

Practical Synthesis of a Cathepsin S Inhibitor: Route Identification, Purification Strategies and Serendipitous Discovery of a Crystalline Salt Form

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Supporting information

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|---|-----|
| 1) General experimental methods | S-2 |
| 2) Chiral HPLC chromatograms of compound 1 | S-3 |
| 3) ¹ H and ¹³ C NMR spectra | S-4 |

General experimental methods:

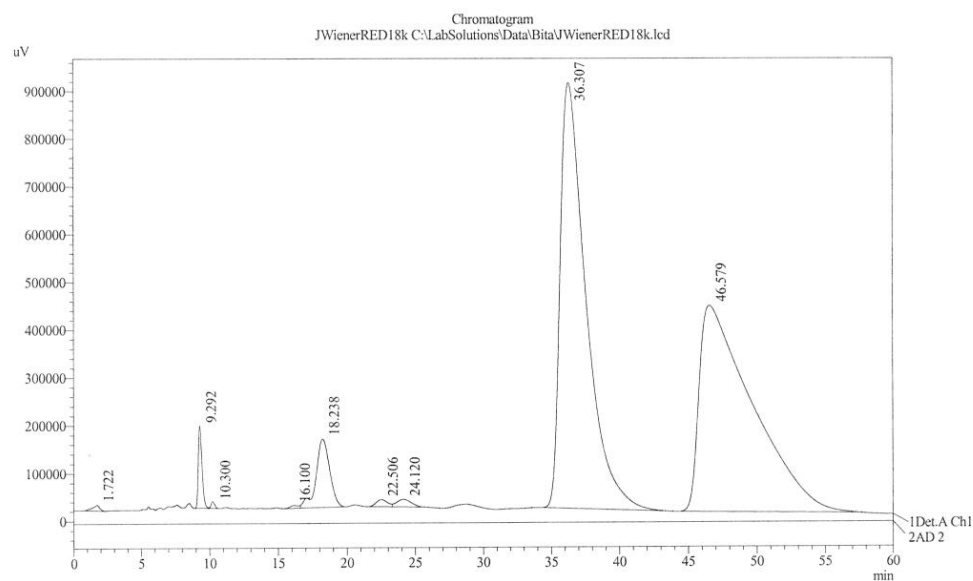
Proton and carbon NMR spectra were recorded at a 500 MHz or a 600 MHz NMR spectrometers. Flash column chromatography was performed using Merck silica gel 60. HRMS (ESI) was performed on a μ Tof apparatus. Powder X-ray Diffraction (PXRD) conditions: Philips X'PERT PRO with X'Celerator Cu detector equipped with a real time multiple strips X-ray detection technology. The samples were scanned from 4° to 40° 2 θ , at a step size 0.0167° 2 θ and a time per step of 29.8450 seconds. The tube voltage and current were 45 kV and 40 mA, respectively. The samples were placed onto zero background holders and analyzed on a spinning stage. The thermogravimetric analyses (TGA) were performed on a TA Instruments model TGA Q 50 under a nitrogen purge. The samples were placed in aluminum pans and scanned at a rate of 10°C/min up to 400 °C. Differential Scanning Calorimetry (DSC) was performed on a DCS Q 100. Samples were scanned with a temperature ramp of 10 °C/min. Samples were placed in aluminum DSC pans with a single hole punched into the lid up to 350 °C. Polarized Light Microscopy (PLM) was performed on a Olympus model BX51 polarizing microscope, equipped with a PAX-IT digital camera. The magnification was 500X.

Unless specified, all the reagents and solvents were purchased from commercial sources and used without further purification.

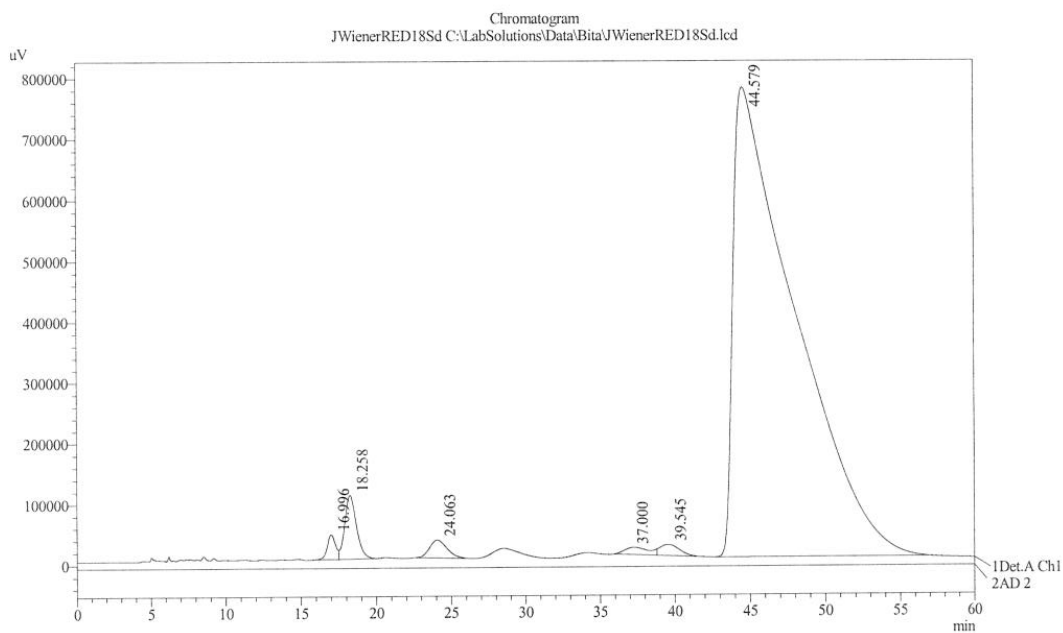
Chiral HPLC chromatograms of compound 1

Chiral separation conditions are: The Chiralpak AD-H 250 x 4.6 mm @ 40 Celsius, 50% MeOH: EtOH (1:1) with 0.1% diethylamine, 50% hexanes @ 0.5mL/min, Detection UV 214nm.

