SUPPORTING INFORMATION

FOR

Total Syntheses of Yingzhaosu A and of its C(14)-Epimer Including the First Evaluation of their Antimalarial and Cytotoxic Activities

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General

Compounds 8 and 15 (as an inseparable mixture) were synthesized and purified as described earlier.¹

¹H, ¹³C and ³¹P NMR as well as DEPT were recorded at 400MHz or 250 MHz NMR. COSY, NOEdifference and HMQC NMR spectra were recorded at 100MHz. Residual CHCl₃ (7.27 ppm for ¹H NMR) and CDCl₃ (77.00 ppm for ¹³C NMR) peaks were used as internal standards. Infrared spectra were measured on a FT-IR instrument as thin films or in KBr pellets. Specific rotations, HRMS and X-ray analysis were performed using commercial instruments and software packages. HRMS were performed using the Desorption Chemical Ionization (DCI) Technique. Melting points are uncorrected. TLC analysis was performed on precoated silica gel plates (UV detection) followed by development with either alkaline KMnO4 or a solution of N,N-dimethyl-*p*-phenylenediamine dihydrochloride in water/methanol/acetic acid (stain specific for peroxides), stabilized by 2–3 mL of conc. HCl per 150 mL of the stain solution.\

Flash chromatography was performed on Silica gel 60 (230-400 mesh). Medium pressure liquid chromatography (MPLC) was performed on glass columns (l = 460 mm, d = 26 or 43 mm, filled with silica gel (15–25 µm particle size). For analytical and semi-preparative HPLC a commercial HPLC instrument with an UV detector and direct-phase columns was used. Retention times ($R\tau$) are given for analytical HPLC. Semi-preparative direct-phase HPLC was performed on a 250 x 10mm column (Si-60 (10 µm); flow rate: 2.5 - 3 mL/min). Analytical direct-phase HPLC was performed on a, 250 x 4 mm column (Si-60 (5 µm); flow rate: 1 mL/min).

All reagents were used as received from the supplier with the following exceptions: Titanium tetrachloride, trimethylsilyl chloride and trimethyl boroxine were distilled immediately prior to use. Pyridine and 2,6-lutidine was distilled over KOH pellets. The purity of MCPBA was determined by iodometric titration (purity noted). Commercial CH_2Cl_2 was distilled over P_2O_5 . THF was distilled from

sodium/benzophenone. All reactions specified as dry were carried out in oven dried glassware under an atmosphere of dry argon.

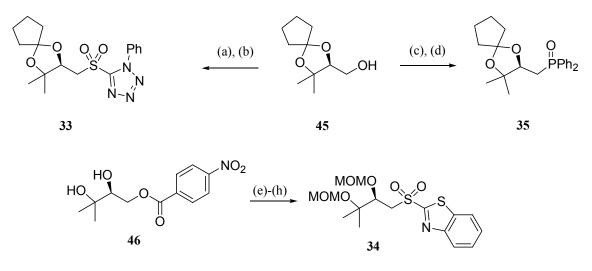
Unless otherwise stated the purity of all the title compounds was estimated to be \ge 95% by ¹H NMR and analytical HPLC determination. The ratios of diastereomers were determined by integration of the relevant separated signals in ¹H NMR spectra.

The purity of all the *in vitro* tested compounds was judged to be $\ge 95\%$ by 400 MHz ¹H NMR. The purity of the *in vivo* tested compounds was $\ge 97\%$ according to the same criteria.

Synthesis of Julia-Kociensky sulfones 33-34 and Horner-Wittig reagent 35

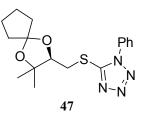
Compounds **33** and **35** were synthesized from the readily available³ alcohol **45** (see Scheme I). Compound **34** was synthesized in four steps from the readily available³ alcohol **46** (Scheme I).

Scheme S1



(a) 1.1 equiv. Ph₃P, 1.1 equiv. DEAD, 1.3 equiv. 1-phenyl-1*H*-tetrazole-5-thiol, THF, 98% yield. (b) cat. $(NH_4)_6Mo_7O_{24}$, 4 equiv. H_2O_2 , $H_2O/EtOH$, 95% yield. (c) 2 equiv. Ph₃P, 4.5 equiv. imidazole, 1.9 equiv. I₂, 1,4-dioxane, 65-70°C, 94% yield. (d) 1.1 equiv. Ph₂PH, 1.1 equiv. *n*-BuLi, THF 0 °C. then exposed to atmospheric air, 49% yield. (e) 4 equiv. MOMBr, 3 equiv. 2,6-di-*tert*-butyl-4-methyl-pyridine, 94% yield. (f) 2 equiv. K₂CO₃, H₂O/EtOH, 87%. (g) 1.1 equiv. Ph₃P, 1.1 equiv. DEAD, 1.3 equiv. benzothiazole-2-thiol, THF, 90% yield. (h) cat. (NH₄)₆Mo₇O₂₄, 1 equiv. NaHCO₃, 4 equiv. H₂O₂, H₂O/EtOH, 86% yield.

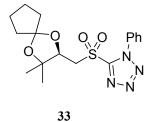
5-[(2S)-3,3-Dimethyl-1,4-dioxaspiro[4.4]non-2-ylsulfanyl]-1-phenyl-1H-tetraazole (47):



To a solution of alcohol **45** (233 mg; 1.25 mmol) in dry THF (11 mL) were added triphenyl phosphine (1.3 equiv.; 426 mg; 1.62 mmol), 1-phenyl-1*H*-tetrazole-5-thiol (1.3 equiv.; 289 mg; 1.62 mmol) and DEAD (1.3 equiv.; 0.26 mL; 1.62 mmol). The reaction mixture was stirred for 45 min., poured into 35 mL saturated sodium bicarbonate and extracted with dichloromethane (4×30 mL). The combined organic extracts were

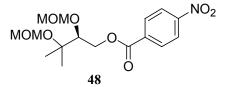
washed with brine, dried (Na₂SO₄) and the solvent removed under reduced pressure. Flash chromatography (15% ethyl acetate/hexane) afforded 426 mg (1.22 mmol, 98% yield) of the desired sulfide **47** as a colourless oil; R_f = 0.21 (15% ethyl acetate/hexane).

5-[(2S)-3,3-Dimethyl-1,4-dioxaspiro[4.4]non-2-ylsulfonyl]-1-phenyl-1H-tetraazole (33):



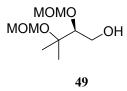
Sulfide **47** (106 mg; 0.31 mmol) was dissolved in ethanol (2.2 mL) and a catalytic amount (0.10 equiv.; 37 mg; 0.03 mmol) ammonium molybdate tetrahydrate was added. The solution was cooled to 0°C and a 30% hydrogenperoxide solution (1.6 equiv.; 0.55 mL) was added and the resulting reaction mixture was stirred for 22 h. at room temperature. The reaction mixture was poured into brine (30 mL) and extracted with 4×20 mL dichloromethane. The combined extracts were dried (Na₂SO₄) and the solvent removed under reduced pressure. Flash chromatography (17% ethyl acetate/hexane) afforded **33** (111 mg; 95% yield) as a white solid. R_f = 0.45 (30% ethyl acetate/hexane); ¹H NMR (400MHz, CDCl₃): δ 1.09 (s, 3H), 1.32 (s, 3H), 1.46-1.80 (comp. m, 8H,), 3.63 (dd, *J*= 2.6, 14.8 Hz, 1H), 3.69 (dd, *J*= 9.8, 14.8 Hz, 1H), 4.13 (dd, *J*= 2.6, 9.8 Hz, 1H), 7.63 (m, 5H, Ar); ¹³C NMR (100 MHz, CDCl₃): δ 22.3 (CH₃), 23.0 (CH₂), 23.1 (CH₂), 25.0 (CH₃), 37.6 (CH₂), 37.8 (CH₂), 56.4 (CH₂), 79.7 (CH-O-), 118.4 (C), 125.5 (CH, Ar), 127.3 (CH, Ar), 129.2 (CH, Ar), 130.7 (CH, Ar), 132.8 (CH, Ar), 135.4 (C, Ar), 153.5 (C=N, tetrazole).

