

A Very Efficient Synthesis of a Mannosyl Orthoester Rotaxane and Mannosidic Rotaxanes

Frédéric Coutrot*, Eric Busseron, and Jean-Louis Montero

Institut des Biomolécules Max Mousseron (IBMM), UMR 5247CNRS-Universités Montpellier 2 et 1, Bâtiment de Recherche Max Mousseron, Ecole Nationale Supérieure de Chimie de Montpellier, 8 rue de l'Ecole Normale, 34296 Montpellier cedex 5, France

frederic.coutrot@univ-montp2.fr

1. General methods.....	S2
2. Complexation study between compound 2 and DB24C8	S2
3. Synthetic procedures and characterization of compounds 1 and 2	S3
3.1 Synthetic procedure of compound 1	S3
3.2 Synthetic procedure and characterization of compound 2	S3
4. General procedure for glycosylation	S4
5. General procedure for isomerization of orthoester 6	S4
6. Characterization of rotaxanes 5a-c , 6	S4
6.1 Characterization of compound 5a	S4
6.2 Characterization of compound 5b	S5
6.3 Characterization of compound 5c	S5
6.4 Characterization of compound 6	S5
7. Characterization of threads 3a-b , 4	S6
7.1 Note about threads.....	S6
7.2 Characterization of deprotonated compound 3a	S6
7.3 Characterization of deprotonated compound 3b	S6
7.4 Characterization of deprotonated compound 4	S7
8. NMR spectra	S8

8.1 NMR spectra of compound 2	S8
8.2 NMR spectra of compound 5a	S12
8.3 NMR spectra of compound 5b	S14
8.4 NMR spectra of compound 5c	S16
8.5 NMR spectra of compound 6	S18
8.6 NMR spectra of deprotonated compound 3a	S20
8.7 NMR spectra of deprotonated compound 3b	S22
8.8 NMR spectra of deprotonated orthoester compound 4	S24
9. References	S26

1. General methods

All reactions were achieved under an atmosphere of argon unless otherwise indicated and all reactions of glycosylation were carried out on a 100 mg scale. Flasks were oven dried and allowed to cool under argon prior to use. All reagents were purchased from Aldrich or Senn Chemical and were used as received without further purification. Dichloromethane was distilled over P₂O₅. Analytical thin-layer chromatography (TLC) was performed on Merck silicagel 60 F254 plates. Compounds were visualized by dipping the plates in an ethanolic solution of 10% sulphuric acid or in an ethanolic solution of 10% ninhydrine, followed by heating. ¹H NMR and ¹³C NMR spectra were obtained on a Bruker DRX-400 spectrometer (respectively at 400.13 MHz and 100.62 MHz). Chemical shifts of ¹H NMR and ¹³C NMR are given by using CHCl₃ as the reference (7.27 ppm for ¹H spectrum and 77 ppm for ¹³C spectrum). Coupling constants (J) are reported in hertz (Hz). Standard abbreviations indicating multiplicity were used as follows: s (singlet), br (broad), d (doublet), t (triplet), q (quartet), m (multiplet). Lettering and numbering of hydrogens and carbons correspond to the assignments indicated in Scheme 1 of the article. The electrospray ionisation spectra were obtained on a Waters 2614 Micromass ZQ mass spectrometer in positive (ESI+) or negative (ESI-) mode. High-resolution mass spectra (HRMS) were recorded on a Q-TOF Micro (water) apparatus.

2. Complexation study between compound **2** and DB24C8

In order to study, by ¹H NMR spectroscopy, the binding of *N*-alkyl anilinium cation **2** with DB24C8, stock solutions of **2** and DB24C8 were made. The 20 mM solution of alcohol **2** was prepared by dissolving 18.5 mg (0.047 mmol) in CDCl₃ (2.340 mL) and the 100 mM solution of DB24C8 by dissolving 44.2 mg (0.099 mmol) in CDCl₃ (0.985 mL). Six samples of thread **2** were made according to table 1. Overlay of ¹H NMR spectra is reported in figure 1 of the article.

Table 1: Dilutions used to study complexation by ¹H NMR.

sample ID	thread 2 mmol	DB24C8 mmol	vol. thread 2 stock solution μ L	vol. DB24C8 stock solution μ L	vol. blank CDCl ₃ μ L
1	10	0	250	0	250
2	10	5	250	25	225
3	10	10	250	50	200
4	10	15	250	75	175
5	10	20	250	100	150
6	10	25	250	125	125

3. Synthetic procedures and characterization of compounds 1 and 2

3.1 Synthetic procedure of compound 1

This compound was synthesised in a two-step sequence according to the procedure described by Vishwakarma et al. from 1,2,3,4,6-penta-*O*-acetyl- α -*D*-mannopyranose.¹

3.2 Synthetic procedure and characterization of compound 2

- First step

To a solution of 4-*tert*-butylaniline (7.4 g, 50 mmol, 3.1 equiv) in toluene (60 mL) under reflux was added dropwise 6-aminohexan-1-ol (3.03 g, 16 mmol, 1 equiv) *via* a syringe pump over a period of 5 hours. The mixture was stirred under reflux overnight. The solvent was then removed and the residue was diluted with AcOEt (75 mL). The organic layer was washed with an aqueous solution of NaOH (2M) until pH 10. Then the aqueous layer was extracted with AcOEt (4 x 75 mL). The organic layers were combined, dried over Na₂SO₄ and concentrated. The oily residue was distilled under reduced pressure to give pure aminoalcohol (2.24 g, 54%, bp = 170 °C, 1.4 mbar) as a pale yellow oil.

R_f 0.52 (20/80 AcOEt/CH₂Cl₂)

¹H NMR (400 MHz, CDCl₃, 298 K): δ ppm 1.29 (s, 9H, H₁₆), 1.42 – 1.45 (m, 4H, H₉ H₁₀), 1.56 – 1.68 (m, 4H, H₈ H₁₁), 3.12 (t, 2H, J = 7.1 Hz, H₁₂), 3.66 (t, 2H, J = 6.7 Hz, H₇), 6.60 (m, 2H, H₁₄), 7.22 (m, 2H, H₁₅).

¹³C NMR (100 MHz, CDCl₃, 298 K): δ ppm 25.5 & 26.9 (C₉ C₁₀), 29.5 (C₁₁), 31.5 (C₁₆), 32.6 (C₈), 33.8 (C_q tBu), 44.3 (C₁₂), 62.8 (C₇), 112.7 (C₁₄), 126.0 (C₁₅), 140.2 & 145.7 (C_q arom).

MS (ESI⁺): 250 [M+H]⁺

- Second step

To a solution of the previous oily compound (2.24 g, 9 mmol, 1 equiv) in Et₂O (10 mL), was introduced dropwise at RT a solution of HCl 2M in Et₂O (22.5 mL, 45 mmol, 5 equiv). The mixture was stirred for 30 min to give a biphasic mixture, which was evaporated under reduced pressure to remove the excess of HCl. Then, Et₂O was added to the residue, the biphasic mixture being stirred afterwards for 5 min. Eventually, the ether organic layer was removed. After having evaporated the residual traces of solvent, the hydrochloride ammonium salt was dissolved in H₂O (10 mL) and NH₄PF₆ (3.7 g, 22 mmol, 2.5 equiv) was introduced. CH₂Cl₂ (15 mL) was added and the resulted bilayer solution was vigorously stirred for 2h. After separation, the aqueous layer was extracted with CH₂Cl₂. The organic layers were combined, dried over Na₂SO₄, and concentrated to obtain the pure compound **2** (3.3 g, 92%) as a pale brown oil.

¹H NMR (400 MHz, CDCl₃, 298 K): δ ppm 1.29 (s, 9H, H₁₆), 1.32-1.39(m, 4H, H₉ H₁₀), 1.52 (m, 2H, H₈), 1.73 – 1.80 (m, 2H, H₁₁), 3.37 (t, 2H, J = 7.8 Hz, H₁₂), 3.62 (t, 2H, J = 6.4 Hz, H₇), 7.37 (d, 2H, J = 8.7 Hz, H₁₅), 7.49 (d, 2H, J = 8.7 Hz, H₁₄).

¹³C NMR (100 MHz, CDCl₃, 298 K): δ ppm 24.5 (C₁₀), 25.1 (C₉), 25.2 (C₁₁), 31.1 (C₁₆), 31.4 (C₈), 34.8 (C_q tBu), 54.0 (C₁₂), 62.7 (C₇), 121.9 (C₁₅), 127.5 (C₁₄), 131.6 & 153.6 (C_q arom).

MS (ESI⁺): 250 [M+H]⁺

MS (ESI⁻): 145 [PF₆]⁻

4. General procedure for glycosylation

A solution of trichloroacetimidate **1** (102 mg, 0.21 mmol, 1 equiv) in dry CH₂Cl₂ (4 mL), in the presence of 4 Å molecular sieves, under argon atmosphere, was cooled until -30 °C. Meanwhile in another flask, DB24C8 (204 mg, 0.45 mmol, 2.2 equiv) was added to a solution of compound **2** (90 mg, 0.23 mmol, 1.1 equiv) in dry CH₂Cl₂ (3 mL) under argon atmosphere. This mixture was stirred for 10 min at RT and was transferred to the first solution. When the temperature was stabilized, TMSOTf (8 µL, 44 µmol, 0.2 equiv) was added and the mixture was stirred for 1 min at the same temperature. The reaction was quenched by addition of triethylamine (6 µL, 44 µmol, 0.2 equiv) and stirred for 5 min. Then the mixture was allowed to reach RT; it was evaporated to dryness and the crude was analysed by ¹H NMR (Table 1 of the article). The residue was purified by chromatography on a silicagel column (solvent gradient elution: 5/95 Acetone/CH₂Cl₂, 10/90, 20/80) to give successively compound **5a** (31 mg, 13%) and compound **6** (145 mg, 60%) as solids.

5. General procedure for isomerization of orthoester **6**

A solution of rotaxane orthoester **6** (40 mg, 34 µmol, 1 equiv) in dry CH₂Cl₂ (8 mL) in the presence of 4 Å molecular sieves under argon atmosphere was cooled to -15 °C. A solution of TMSOTf (25 µL, 7 µmol, 0.2 equiv, C = 0.28 mol.L⁻¹) in CH₂Cl₂ was then added and the reaction mixture was stirred at this temperature for 5 min. The reaction was quenched by addition of a solution of triethylamine (19 µL, 7 µmol, 0.2 equiv, C = 0.36 mol.L⁻¹) in CH₂Cl₂ and stirred for another 5 min. Then the mixture was allowed to reach RT and the solvent was removed. The crude residue was analysed by ¹H NMR (Table 2 of the article) and purified by chromatography on a silicagel column (solvent gradient elution: 2/98 Acetone/CH₂Cl₂, 5/95, 10/90) to give **5a** (9 mg; 22%) as a solid.