Racemic sample



Compound 1



¹H NMR of compound **2**, DMSO-d₆, 600 MHz

8.16
7.65
7.64
7.58

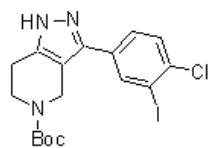
4.55

3.64
3.63
3.62

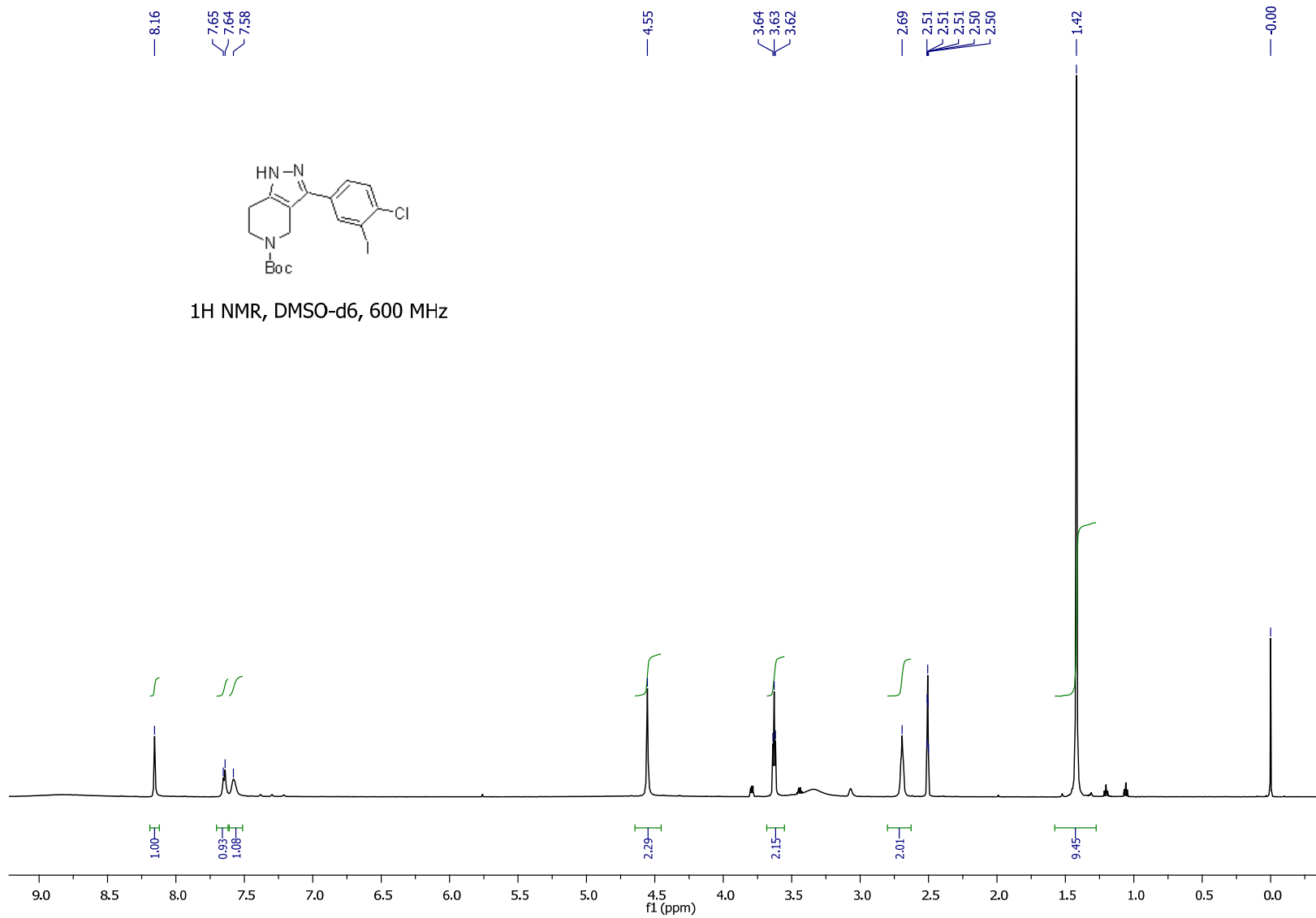
2.69
2.51
2.51
2.51
2.50
2.50

1.42

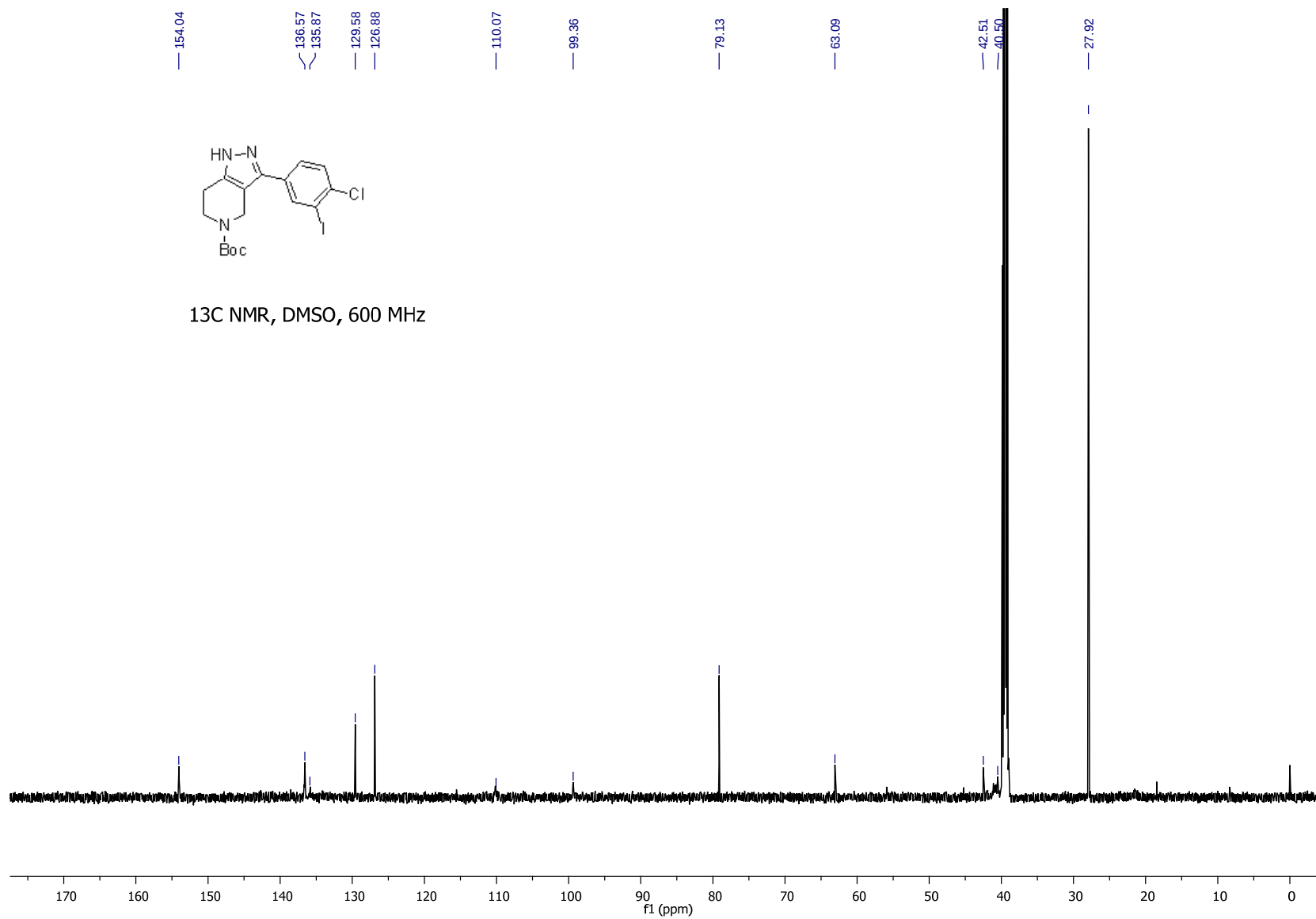
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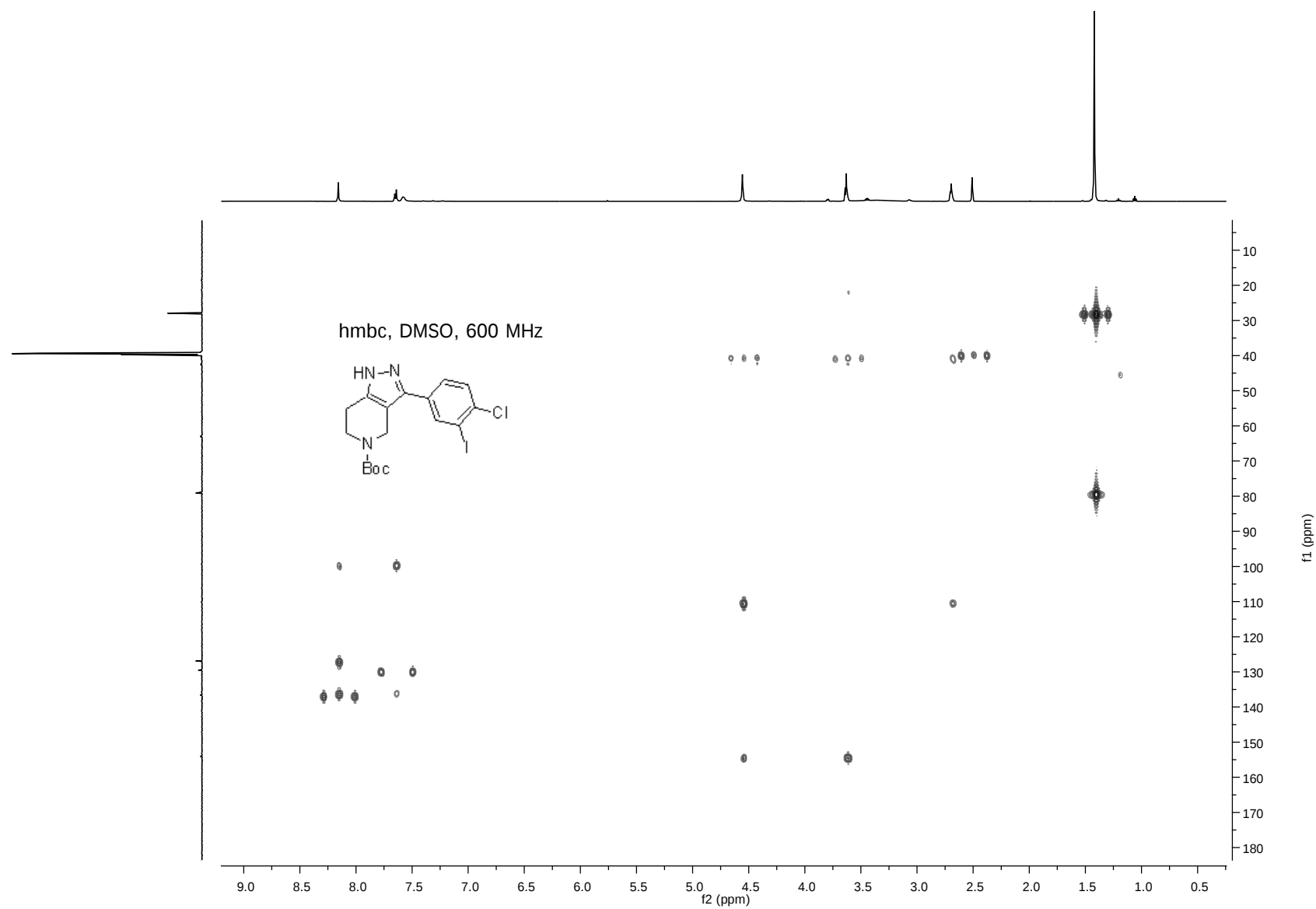