(2S)-2,3-Di(methoxymethoxy)-3-methylbutyl 4-nitrobenzoate (48):



Diol **46** (578 mg; 2.14 mmol) and 4-methyl-2,6-di-*t*-butyl-pyridine (3 equiv.; 1.37 g; 6.44 mmol) was dissolved in freshly dried chloroform (22 mL). Methoxymethylbromide (4 equiv.; 0.7 mL; 8.6 mmol) was added to the solution drop wise over 5 min. The resulting reaction mixture was stirred for 4 h. Poured into water (50 mL) at 4°C and extracted with 4×20 mL dichloromethane. The combined extracts were washed with saturated sodium bicarbonate (45 mL) and dried (Na₂SO₄). The solvent was removed under reduced pressure. Flash chromatography (gradient: 15% to 35% ethyl acetate/hexane) afforded the title compound **48** (728 mg; 94 % yield) as a brownish oil: R_f = 0.66 (50% ethyl acetate/hexane); ¹H NMR (250 MHz, CDCl₃): δ 1.33 (s, 3H), 1.38 (s, 3H), 3.35 (s, 3H), 3.38 (s, 3H), 3.80 (dd, *J* = 2.5, 7.3 Hz, 1H), 4.44 (dd, *J*= 7.3, 11.8 Hz, 1H), 4.73 (d, *J*= 6.7 Hz, 1H), 4.75 (d, *J*= 7.3 Hz, 1H), 4.78 (dd, *J* = 2.5, 11.8 Hz, 1H), 4.82 (d, *J*= 7.3 Hz, 1H), 4.84 (d, *J*= 6.7 Hz, 1H), 8.28 (m, 4H); ¹³C NMR (62.5 MHz, CDCl₃): δ 22.1 (CH₃), 24.0 (CH₃), 55.4 (CH₃), 56.1 (CH₃), 66.5 (CH₂), 82.0 (CH), 91.2 (CH₂), 97.7 (CH₂), 123.6 (2×CH, Ar), 130.7 (2×CH, Ar), 164.7 (C=O, ester).

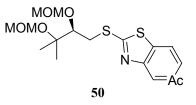
(2S)-2,3-Di(methoxymethoxy)-3-methylbutan-1-ol (49):



Nitrobenzoate **48** (716 mg; 2 mmol) was dissolved in ethanol (16.5 mL) and potassium carbonate (2 equiv.; 550 mg; 4 mmol) was added. The solution was stirred for 6 h. Potassiumhydrogensulfate (2 equiv.; 542 mg; 4mmol) was added followed by 100 mL dichloromethane and drying material (Na₂SO₄). The resulting heterogeneous mixture was stirred for 12 h. and filtered. The solvent was removed under reduced pressure.

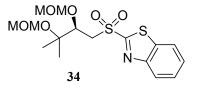
Flash chromatography (gradient: 15% to 40% ethyl acetate/hexane) afforded hygroscopic alcohol **49** as an yellowish oil (317 mg; 87 % yield): $R_f = 0.35$ (30% ethyl acetate/hexane), which was used without further purification but after careful drying in a desiccator over phosphorouspentoxide under vacuum.

2-[(2R)-2,3-Di(methoxymethoxy)-3-methylbutylsulfanyl]-1,3-benzothiazole (50):



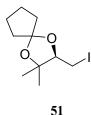
To a solution of alcohol **49** (361 mg; 1.74 mmol) in dry THF (18 mL) was added triphenyl phosphine (1.3 equiv.; 593 mg; 2.2 mmol), benzothiazolemercaptane (1.5 equiv.; 435 mg; 2.6 mmol) and DEAD (1.3 equiv.; 0.4 mL; 2.2 mmol). The reaction mixture was stirred for 2 h., poured into saturated sodium bicarbonate (100 mL) and extracted with dichloromethane (5×30 mL). The combined extracts were dried (Na₂SO₄) and the solvent removed under reduced pressure. Two flash chromatographies (gradient: 6% to 20% ethyl acetate/hexane) afforded 561 mg (1.56 mmol, 90% yield) of benzothiazole sulfide **50** as an oil: R_f = 0.53 (30% ethyl acetate/hexane); ¹H NMR (250 MHz, CDCl₃): δ 1.36 (s, 3H), 1.38 (s, 3H), 3.38 (dd, *J*= 7.9, 13.3 Hz, 1H), 3.41 (s, 3H), 3.44 (s, 3H), 3.80 (dd, *J* = 3.3, 7.9 Hz, 1H), 3.96 (dd, *J* = 3.3, 13.3 Hz, 1H), 4.76 (d, *J*= 6.7 Hz, 1H), 4.79 (d, *J*= 7.2 Hz, 1H), 4.85 (d, *J*= 7.3 Hz, 1H), 4.87 (d, *J*= 6.7 Hz, 1H), 7.29 (m, 1H), 7.41 (m, 1H), 7.75 (m, 1H), 7.83 (m, 1H).

2-[(2R)-2,3-Di(methoxymethoxy)-3-methylbutylsulfonyl]-1,3-benzothiazole (34):



Sulfide **50** (551 mg; 1.54 mmol) was dissolved in ethanol (14 mL) and a catalytic amount (0.10 equiv.; 190 mg; 0.15 mmol) ammonium molybdate tetrahydrate was added followed by sodium bicarbonate (80 mg) and a 30% hydrogen peroxide solution (1.6 equiv.; 3.4 mL). The solution became yellow then orange. The reaction mixture was stirred for 24 h. To the resulting yellowish solution was added a 25% solution of ethyl acetate in hexane (100 mL). The solution was washed with saturated sodium bicarbonate. The water phase was extracted with dichloromethane (4×25 mL). The combined organic solutions were dried (MgSO₄) and the solvent removed under reduced pressure. Flash chromatography (30% ethyl acetate/hexane) afforded the desired sulfone **34** (518 mg; 86% yield) as a clear oil. R_f = 0.28 (30% ethyl acetate/hexane); ¹H NMR (400MHz, CDCl₃): δ 1.23 (s, 3H), 1.33 (s, 3H), 3.24 (s, 3H), 3.62 (s, 3H), 3.38 (dd, *J*= 8.0, 15.5 Hz, 1H), 4.13 (dd, *J*= 2.3, 15.5 Hz, 1H), 4.17 (dd, *J*= 2.3, 8.0 Hz, 1H), 4.60 (d, *J*= 7.4 Hz, 1H), 4.64 (d, *J*= 7.4 Hz, 1H), 4.72 (d, *J*= 6.6 Hz, 1H), 4.95 (d, *J*= 6.6 Hz, 1H), 7.59 (m, 1H), 7.64 (m, 1H), 8.02 (m, 1H), 8.22 (m, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 21.2 (CH₃), 23.9 (CH₃), 55.3 (CH₃), 56.6 (CH₃), 57.5 (CH₂), 77.7 (C), 78.3 (CH), 91.1 (CH₂), 98.1 (CH₂), 122.3 (CH), 125.4 (CH), 127.6 (CH), 128.0 (CH), 136.8 (C), 152.7 (C), 166.4 (C=N, thiazole).

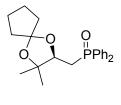
(2*R*)-3,3-(Dimethyl-1,4-dioxaspiro[4.4]non-2-yl)-methyl iodide (51):



Alcohol **45** (242 mg; 1.3 mmol), triphenylphosphine (2.1 equiv.; 716 mg; 2.7 mmol) and imidazole (4.6 equiv.; 411 mg; 5.98 mmol) were dissolved in dry 1,4-dioxane (5 mL). Upon addition of iodine (2.0 equiv.; 660 mg; 2.6 mmol) a weakly exothermic reaction was observed. The mixture was stirred at room temperature overnight and then heated to 70°C for 1.5 h. The reaction mixture was poured into cold water (50 mL) and extracted with ether (2×50 mL). The combined extracts were washed with brine (50 mL) and dried (MgSO₄).