6. Characterization of rotaxanes **5a-c**, **6**

6.1 Characterization of compound **5a**

R_f 0.76 (15/85 Acetone/CH₂Cl₂)

¹H NMR (400 MHz, CDCl₃, 298 K): δ ppm 0.94-1.03 (m, 4H, H₉ H₁₀), 1.17-1.23 (m, 2H, H₈), 1.26 (s, 9H, H₁₆), 1.40-1.48 (m, 2H, H₁₁), 2.01 & 2.06 & 2.11 & 2.17 (4s, 12H, OAc), 3.16-3.23 (m, 1H, H_{7a}), 3.28-3.34 (m, 4H, H_E), 3.47-3.53 (m, 1H, H_{7b}), 3.56-3.62 (m, 4H, H_{E'}), 3.78-3.87 (m, 8H, H_D), 3.90-3.99 (m, 3H, H₅ H₁₂), 4.08-4.13 (m, 5H, H_{6b} H_C), 4.20-4.26 (m, 4H, H_{C'}), 4.29 (dd, 1H, J = 12.3 Hz, J = 5.2 Hz, H_{6a}), 4.73 (d, 1H, J = 1.3 Hz, H_I), 5.20-5.22 (m, 1H, H₂), 5.26-5.33 (m, 2H, H₃ H₄), 6.85-6.92 (m, 8H, H_A H_B), 7.35 (d, 2H, J = 8.5 Hz, H₁₅), 7.54 (d, 2H, J = 8.5 Hz, H₁₄), 8.52-8.61 (br s, 2H, H₁₃).

¹³C NMR (100 MHz, CDCl₃, 298 K): δ ppm 20.7 & 20.7 & 20.8 & 20.9 (CH₃CO), 25.5 & 25.9 (C₉ C₁₀), 27.1 (C₁₁), 28.8 (C₈), 31.2 (C₁₆), 34.7 (C_q tBu), 50.5 (C₁₂), 62.4 (C₆), 66.1 (C₃), 68.0 (C₇), 68.1 (C_C), 68.3 (C₅), 69.1 (C₄), 69.6 (C₂), 70.1 (C_D), 70.6 (C_E), 97.4 (C_I), 112.3 & 121.6 (C_A C_B), 122.5 (C₁₄), 126.1 (C₁₅), 132.9 & 152.8 (C_q arom thread), 147.3 (C_q DB24C8), 169.7 & 170.0 & 170.2 & 170.7 (COCH₃).

HRMS (ESI): [M-PF₆]⁺ calcd for C₅₄H₇₈NO₁₈: 1028.5219, found 1028.5200

6.2 Characterization of compound 5b

R_f 0.30 (15/85 Acetone/CH₂Cl₂)

¹H NMR (400 MHz, CDCl₃, 298 K): δ ppm 0.94-1.03 (m, 4H, H₉ H₁₀), 1.17-1.23 (m, 2H, H₈), 1.25 (s, 9H, H₁₆), 1.40-1.48 (m, 2H, H₁₁), 2.02 & 2.07 & 2.08 (3s, 9H, OAc), 3.17-3.23 (m, 1H, H_{7a}), 3.27-3.33 (m, 4H, H_E), 3.45-3.50 (m, 1H, H_{7b}), 3.56-3.62 (m, 4H, H_{E'}), 3.77-3.88 (m, 9H, H₅ H_D), 3.89-4.00 (m, 2H, H₁₂), 4.03-4.05 (m, 1H, H₂), 4.07-4.14 (m, 5H, H_{6a} H_C), 4.19-4.25 (m, 5H, H_{6b} H_{C'}), 4.79 (d, 1H, J = 1.4 Hz, H₁), 5.22 (dd, 1H, J = 10 Hz, J = 3.2 Hz, H₃), 5.33 (t, 1H, J = 10 Hz, H₄), 6.84-6.92 (m, 8H, H_A H_B), 7.32 (d, 2H, J = 8.6 Hz, H₁₅), 7.52 (d, 2H, J = 8.6 Hz, H₁₄), 8.48-8.61 (br s, 2H, H₁₃).

¹³C NMR (100 MHz, CDCl₃, 298 K): δ ppm 20.7 & 20.7 & 20.9 (CH₃CO), 25.4 & 25.8 (C₉ C₁₀), 27.1 (C₁₁), 28.7 (C₈), 31.1 (C₁₆), 34.7 (C_q tBu), 50.4 (C₁₂), 62.6 (C₆), 66.6 (C₄), 67.5 (C₇), 68.0 (C_C), 68.2 & 68.9 (C₂ C₅), 70.0 (C_D), 70.6 (C_E), 71.5 (C₃), 99.6 (C_I), 112.3 & 121.6 (C_A C_B), 122.5 (C₁₄), 126.0 (C₁₅), 132.8 & 152.8 (C_q arom thread), 147.2 (C_q DB24C8), 169.7 & 170.3 & 170.8 (COCH₃).

HRMS (ESI): [M-PF₆]⁺ calcd for C₅₂H₇₆NO₁₇: 986.5113, found: 986.5109

6.3 Characterization of compound 5c

R_f 0.31 (5/95 MeOH/CH₂Cl₂)

¹H NMR (400 MHz, CDCl₃, 298 K): δ ppm 0.14 (s, 9H, (CH₃)₃Si), 0.94-1.03 (m, 4H, H₉ H₁₀), 1.17-1.23 (m, 2H, H₈), 1.27 (s, 9H, H₁₆), 1.40-1.47 (m, 2H, H₁₁), 2.04 & 2.07 & 2.09 (3s, 9H, OAc), 3.17-3.23 (m, 1H, H_{7a}), 3.28-3.34 (m, 4H, H_E), 3.43-3.51 (m, 1H, H_{7b}), 3.57-3.63 (m, 4H, H_{E'}), 3.80-3.89 (m, 9H, H₅ H_D), 3.89-3.95 (m, 2H, H₁₂), 3.99 (dd, 1H, J = 3 Hz, J = 1.8 Hz, H₂), 4.09-4.15 (m, 5H, H_{6a} H_C), 4.21-4.28 (m, 5H, H_{6b} H_{C'}), 4.59 (d, 1H, J = 1.8 Hz, H₁), 5.17 (dd, 1H, J = 10 Hz, J = 3 Hz, H₃), 5.35 (t, 1H, J = 10 Hz, H₄), 6.85-6.98 (m, 8H, H_A H_B), 7.35 (d, 2H, J = 8.5 Hz, H₁₅), 7.54 (d, 2H, J = 8.5 Hz, H₁₄), 8.50-8.69 (br s, 2H, H₁₃).

¹³C NMR (100 MHz, CDCl₃, 298 K): δ ppm 0.0 ((CH₃)₃Si), 20.8 & 20.8 & 21.0 (CH₃CO), 25.6 & 26.1 (C₉ C₁₀), 27.2 (C₁₁), 29.0 (C₈), 31.2 (C₁₆), 34.7 (C_q tBu), 50.5 (C₁₂), 62.7 (C₆), 66.5 (C₄), 67.8 (C₇), 68.1 (C_C), 68.7 & 69.8 (C₂ C₅), 71.4 (C₃), 70.1 (C_D), 70.6 (C_E), 100.4 (C_I), 112.3 & 121.6 (C_A C_B), 122.5 (C₁₄), 126.1 (C₁₅), 132.9 & 152.8 (C_q arom thread), 147.3 (C_q DB24C8), 169.6 (COCH₃).

HRMS (ESI): [M-PF₆]⁺ calcd for C₅₅H₈₄NO₁₇Si: 1058.5509, found: 1058.5498

6.4 Characterization of compound 6

R_f 0.60 (20/80 Acetone/CH₂Cl₂)

¹H NMR (400 MHz, CDCl₃, 298 K): δ ppm 0.94-1.03 (m, 4H, H₉ H₁₀), 1.17-1.21 (m, 2H, H₈), 1.23 (s, 9H, H₁₆), 1.38-1.47 (m, 2H, H₁₁), 1.70 (s, 3H, CH₃ orthoester), 2.03 & 2.06 & 2.09 (3s, 9H, OAc), 3.26-3.35 (m, 6H, H_{7a} H_{7b} H_E), 3.54-5.60 (m, 4H, H_{E'}), 3.70 (ddd, 1H, J = 9.8 Hz, J = 4.7 Hz, J = 2.7 Hz, H₅), 3.78-3.85 (m, 8H, H_D), 3.91-3.99 (m, 1H, H₁₂), 4.06-4.12 (m, 5H, H_{6b} H_C), 4.16-4.26 (m, 5H, H_{6a} H_{C'}), 4.58 (dd, 1H, J = 4 Hz, J = 2.4 Hz, H₂), 5.14 (dd, 1H, J = 9.8 Hz, J = 4 Hz, H₃), 5.27 (t, 1H, J = 9.8 Hz, H₄), 5.48 (d, 1H, J = 2.4 Hz, H₁), 6.81-6.89 (m, 8H, H_A H_B), 7.29 (d, 2H, J = 8.6 Hz, H₁₅), 7.49 (d, 2H, J = 8.6 Hz, H₁₄), 8.41-8.61 (br, 2H, H₁₃).

¹³C NMR (100 MHz, CDCl₃, 298 K): δ ppm 20.6 & 20.7 & 20.7 (CH₃CO), 24.8 (CH₃ orthoester), 25.5 & 25.9 (C₉ C₁₀), 27.2 (C₁₁), 29.0 (C₈), 31.1 (C₁₆), 34.7 (C_q tBu), 50.4 (C₁₂), 62.0 (C₇), 62.3 (C₆), 65.5 (C₄), 68.0 (C_C), 70.0 (C_D), 70.6 (C_E), 70.7 & 71.0 (C₃ C₅), 76.4 (C₂), 97.3 (C_I), 112.2 & 121.6 (C_A C_B), 122.4 (C₁₄), 124.0 (C_q orthoester), 126.0 (C₁₅), 132.7 & 152.7 (C_q arom thread), 147.2 (C_q DB24C8), 169.4 & 170.2 & 170.6 (COCH₃).

HRMS (ESI): $[M-PF_6]^+$ calcd for $C_{54}H_{78}NO_{18}$: 1028.5219, found: 1028.5193

7. Characterization of threads 3a-b, 4

7.1 Note about threads.

Although 0.2 equiv of triethylamine (necessary amount to quench the 0.2 equiv of TMSOTf) was added at the end of reaction, we noticed that every thread was totally deprotonated after purification by chromatography. We could suppose that deprotonation occurred during the purification. Equilibrium between anilinium and aniline thread has already been described by S.J. Loeb.²

7.2 Characterization of deprotonated compound 3a

R_f 0.72 (50/50 AcOEt/Petroleum ether).

¹H NMR (400 MHz, CDCl₃, 298 K): δ ppm 1.28 (s, 9H, H₁₆), 1.39-1.47 (m, 4H, H₉ H₁₀), 1.60-1.67 (m, 4H, H₈ H₁₁), 2.04 & 2.05 & 2.11 & 2.17 (4s, 12H, OAc), 3.11 (t, 2H, J = 7.1 Hz, H₁₂), 3.43-3.49 (m, 1H, H_{7a}), 3.66-3.72 (m, 1H, H_{7b}), 3.99 (ddd, 1H, J = 9.9 Hz, J = 5.3 Hz, J = 2.4 Hz, H₅), 4.11 (dd, 1H, J = 12.2 Hz, J = 2.4 Hz, H_{6a}), 4.29 (dd, 1H, J = 12.2 Hz, J = 5.3 Hz, H_{6b}), 4.81 (d, 1H, J = 1.4 Hz, H₁), 5.24 (dd, 1H, J = 3.3 Hz, J = 1.4 Hz, H₂), 5.29 (t, 1H, J = 9.9 Hz, H₄), 5.36 (dd, 1H, J = 9.9 Hz, J = 3.3 Hz, H₃), 6.58 (d, 2H, J = 8.6 Hz, H₁₄), 7.21 (d, J = 8.6 Hz, H₁₅).