¹H NMR, DMSO-d₆, 600 MHz

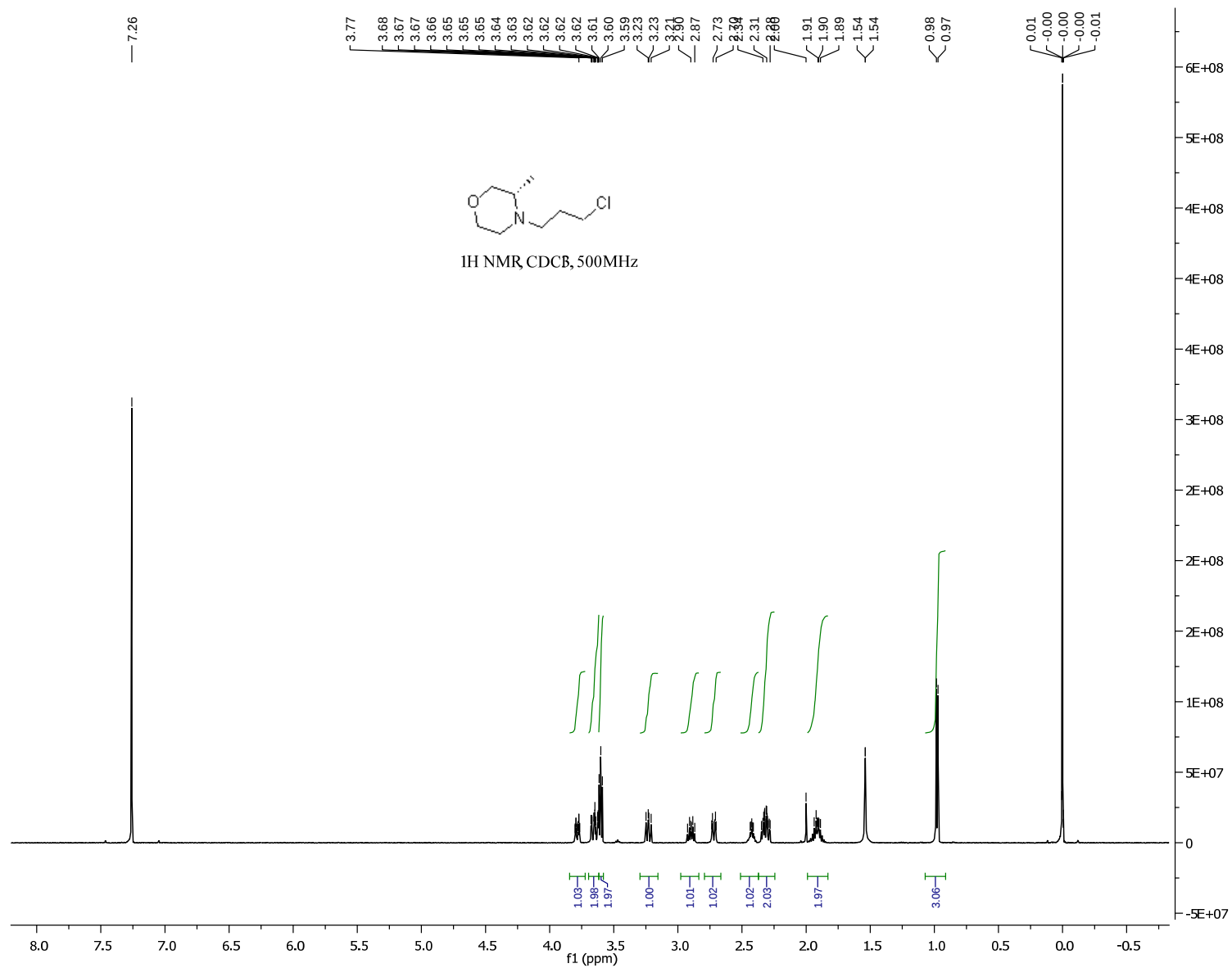


^{13}C NMR of compound 2, DMSO-d₆, 600 MHz

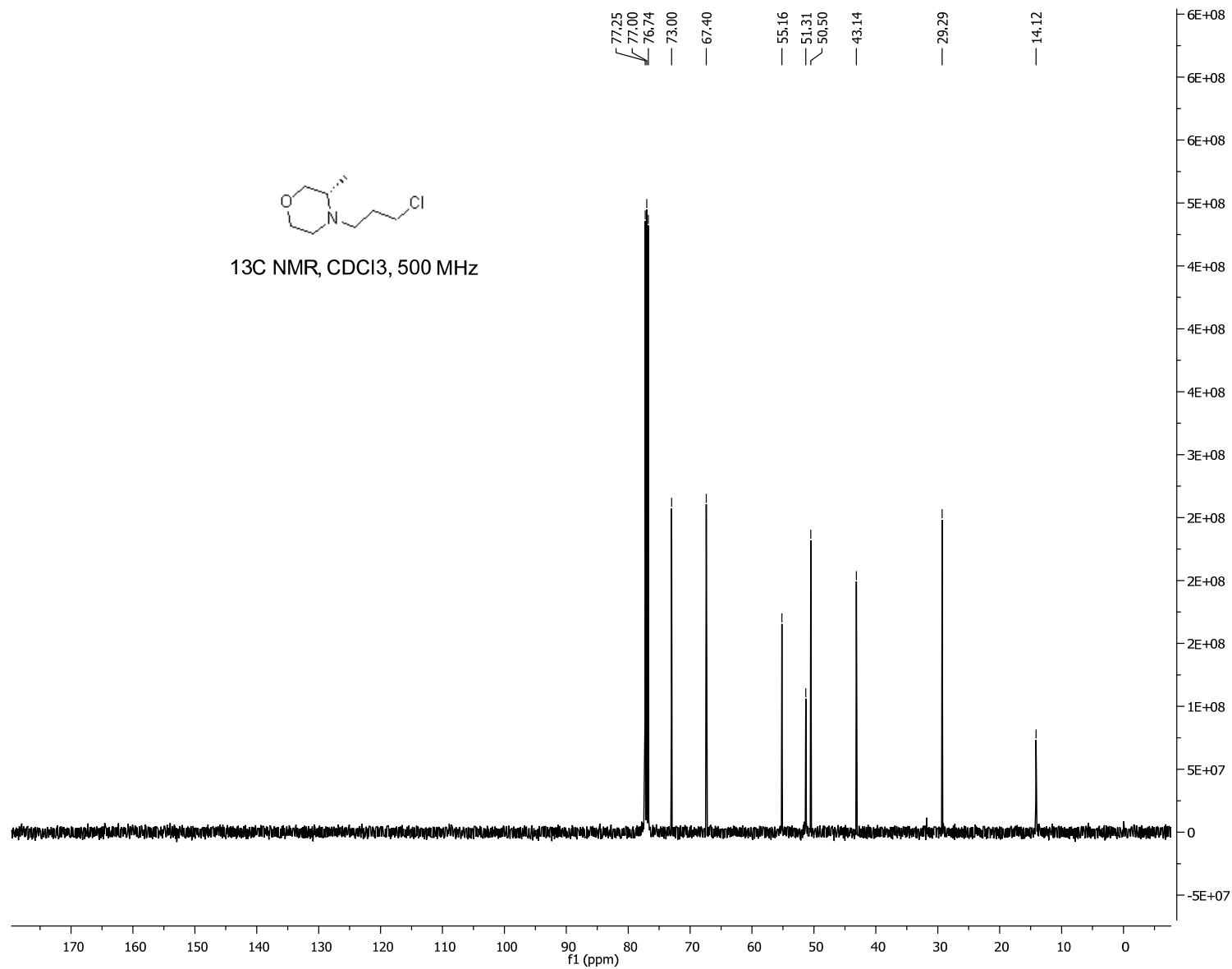


hmbc of compound 2, DMSO-d₆, 600 MHz

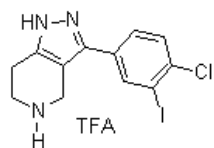
^1H NMR of compound **3**, CDCl_3 , 500 MHz



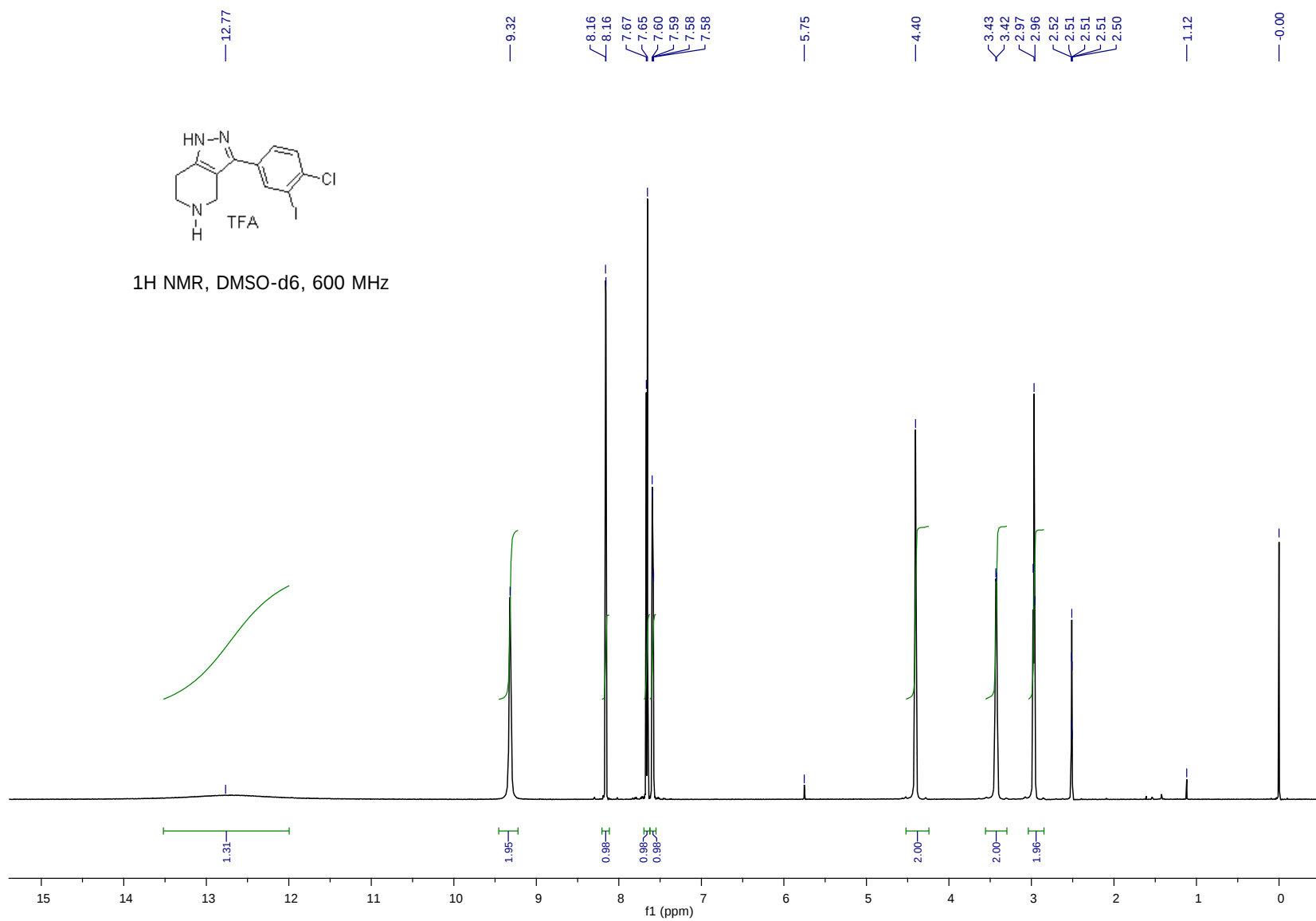
^{13}C NMR of compound **3**, CDCl_3 , 500 MHz



