1,2-*Trans*-Selective Synthesis of Glycosyl Boranophosphates and Their Utility as Building Blocks for the Synthesis of Phosphodiester-linked Disaccharides

Kazuki Sato, Natsuhisa Oka, Shoichi Fujita, Fumiko Matsumura and Takeshi Wada*

Department of Medical Genome Sciences, Graduate School of Frontier Sciences, The University of Tokyo, Bioscience Building 702, 5-1-5 Kashiwanoha, Kashiwa, Chiba 277-8562, Japan

wada@k.u-tokyo.ac.jp

Supporting Information

Table of contents

General information	S 1
Experimental procedures 1. Preparation of per- <i>O</i> -acyl glycosides	S1
2. Synthesis of authentic sample of dimethyl 2,3,4,6-tetra- O -benzoyl- α -D-mannopyranosyl boranophosphate	S4
References	S5
¹ H, ¹³ C and ³¹ P NMR spectra	S 6

General information

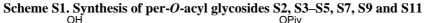
All the reactions were conducted under Ar atmosphere. Dry organic solvents were prepared by appropriate procedures. ¹H NMR spectra were recorded at 300 MHz with tetramethylsilane (δ 0.0) as an internal standard in CDCl₃. ¹³C NMR spectra were recorded at 75.5 MHz with CDCl₃ (δ 77.0) as an internal standard in CDCl₃. ³¹P NMR spectra were recorded at 121.5 MHz with H₃PO₄ (δ 0.0) as an external standard. COSY and HMQC were used to confirm the NMR peak assignments. Silica gel column chromatography was carried out using spherical, neutral, 63–210 or 40–50 µm silica gel unless otherwise noted. Analytical TLC was performed on commercial glass plates bearing 0.25 mm layer of silica gel.

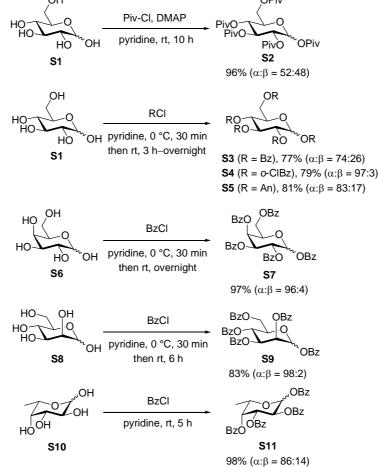
Experimental procedures

1. Preparation of per-O-acyl glycosides

1,2,3,4,6-Penta-O-pivaloyl-D-glucopyranoside (S2).

Pivaloyl chloride (12.0 mL, 99 mmol) and catalytic amount of DMAP were added to a solution of D-glucose (**S1**, 1.80 g, 10 mmol) in dry pyridine (40 mL) at rt while stirring. After 10 h, the solvent was evaporated under reduced pressure, and the residue was diluted with CH₂Cl₂ (120 mL). The mixture was washed with saturated NaHCO₃ aqueous solutions (3×100 mL), dried over Na₂SO₄, filtered and concentrated under reduced pressure. The residue was recrystallized from methanol and purified by silica gel column chromatography using AcOEt–hexane (1:4, v/v) as an eluent to give **S2** as a colorless foam (5.73 g, 0.95 mmol, α : β = 52:48, 96%). The ¹H NMR spectrum was identical to the reported data.¹





1,2,3,4,6-Penta-O-benzoyl-D-glucopyranoside (S3).

Benzoyl chloride (13.9 mL, 120 mmol) was added dropwise to a solution of D-glucose (**S1**, 3.60 g, 20 mmol) in dry pyridine (24 mL) at 0 °C while stirring. After being kept at 0 °C for 30 min, the mixture was allowed to warm to rt and stirred for 3 h. The reaction was quenched by adding a saturated NaHCO₃ aqueous solution (100 mL) and methanol (10 mL), and CH₂Cl₂ (100 mL) was added to the mixture. The organic layer was separated, and washed with saturated NaHCO₃ aqueous solutions (3 × 150 mL). The aqueous layers were combined and back-extracted with CH₂Cl₂ (150 mL). The organic layers were combined, dried over Na₂SO₄, filtered and concentrated under reduced pressure. The residue was recrystallized from ethanol (75 mL) to give **S3** as a colorless solid (10.8 g, 15 mmol, α : β = 74:26, 77%). The ¹H NMR spectrum was identical to the reported data.²

1,2,3,4,6-Penta-O-(2-chlorobenzoyl)-D-glucopyranoside (S4).

2-Chlorobenzoyl chloride (1.52 mL, 12 mmol) was added dropwise to a solution of D-glucose (**S1**, 0.362 g, 2.0 mmol) in dry pyridine (10 mL) at 0 °C while stirring. After being kept at 0 °C for 30 min, the mixture was allowed to warm to rt. After being stirred for 4.5 h at rt, the reaction was quenched with methanol (1 mL), and the mixture was diluted with CHCl₃ (10 mL). The mixture was washed with saturated NaHCO₃ aqueous solutions (3 × 15 mL). The aqueous layers were combined and back-extracted with CHCl₃ (30 mL). The organic layers were combined, dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The residue was recrystallized from ethanol (50 mL) to give **S4** as a colorless solid (1.39 g, 1.6 mmol, α : β = 97:3, 79%). IR (KBr, cm⁻¹) 3071, 3033, 1750, 1592, 1571, 1474, 1437, 1253, 1107, 1020, 919, 858, 789, 744, 688, 650, 604, 515, 476. ¹H NMR (CDCl₃) δ 7.99–7.19 (m, 20H, Ar), 6.90 (d, *J* = 3.3 Hz, 1H, H-1), 6.29 (t, *J* = 9.9 Hz, 1H, H-3), 5.84 (t, *J* = 9.8 Hz, 1H, H-4), 5.68 (dd, *J* = 3.3, 10.2 Hz, 1H, H-2), 4.70–4.56 (m, 3H, H-5,6). ¹³C NMR (CDCl₃) δ 165.0, 164.4, 164.2, 164.1, 163.6, 134.4, 134.1, 133.9, 133.8, 133.5, 133.3, 133.1, 132.8, 132.1, 131.8, 131.6, 131.5, 131.4, 131.2, 131.1, 131.0, 129.3, 128.7, 128.6, 128.0, 126.9,

126.8, 126.7 (C=O, Ar), 90.2 (C-1), 70.5, 70.4 (C-2,3,5), 68.8 (C-4), 62.7 (C-6). HRMS (ESI): calcd for $C_{41}H_{27}Cl_5O_{11}Na$ [M+Na]⁺: 892.9894; found: 892.9906.

1,2,3,4,6-Penta-O-(4-methoxybenzoyl)-D-glucopyranoside (S5).

4-Methoxybenzoyl chloride (5.13 g, 30 mmol) was added dropwise to a solution of D-glucose (**S1**, 0.903 g, 5.0 mmol) in dry pyridine (25 mL) at 0 °C while stirring. After being kept at 0 °C for 30 min, the mixture was allowed to warm to rt and stirred overnight. The reaction was quenched with ethanol (3 mL) and the mixture was diluted with CHCl₃ (30 mL). The mixture was washed with saturated NaHCO₃ aqueous solutions (3 × 30 mL). The aqueous layers were combined and back-extracted with CHCl₃ (30 mL). The organic layers were combined, dried over Na₂SO₄, filtered and concentrated under reduced pressure. The residue was recrystallized from ethanol (80 mL) and further purified by silica gel column chromatography using AcOEt–hexane (1:1–3:2, v/v) as an eluent to give **S5** as a colorless foam (3.47 g, 4.1 mmol, $\alpha:\beta = 83:17, 81\%$). IR (KBr, cm⁻¹) 3007, 2961, 2840, 1729, 1606, 1512, 1460, 1421, 1318, 1262, 1169, 1096, 922, 846, 767, 695, 614, 509. ¹H NMR (CDCl₃) δ 8.19–7.80 (m, 10H, Ar), 7.04–6.72 (m, 11H, Ar, H-1), 6.24 (t, *J* = 10.1 Hz, 1H, H-3), 5.78 (t, *J* = 10.1 Hz, 1H, H-4), 5.61 (dd, *J* = 3.6, 10.2 Hz, 1H, H-2), 4.62–4.51 (m, 2H, H-5.6), 4.41 (dd, *J* = 4.4, 12.2 Hz, 1H, H-6), 3.91, 3.86, 3.81, 3.78, 3.78 (s, 5 × 3H, Ar-OC<u>H₃</u>). ¹³C NMR (CDCl₃) δ 165.8, 165.5, 165.0, 164.8, 164.1, 164.0, 163.7, 163.5, 163.4, 132.2, 132.0, 131.9, 131.8, 122.0, 121.3, 121.1, 121.0, 114.0, 113.6 (C=O, Ar), 89.9 (C-1), 70.4, 70.2, 70.1 (C-2,3,5), 68.6 (C-4), 62.4 (C-6), 55.5, 55.4 (Ar-O<u>C</u>H₃). HRMS (ESI): calcd for C₄₆H₄₂O₁₆Na [M+Na]⁺: 873.2371; found: 873.2331.

1,2,3,4,6-Penta-O-benzoyl-D-galactopyranoside (S7).

Benzoyl chloride (3.5 mL, 30 mmol) was added dropwise to a solution of D-galactose (**S6**, 0.90 g, 5.0 mmol) in dry pyridine (25 mL) at 0 °C while stirring. After being kept at 0 °C for 30 min, the mixture was allowed to warm to rt and stirred overnight. Then, the reaction was quenched with methanol (3 mL), and CHCl₃ (18 mL) was added. The mixture was washed with saturated NaHCO₃ aqueous solutions (3 × 20 mL). The aqueous layers were combined and back-extracted with CHCl₃ (30 mL). The organic layers were combined, dried over Na₂SO₄, filtered and concentrated under reduced pressure. The residue was purified by silica gel column chromatography using AcOEt–hexane (1:3, v/v) as an eluent to give **S7** as a colorless foam (3.39 g, 4.8 mmol, α : β = 96:4, 97%). The ¹H NMR spectrum was identical to the reported data.²

1,2,3,4,6-Penta-O-benzoyl-D-mannopyranoside (S9).

Benzoyl chloride (6.3 mL, 54 mmol) was added dropwise to a solution of D-mannose (**S8**, 1.61 g, 8.9 mmol) in dry pyridine (30 mL) at 0 °C while stirring. After being kept at 0 °C for 30 min, the mixture was allowed to warm to rt and stirred for 6 h. Then, the reaction was quenched with methanol (6 mL), and CHCl₃ (30 mL) was added to the mixture. The mixture was washed with saturated NaHCO₃ aqueous solutions (3×30 mL). The aqueous layers were combined and back-extracted with CHCl₃ (2×60 mL). The organic layers were combined, dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The residue was recrystallized from ethanol (260 mL) to give **S9** as a colorless solid (5.17 g, 2.3 mmol, $\alpha:\beta = 98:2, 83\%$). The ¹H NMR spectrum was identical to the reported data.²

1,2,3,4-Tetra-O-benzoyl-L-fucopyranoside (S11).

Benzoyl chloride (3.0 mL) was added to a solution of L-fucose (**S10**, 49 mg, 3.0 mmol) in dry pyridine (10 mL) at rt while stirring. After 5 h, the mixture was diluted with CH_2Cl_2 (50 mL). The organic layer was washed with saturated NaHCO₃ aqueous solutions (5 × 50 mL), dried over Na₂SO₄, filtered and concentrated under reduced pressure. The residue was purified by silica gel column chromatography using AcOEt–hexane (1:7, v/v) as an eluent to give **S11** as a colorless foam (1.71 g, 2.9 mmol, α : β = 86:14, 98%). The ¹H NMR spectrum was identical to the reported data.³

2. Synthesis of authentic sample of dimethyl 2,3,4,6-tetra-O-benzoyl-a-D-mannopyranosyl boranophosphate .

Authentic sample of **12** was prepared from 1,2,3,4,6-penta-*O*-benzoyl-D-mannopyranoside (**S9**, α : β = 98:2) in 4 steps (Scheme S2).

