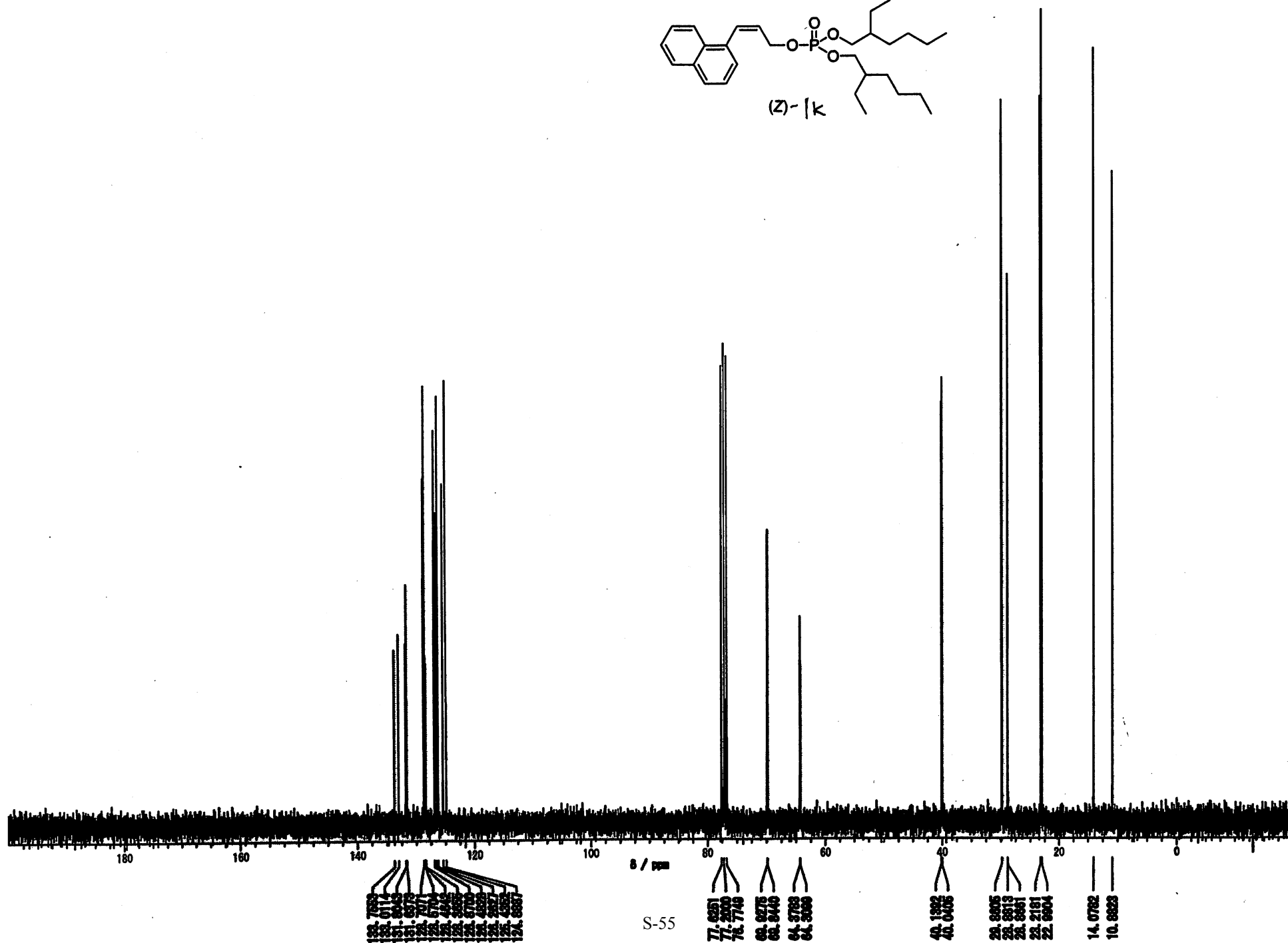
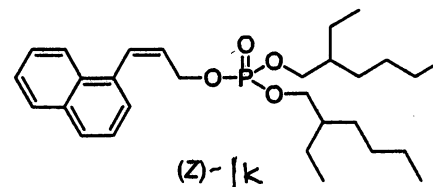