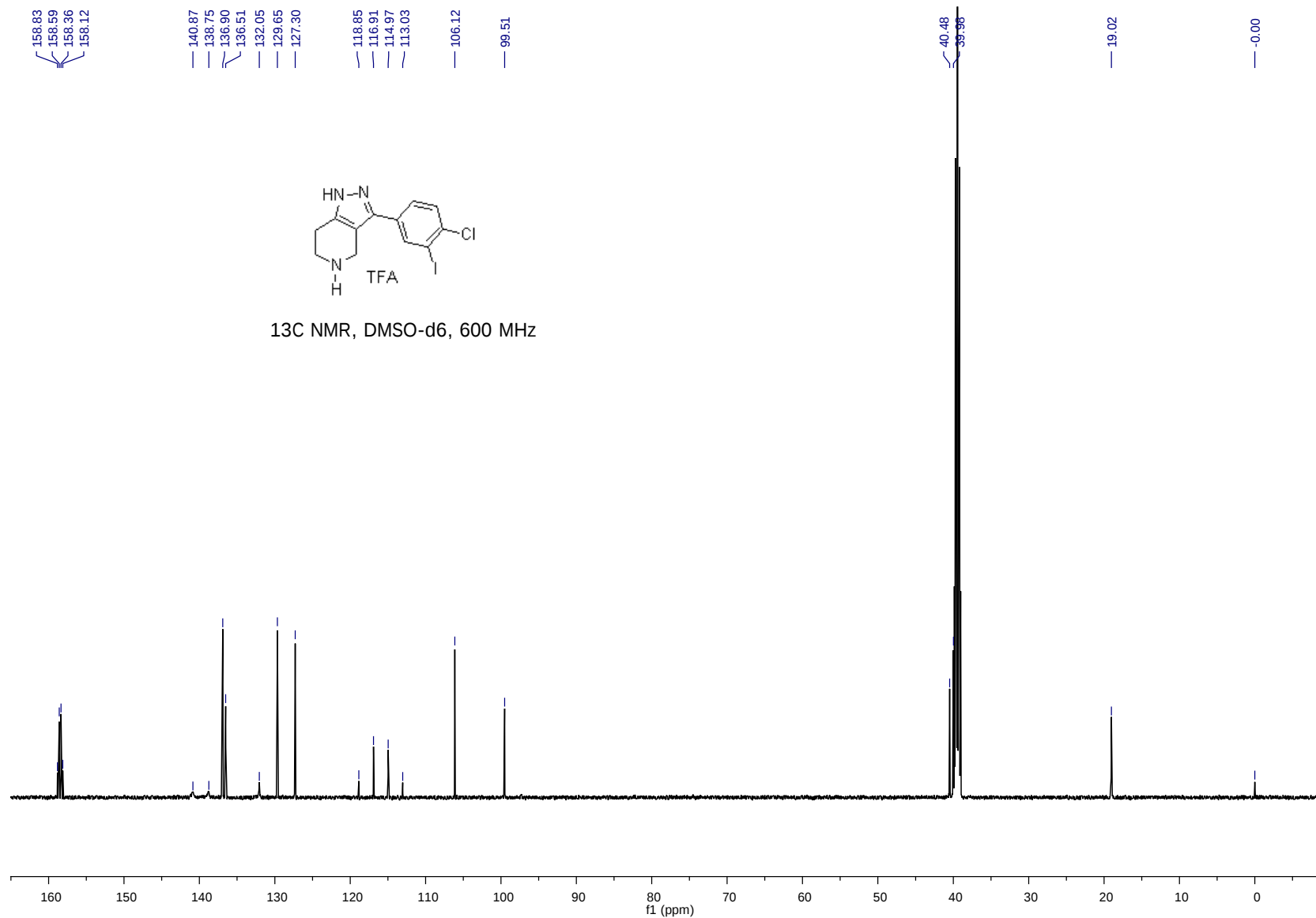
^1H NMR of compound **8**, TFA salt, DMSO- d_6 , 600 MHz