¹³C NMR (100 MHz, CDCl₃, 298 K): δ ppm 20.7 & 20.9 ($\underline{CH_3CO}$), 26.0 & 26.9 (C₉ C₁₀), 29.2 & 29.5 (C₈ C₁₁), 31.5 (C₁₆), 33.7 (C_q tBu), 44.0 (C₁₂), 62.5 (C₆), 66.2 (C₄), 68.3 (C₅), 68.4 (C₇), 69.0 (C₃), 69.7 (C₂), 97.5 (C₁), 112.3 (C₁₄), 125.9 (C₁₅), 139.8 & 146.1 (C_q arom), 169.7 & 169.9 & 170.1 & 170.6 ($\underline{COCH_3}$).

HRMS (ESI): $[M+H]^+$ calcd for $C_{30}H_{46}NO_{10}$: 580.3122, found: 580.3138

7.3 Characterization of deprotonated compound 3b

R_f 0.65 (20/80 Acetone/CH₂Cl₂).

¹H NMR (400 MHz, CDCl₃, 298 K): δ ppm 1.28 (s, 9H, H₁₆), 1.39-1.47 (m, 4H, H₉ H₁₀), 1.60-1.67 (m, 4H, H₈ H₁₁), 2.04 & 2.10 (2s, 9H, OAc), 3.11 (t, 2H, J = 7.1 Hz, H₁₂), 3.43-3.50 (m, 1H, H_{7a}), 3.67-3.74 (m, 1H, H_{7b}), 3.97 (ddd, 1H, J = 9.5 Hz, J = 4.8 Hz, J = 2.4 Hz, H₅), 4.04 (dd, 1H, J = 2.8 Hz, J = 1.7 Hz, H₂), 4.11 (dd, 1H, J = 12.2 Hz, J = 2.4 Hz, H_{6a}), 4.29 (dd, 1H, J = 12.2 Hz, J = 4.8 Hz, H_{6b}), 4.87 (d, 1H, J = 1.7 Hz, H₁), 5.27-5.36 (m, 2H, H₃ H₄), 6.58 (d, 2H, J = 8.6 Hz, H₁₄), 7.21 (d, J = 8.6 Hz, H₁₅).

¹³C NMR (100 MHz, CDCl₃, 298 K): δ ppm 20.7 & 20.8 & 20.9 ($\underline{CH_3CO}$), 26.0 & 26.9 (C₉ C₁₀), 29.3 & 29.6 (C₈ C₁₁), 31.5 (C₁₆), 33.7 (C_q tBu), 44.2 (C₁₂), 62.5 (C₆), 66.3 (C₃), 68.2 (C₅), 68.3 (C₇), 69.5 (C₂), 71.7 (C₄), 99.4 (C₁), 112.5 (C₁₄), 126.0 (C₁₅), 139.9 & 146.2 (C_q arom), 169.9 & 170.8 ($\underline{COCH_3}$).

HRMS (ESI): $[M+H]^+$ calcd for $C_{28}H_{44}NO_9$: 538.3016, found: 538.3008

7.4 Characterization of deprotonated compound **4**

R_f 0.70 (50/50 AcOEt/Petroleum ether).

¹H NMR (400 MHz, CDCl₃, 298 K): δ ppm 1.27 (s, 9H, H₁₆), 1.36-1.42 (m, 4H, H₉ H₁₀), 1.52-1.65 (m, 4H, H₈ H₁₁), 1.76 (s, 3H, CH₃ orthoester), 2.05 & 2.08 & 2.12 (3s, 9H, OAc), 3.11 (t, 2H, J = 7.1 Hz, H₁₂), 3.45-3.53 (m, 2H, H₇), 3.67 (ddd, 1H, J = 9.7 Hz, J = 4.9 Hz, J = 2.6 Hz, H₅), 4.14 (dd, 1H, J = 12.2 Hz, J = 2.6 Hz, H_{6a}), 4.24 (dd, 1H, J = 12.2 Hz, J = 4.9 Hz, H_{6b}), 4.60 (dd, 1H, J = 3.8 Hz, J = 2.6 Hz, H₂), 5.14 (dd, 1H, J = 9.7 Hz, J = 3.8 Hz, H₃), 5.29 (t, 1H, J = 9.7 Hz, H₄), 5.45 (d, 1H, J = 2.6 Hz, H₁), 6.58 (d, 2H, J = 8.6 Hz, H₁₄), 7.21 (d, J = 8.6 Hz, H₁₅).

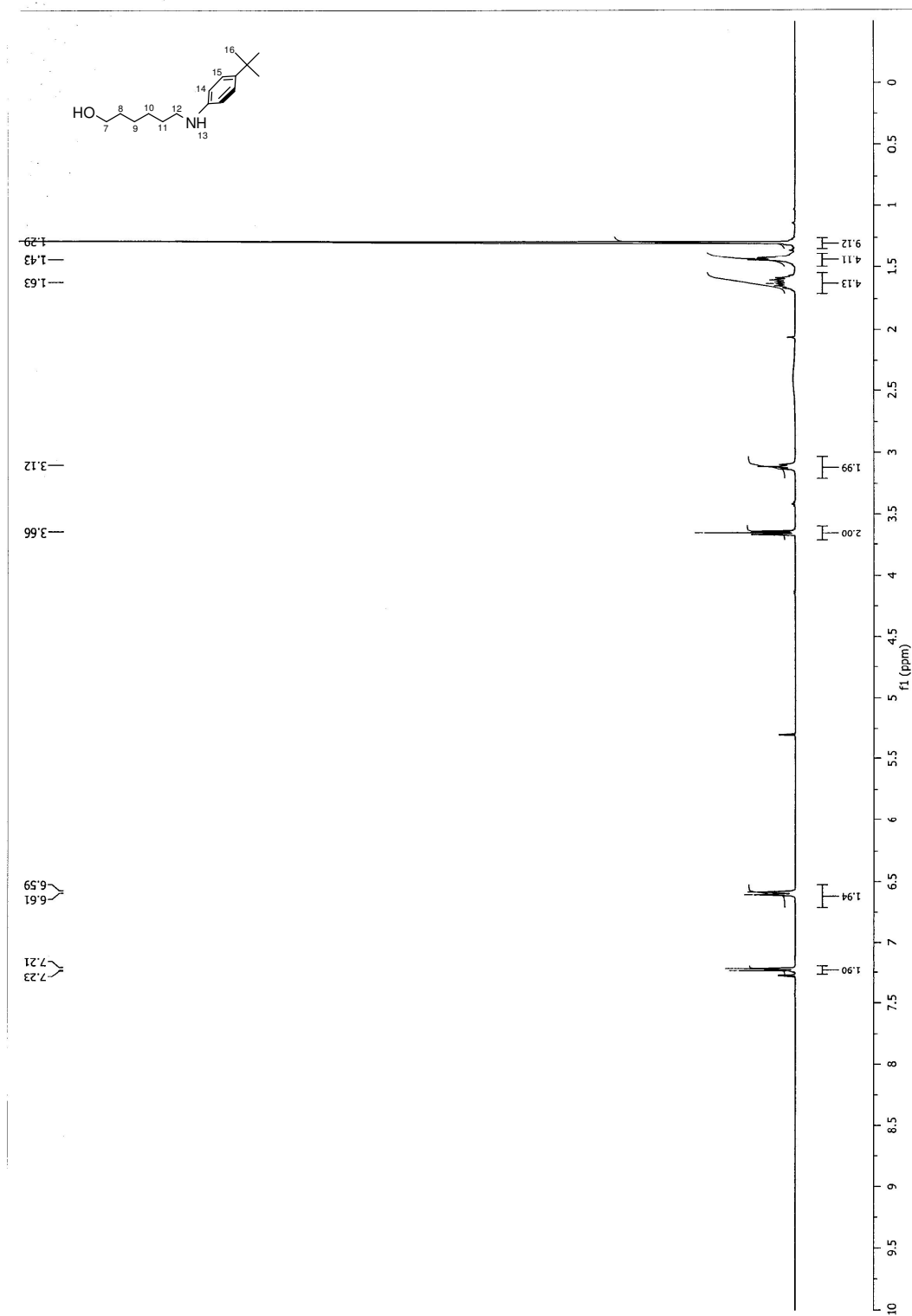
¹³C NMR (100 MHz, CDCl₃, 298 K): δ ppm 20.7 & 20.7 & 20.8 (CH₃CO), 24.6 (CH₃ orthoester), 25.8 & 26.8 (C₉ C₁₀), 29.3 & 29.5 (C₈ C₁₁), 31.5 (C₁₆), 33.7 (C_q tBu), 44.0 (C₁₂), 62.2 (C₆), 62.4 (C₇), 65.5 (C₄), 70.6 (C₃), 71.3 (C₅), 76.3 (C₂), 97.3 (C₁), 112.4 (C₁₄), 124.1 (C_q orthoester), 125.9 (C₁₅), 139.8 & 146.1 (C_q arom), 169.4 & 170.4 & 170.6 (COCH₃).

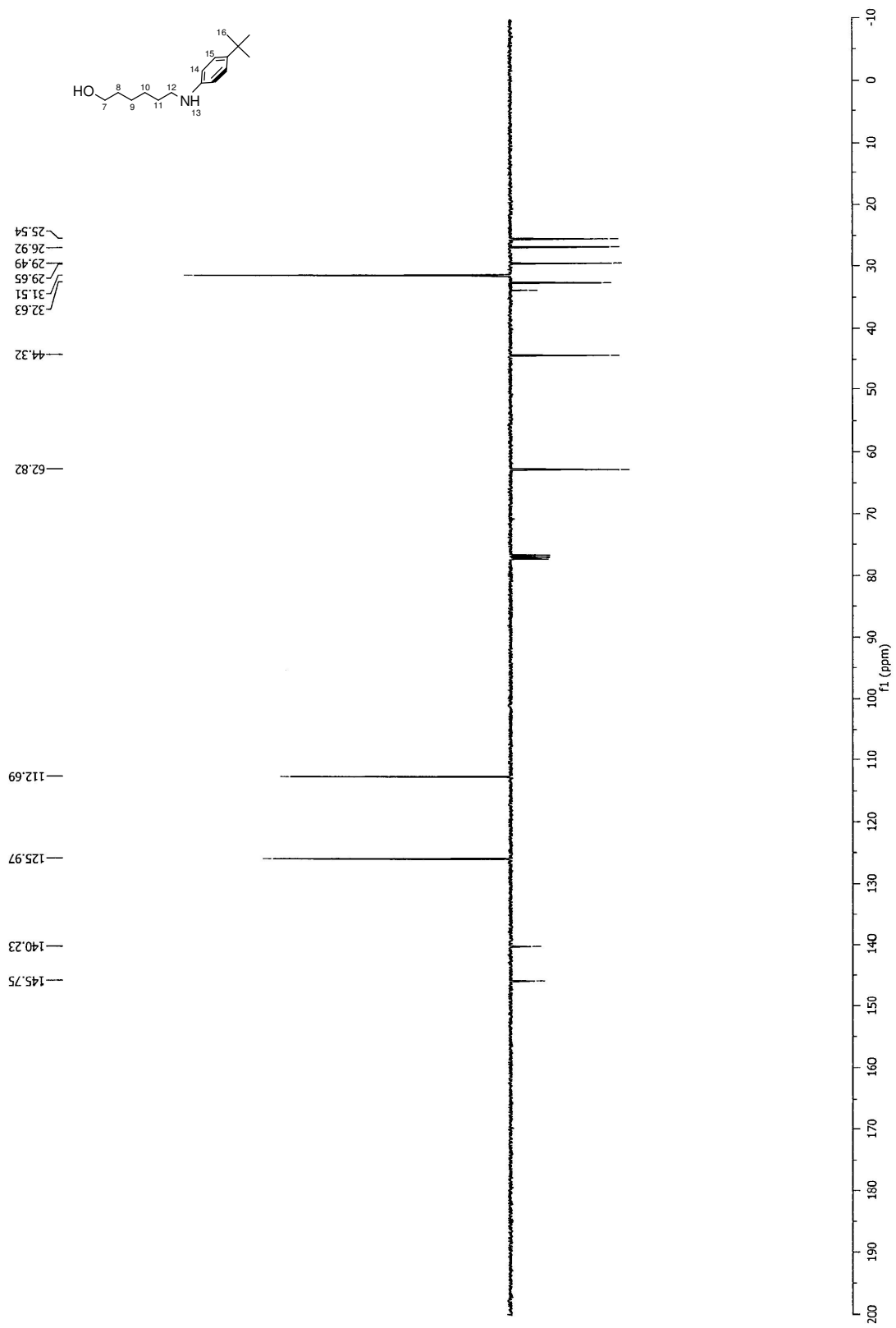
HRMS (ESI): [M+H]⁺ calcd for C₃₀H₄₆NO₁₀: 580.3122, found: 580.3130