The solvent was removed under reduced pressure. Two flash chromatographies (10% ether/hexane) afforded iodide **51** (362 mg; 94%) as an oil. $R_f = 0.41$ (10% ether/hexane); ¹H NMR (250 MHz, CDCl₃): δ 1.12 (s, 3H), 1.32 (s, 3H), 1,56-1.87 (comp m, 8H) 3.07 (dd, J = 5.8, 10.3 Hz, 1H), 3.19 (dd, J = 7.9, 10.3 Hz, 1H), 3.90 (dd, J = 5.8, 7.9 Hz, 1H); ¹³C NMR (62.5 MHz, CDCl₃): δ 1.1 (CH₂), 21.7 (CH₃), 23.3 (CH₂), 23.7 (CH₂), 26.7 (CH₃), 38.1 (CH₂), 38.2 (CH₂), 79.8 (C), 83.7 (CH), 116.7 (C).

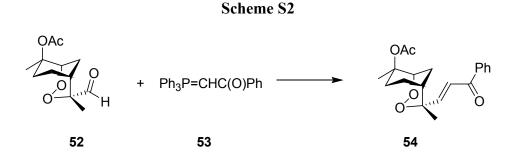
(3S)-3-(Diphenyl-phosphinoylmethyl)-2,2-dimethyl-1,4-dioxa-spiro[4.4]nonane (35):



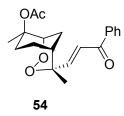
35

Diphenylphosphine (1.1 equiv.; 251 mg; 1.348 mmol) was dissolved in dry THF (3 mL) and the solution cooled to 0°C. A hexane solution of *n*-butyllithium (1.1 equiv.; *ca.* 1.4 M; 0.96 mL; 1.348 mmol) was added drop wise affording a deep orange solution. The reaction mixture was stirred for 30 min. at 0°C and then a solution of iodide **51** in dry THF (1.5 mL + 2×0.5 mL) was added via canula. The colour changed to pale yellow. The reaction mixture was stirred for 30 min at 0°C and then for 2.5 h. at room temperature. The reaction mixture was poured into water (50 mL) and extracted with chloroform (100 mL). The organic phase was washed with water and the solvent removed under reduced pressure. Under the entire workup no special precautions were taken to exclude atmospheric oxygen. Four flash chromatographies (25% ethyl acetate/hexane) afforded Phosphinate **35** (170 mg; 49% yield) as a colourless solid. R_f = 0.25 (25% ethyl acetate/hexane); ¹H NMR (250 MHz, CDCl₃): δ 1.14 (s, 3H), 1.24 (s, 3H), 1,29-1.80 (comp m, 8H) 2.30 (ddd, *J* = 3.2, 15.0, 16.8 Hz, 1H), 2.58 (ddd, *J* = 7.4, 9.4, 16.8 Hz, 1H), 3.94 (dd, *J* = 3.2, 9.4 Hz, 1H). ³¹P NMR (101.25 MHz, CDCl₃): δ 30.5.

Model studies; Wittig olefination of aldehyde 52



(1*R*,4*R*,5*R*,8*R*)-8-Acetoxy-4,8-dimethyl-4-(*E*)-(3-oxo-3-phenyl-1-propenyl)-2,3-dioxabicyclo[3.3.1]nonane (54).



A mixture of (1R,4R,5R,8R)-8-acetoxy-4,8-dimethyl-4-formyl-2,3-dioxabicyclo[3.3.1]nonane (**52**)^{4,5} (16.0 mg, 0.066 mmol) and benzoylmethylene)triphenylphosphorane (**53**) (150 mg, 0.396 mmol) in dry CH₂Cl₂ (1.5 ml) was stirred at rt under argon for 30 days. At that time TLC analysis revealed approximately 90% consumption of aldehyde **52**. The mixture was directly subjected to flash chromatography (elution with hexane-benzene-ethyl acetate 4:2:1) to give the title compound **54** (10.8 mg, 48%) as a yellowish oil, R_f = 0.43 (in ethyl acetate/benzene/hexane 3:6:11); ¹H NMR (250 MHz, CDCl₃): δ 1.32 (s, Me(11), 3H), 1.72 (s, Me(10), 3H), 1.78-1.90 (m, 4H), 2.01-2.15 (m, 1H), 2.04 (s, MeC(O), 3H), 2.18-2.42 (m, 2H), 4.44 (br s, H(1), 1H), 7.06 (d, *J* = 15.6 Hz, =C(14)HC(O), 1H), 7.37 (d, *J* = 15.6 Hz, =C(13)H, 1H), 7.47-7.53 (m, 2H), 7.54-7.62 (m, 1H), 8.03 (ddd, *J* = 7.0, 1.1, 1.1 Hz, 2H); ¹³C NMR (100MHz, C₆D₆): δ 21.8 (C(11)H₃), 21.8 (C(6)H₂), 22.9(CH₃C(O)), 23.4 (C(10)H₃), 25.2 (C(6)H₂), 31.2 (C(5)H), 33.6 (C(7)H₂), 78.0 (C(1)H), 82.4 (C(4)), 84.4 (C(8)), 125.3 (=C(13)H), 128.8 (2CH), 128.9 (2CH), 132.7 (CH), 150.4 (=C(14)HC(O)), 189.1 (C=O). A small amount of the

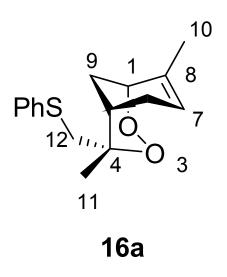
starting aldehyde **52** [0.5 mg, 5%, $R_f = 0.32$ (10% ethyl acetate in hexane)] was recovered by an additional flash chromatography of the less polar fractions.

References for Supporting Information

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- (2) (a) Mair, R. D.; Hall, R. T. In *Organic Peroxides*; Swern, D., Ed.; Wiley Interscience: London, 1971;
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- (3) A. B. Smith III, J. Kingerywood, T. L. Leenay, E. G. Nolen and T. Sunazuka, J. Am. Chem. Soc.
 1992, 11 4, 1438-1449
- (4) The aldehyde 52 was prepared in 92% yield from (1*R*,4*R*,5*R*,8*R*)-8-acetoxy-4,8-dimethyl-4phenylsulfinylmethyl-2,3-dioxabicyclo[3.3.1]nonane² through a Pummerer reaction with TFAA and 2,6-lutidine in conditions which are very similar to the described for the synthesis of unsaturated aldehydes 21-24 (see Scheme 4 and Experimental Section of this paper).
- Bachi, M. D.; Korshin, E. E.; Hoos, R.; Szpilman, A. M.; Cumming, J. N.; Plyopradith, P.; Xie, S. J.;
 Shapiro, T. A.; Posner, G. H. *J. Med. Chem.* 2003, *46*, 2516-2533.

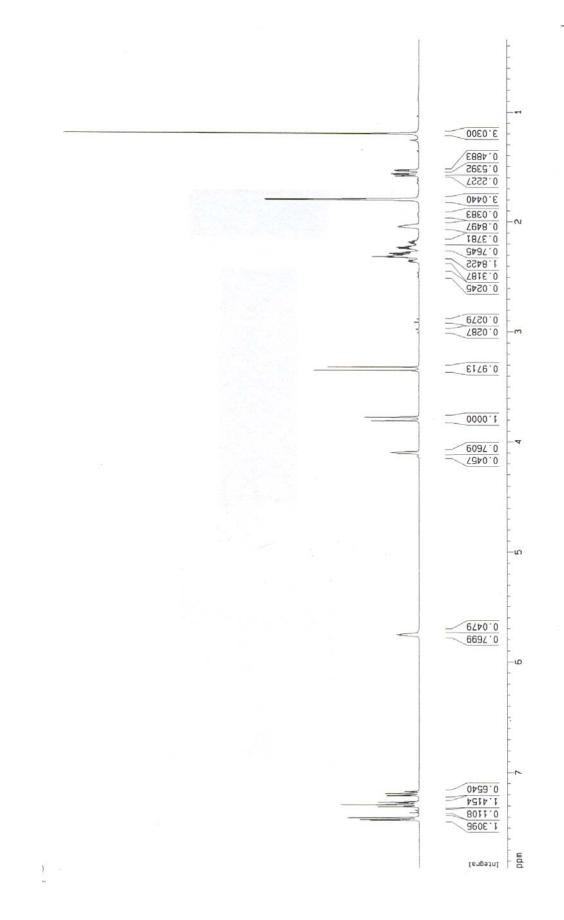
Copies of ¹H, ¹³C, DEPT, COSY and HMQC NMR spectra