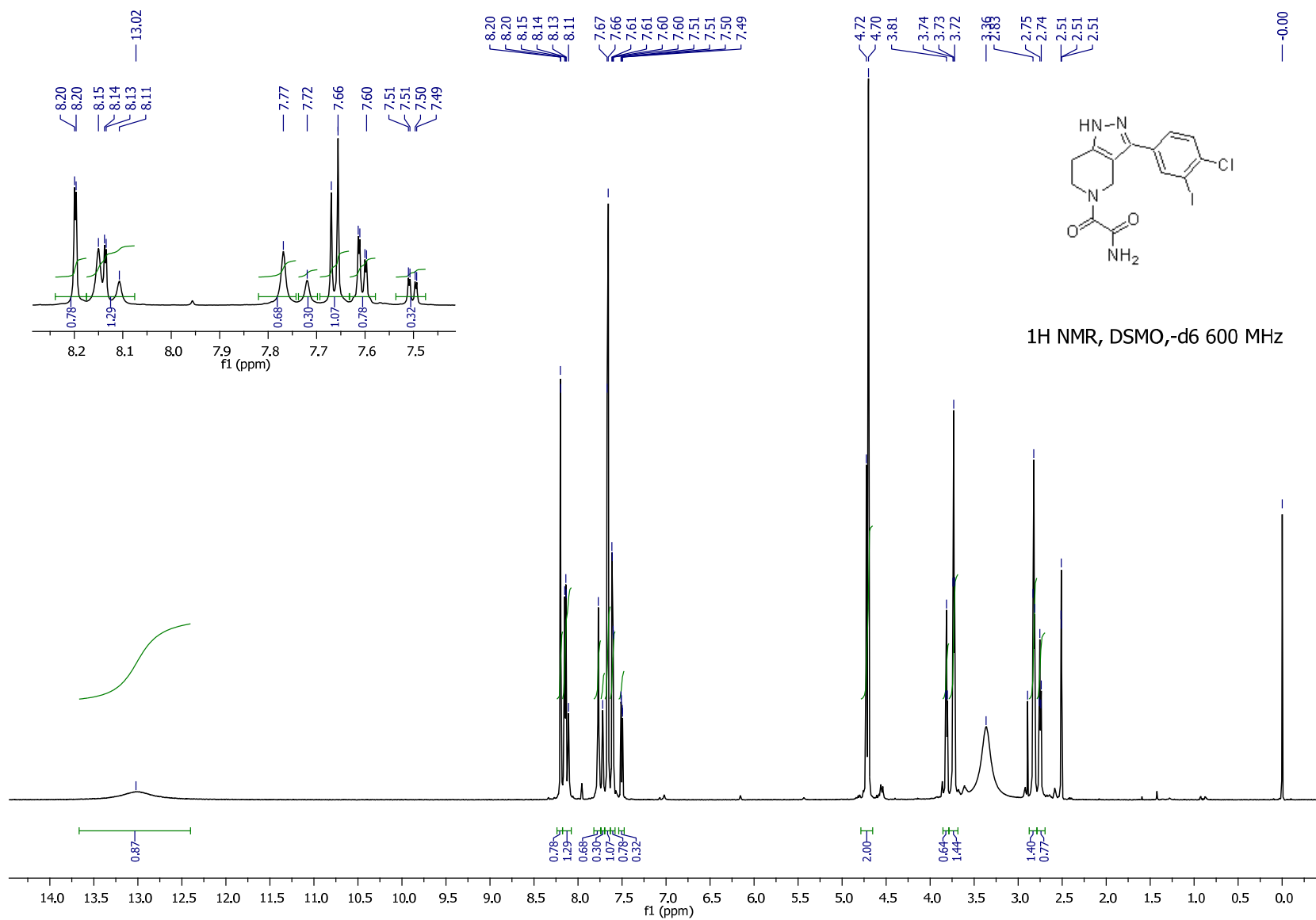
^1H NMR, DMSO- d_6 , 600 MHz



^{13}C NMR of compound **8**, TFA salt, DMSO- d_6 , 600 MHz



^1H NMR of compound **9**, DMSO- d_6 , 600 MHz



^{13}C NMR of compound **9**, DMSO- d_6 , 600 MHz

165.61
165.29
164.48
163.84

136.70
136.60
136.13
136.02

129.63
126.98
126.90

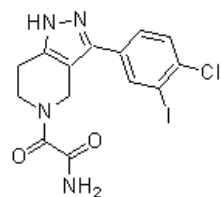
109.65
109.17

99.45
99.37