8. NMR spectra

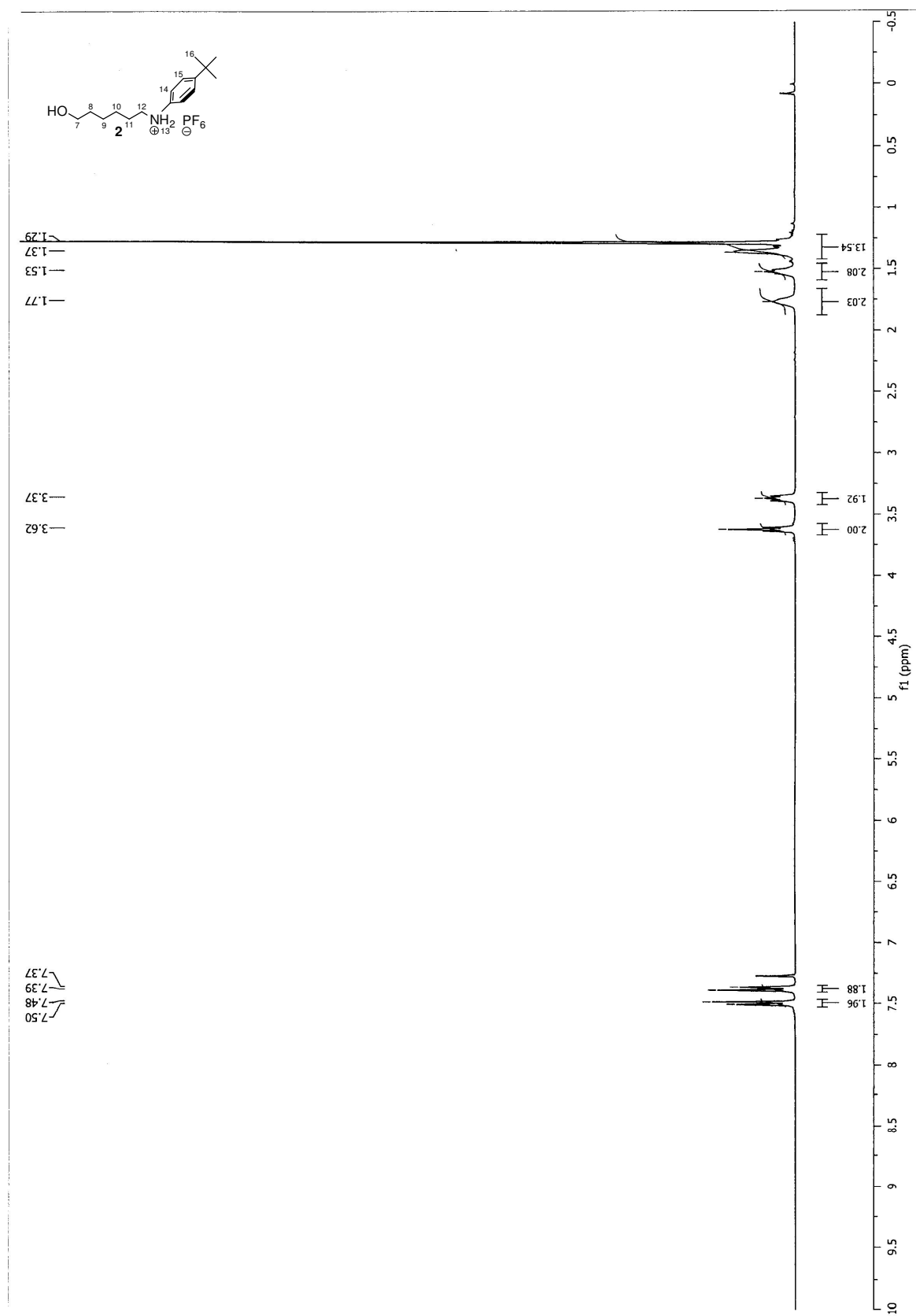
8.1 NMR spectra of compound 2

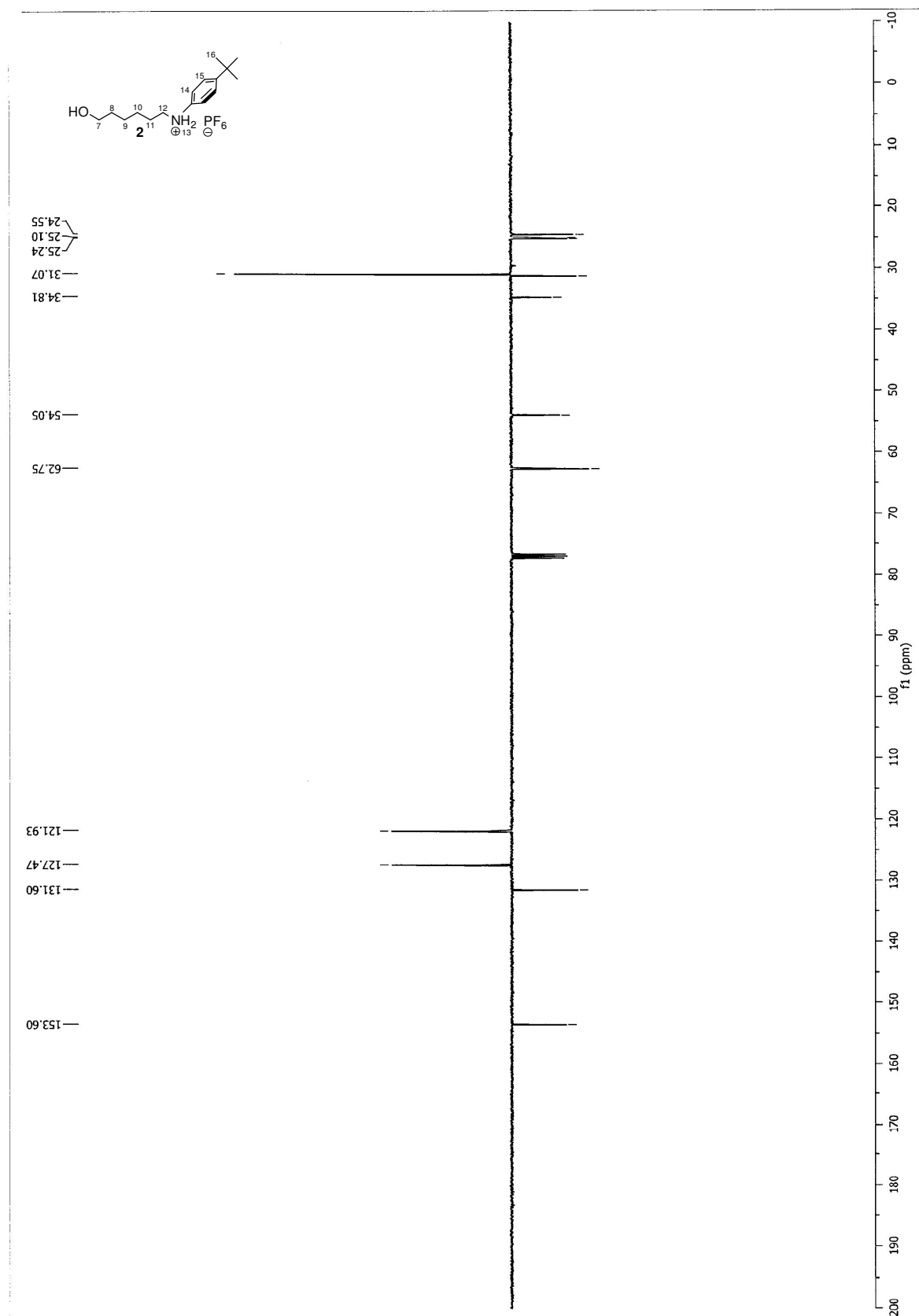
a. NMR spectra of the intermediary deprotonated aminoalcohol (first step)