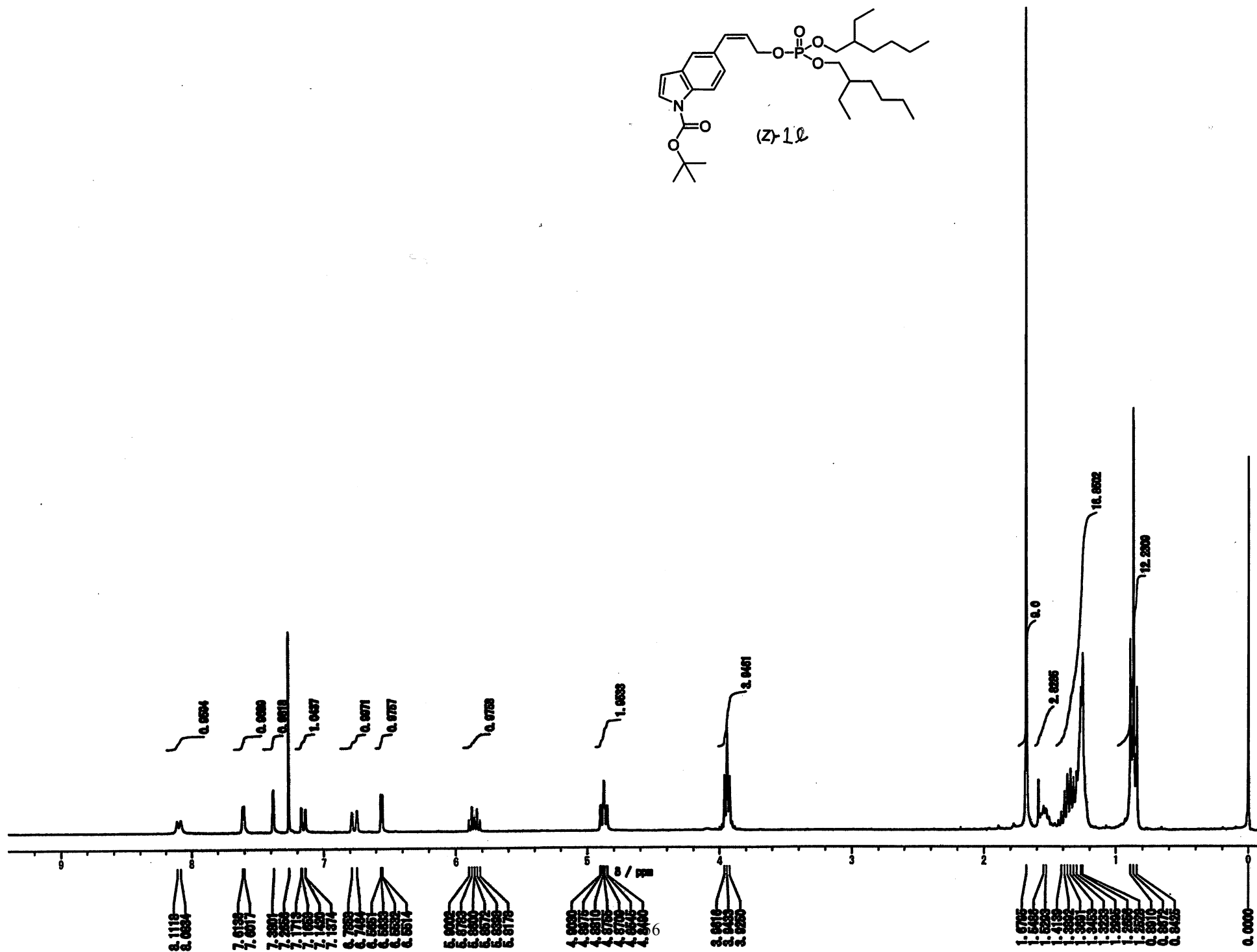
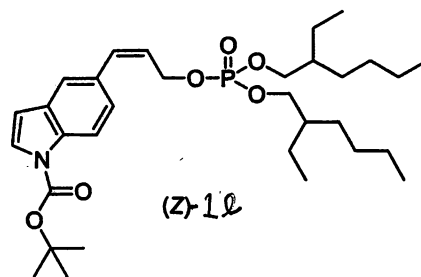


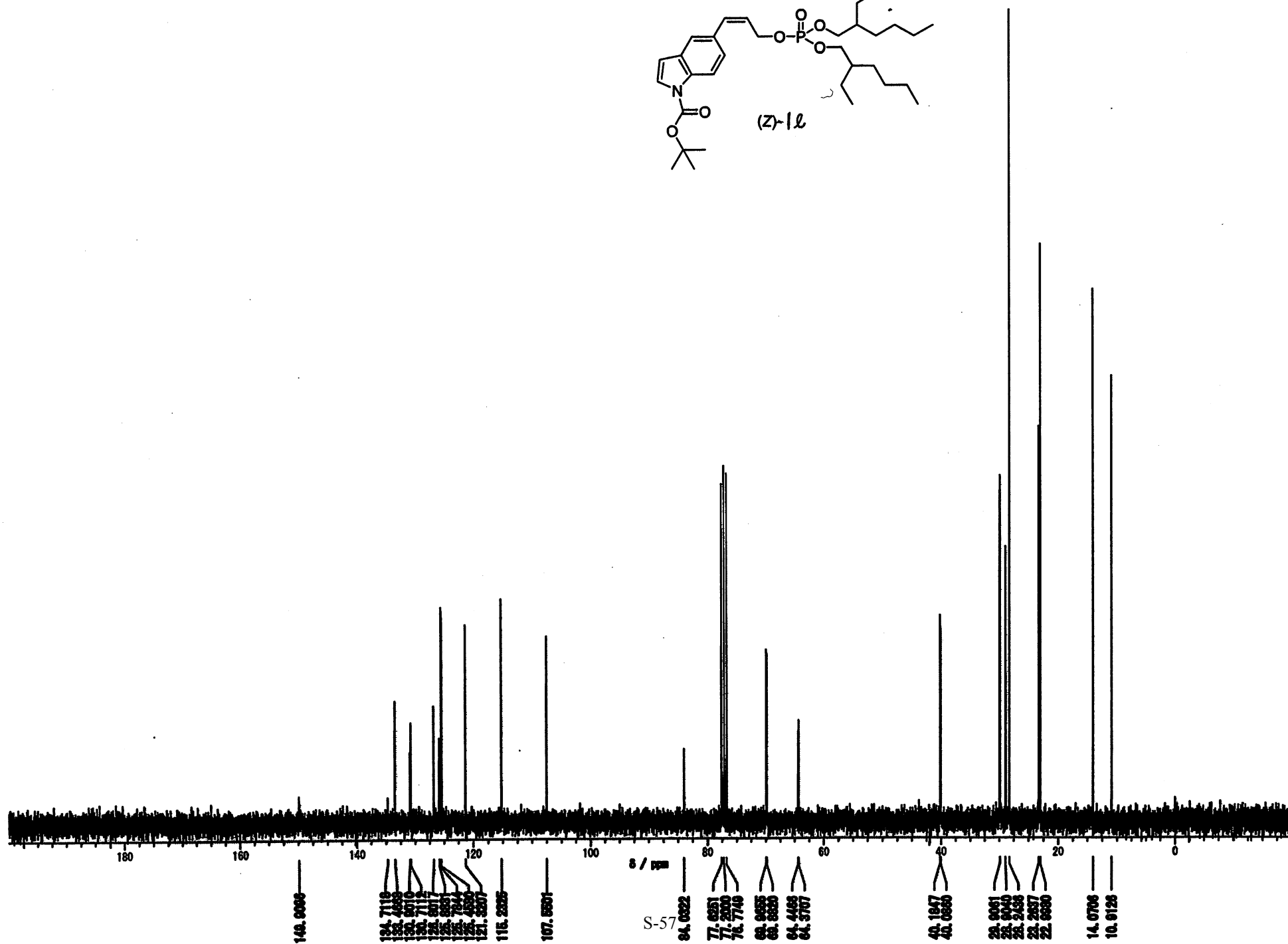