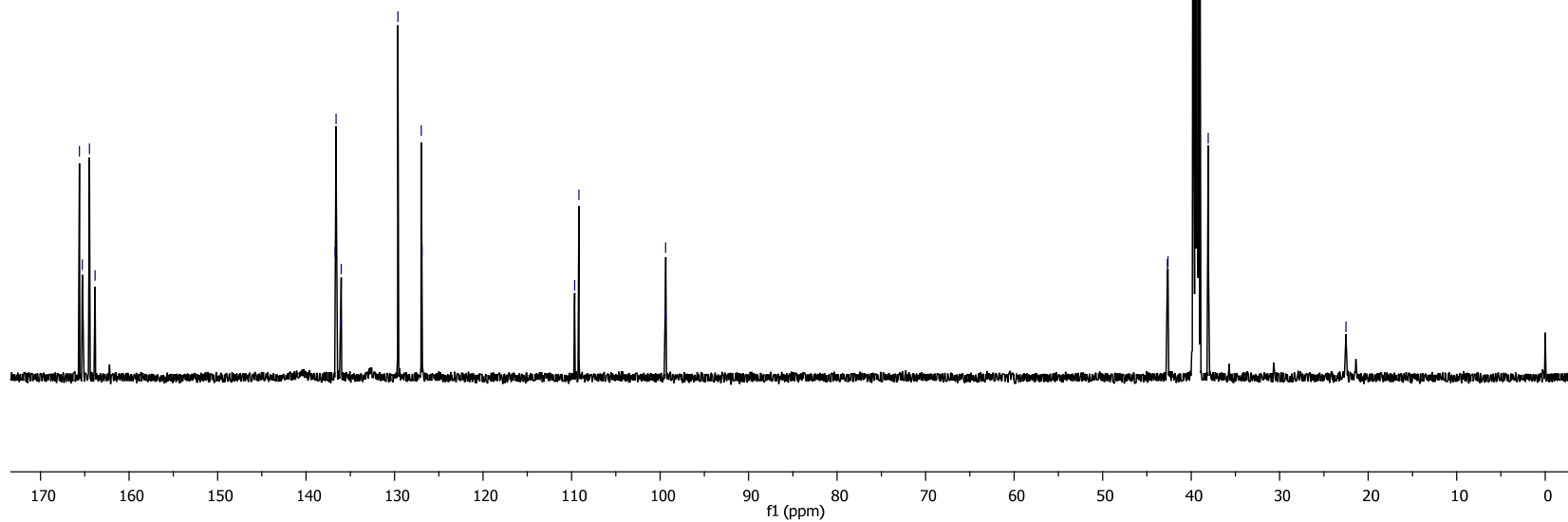
42.69
42.62

39.41
39.27
39.13
38.99
38.06

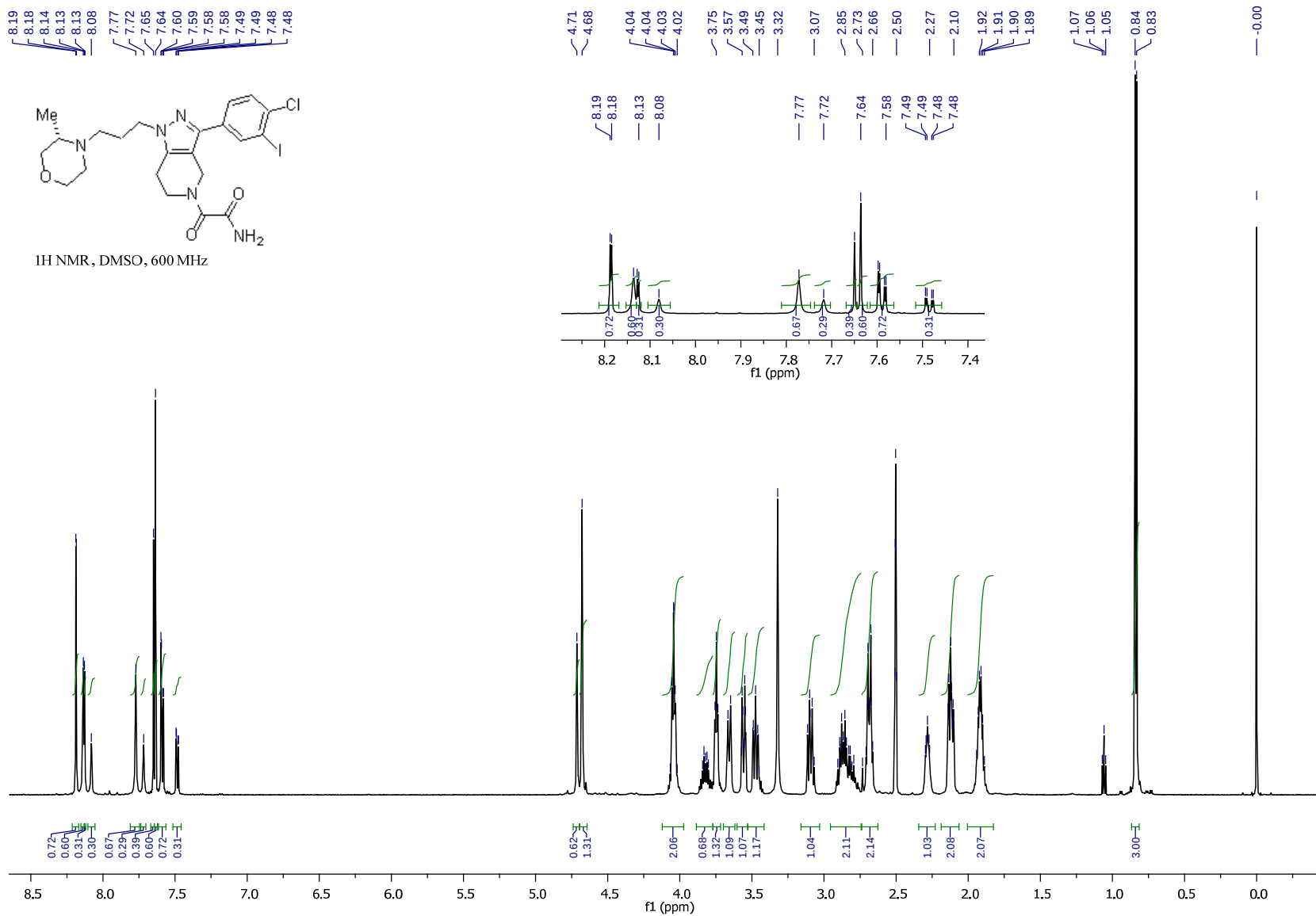
22.51



^{13}C NMR, DMSO- d_6 , 600 MHz



^1H NMR of compound **10**, DMSO- d_6 , 600 MHz



^{13}C NMR of compound **10**, DMSO- d_6 , 600 MHz

165.46
165.11
164.31
163.71

142.02
141.74

136.60
135.83

133.65

129.59

126.94
126.84

110.21
109.70

99.35
99.27

72.03
72.01

66.54