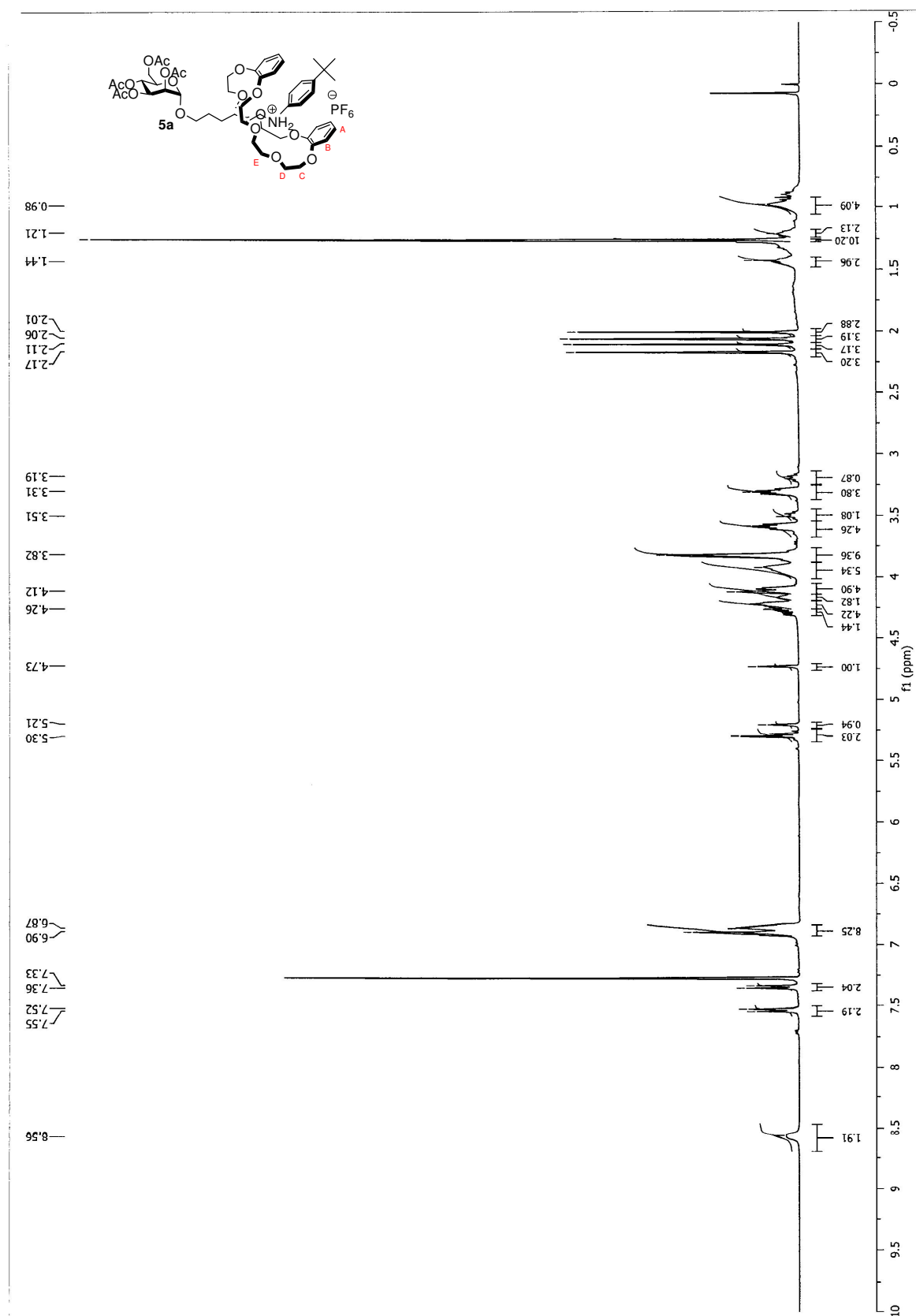


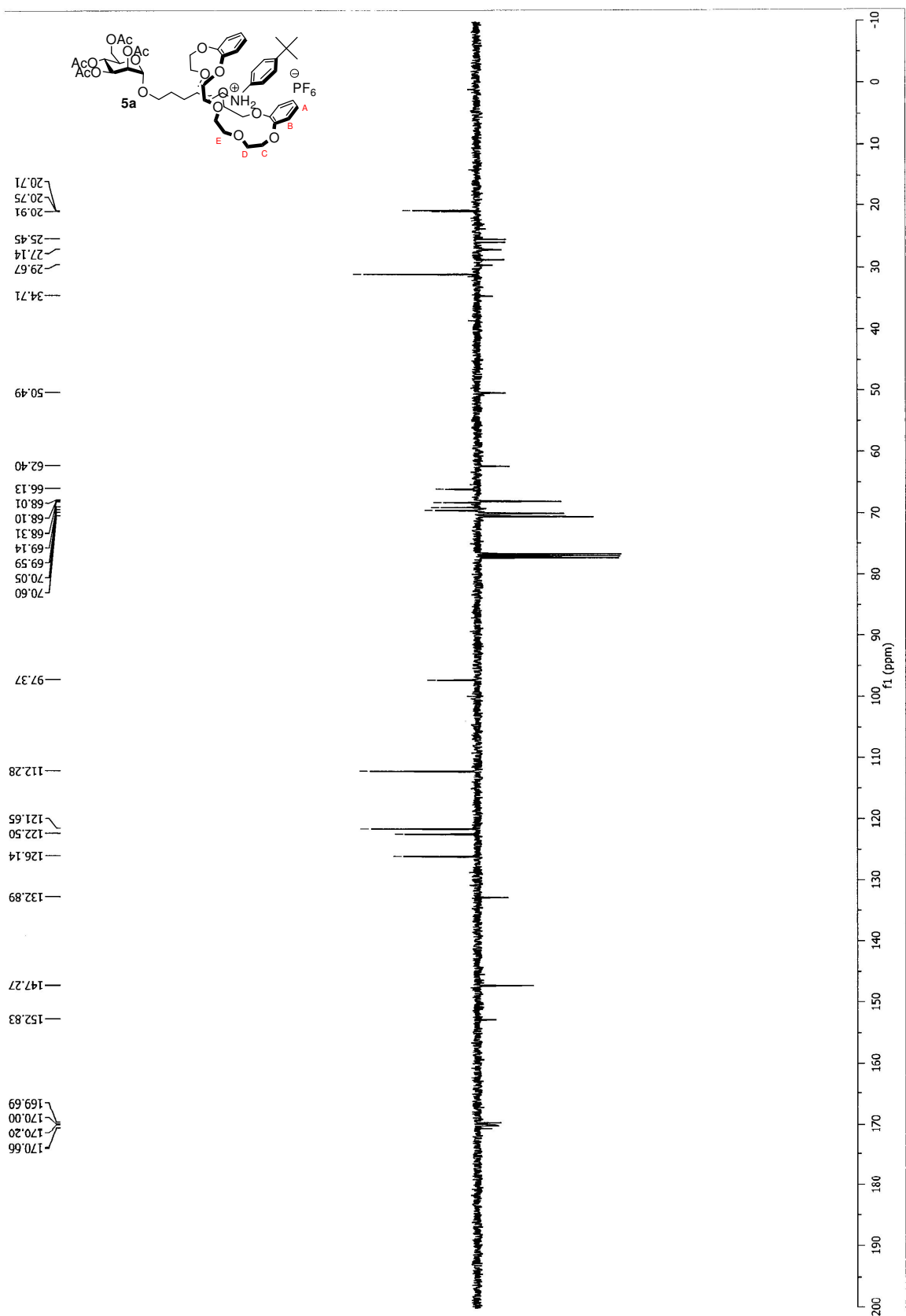
b. NMR spectra of compound 2 (second step)