Scheme S2. Synthesis of dimethyl 2,3,4,6-tetra-O-benzoyl-α-D-mannopyranosyl boranophosphate 12. (*i*-Pr₂N)₂PCl BzO OBz BzC BzC OBz MeNH₂/MeOH i-Pr₂NEt BzO 0 BzO BzO B₇C Bz∩ BzC NPr₂ MeCN, rt, 2 h CH₂Cl₂, rt, 24 h ÓН NPr₂ **S**9 S12 S13 $(\alpha:\beta = 98:2)$ 58% (α : β = 92:8) 45% (α:β = 98:2) BzO OBz BzO OBz MeOH 0 B₇O 0 BzO BZO 1H-tetrazole BH3.THF/THF BzO OMe BH₃ MeCN, rt, 6.5 h MeCN, rt, 2 h `OMe MeO ÒMe S14

2,3,4,6-Tetra-O-benzoyl-α-D-mannopyranoside (S12).

This material was prepared according to the procedure to synthesize 2,3,4,6-tetra-*O*-benzoyl-D-glucopyranoside reported by Fukase *et al.*⁴ A 40% methylamine solution in methanol (0.3 mL) was added to a solution of 1,2,3,4,6-penta-*O*-benzoyl- α -D-mannopyranoside (**S9**, 0.383 g, 0.55 mmol) in dry CH₃CN (3 mL) at rt while stirring. After 2 h, the mixture was concentrated under reduced pressure. CHCl₃ (10 mL) was added to the residue and the organic layer was washed with brine (10 mL). The aqueous layers were combined and back-extracted with CHCl₃ (2 × 10 mL). The organic layers were combined, dried over Na₂SO₄, filtered and concentrated under reduced pressure. The residue was purified by silica gel column chromatography using AcOEt–hexane (1:2, v/v) as an eluent to give **S12** as a yellow foam (0.190 g, 0.32 mmol, α : β = 92:8, 58%). The ¹H NMR spectrum was identical to the reported data.⁵

12 61% from **S13** (α:β = 98:2)

2,3,4,6-Tetra-O-benzoyl-a-D-mannopyranosyl-N,N,N',N'-tetraisopropylphosphorodiamidite (S13).

This material was prepared according to the procedure to synthesize 2,3,4,6-tetra-*O*-acetyl- α -D-mannopyranosyl)-*N,N,N',N'*-tetraisopropylphosphorodiamidite reported by Boons *et al.*⁶ Chlorobis(*N,N*-diisopropylamino)phosphine (32 mg, 0.12 mmol) was added to a solution of 2,3,4,6-tetra-*O*-benzoyl- α -D-mannopyranoside (**S12**, 77 mg, 0.13 mmol) in dry CH₂Cl₂ (0.75 mL). The mixture was allowed to stir at rt for 10 min. Diisopropylethylamine (0.23 mL) was added dropwise to the mixture. After 24 h, the solvent was removed under reduced pressure and the residue was purified by silica gel column chromatography on NH silica gel (100–200 mesh) using AcOEt–hexane–Et₃N (12:83:15, v/v/v) as an eluent to give **S13** as a colorless oil (45 mg, 55 µmol, α : β = 98:2, 45%). IR (KBr, cm⁻¹) 3065, 2971, 1730, 1603, 1492, 1452, 1364, 1268, 1181, 1108, 1069, 1027, 958, 870, 816, 710, 646, 528. ¹H NMR (CDCl₃) δ 8.16–7.23 (m, 20H, Ar), 6.13 (t, *J* = 9.9 Hz, 1H, H-4), 6.02 (dd, *J* = 3.2, 10.1 Hz, 1H, H-3), 5.66–5.62 (m, 1H, H-2), 5.39 (dd, *J* = 2.0, 11.0 Hz, 1H, H-1), 4.68–4.56 (m, 2H, H-5.6), 4.49 (dd, *J* = 4.4, 11.6 Hz, 1H, H-6), 3.79–3.54 (m, 4H, -N-C<u>H</u>-(CH₃)₂), 1.35–1.21 (m, 24H, -N-CH-(C<u>H</u>₃)₂). ¹³C NMR (CDCl₃) δ 166.3, 165.6, 165.5, 165.4 (C=O), 133.3, 133.1, 132.9, 130.0, 129.9, 129.8, 129.7, 129.5, 129.1, 129.0, 128.5, 128.4, 128.3 (Ar), 93.0 (C-1), 71.5 (C-2), 70.3 (C-3), 69.2 (C-5), 67.1 (C-4), 63.1 (C-6), 45.5, 45.3, 45.0, 44.8 (-N-<u>C</u>H-(CH₃)₂), 24.5, 24.4, 24.3, 24.2 (-N-CH-(<u>C</u>H₃)₂). ³¹P NMR (CDCl₃) δ 116.2. HRMS (ESI): calcd for C₄₆H₅₅N₂O₁₀PK [M+K]⁺: 865.3231; found: 865.3257.

Authentic sample of dimethyl 2,3,4,6-tetra-O-benzoyl-α-D-mannopyranosyl boranophosphate (12).

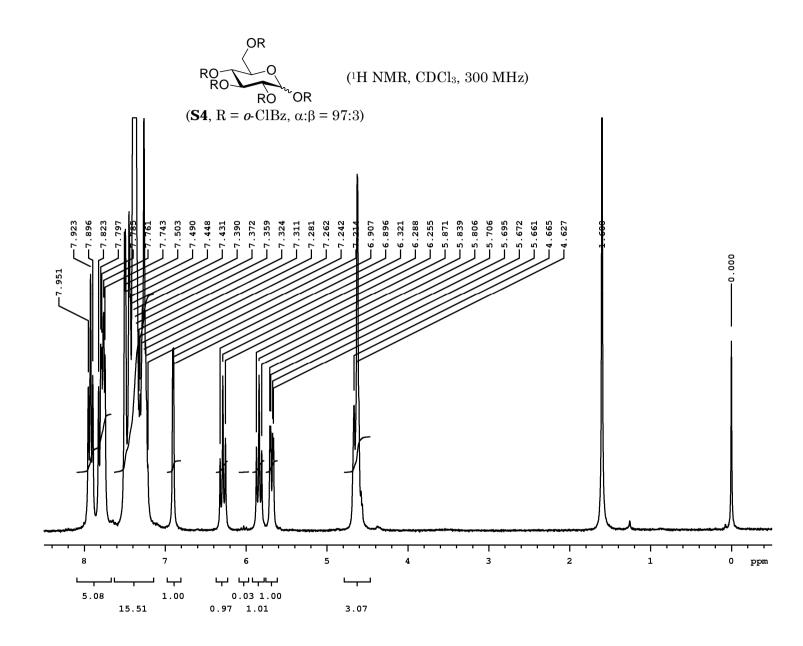
2,3,4,6-Tetra-*O*-benzoyl- α -D-mannopyranosyl-*N*,*N*,*N*,'*N*'-tetraisopropylphosphorodiamidite (**S13**, 45 mg, 55 µmol) was dried by repeated coevaporation with dry toluene and dry CH₃CN. The residue was treated with dry MeOH (9 µL, 0.22 mmol) and a solution of 1*H*-tetrazole (0.16 mg, 22 mmol) in dry CH₃CN (2.2 mL), which was dried over MS 3A overnight. The mixture was allowed to stir for 30 min, and dry MeOH (30 µL, 0.74 mmol) was added to the mixture. After being stirred for 6 h, the mixture was diluted with CHCl₃ (10 mL), and the mixture was washed with saturated NaHCO₃ aqueous solutions (3 × 10 mL). The aqueous layers were combined and back-extracted with CHCl₃ (20 mL). The organic layers were combined, and dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The

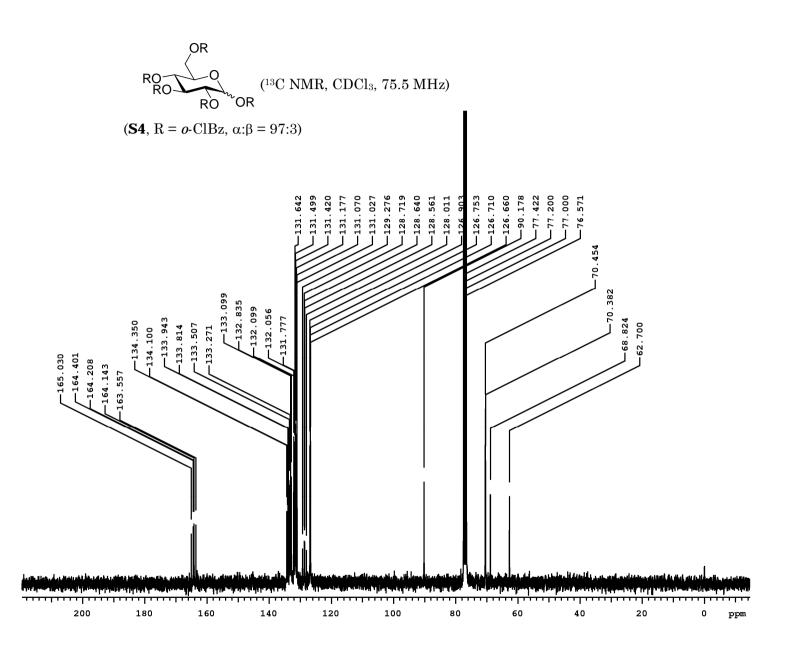
residue was dissolved in dry CH₃CN, then 0.99 M BH₃·THF/THF (0.25 mL, 0.25 mmol) was added dropwise. The mixture was allowed to stir for 2 h, washed with a saturated NaHCO₃ aqueous solution (7 mL), and extracted with toluene (6 mL). The organic layer was separated and washed with saturated NaHCO₃ aqueous solutions (2 × 7 mL). The aqueous layers were combined and back-extracted with toluene (30 mL). The organic layers were combined, dried over Na₂SO₄, filtered and concentrated under reduced pressure. The residue was purified by silica gel column chromatography using AcOEt–hexane (1:3, v/v) as an eluent to give **12** as a colorless foam (24 mg, 34 µmol, α : β = 98:2, 61% from **S13**).

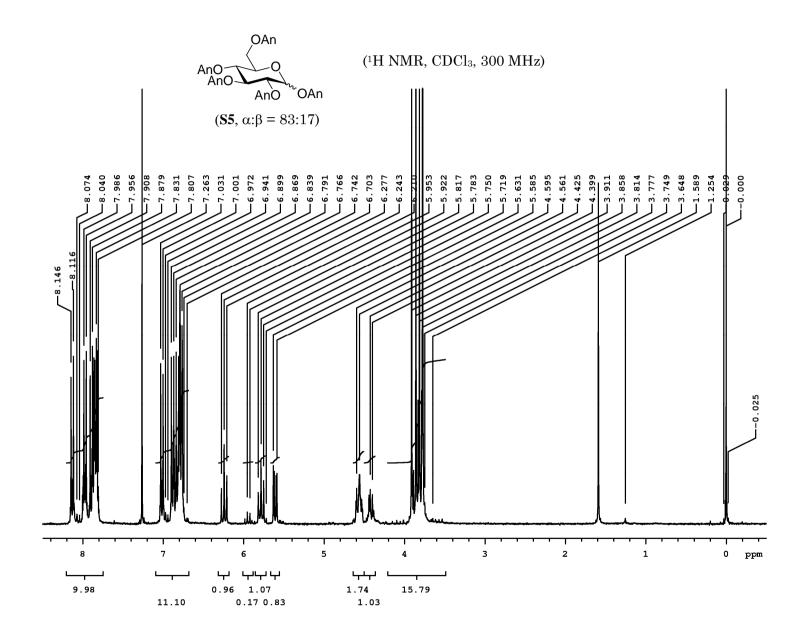
References for Supporting Information

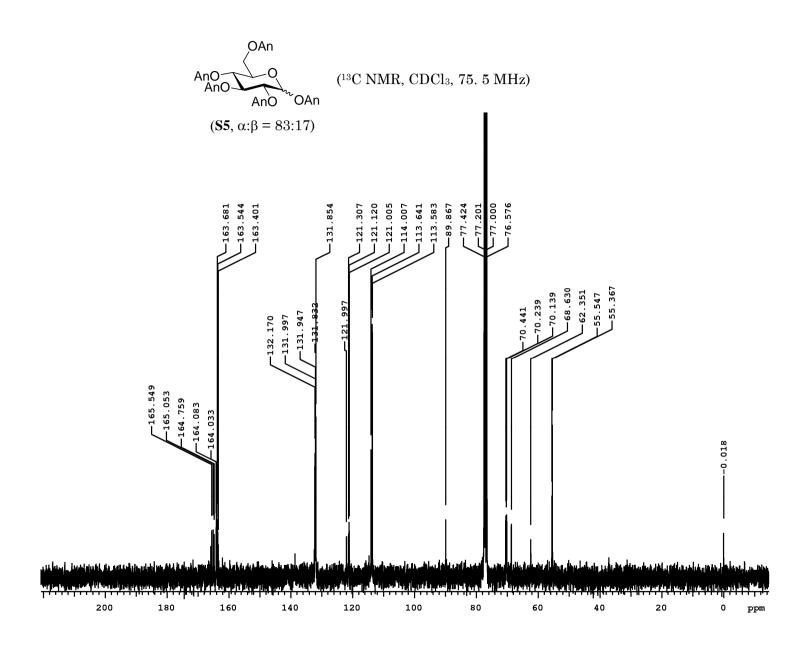
1) Haines, A. H.; Hughes, D. L. Carbohydr. Res. 2007, 342, 2264-2269.