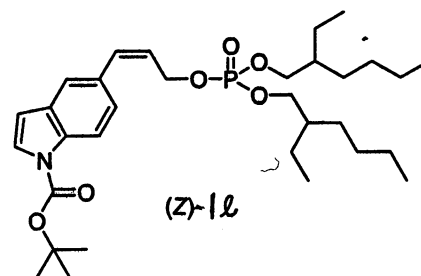


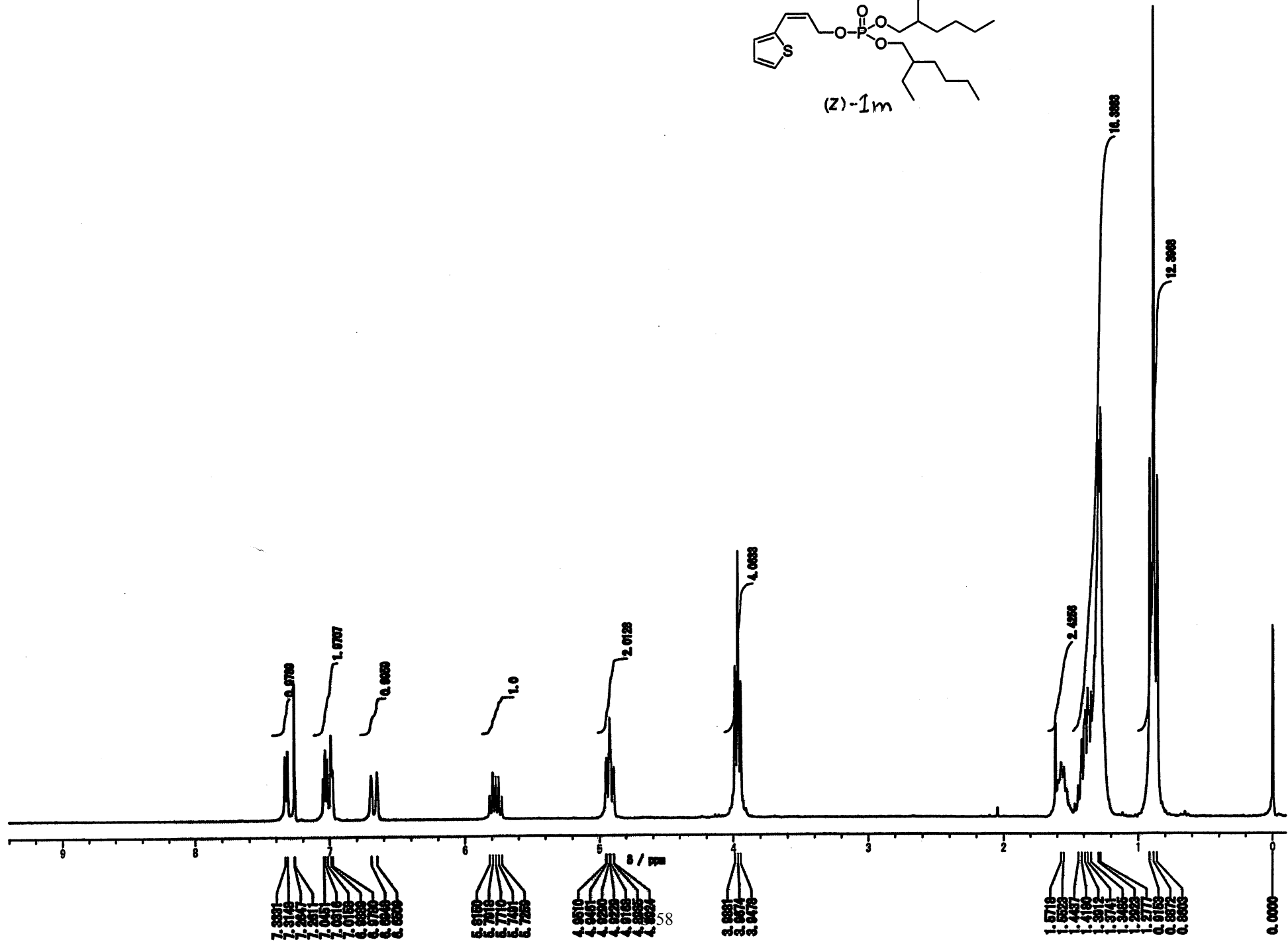