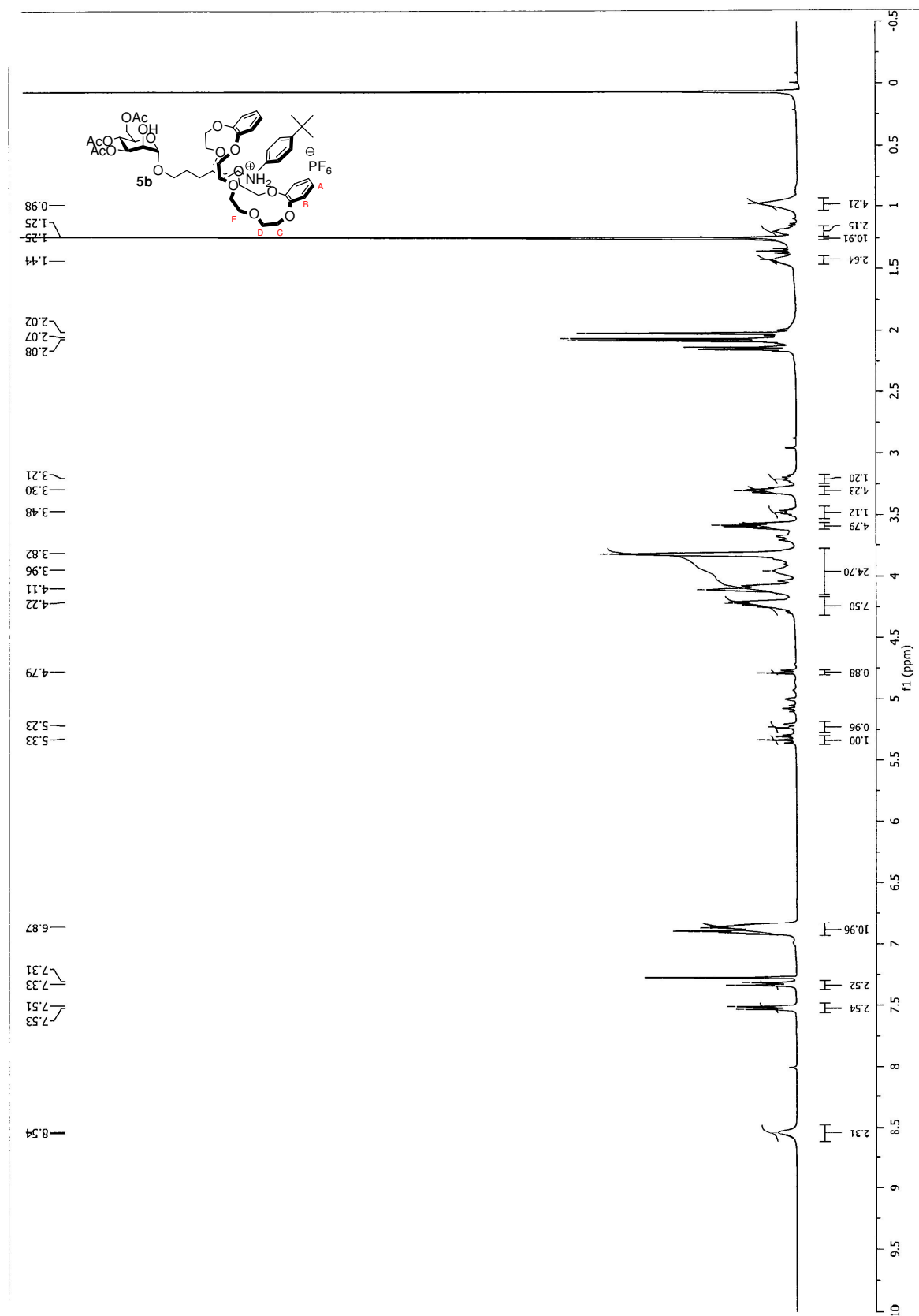


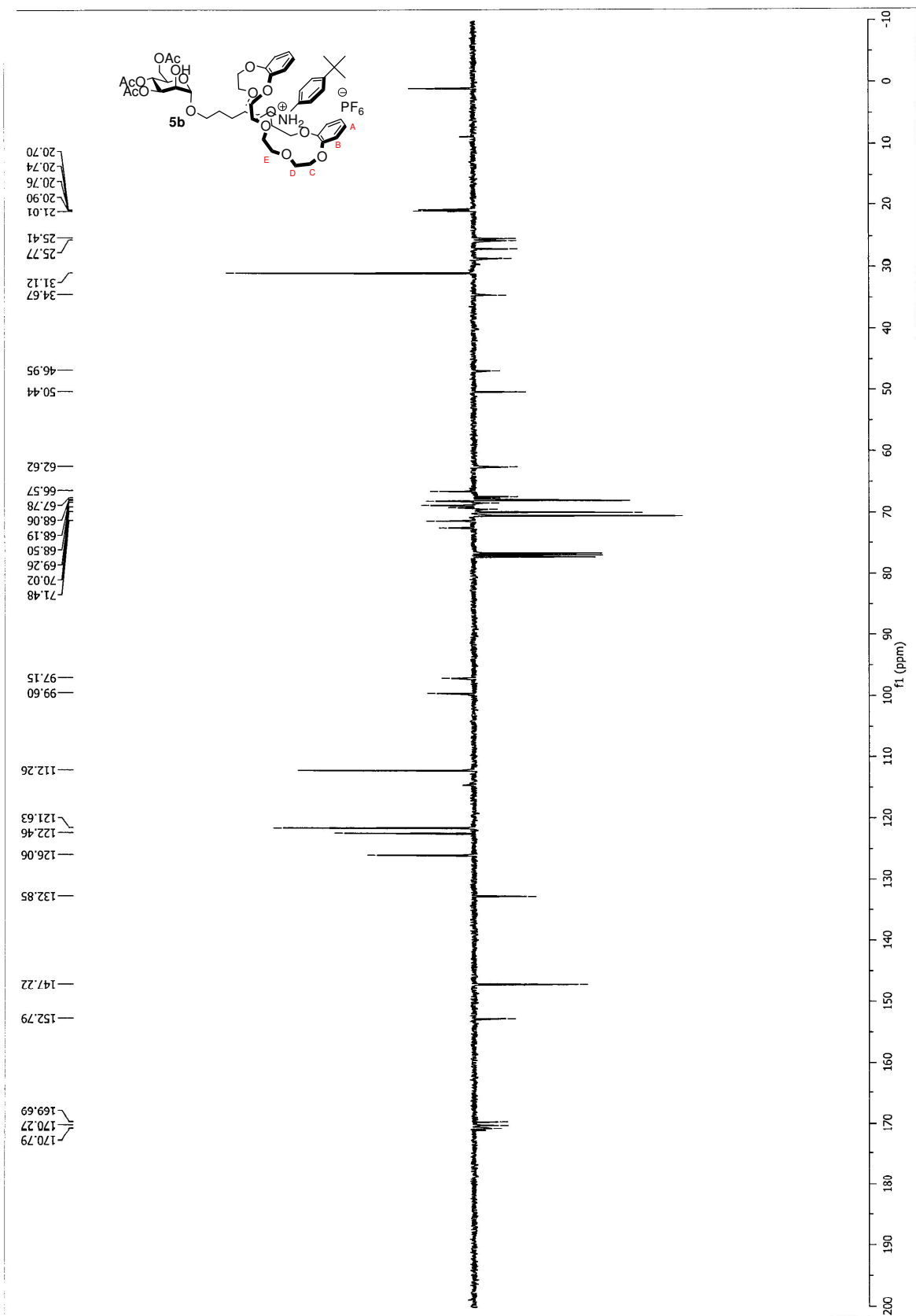
8.2 NMR spectra of compound **5a**



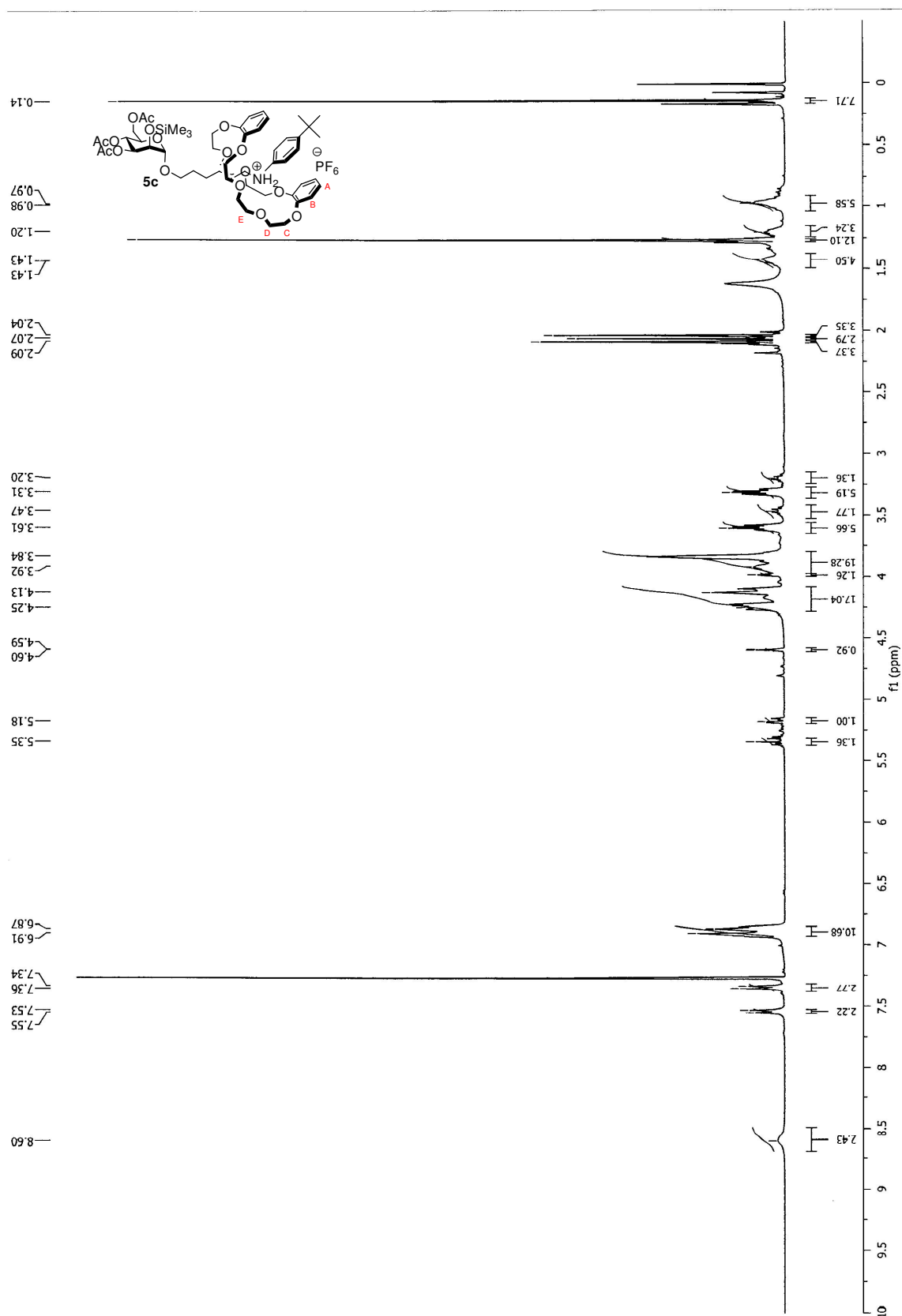


8.3 NMR spectra of compound **5b**



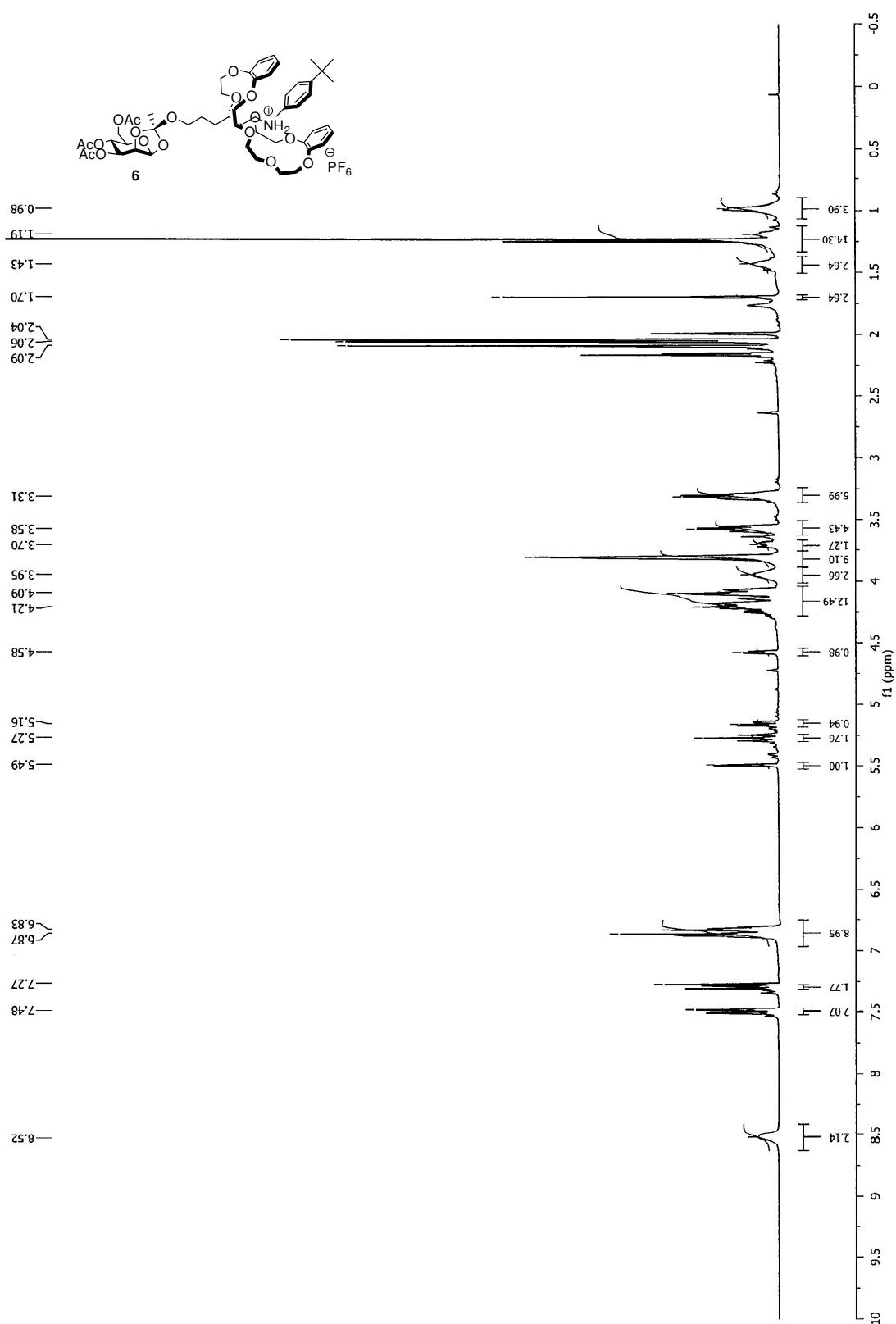


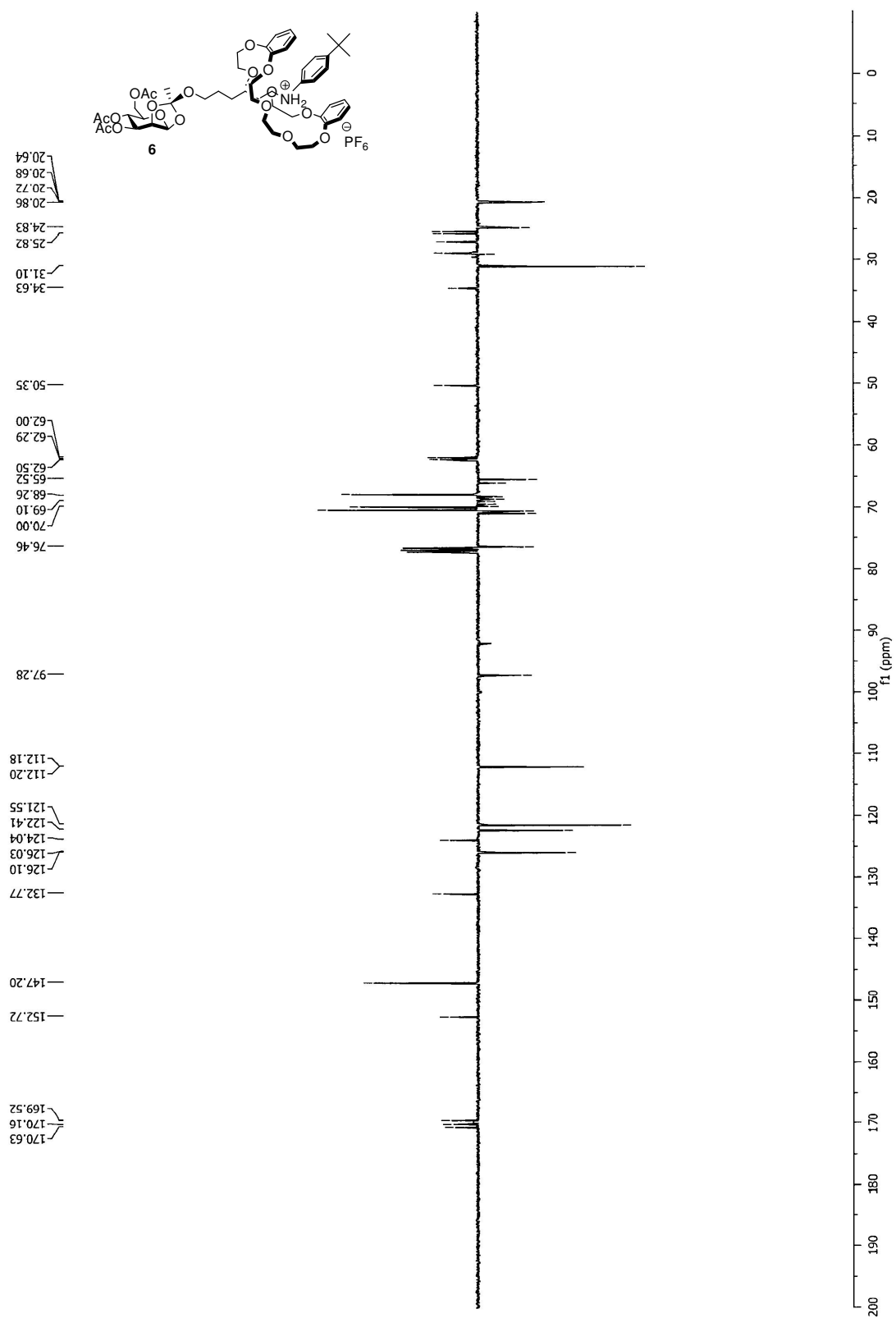
8.4 NMR spectra of compound 5c