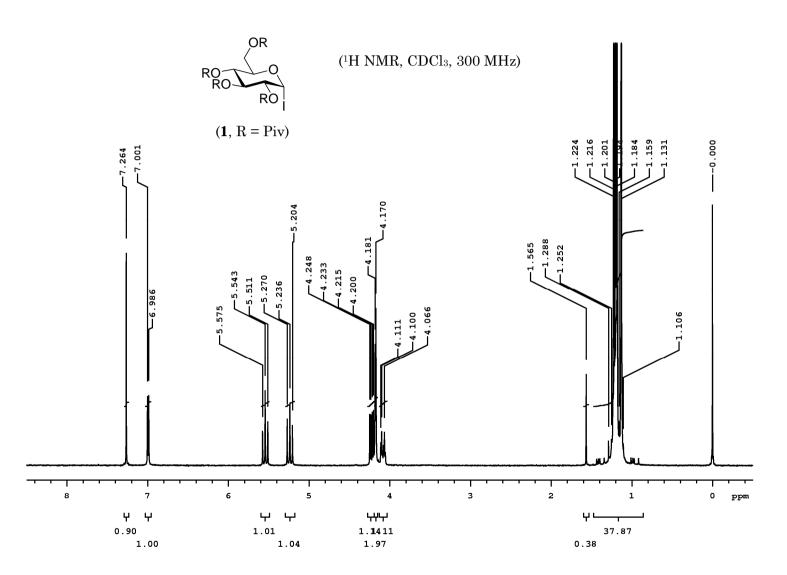
- 2) D'Accorso, N. B.; Thiel, I. M. E.; Schüller, M. Carbohydr. Res. 1983, 124, 177-184.
- 3) Timmons, S. C.; Mosher, R. H.; Knowles, S. A.; Jakeman, D. L. Org. Lett. 2007, 9, 857-860.
- 4) Egusa, K.; Kusumoto, S.; Fukase, K. Eur. J. Org. Chem. 2003, 3435–3445.
- 5) Mbadugha, B. N. A.; Menger, F. M. Org. Lett. 2003, 5, 4041-4044.
- 6) Majumdar, D.; Elsayed, G. A.; Buskas, T.; Boons, G.-J. J. Org. Chem. 2005, 70, 1691-1697.