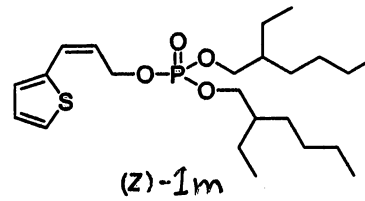


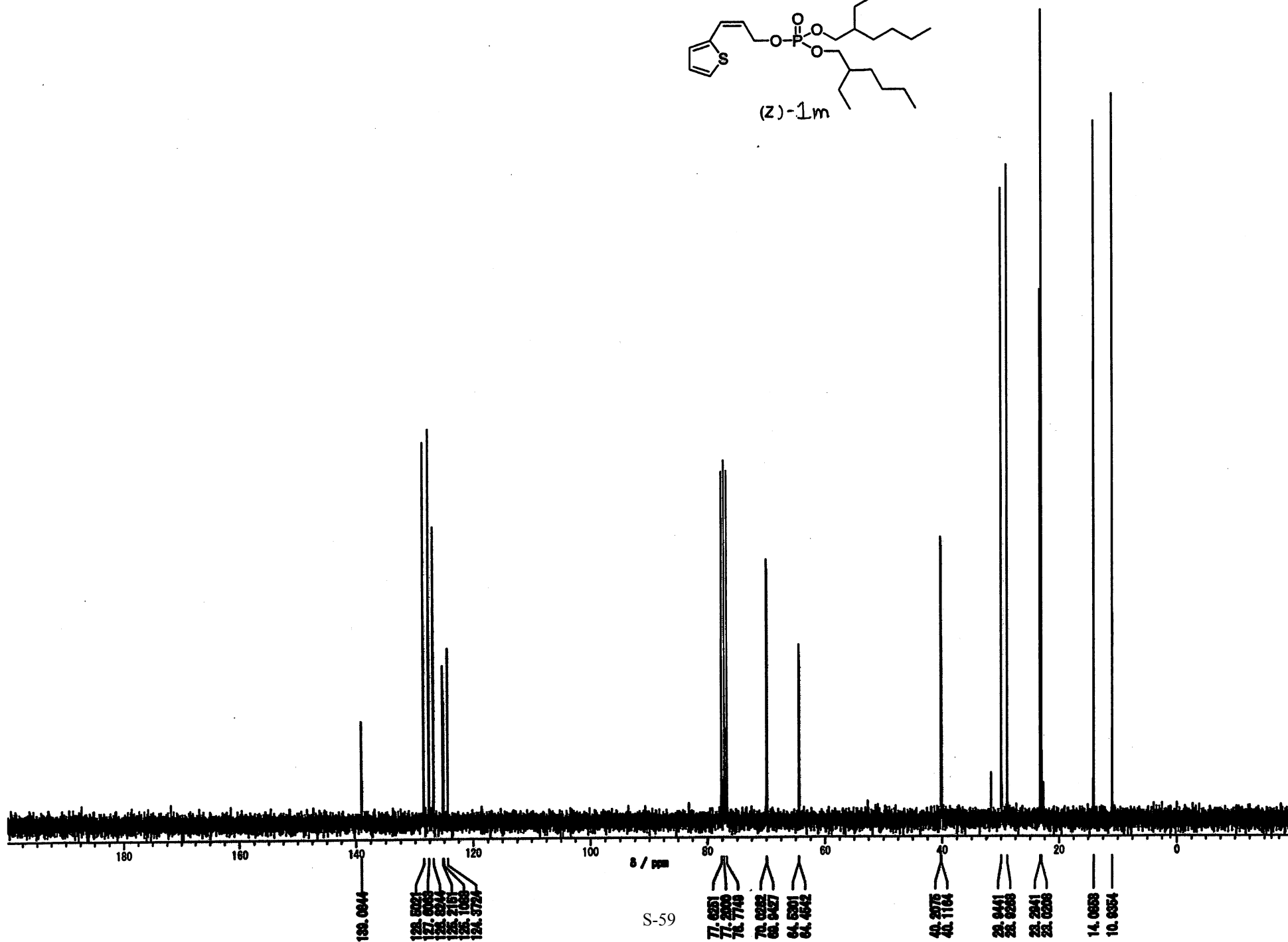