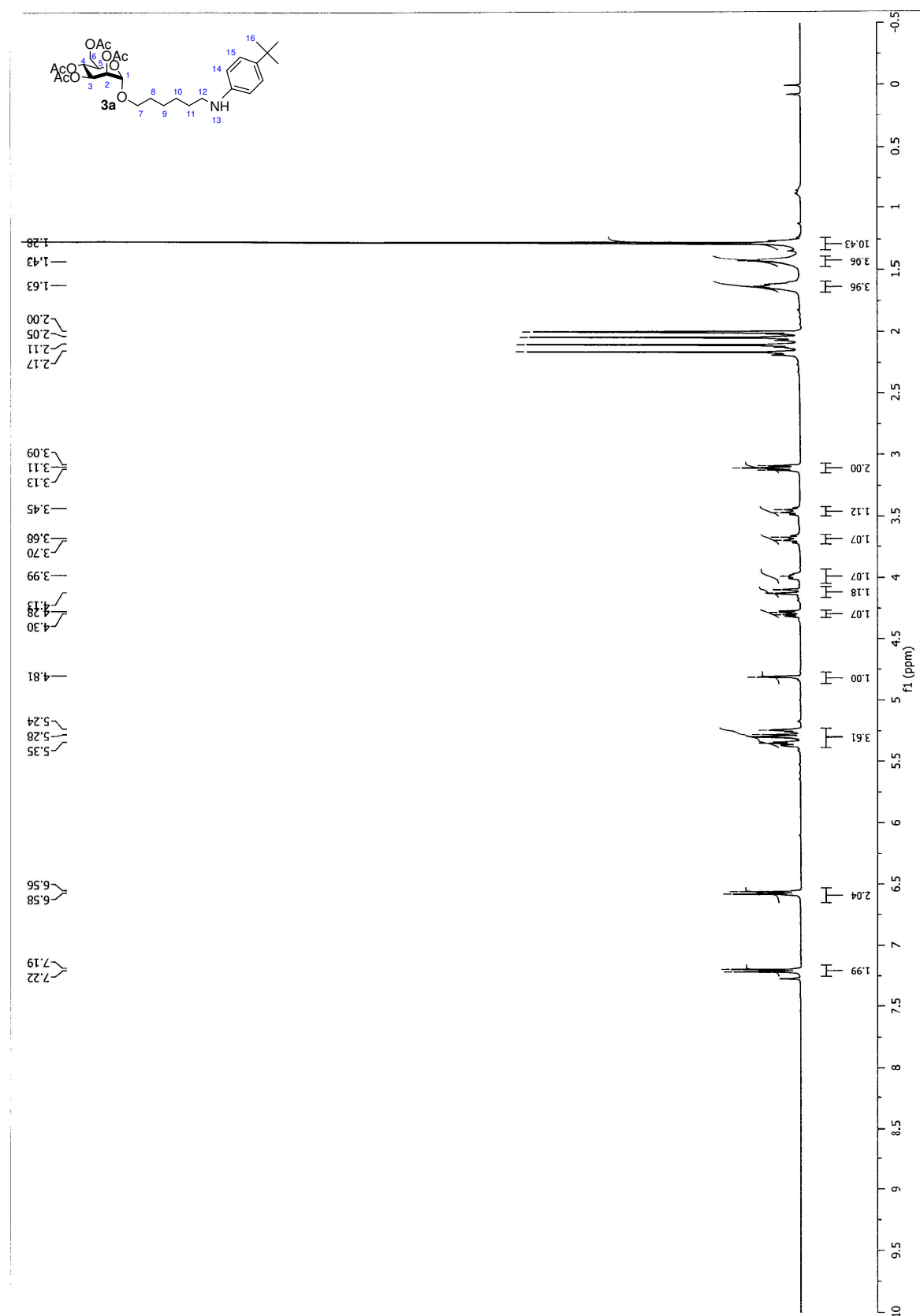


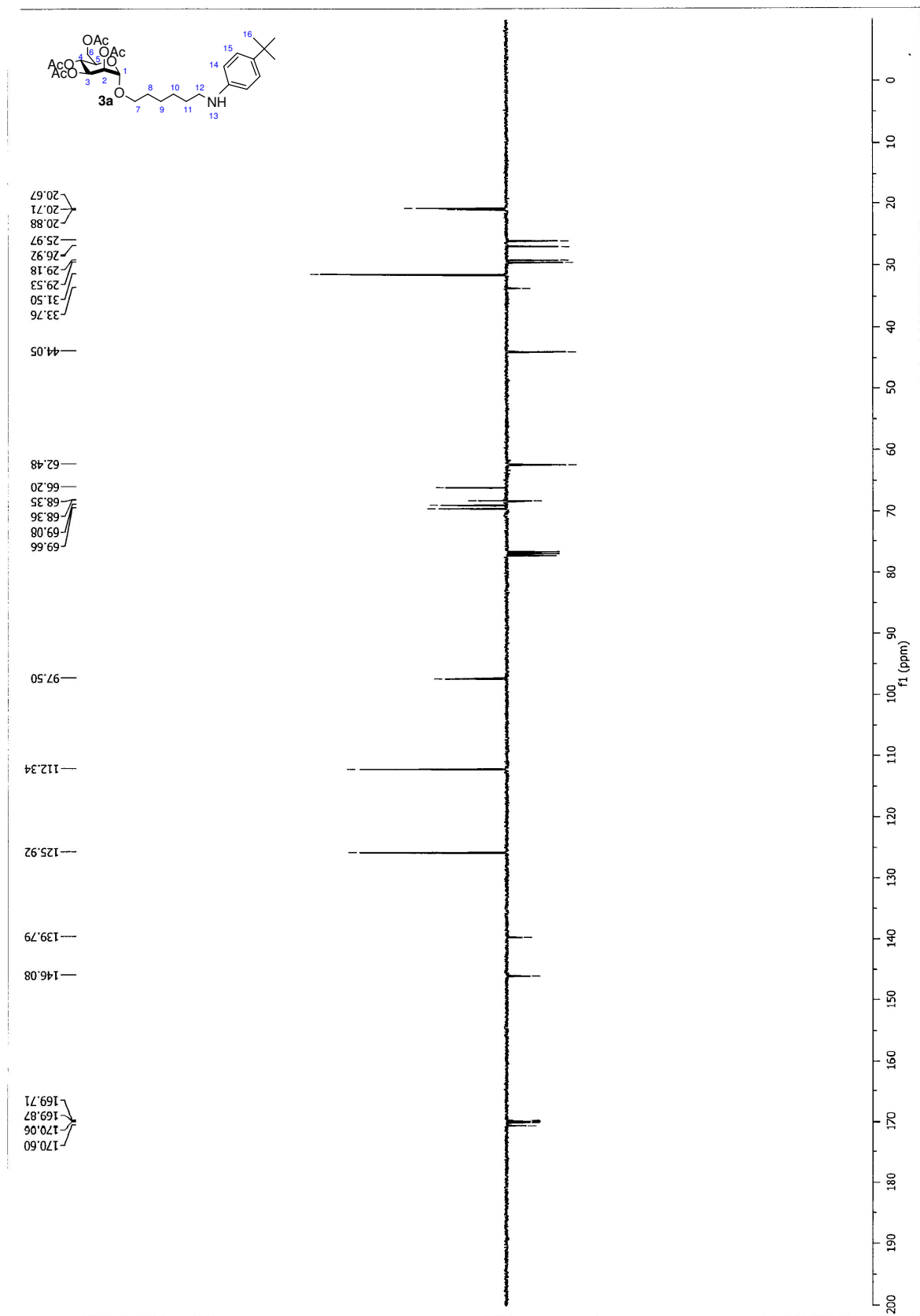
8.5 NMR spectra of compound **6**



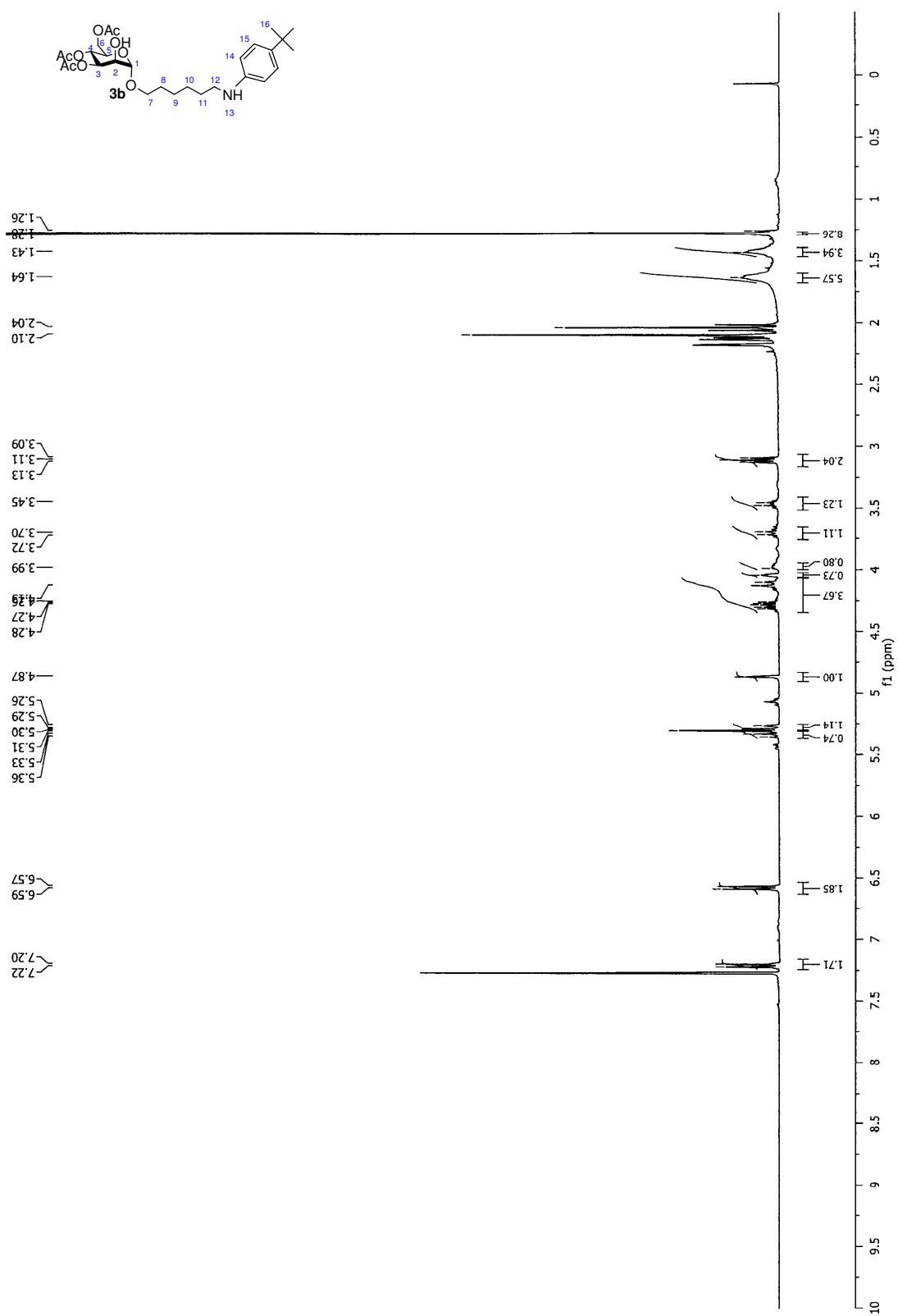


8.6 NMR spectra of deprotonated compound **3a**

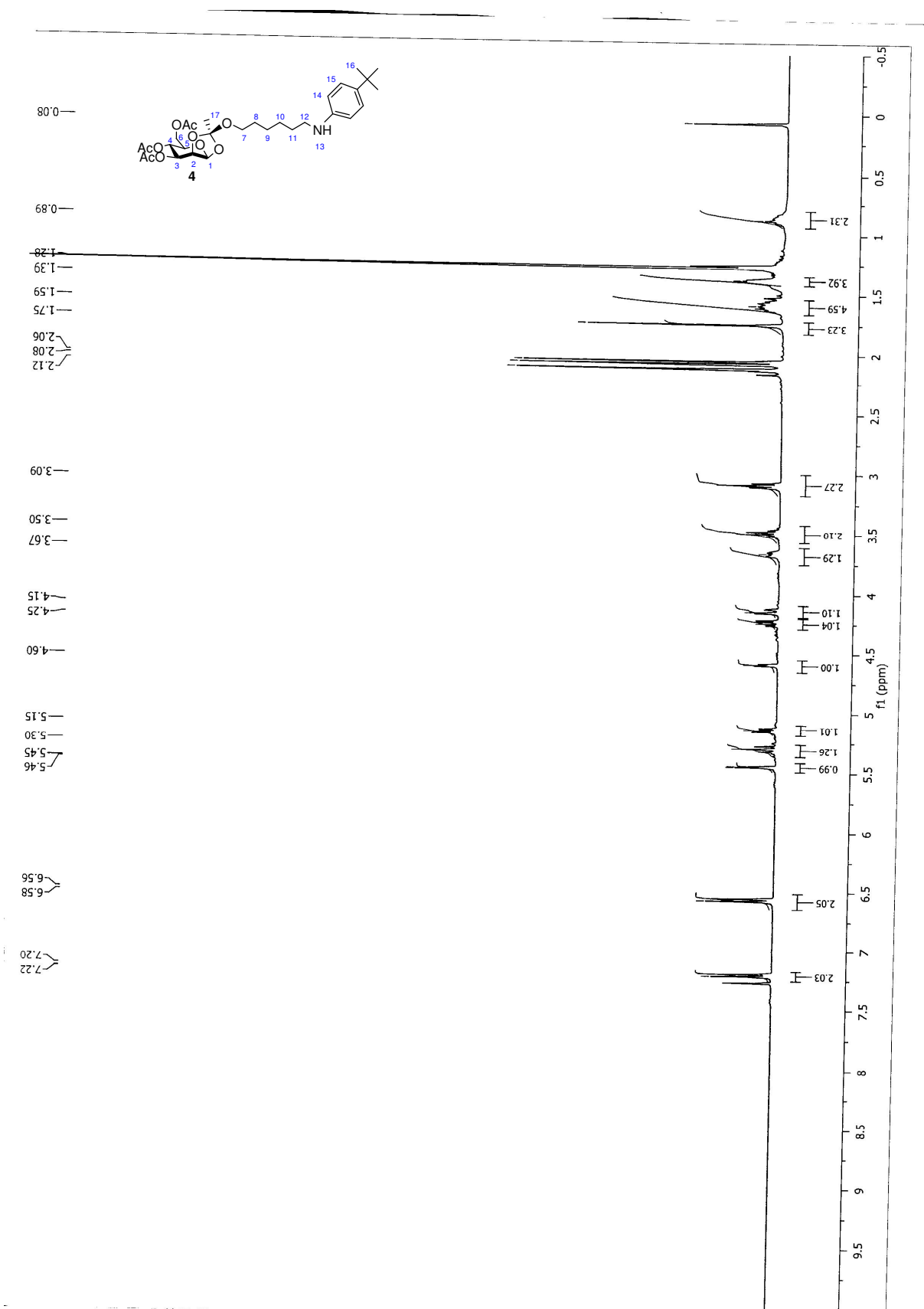


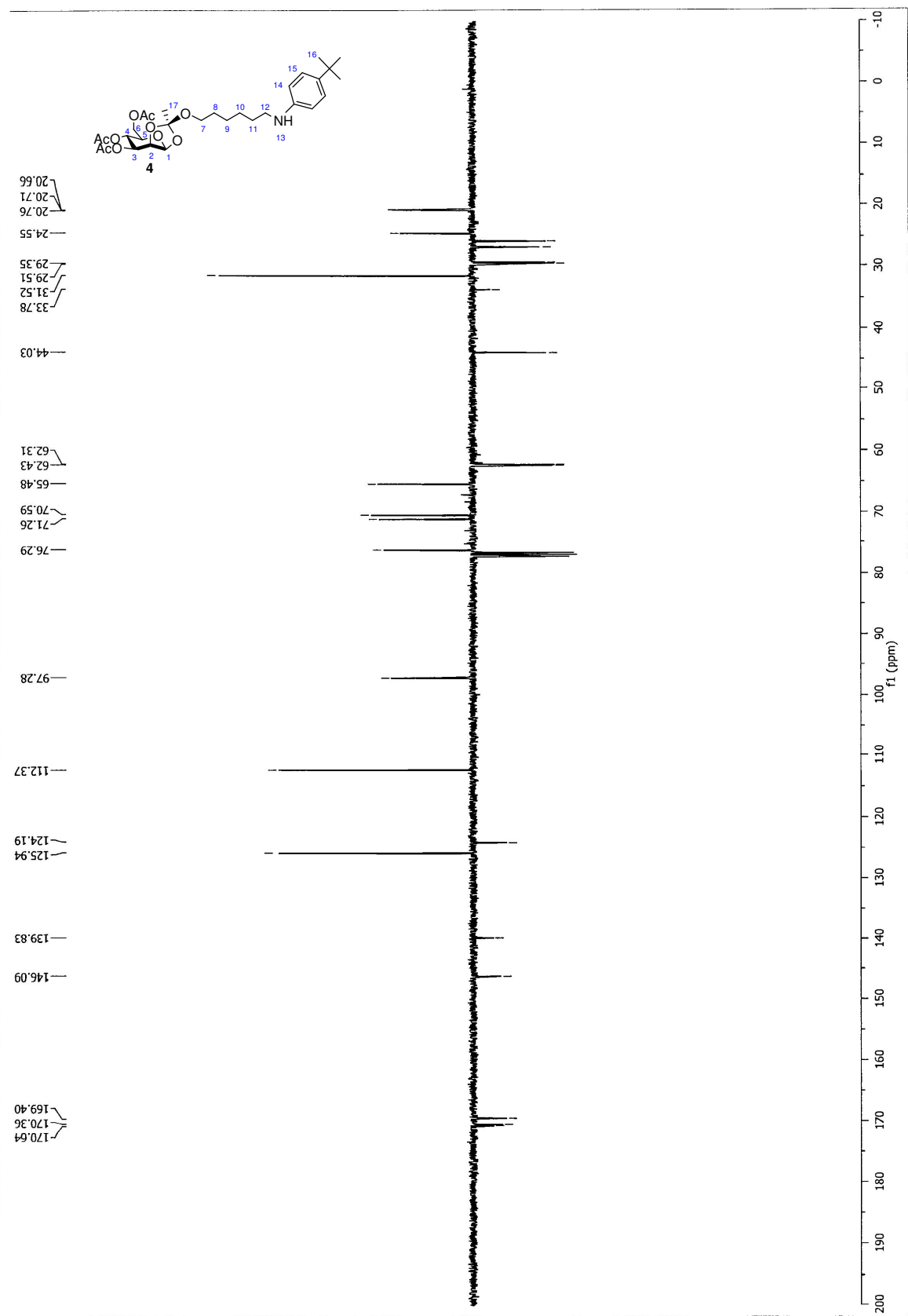


8.7 NMR spectra of deprotonated compound **3b**



8.8 NMR spectra of deprotonated orthoester compound **4**





9. References

- (1) Upreti, M.; Ruhela, D.; Vishwakarma, R. A. *Tetrahedron* **2000**, 56 (35), 6577-6584.
- (2) Loeb, S. J.; Tiburcio, J.; Vella, S. J. *Org. Lett.* **2005**, 7 (22), 4923-4926.