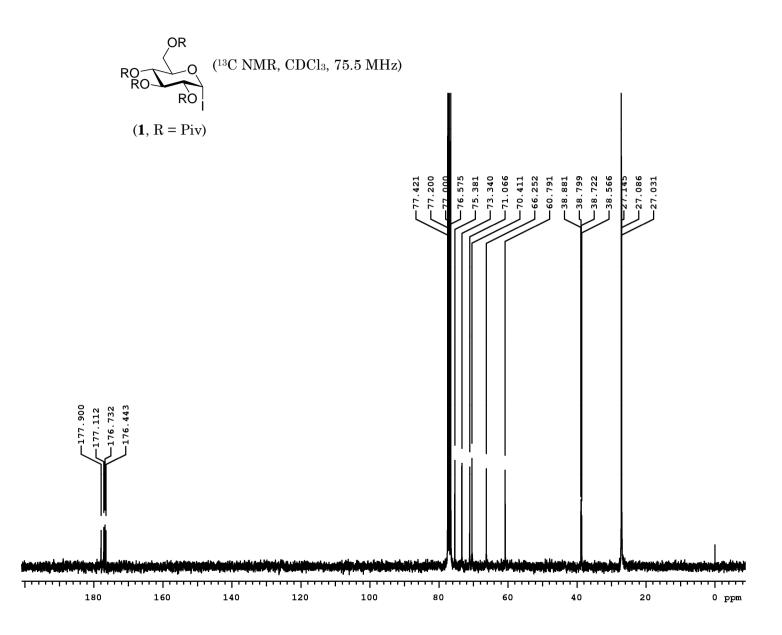


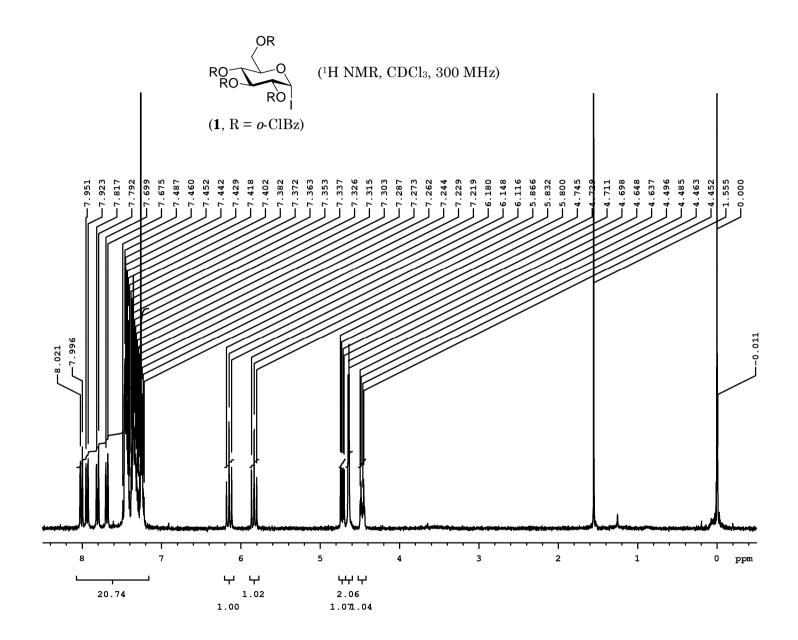


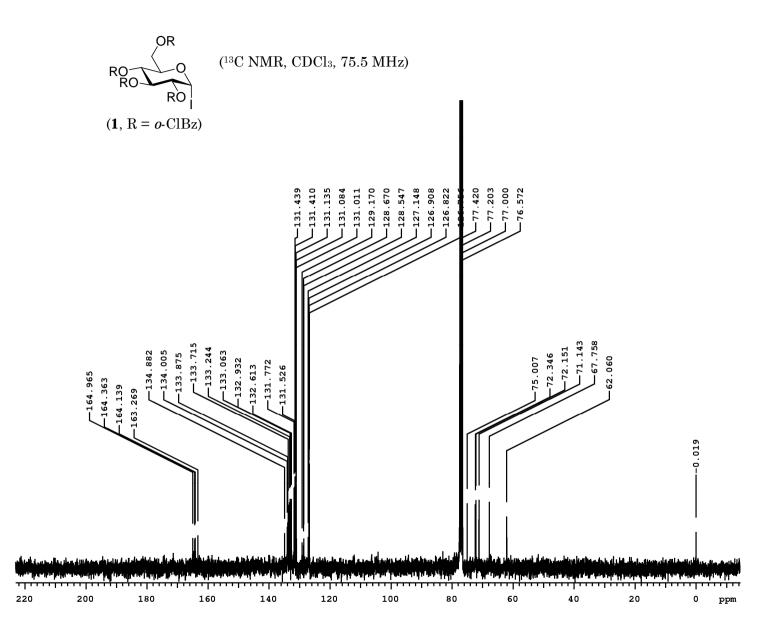


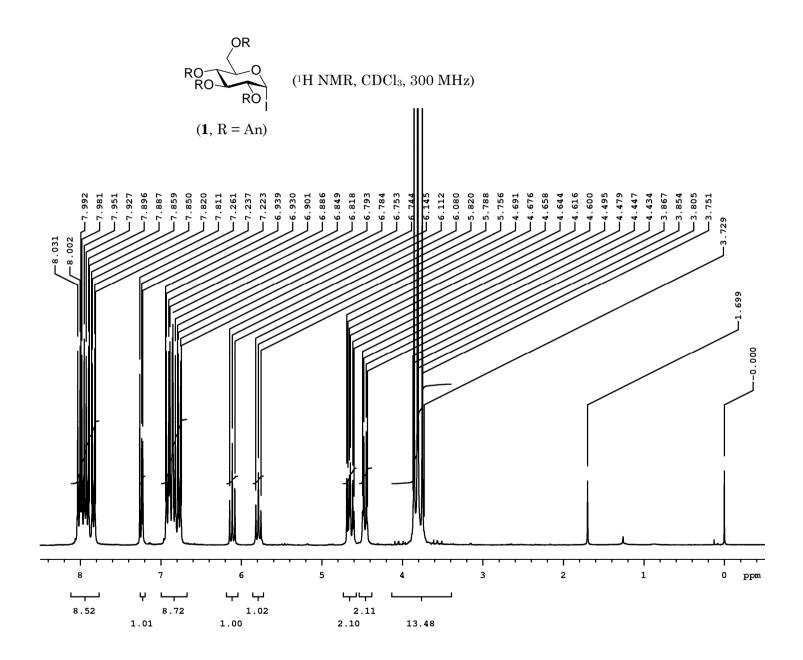


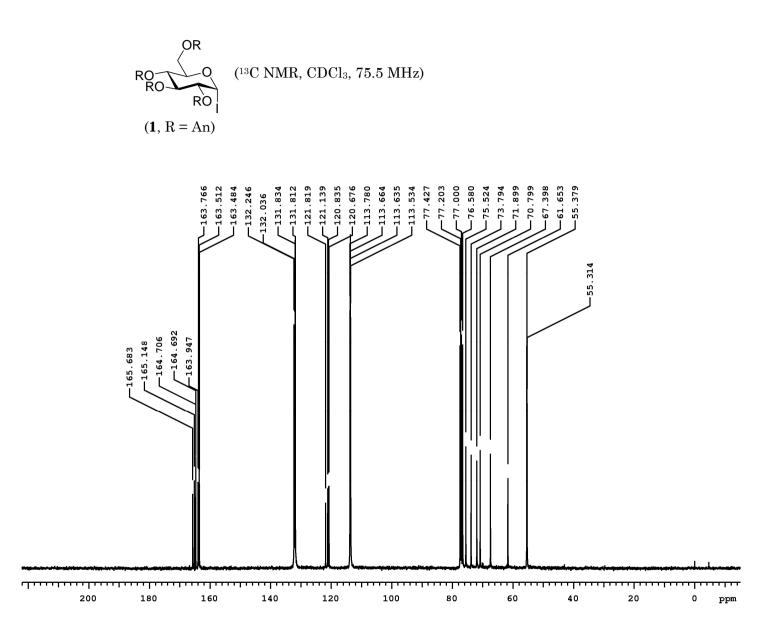


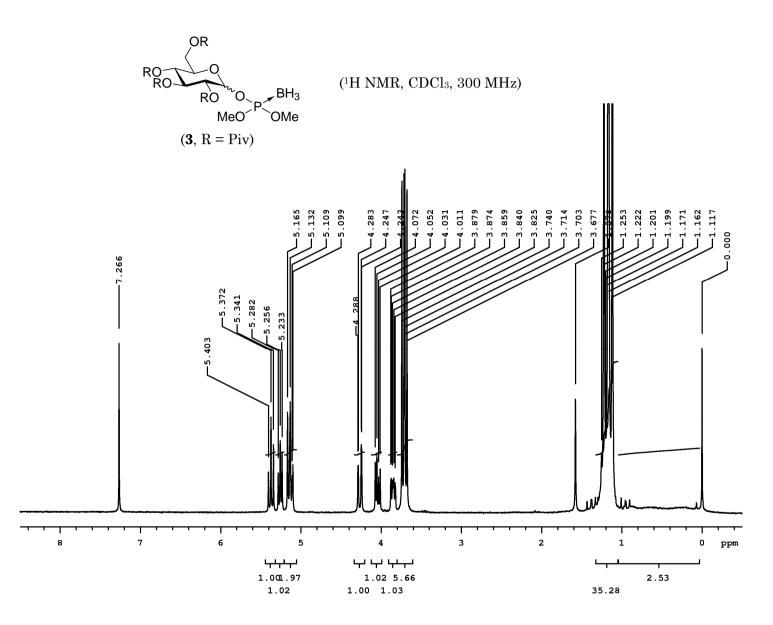


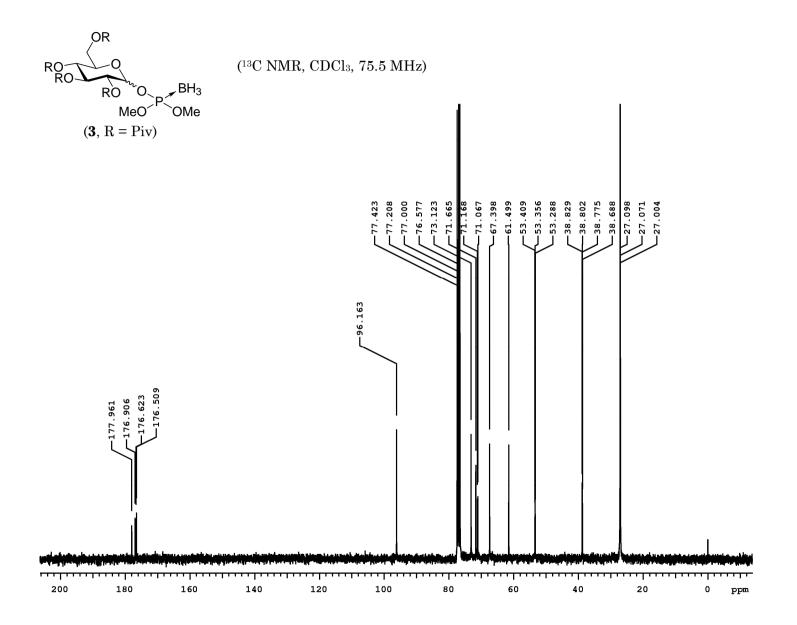


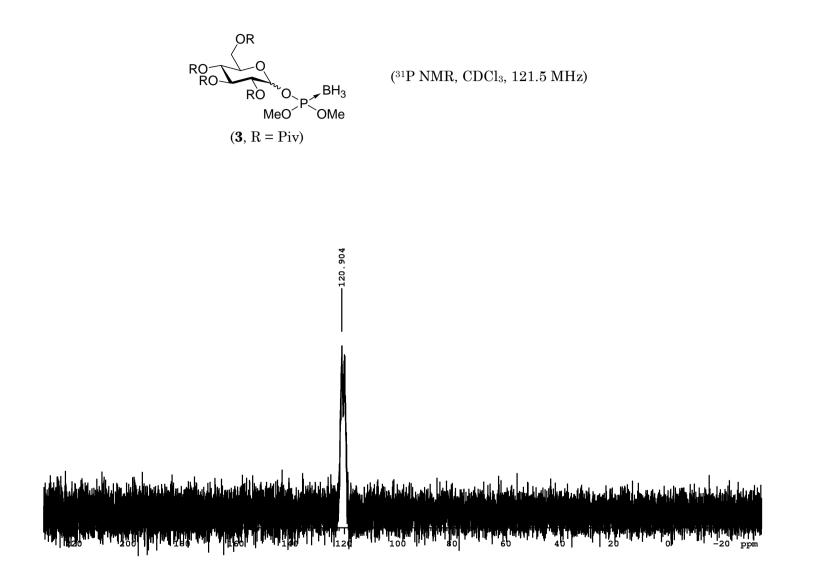


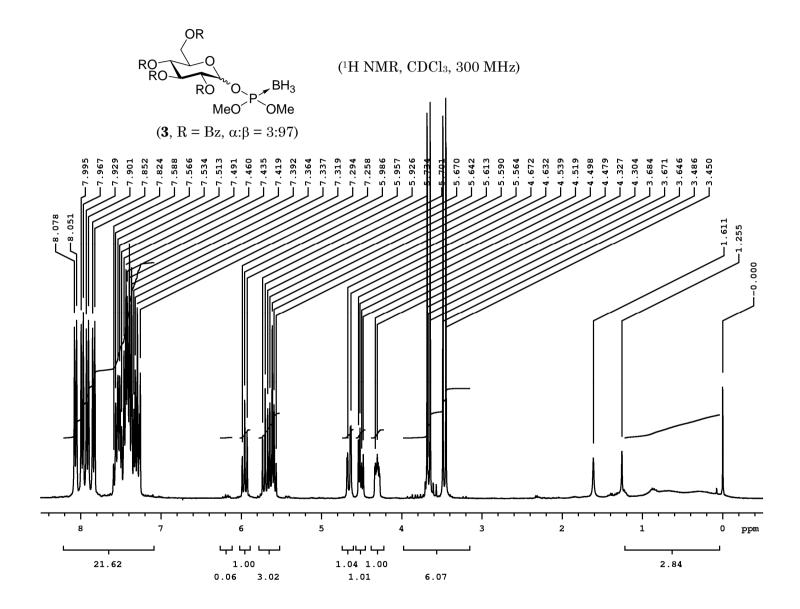


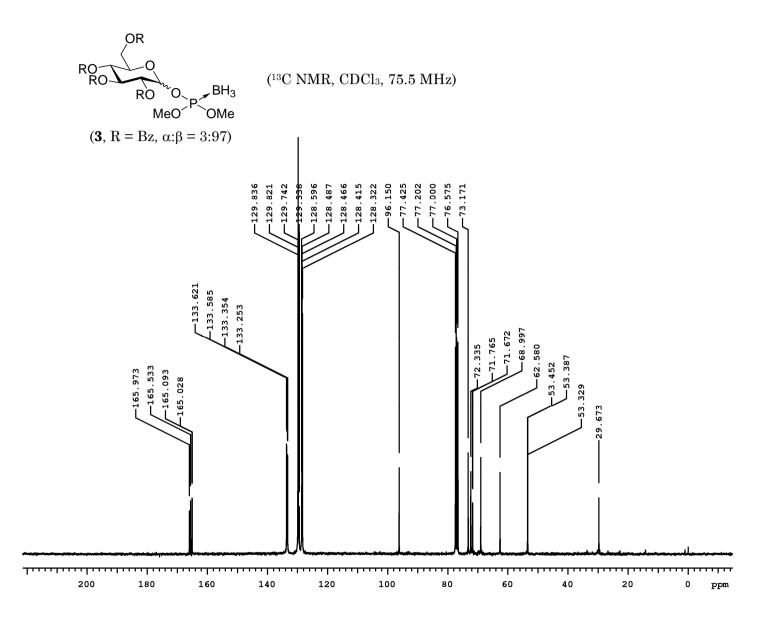


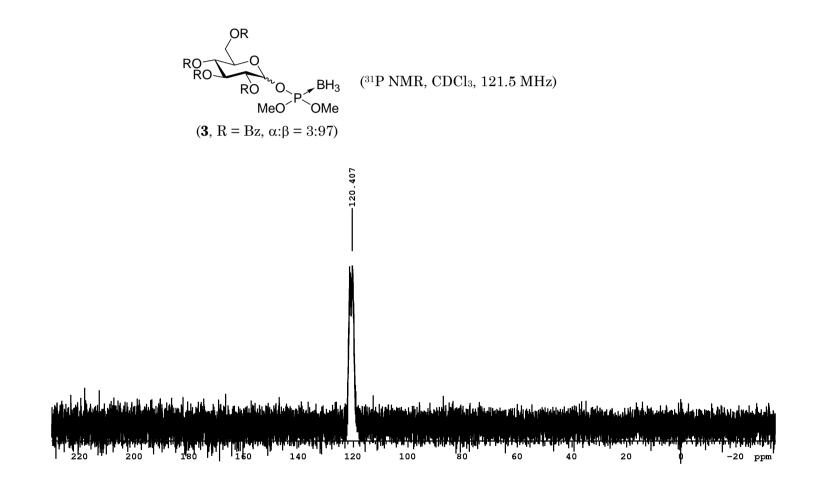


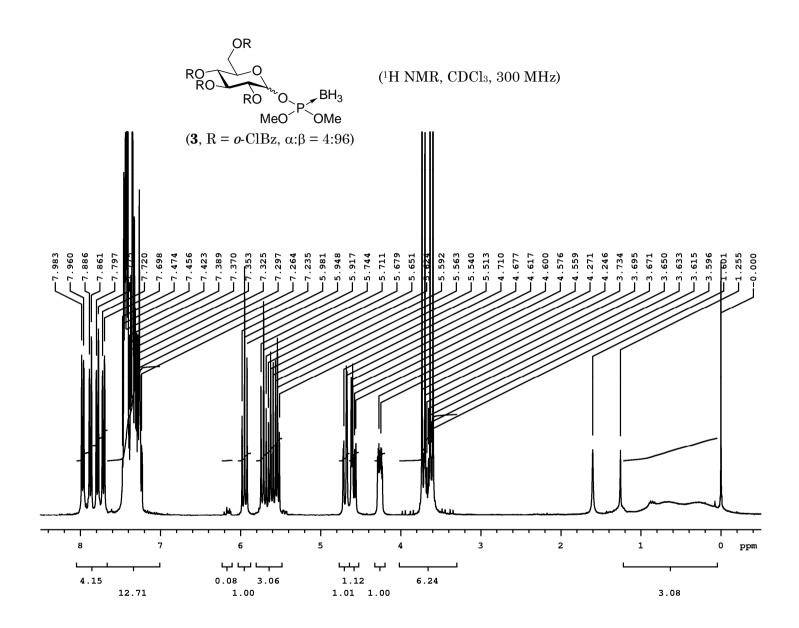