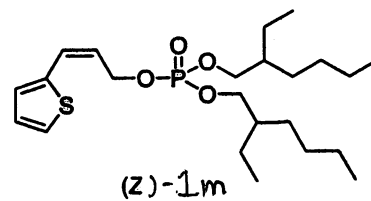




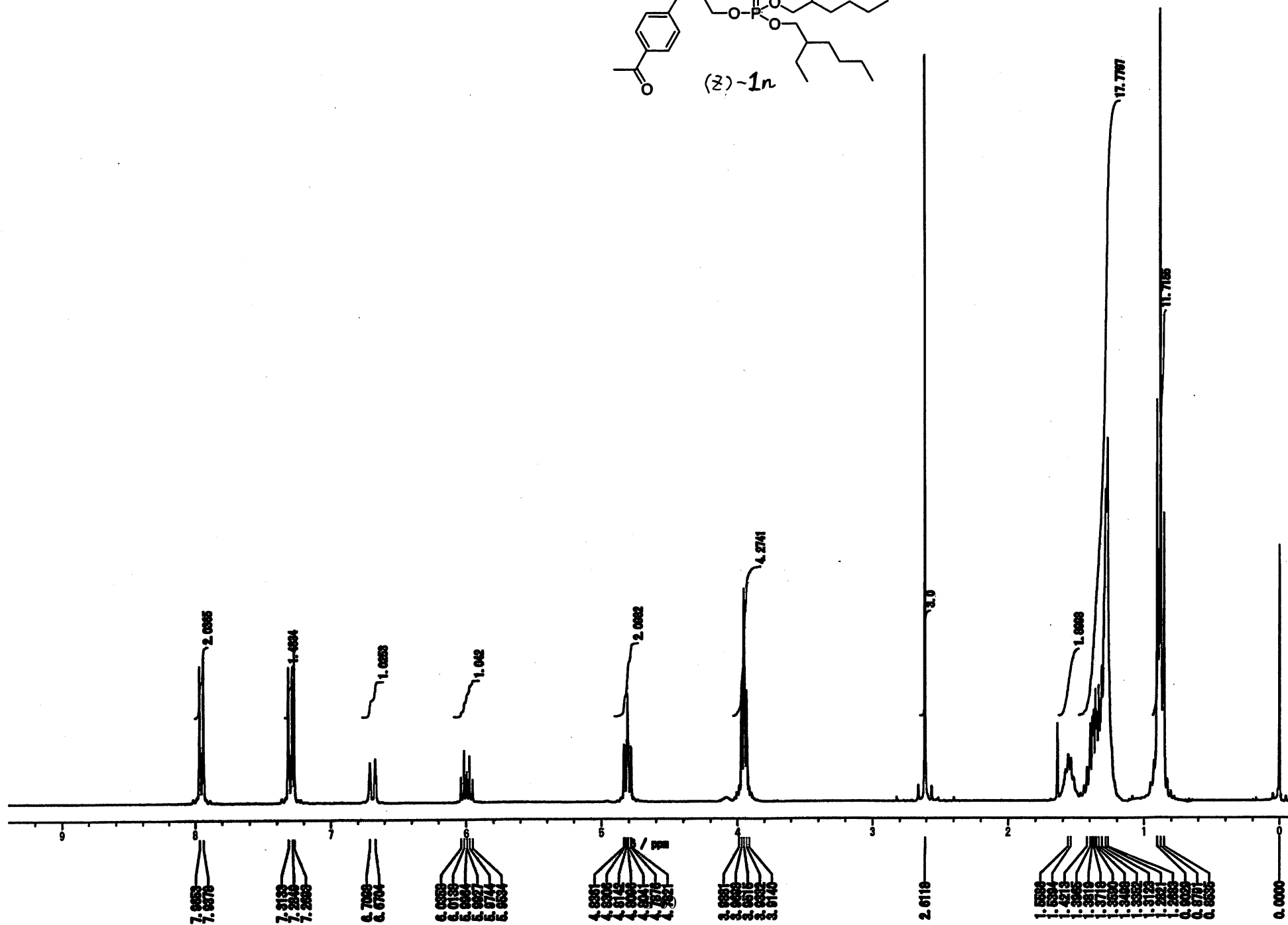
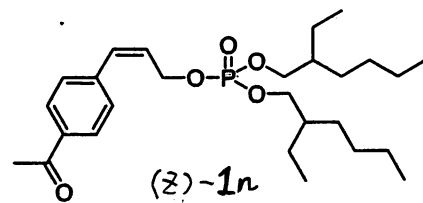