54.61

50.21
49.62

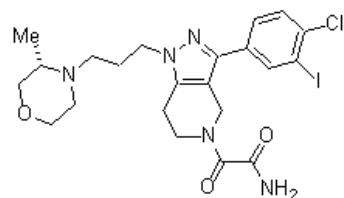
42.62
42.29

38.03
37.71

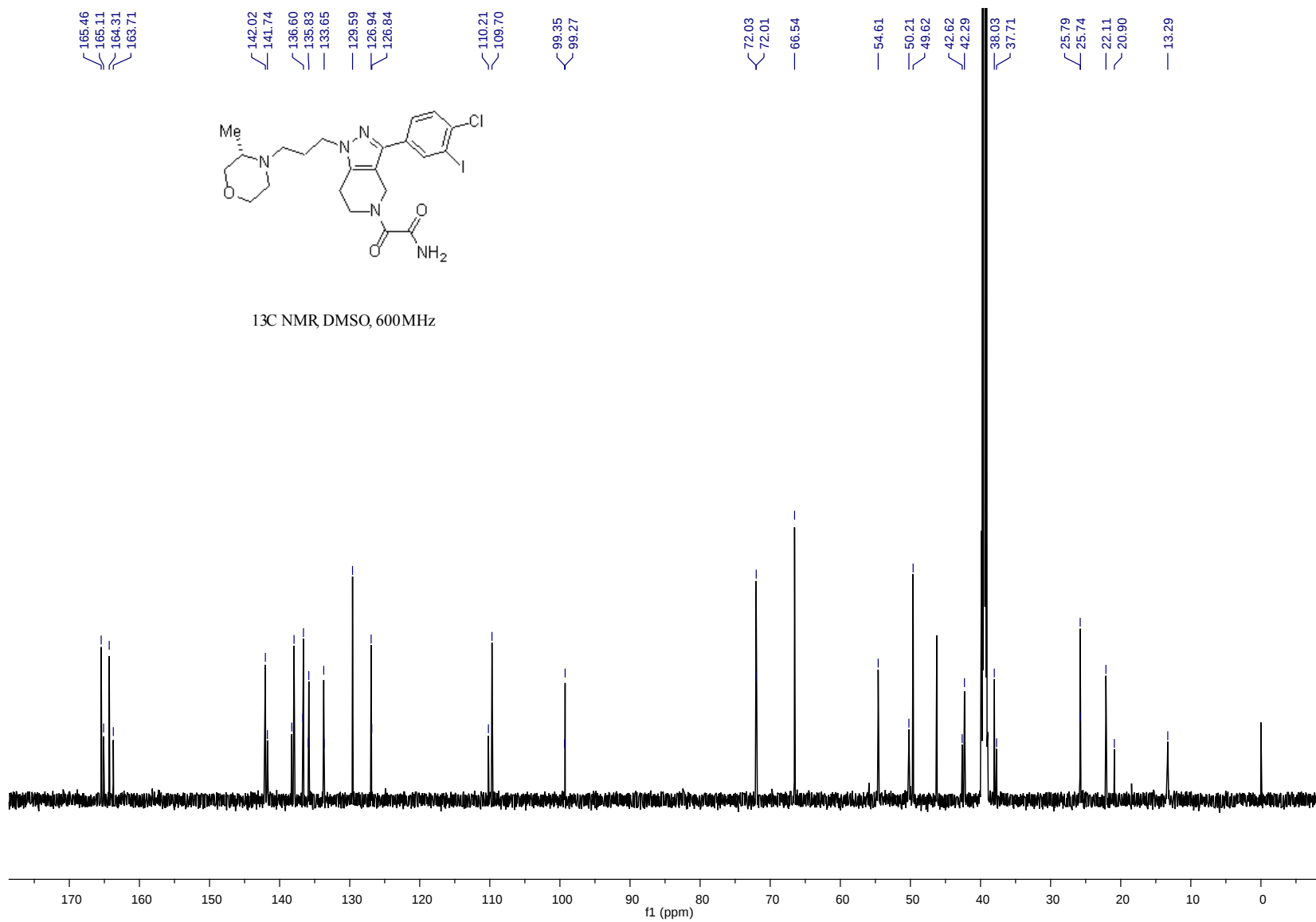
25.79
25.74

22.11
20.90

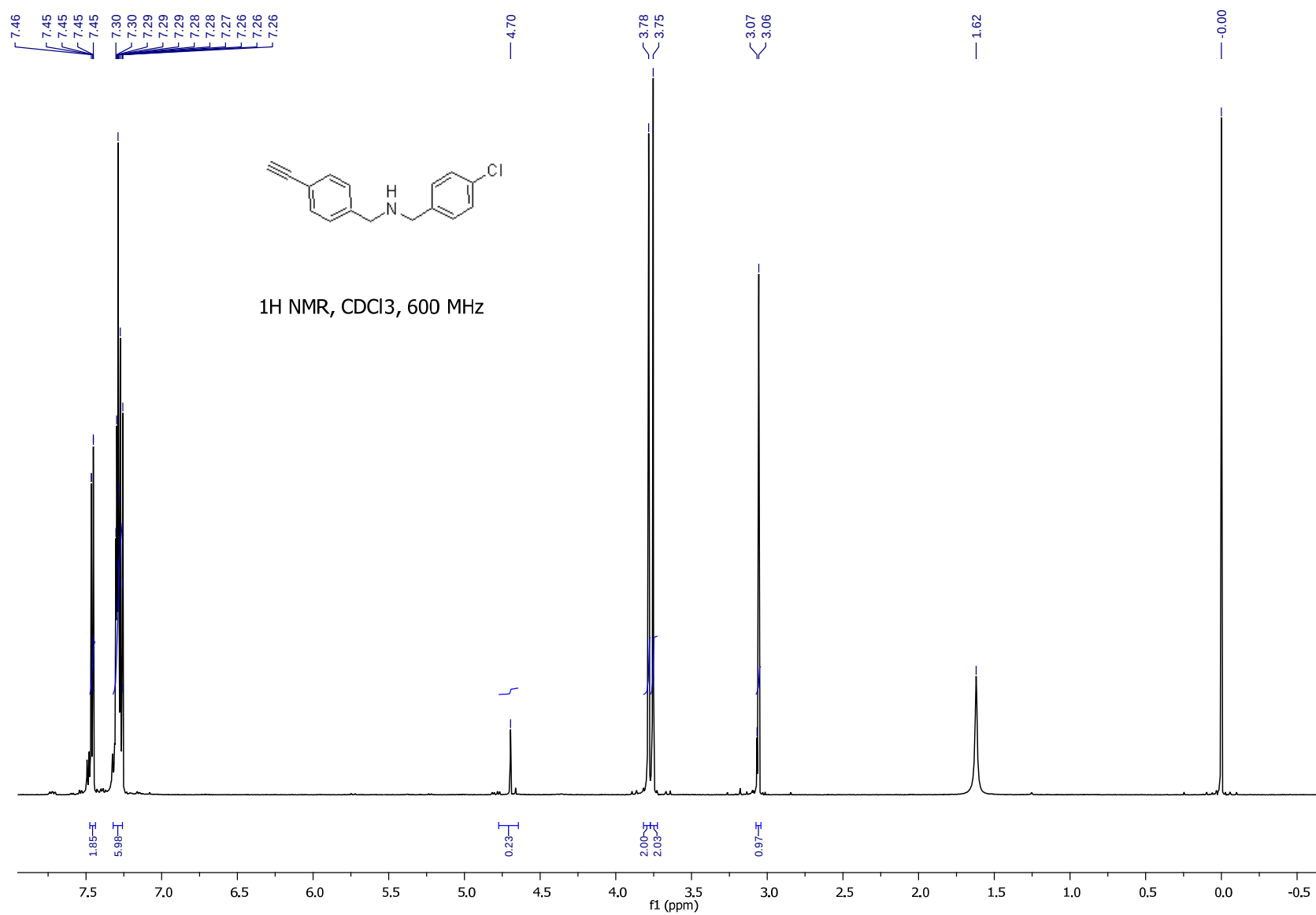
13.29



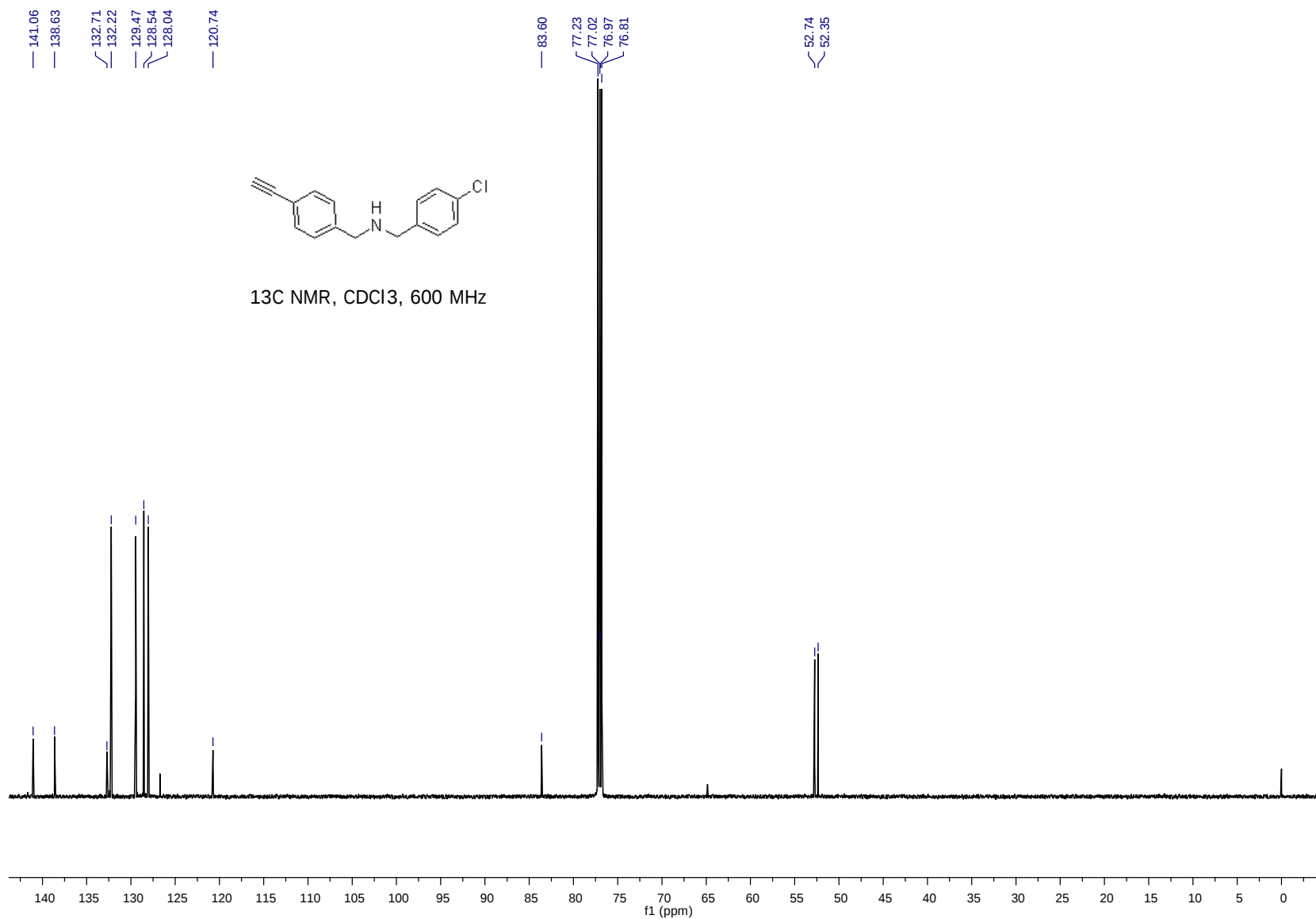
^{13}C NMR, DMSO, 600MHz

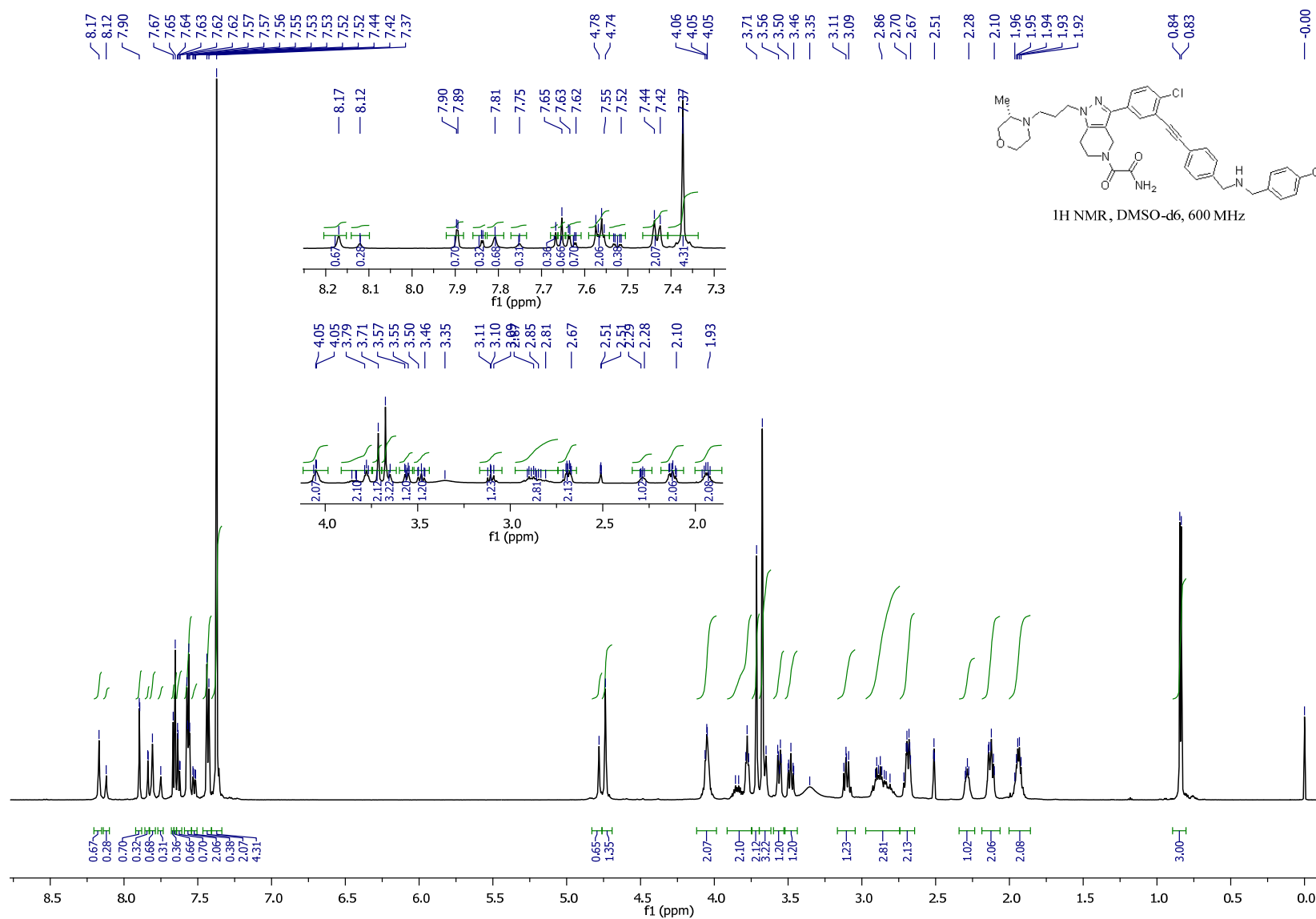


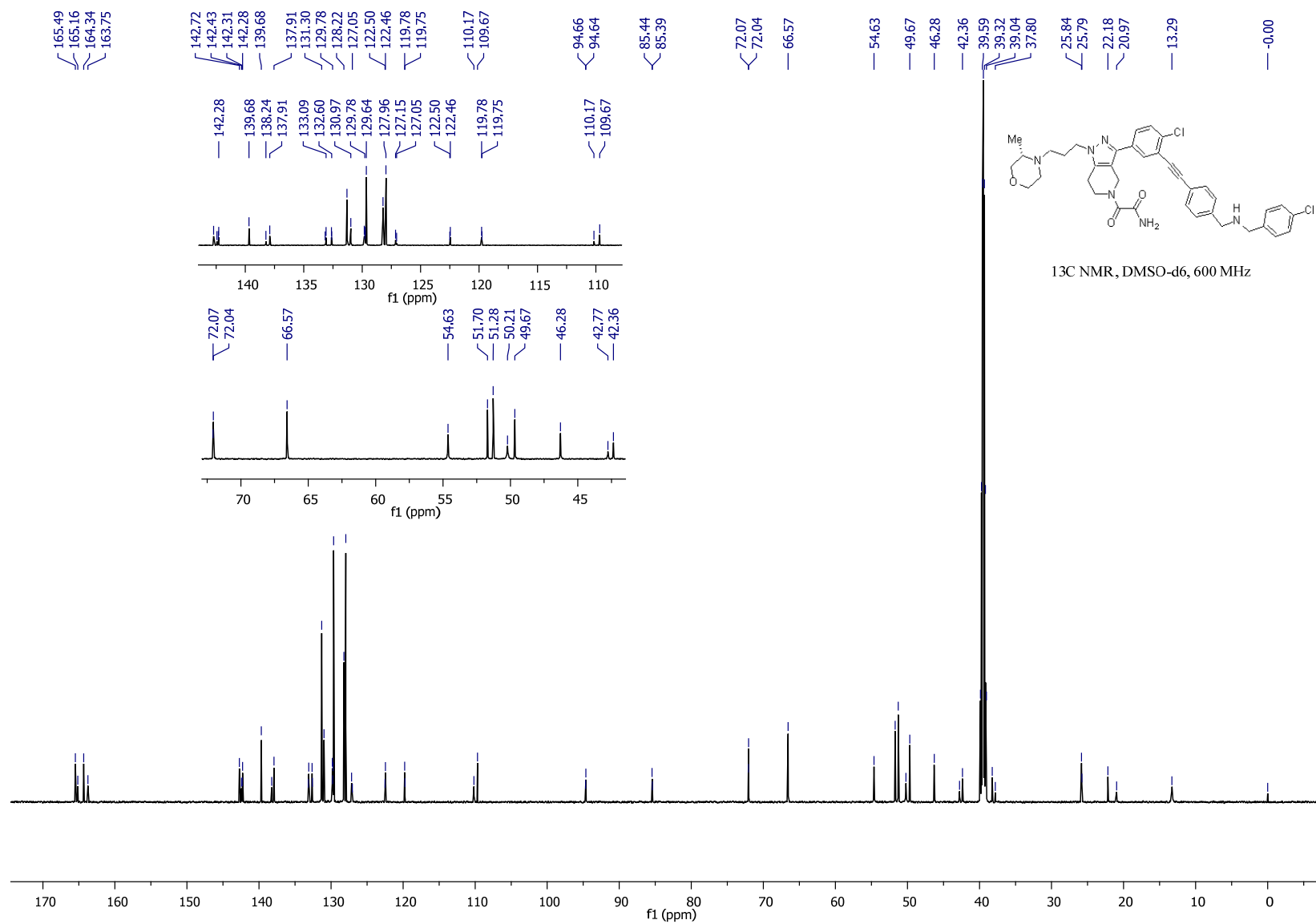
^1H NMR of compound **4**, CDCl_3 , 600 MHz