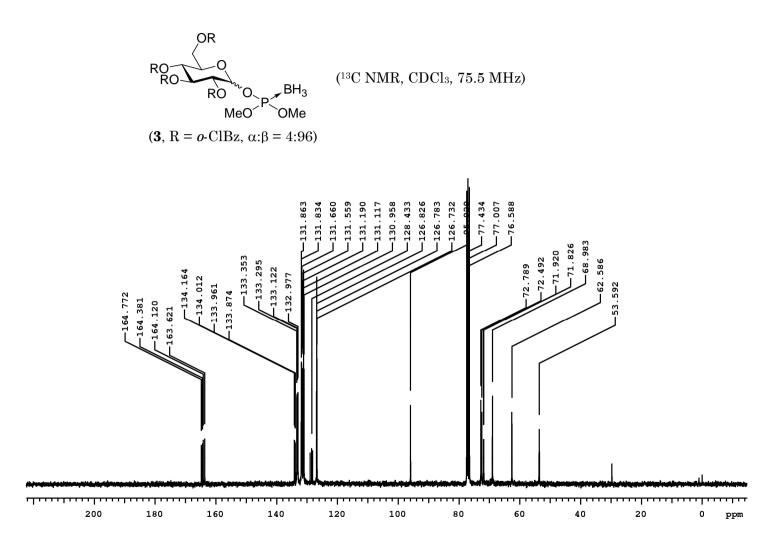


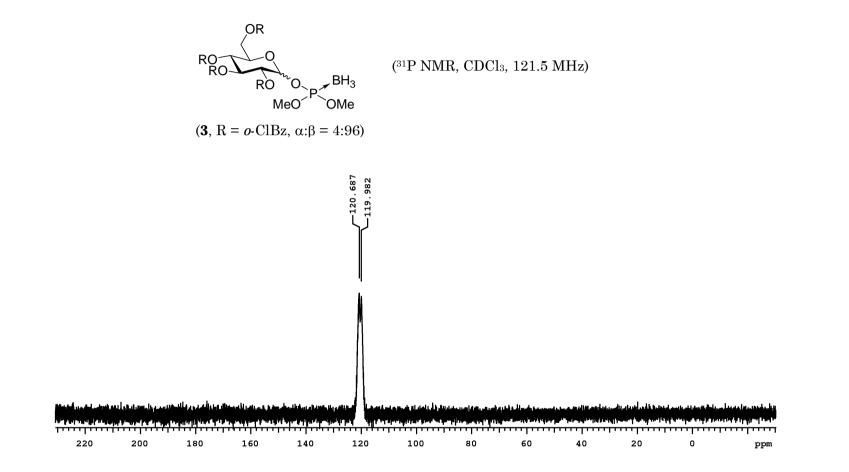


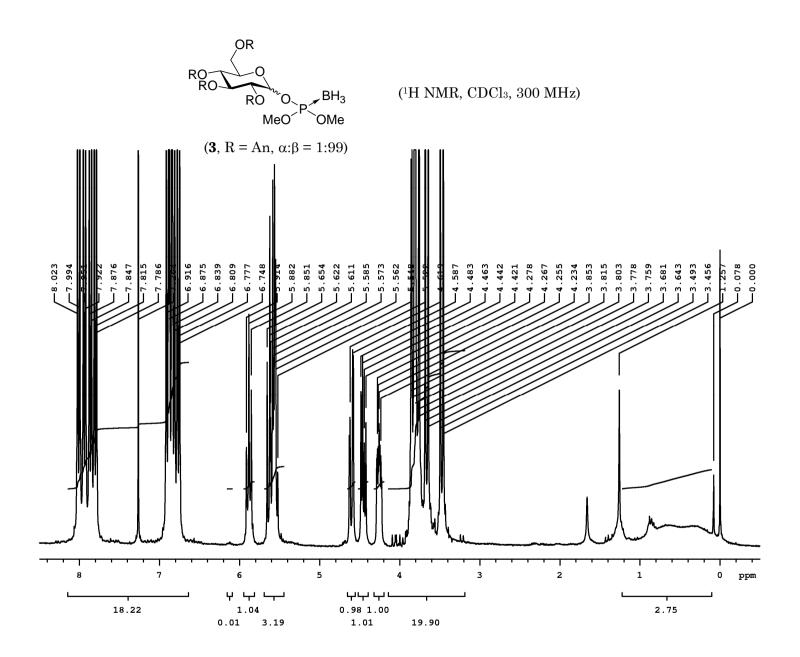


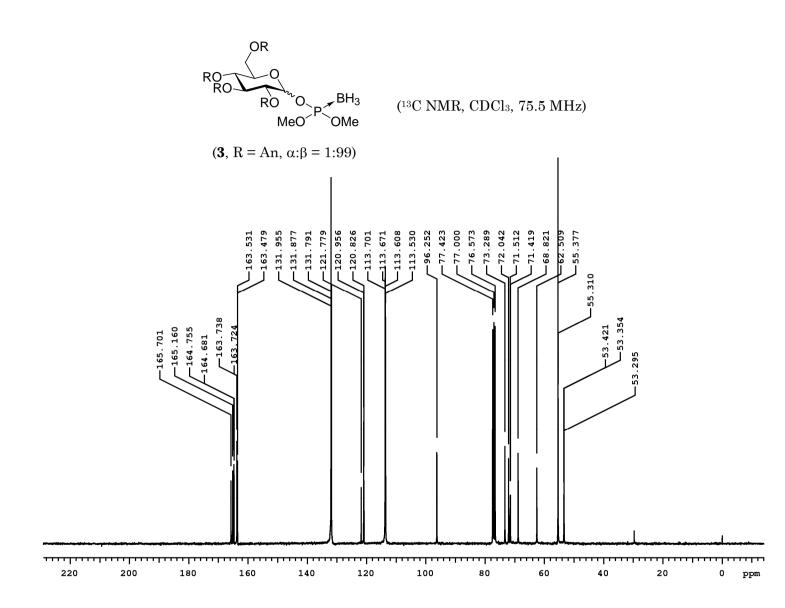


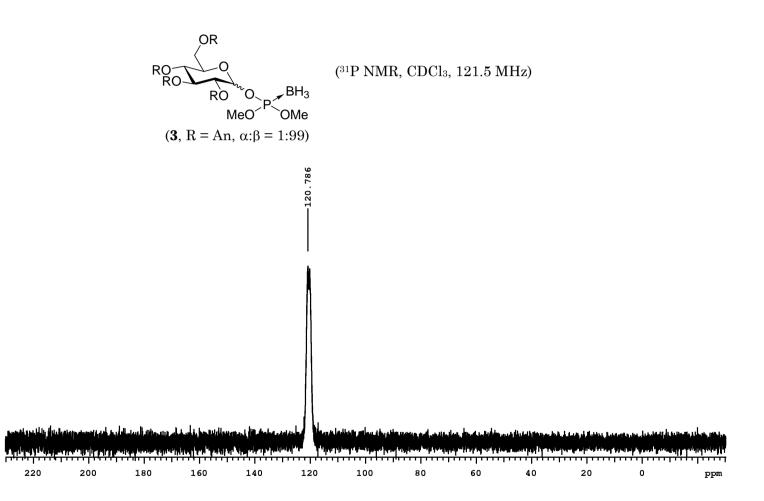


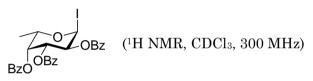




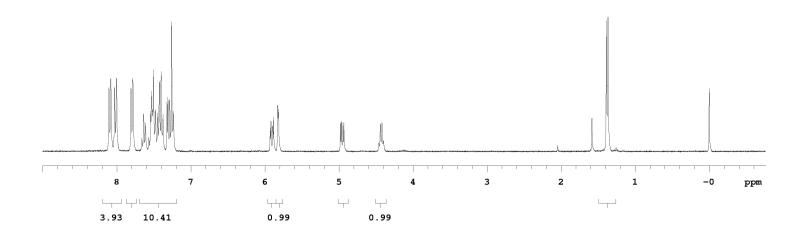


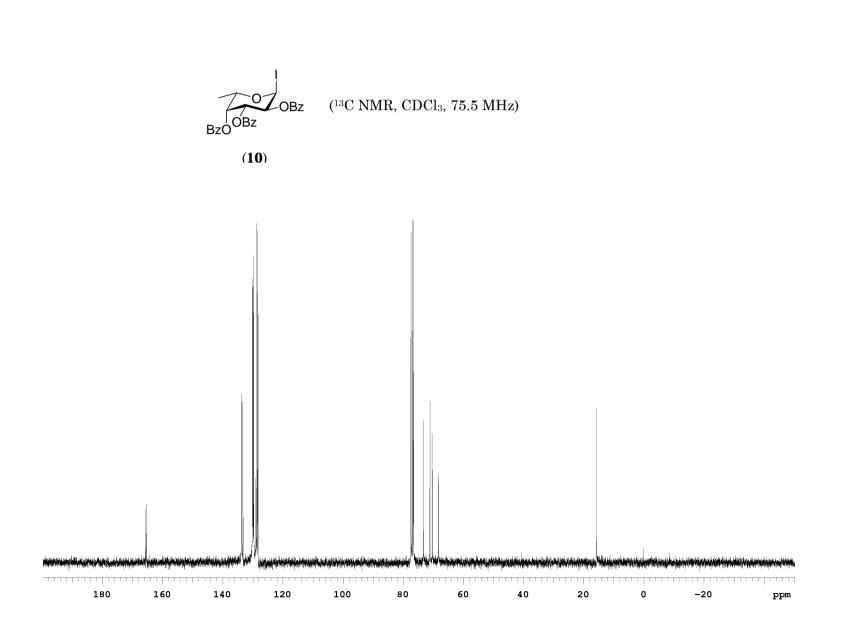


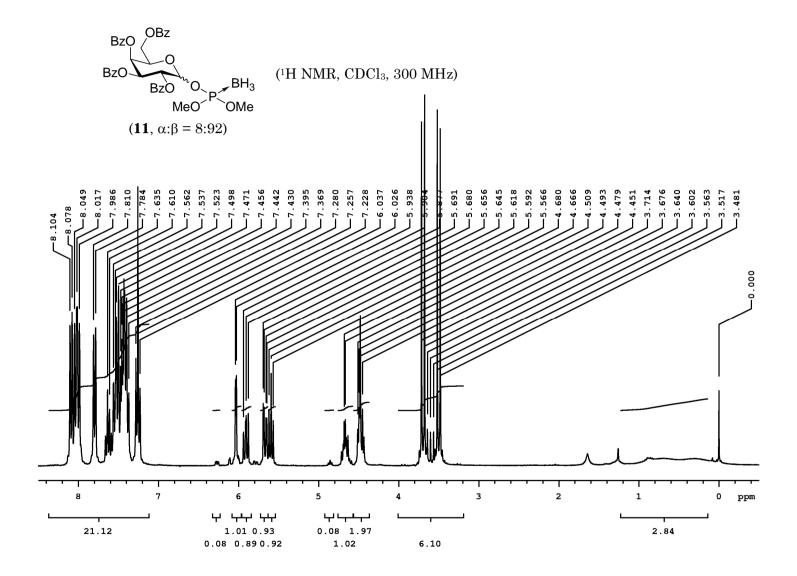


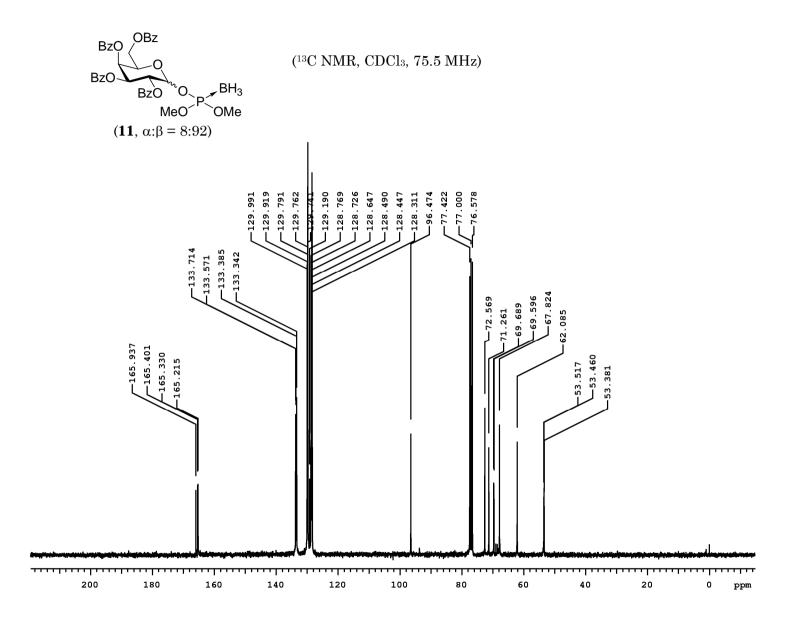


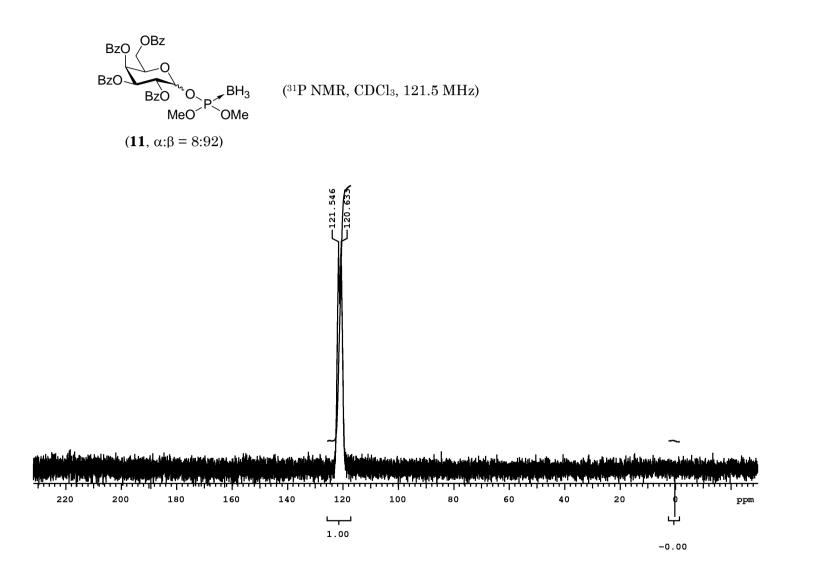
(**10**)