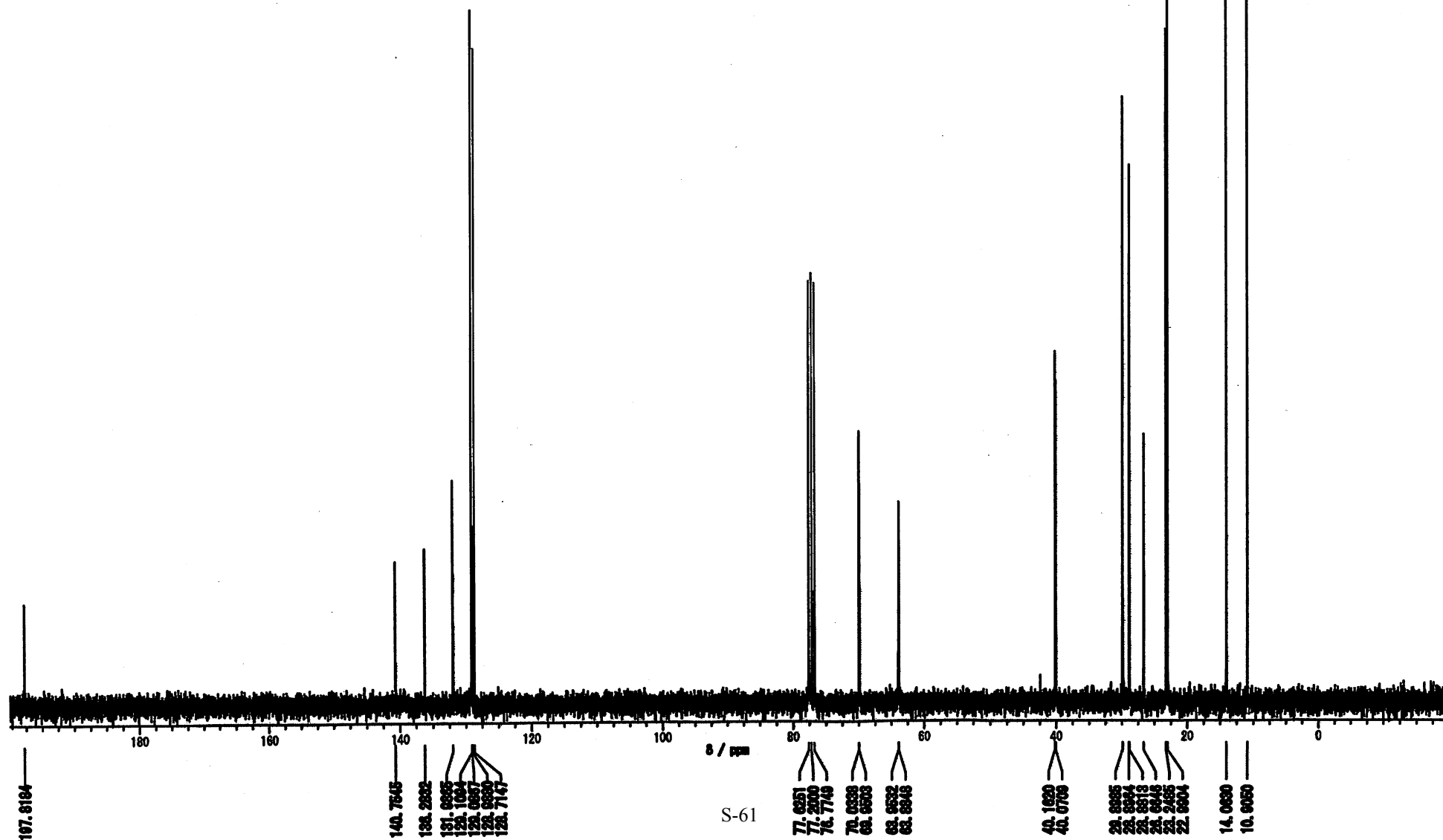
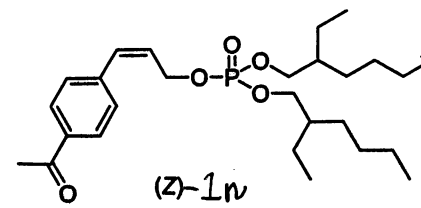


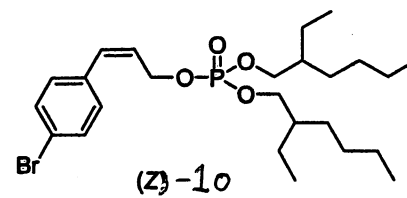




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