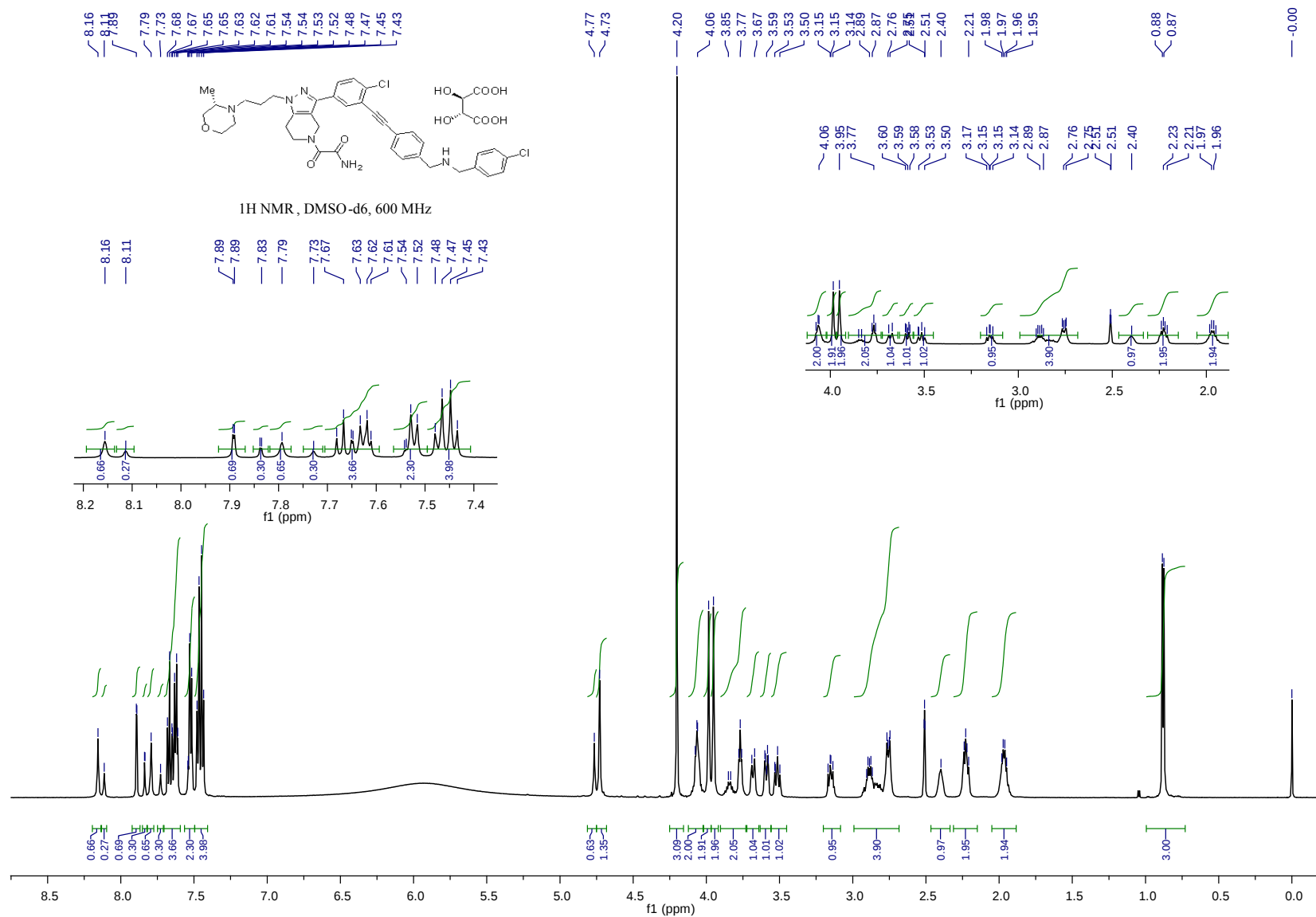


^{13}C NMR of compound **4**, CDCl_3 , 600 MHz

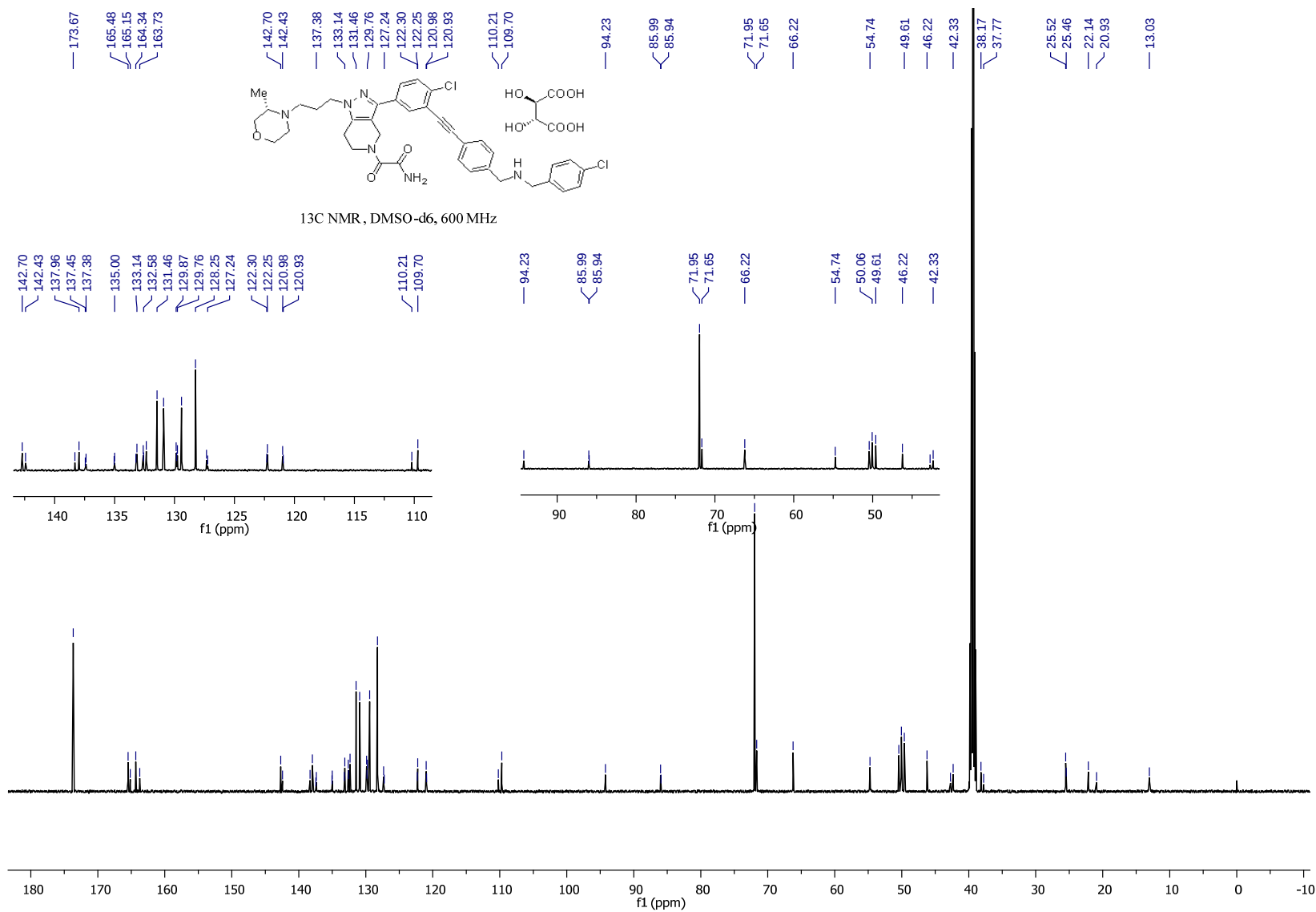


¹H NMR of compound **1**, DMSO-d₆, 600 MHz

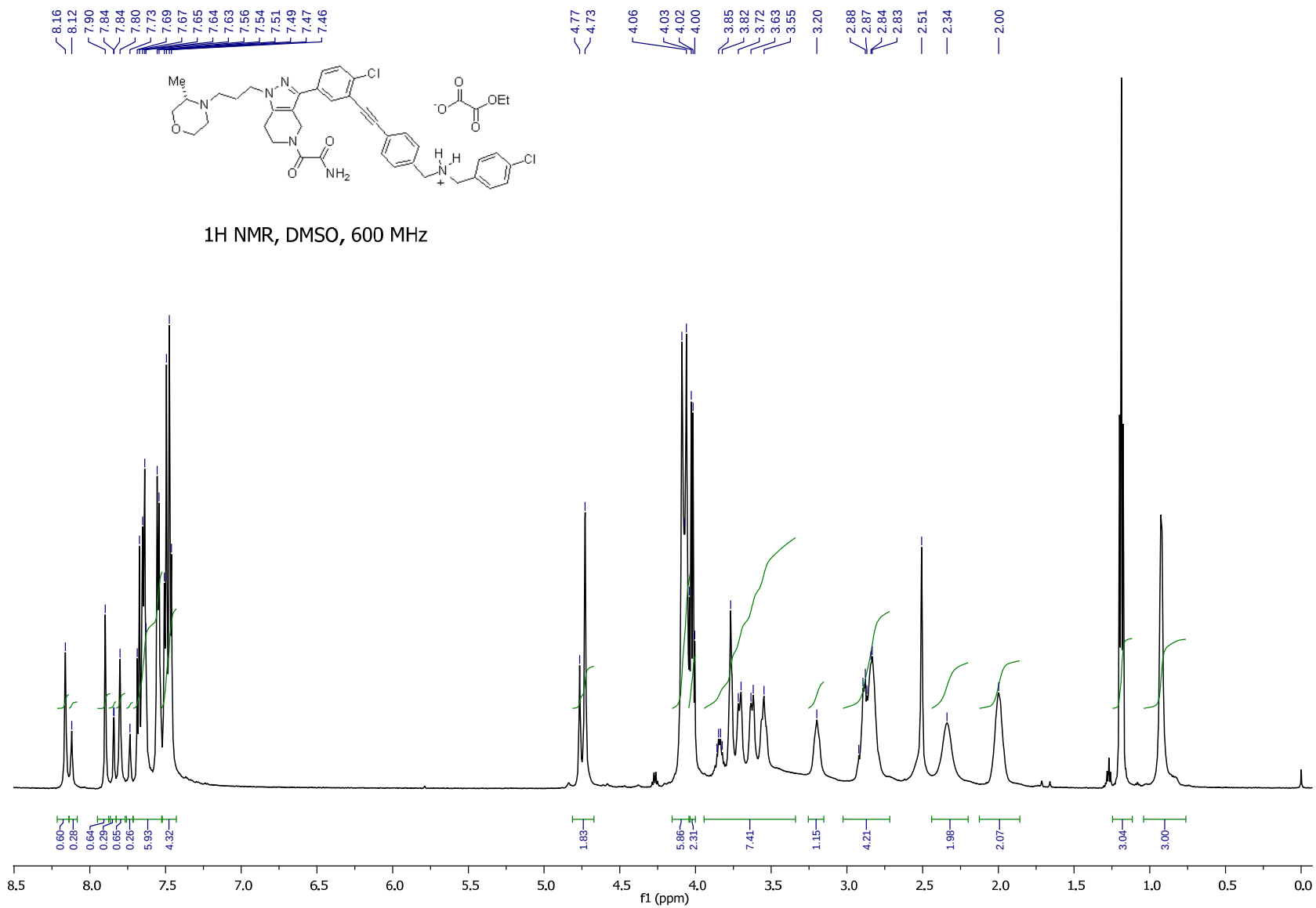
^{13}C NMR of compound **1**, DMSO-d₆, 600 MHz

^1H NMR of compound **1**, L-tartrate, DMSO- d_6 , 600 MHz

^{13}C NMR of compound **1**, L-tartrate, DMSO- d_6 , 600 MHz



^1H NMR of compound 12, DMSO- d_6 , 600 MHz



^{13}C NMR of compound **12**, DMSO- d_6 , 600 MHz