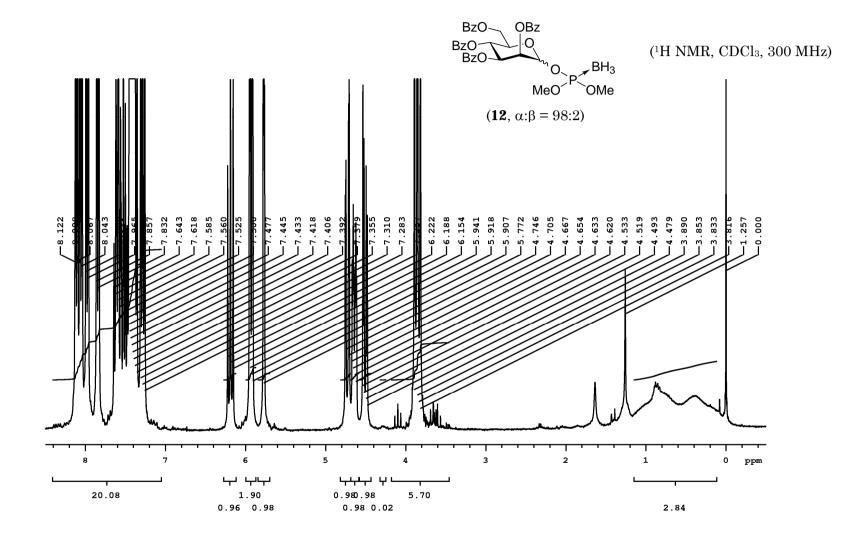


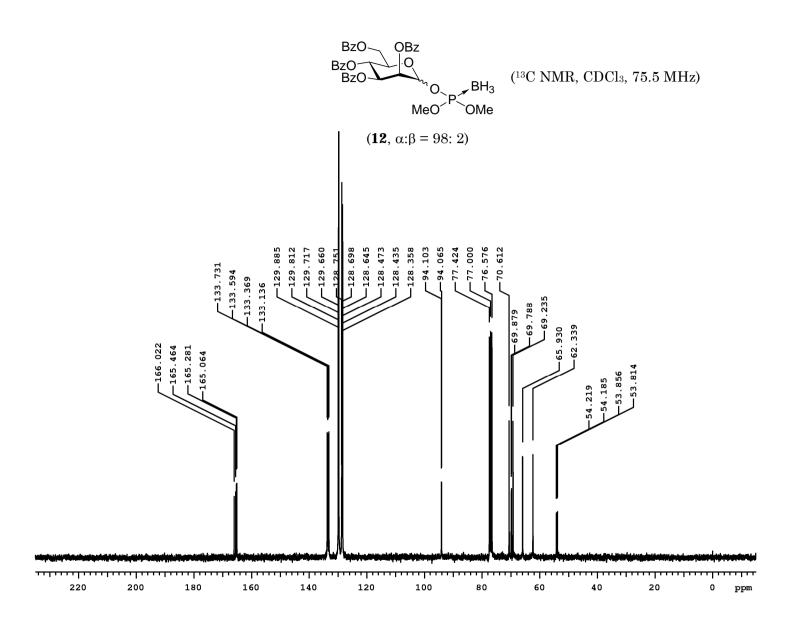




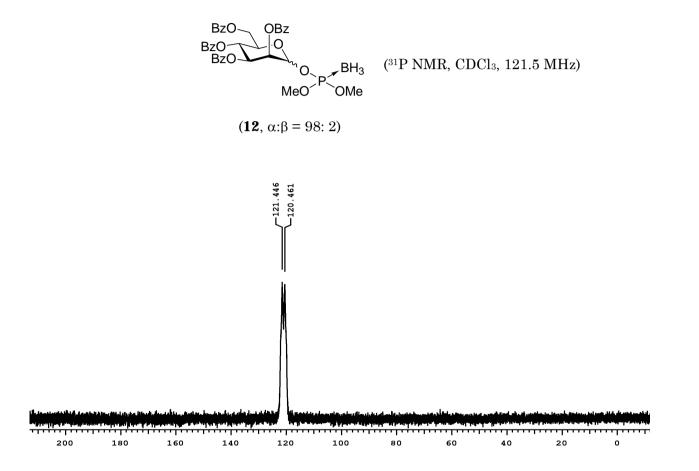


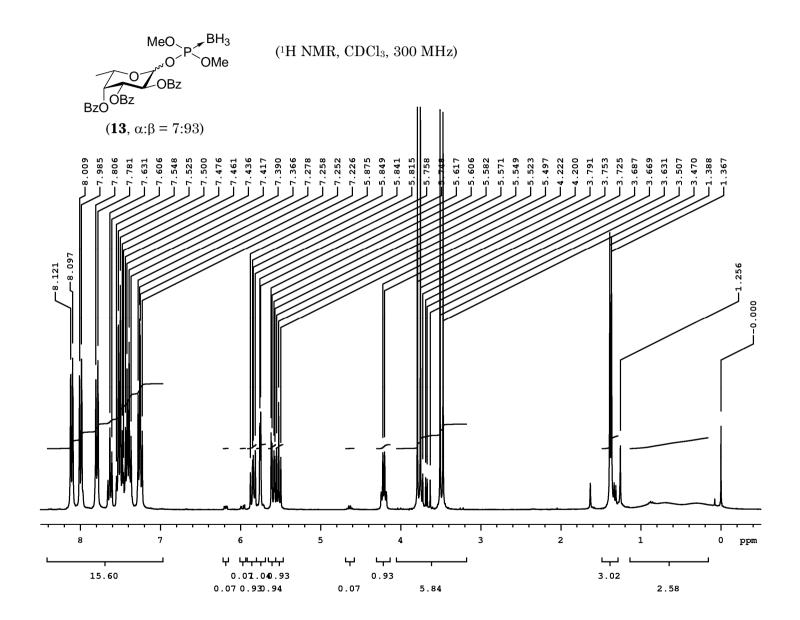


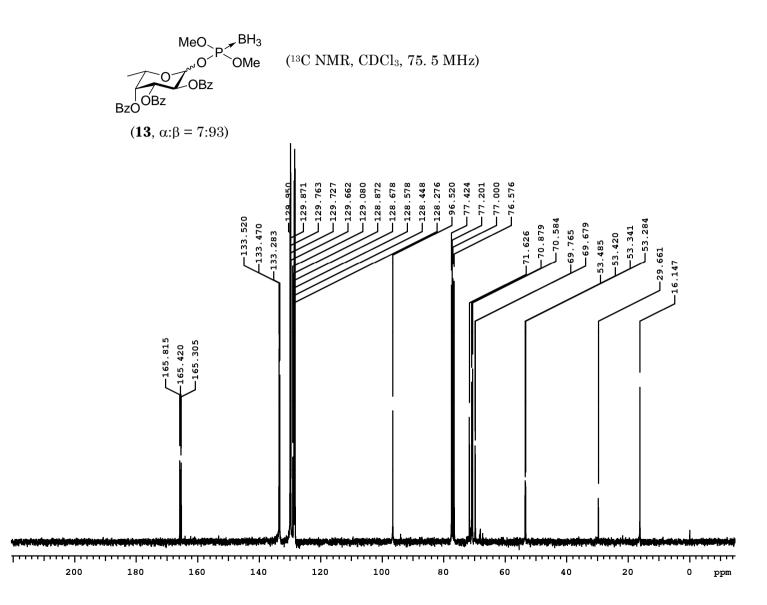


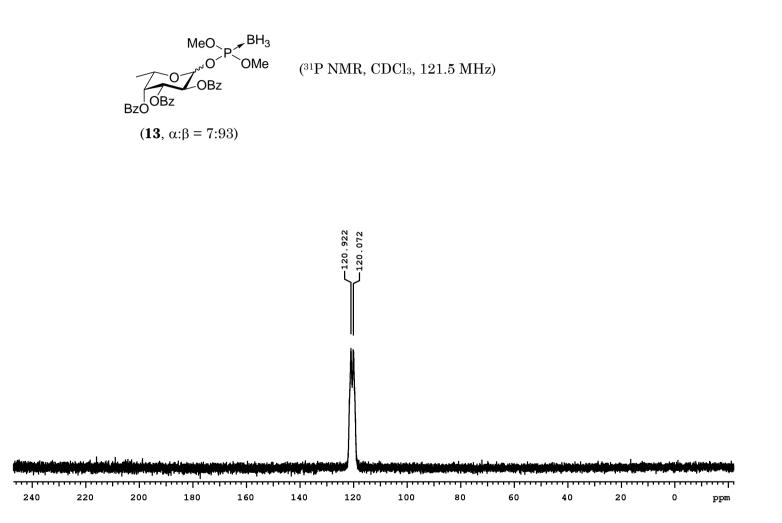


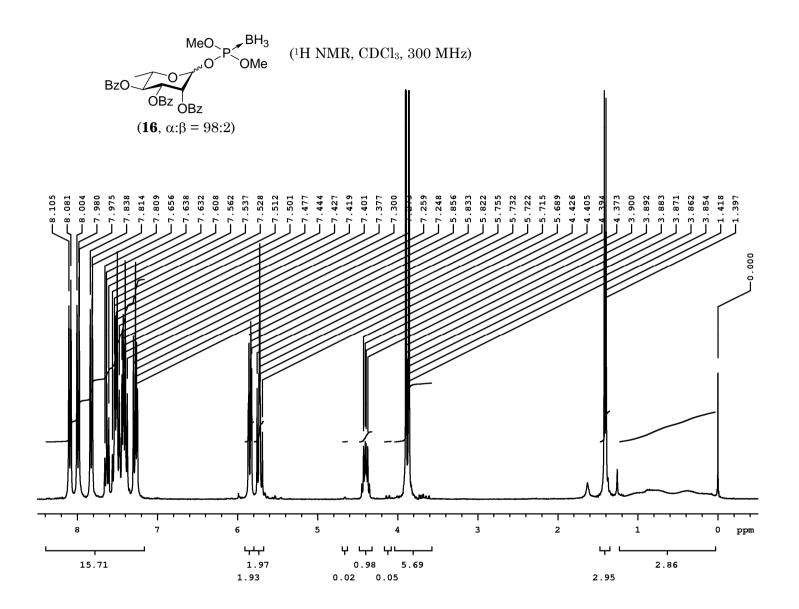
S34

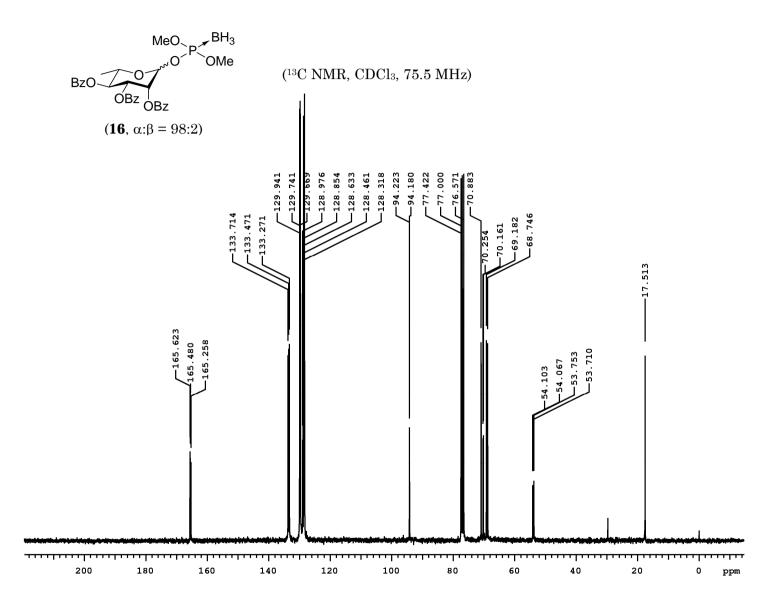


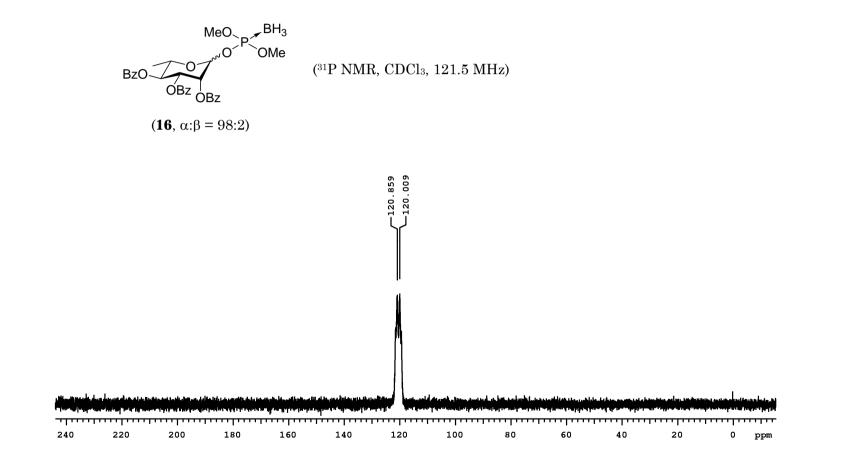


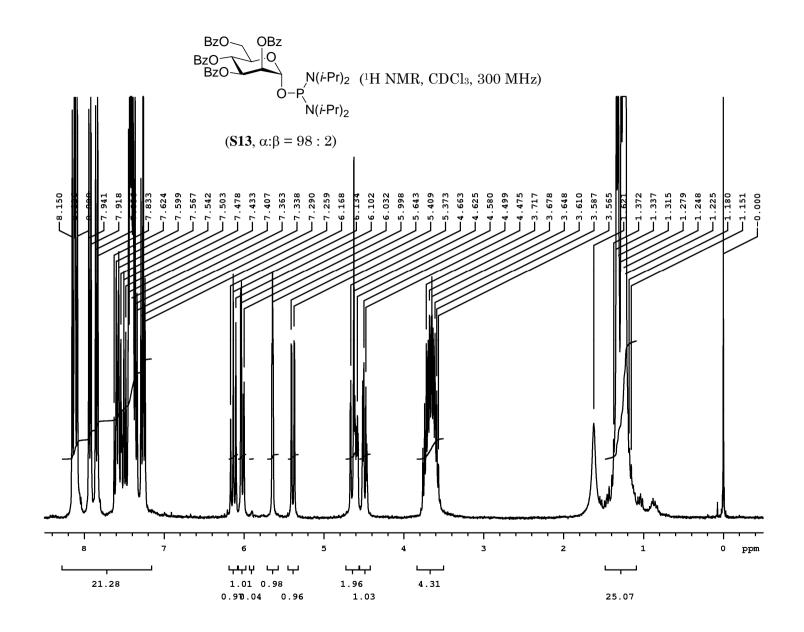


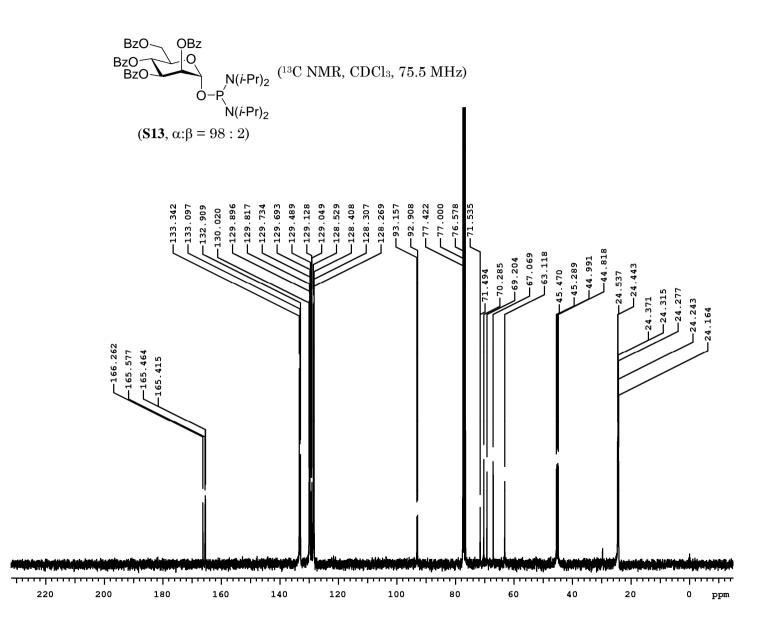


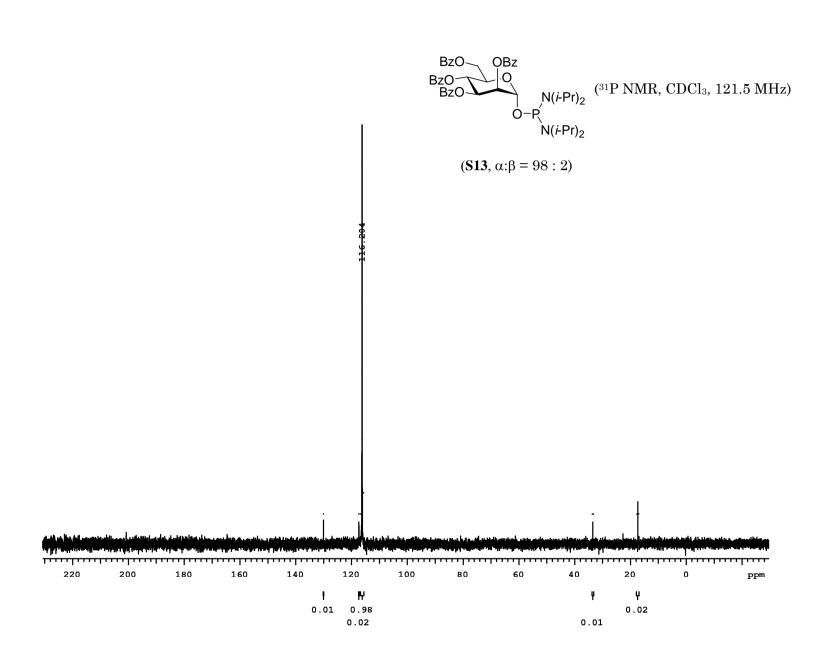


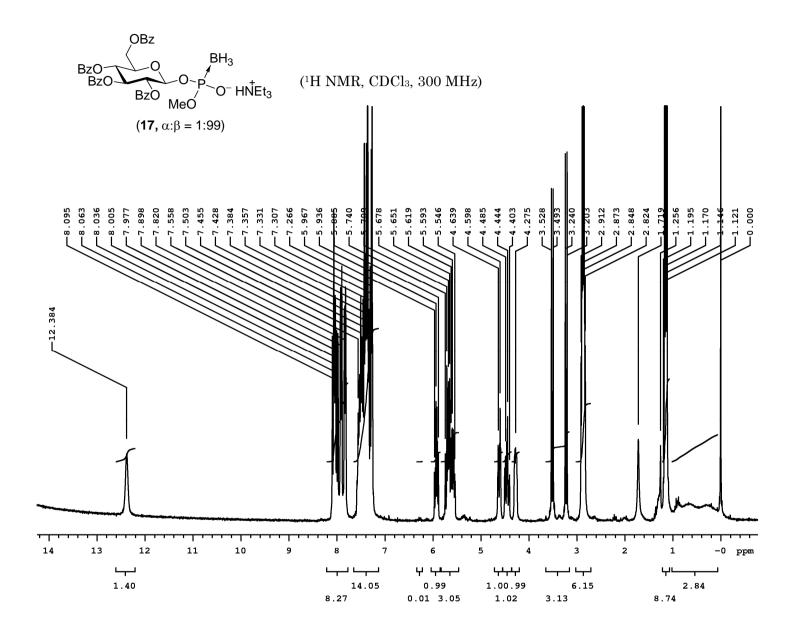


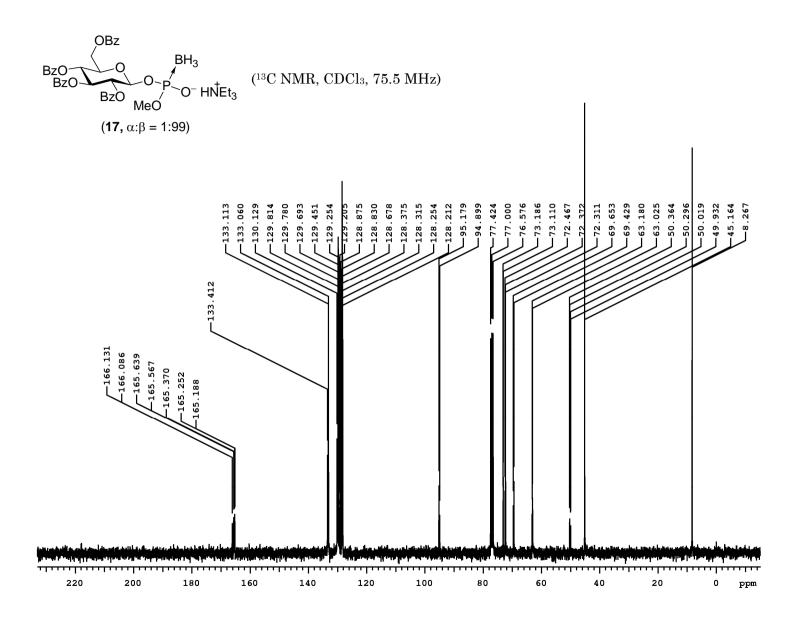


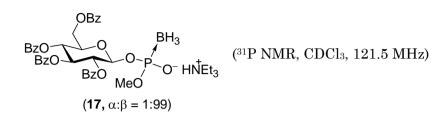


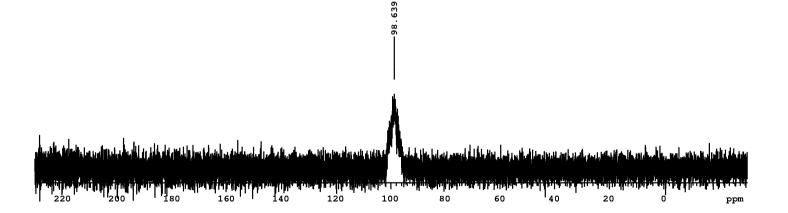


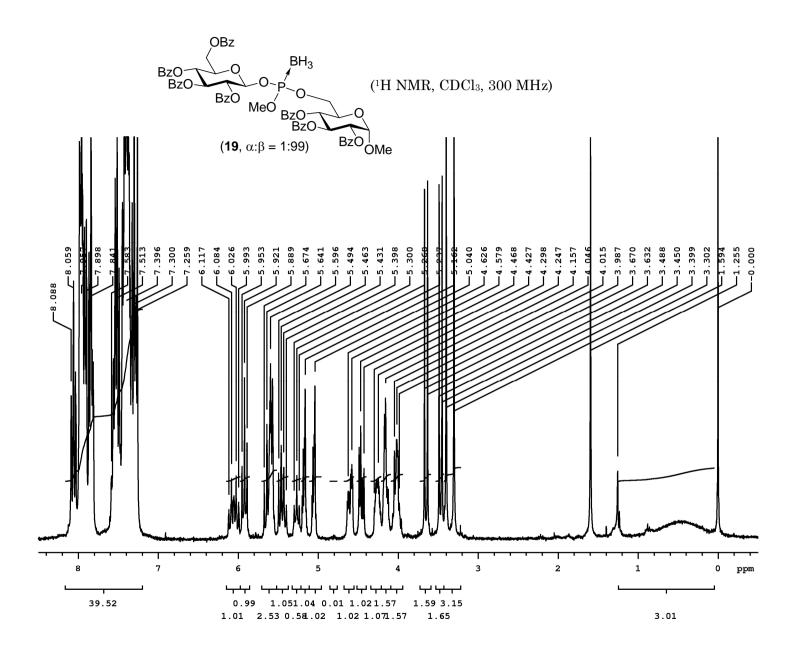


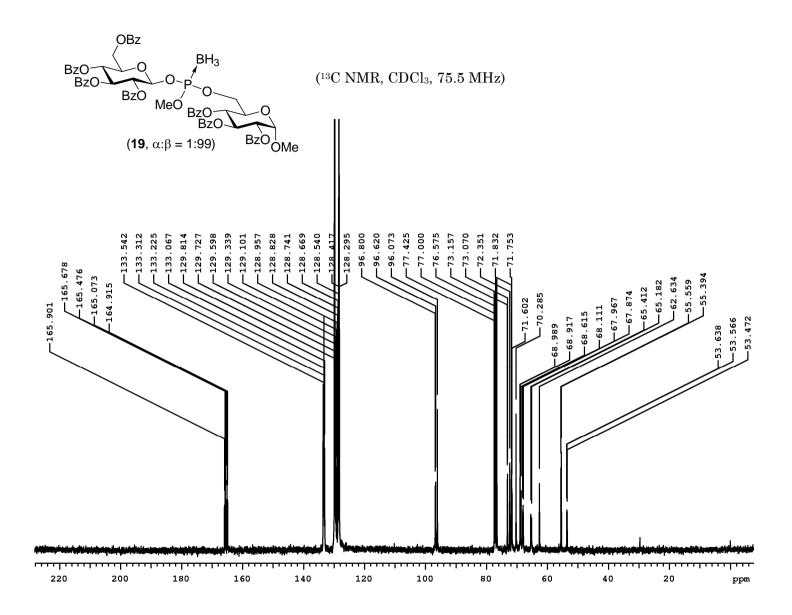


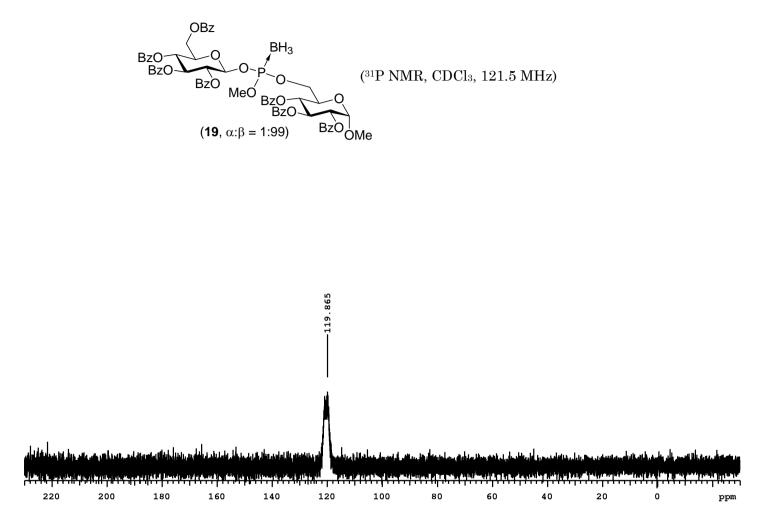


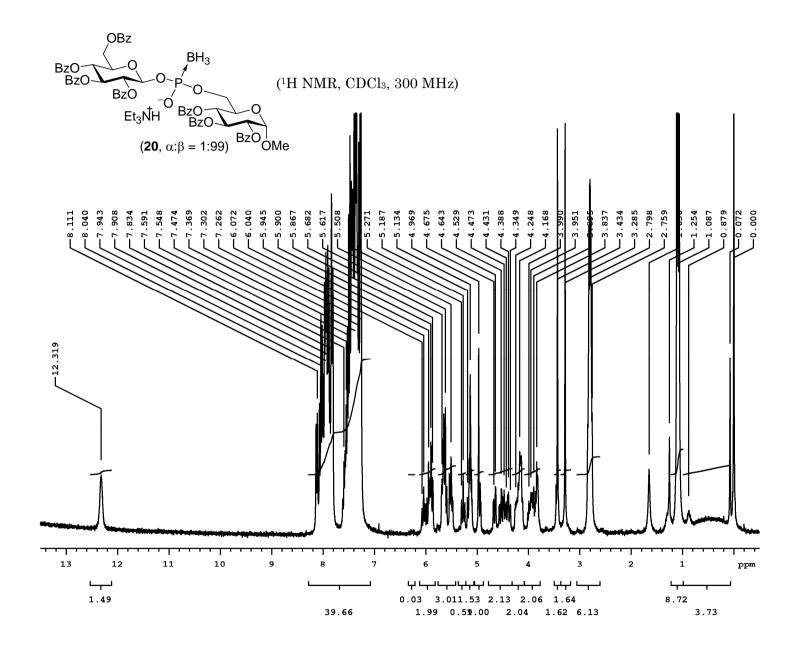


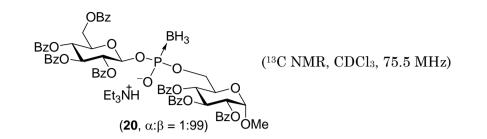


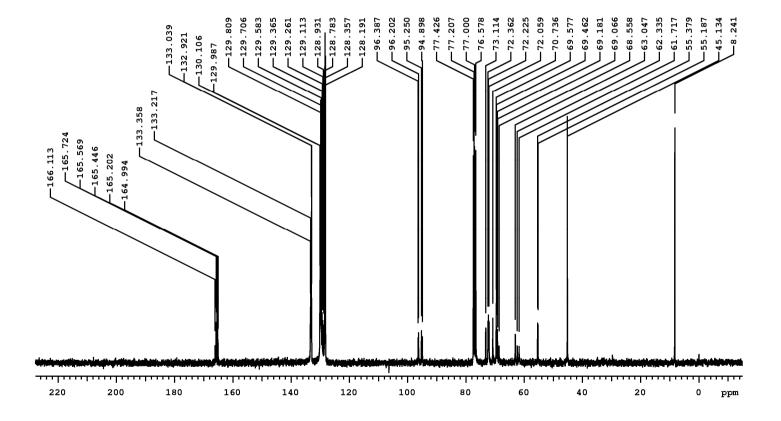


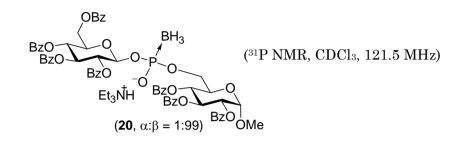


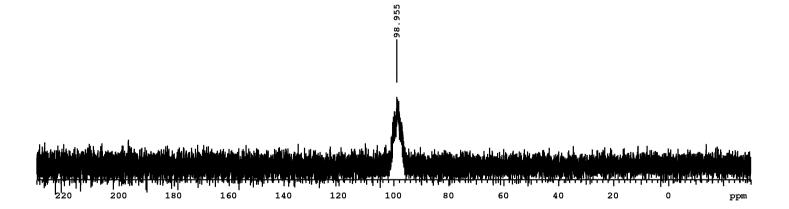


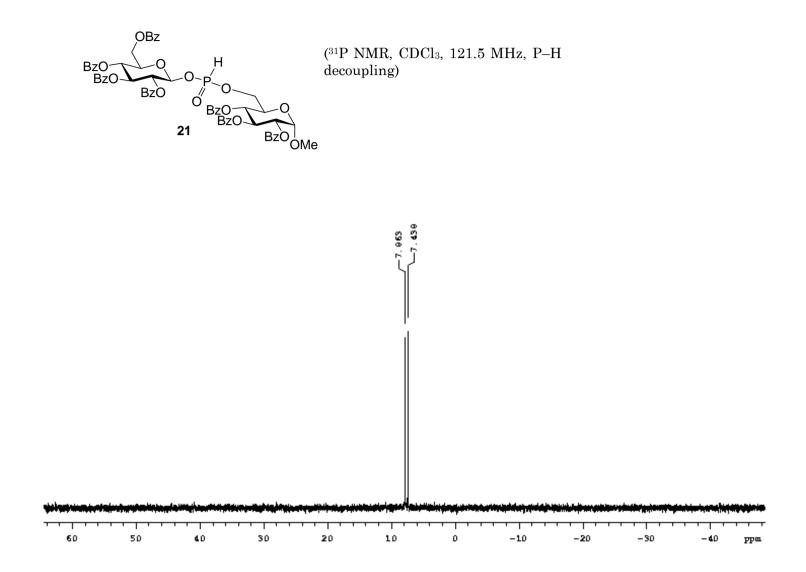


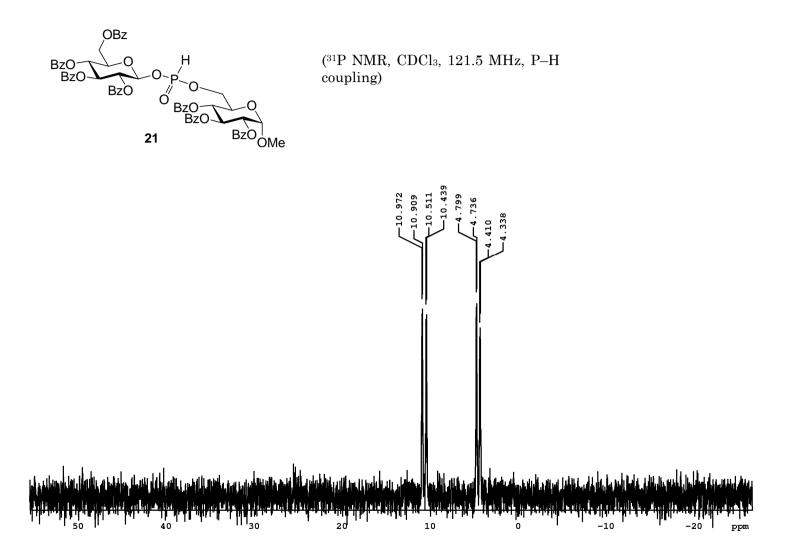


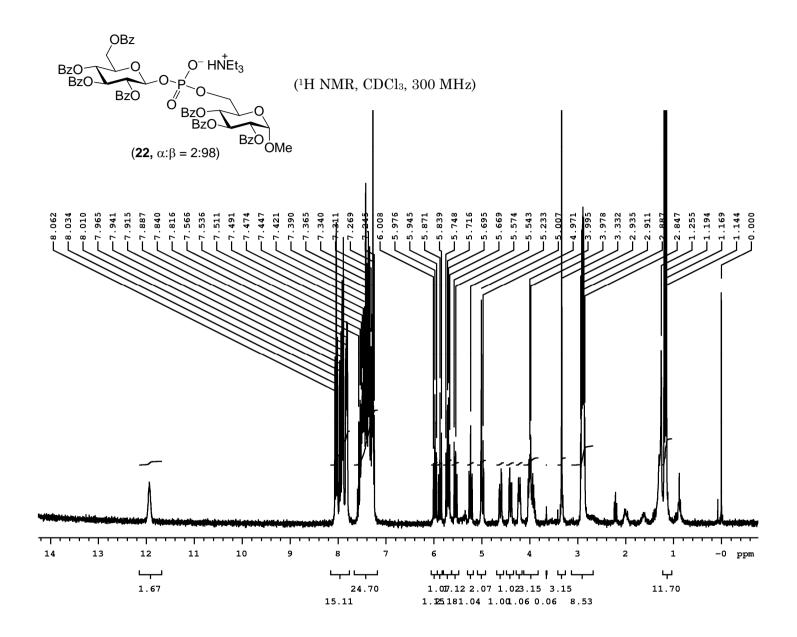


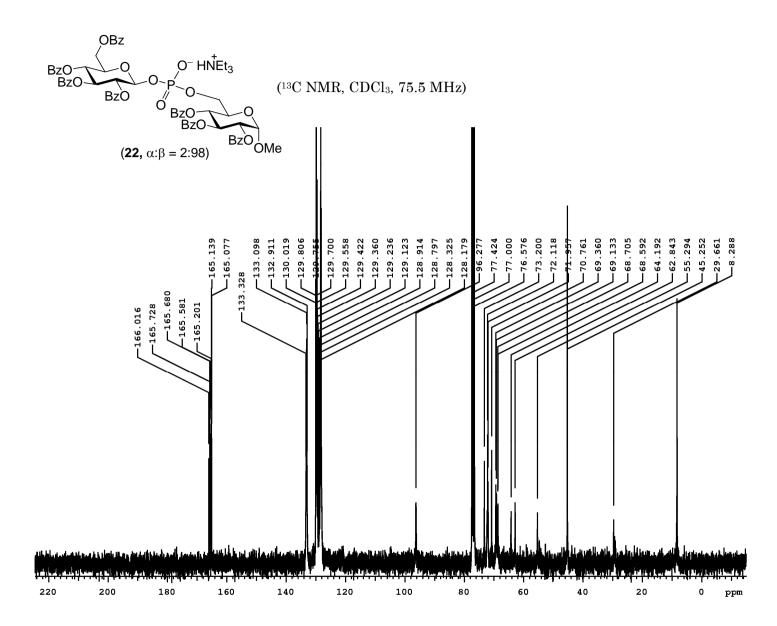


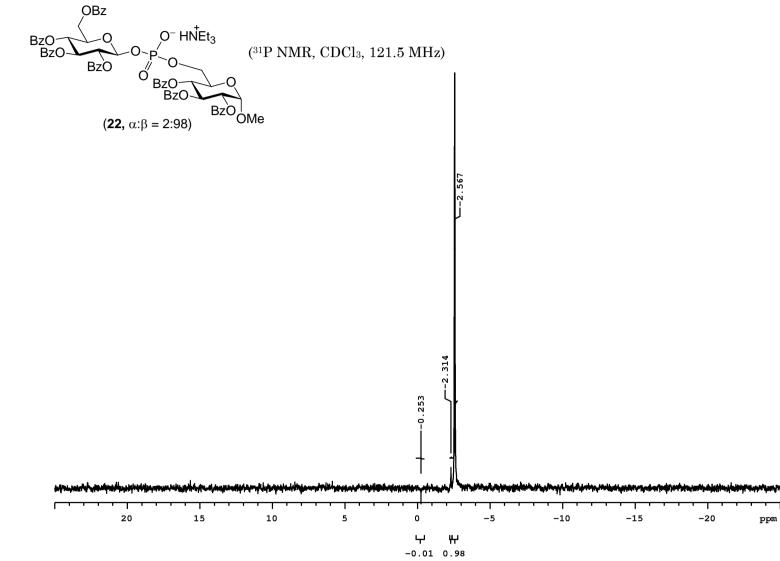












0.02

S58