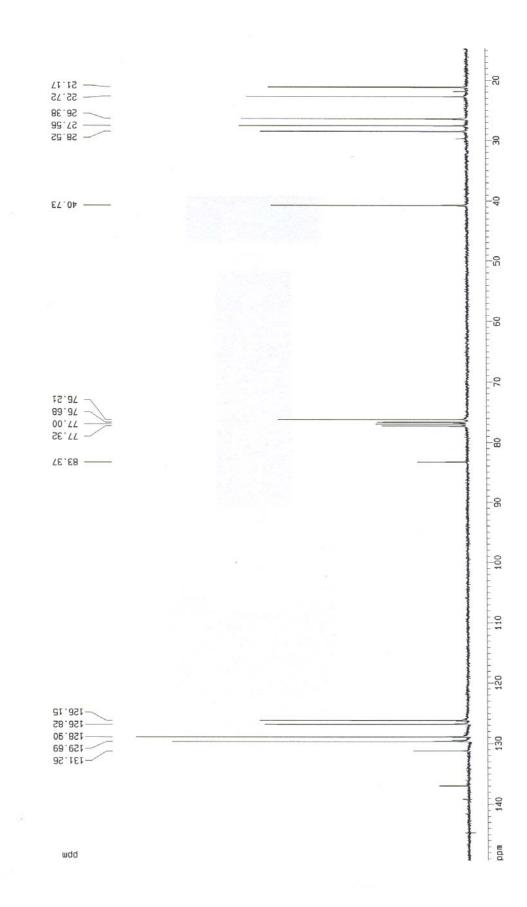
Unsaturated sulfide 16a:



Recorded at 400 MHz(¹H) and 100 MHz (¹³C) in CDCl₃: ¹H NMR spectra

¹³C NMR spectra

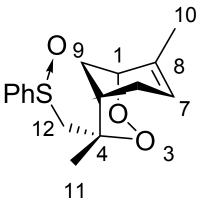




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- S16 -

Unsaturated sulfoxide 18b:



18b

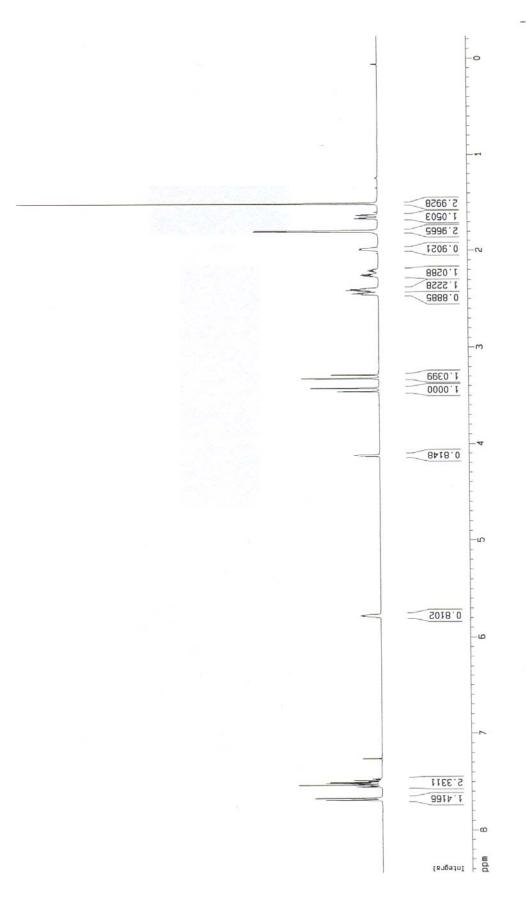
Recorded at 400 MHz(¹H) and 100 MHz (¹³C) in CDCl₃:

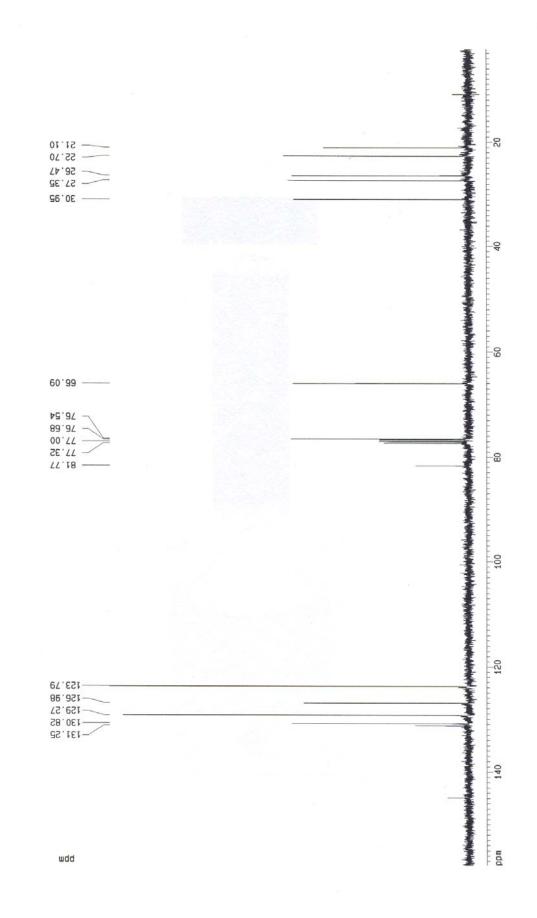
¹H NMR spectra

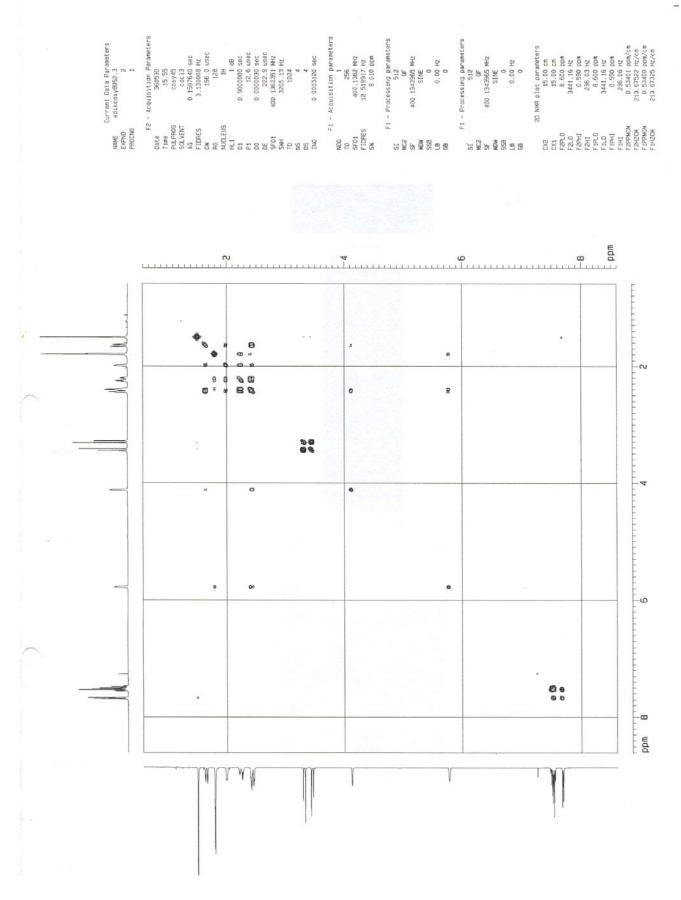
¹³C NMR spectra

COSY spectra

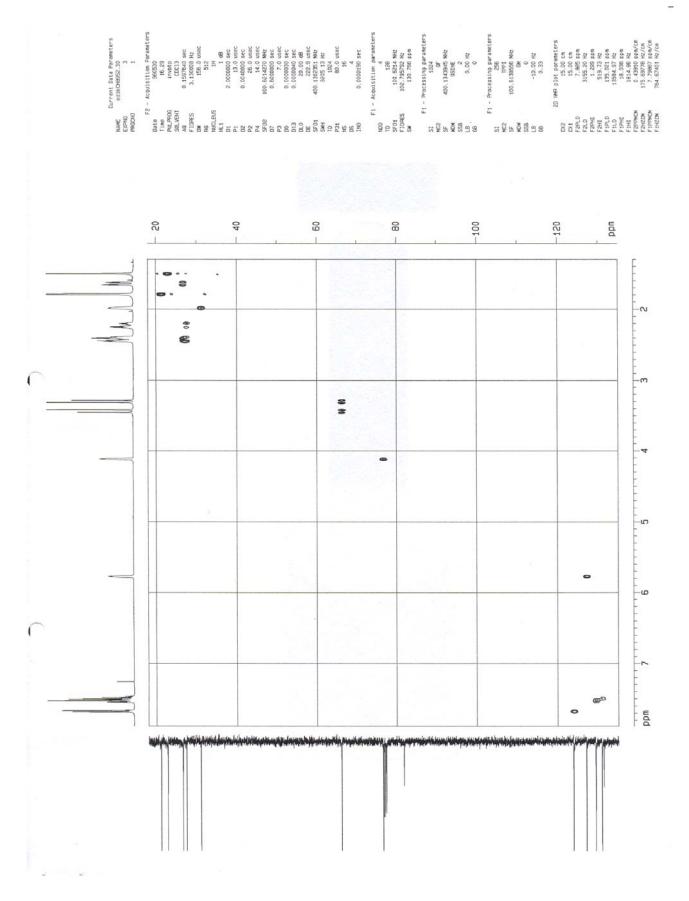
HMQC spectra



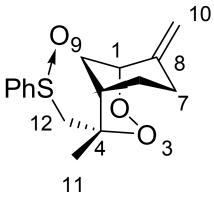




- S20 -



Unsaturated sulfoxide 19b:



19b

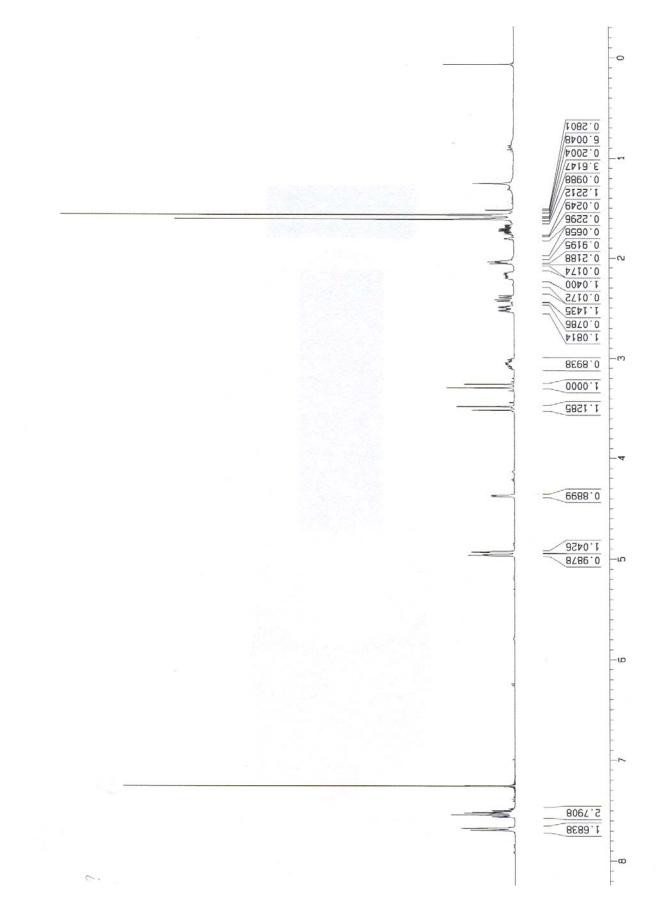
Recorded at 400 MHz(¹H) and 100 MHz (¹³C) in CDCl₃:

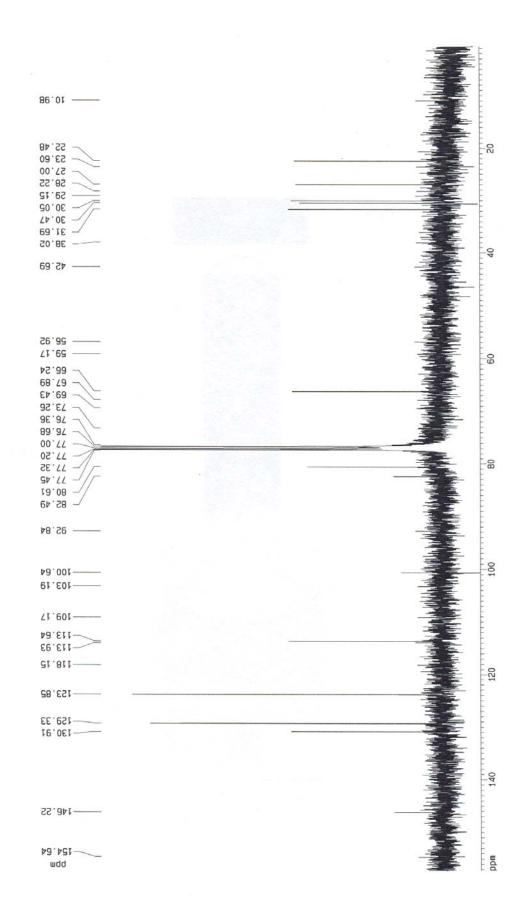
¹H NMR spectra

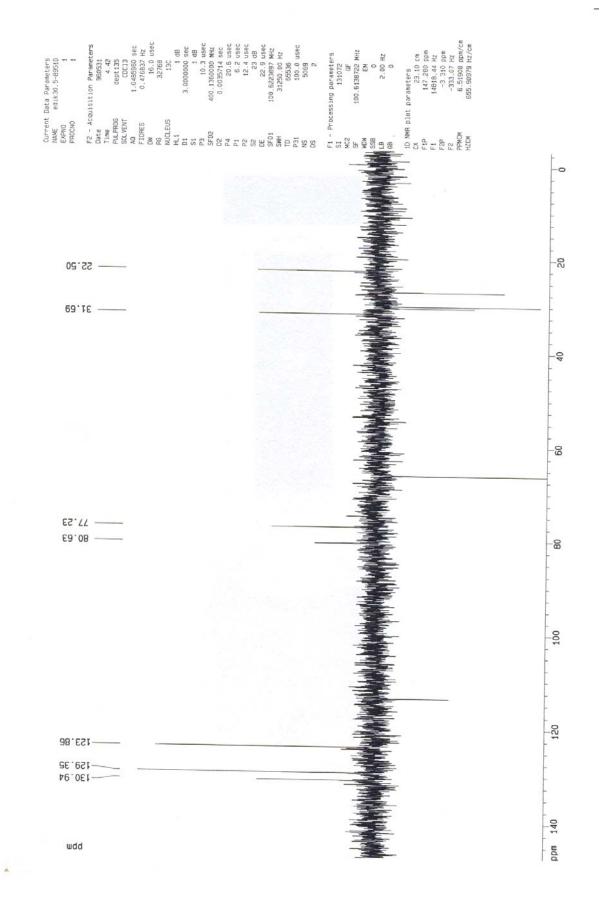
¹³C NMR spectra

DEPT spectra

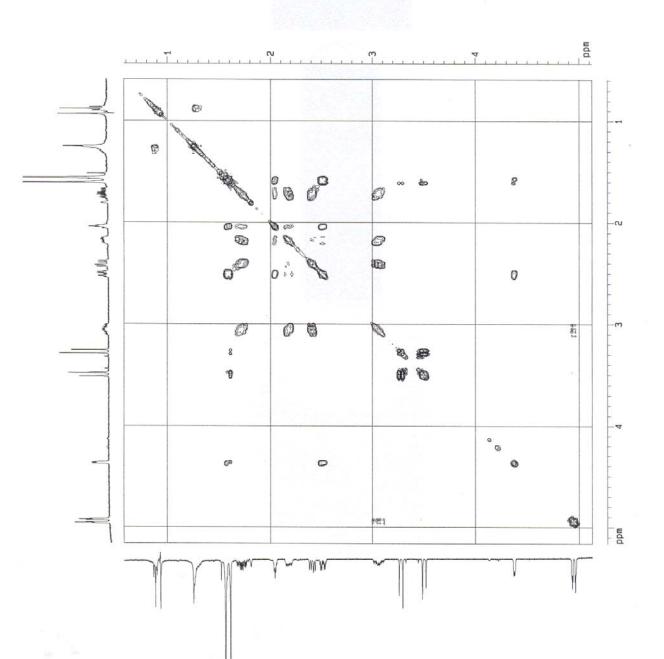
COSY spectra



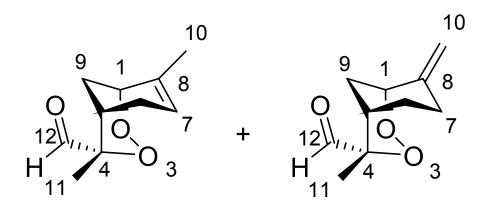








Mixture of unsaturated aldehyde **21** and **22**:



21

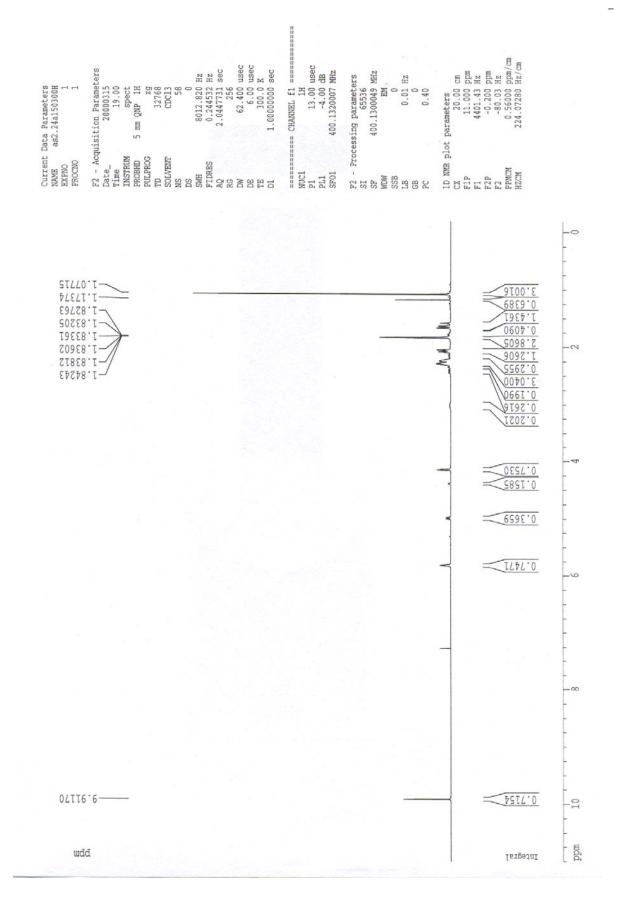
22

Recorded at 400 MHz(1 H) and 100 MHz (13 C) in CDCl₃:

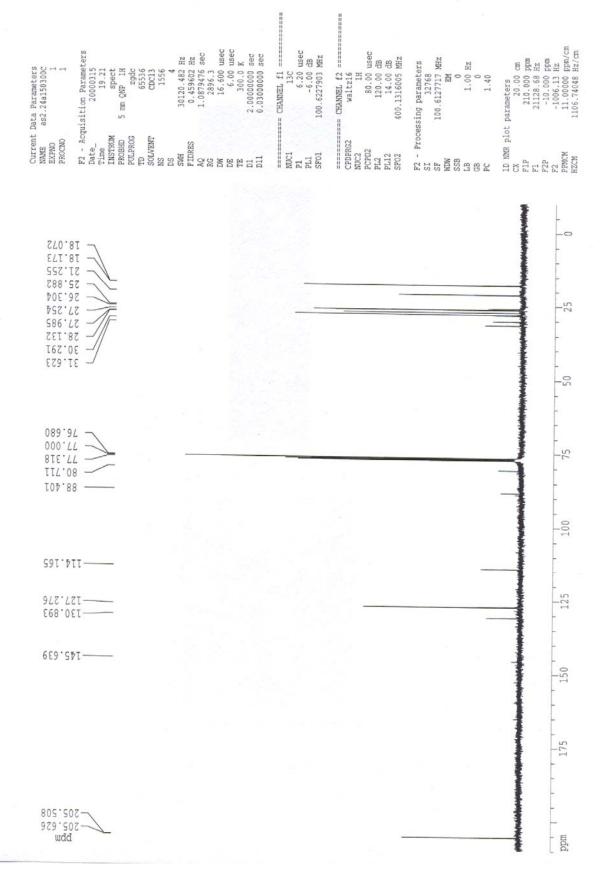
¹H NMR spectra

¹³C NMR spectra

COSY spectra

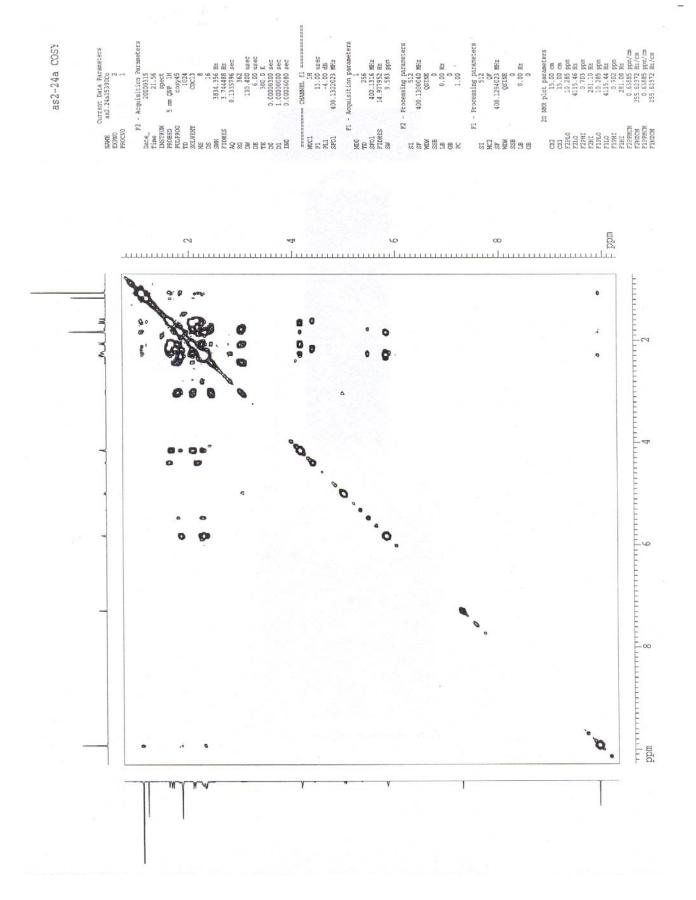


- S28 -

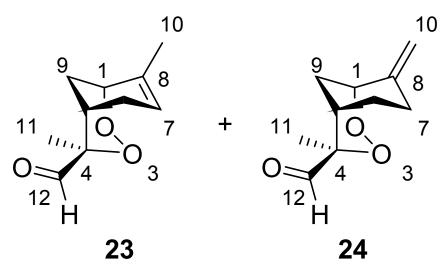


- S29 -

100



Mixture of unsaturated aldehydes 23 and 24:



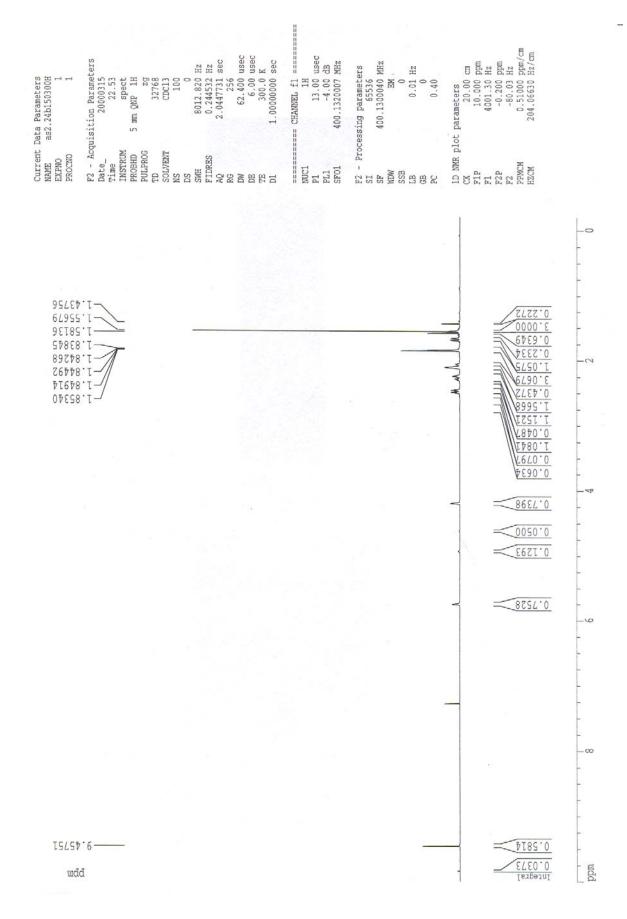
Recorded at 400 MHz(¹H) and 100 MHz (¹³C) in CDCl₃:

¹H NMR spectra

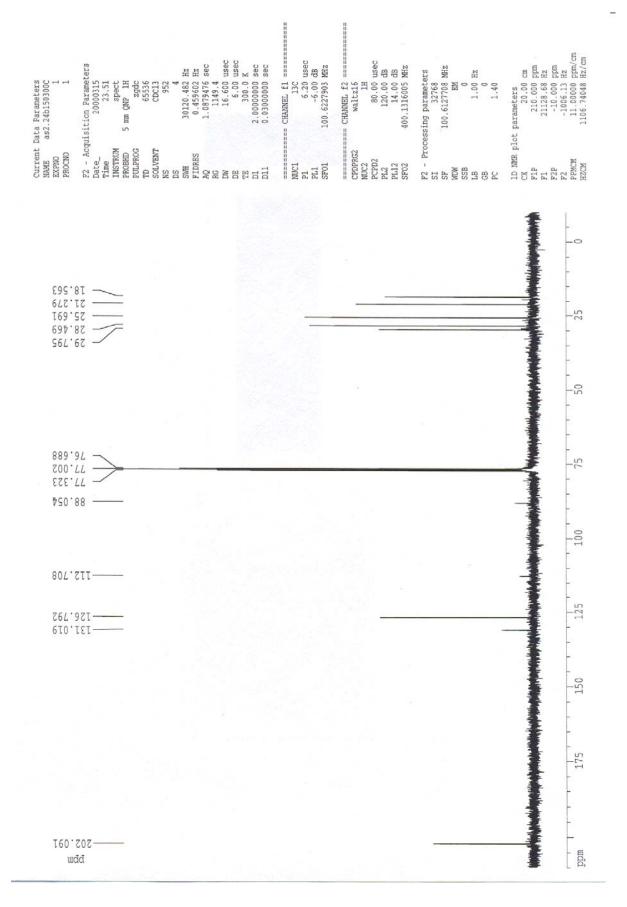
¹³C NMR spectra

DEPT spectra

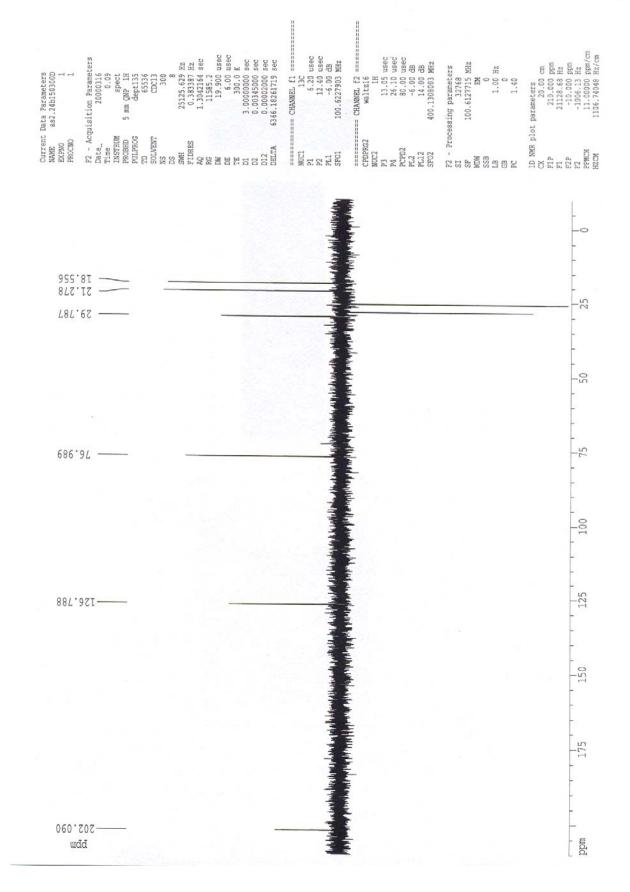
COSY spectra



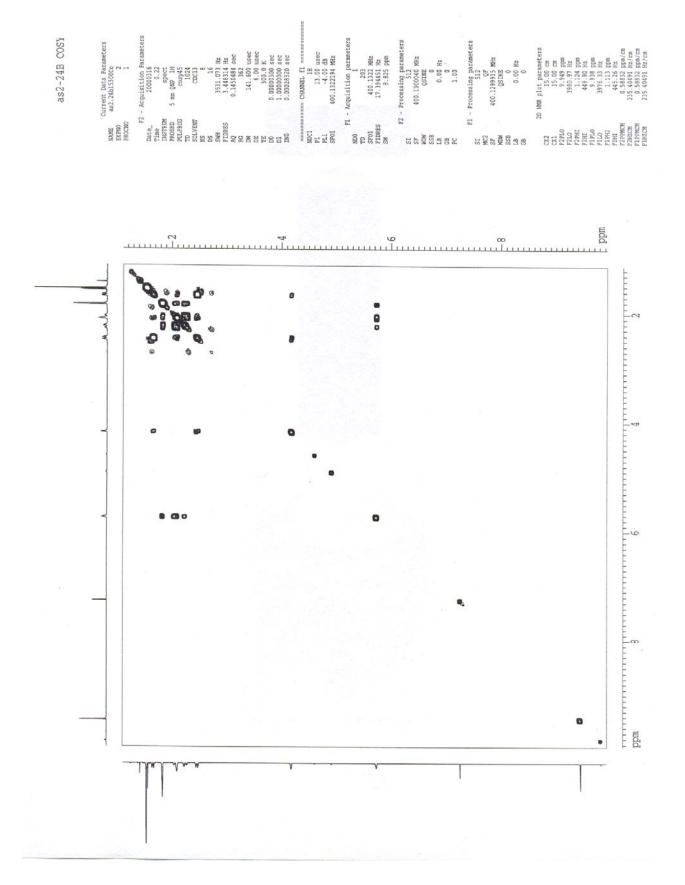
- S32 -



- S33 -



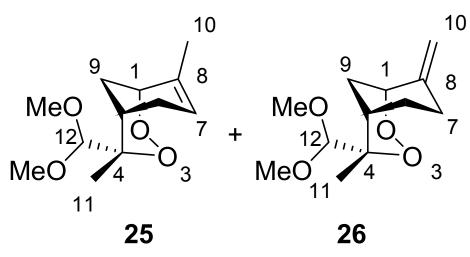
- S34 -



- S35 -

17

Mixture of unsaturated acetals 25 and 26:

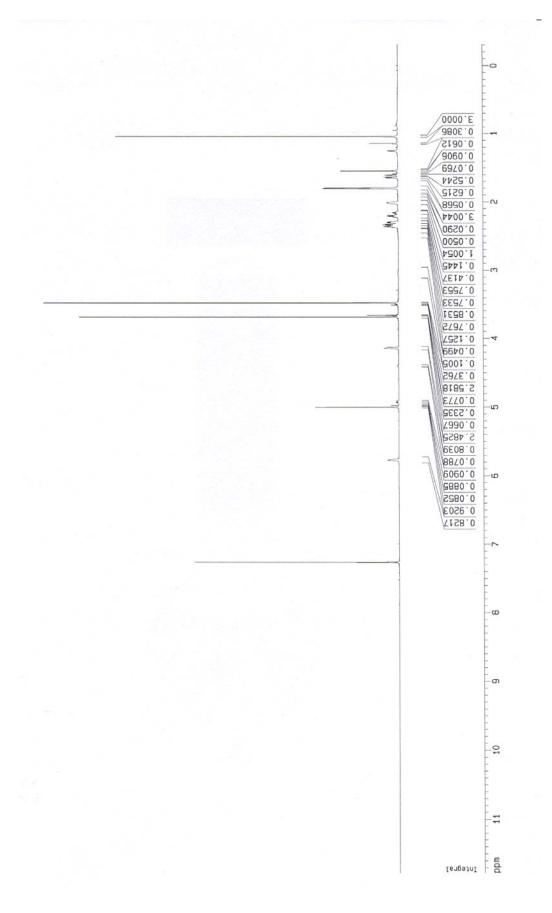


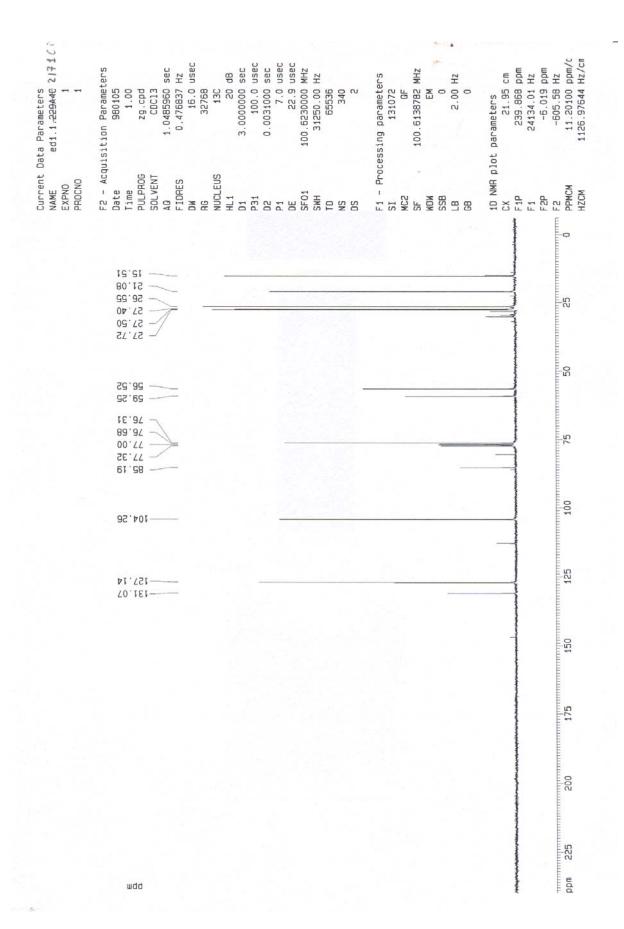
Recorded at 400 MHz(¹H) and 100 MHz (¹³C) in CDCl₃:

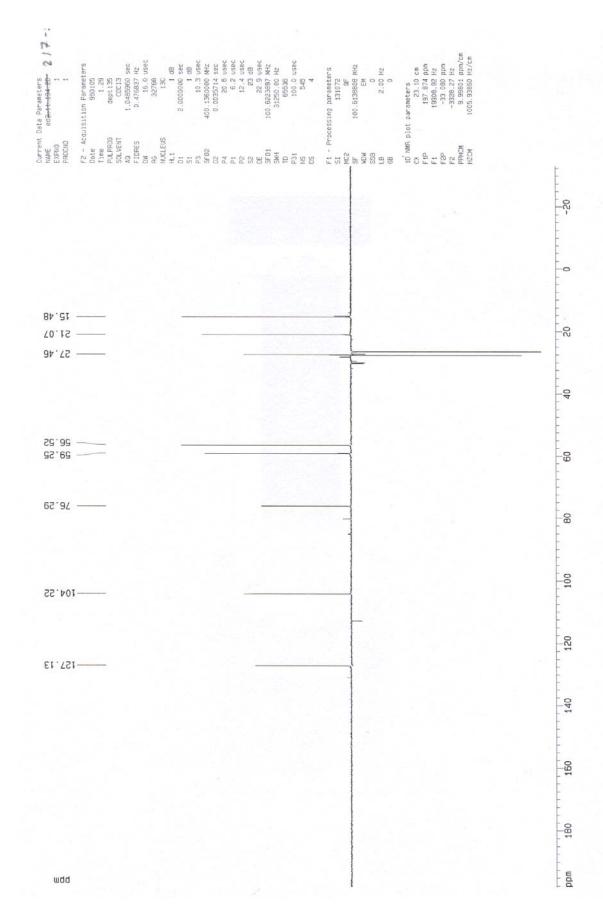
¹H NMR spectra

¹³C NMR spectra

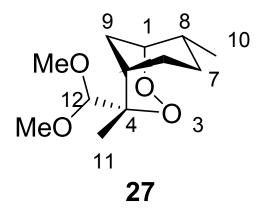
DEPT spectra







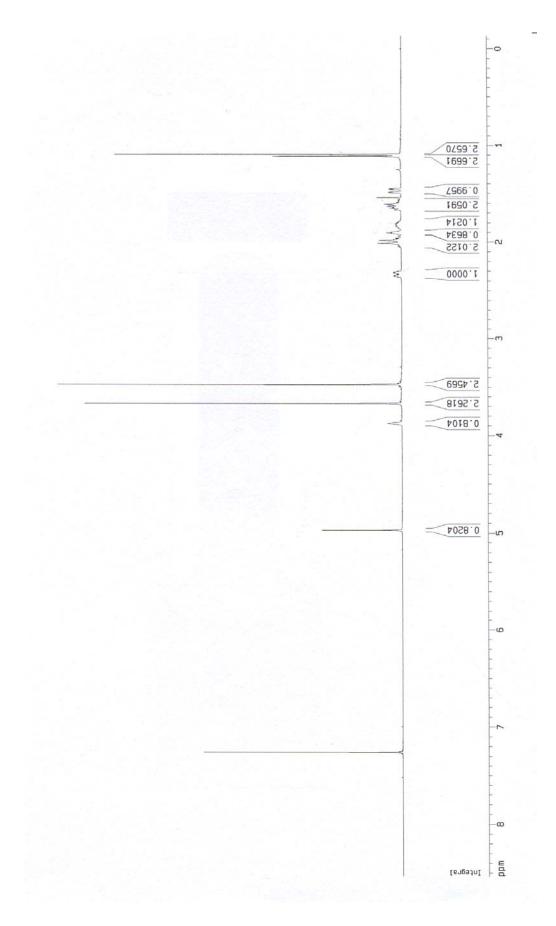
Saturated acetal **27**:

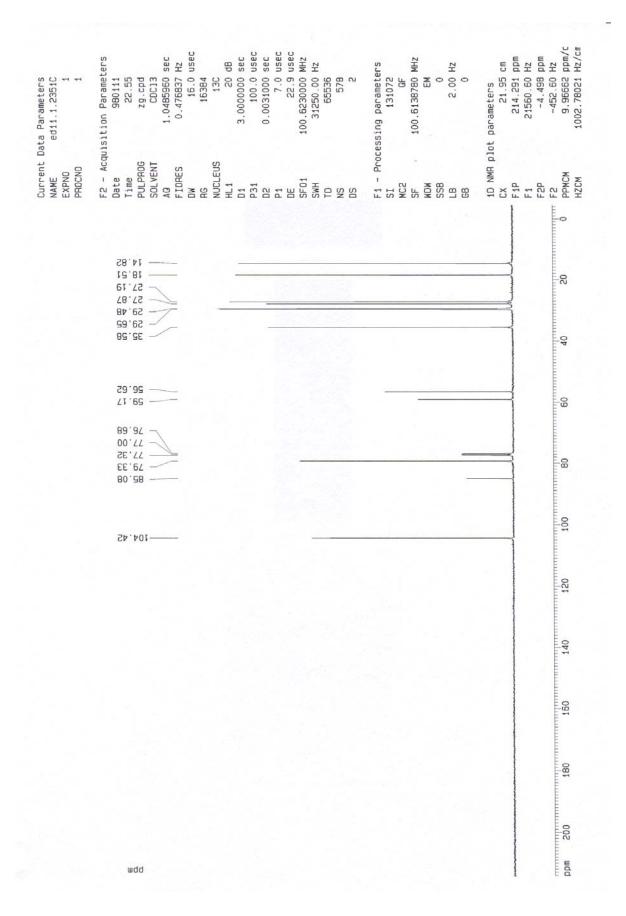


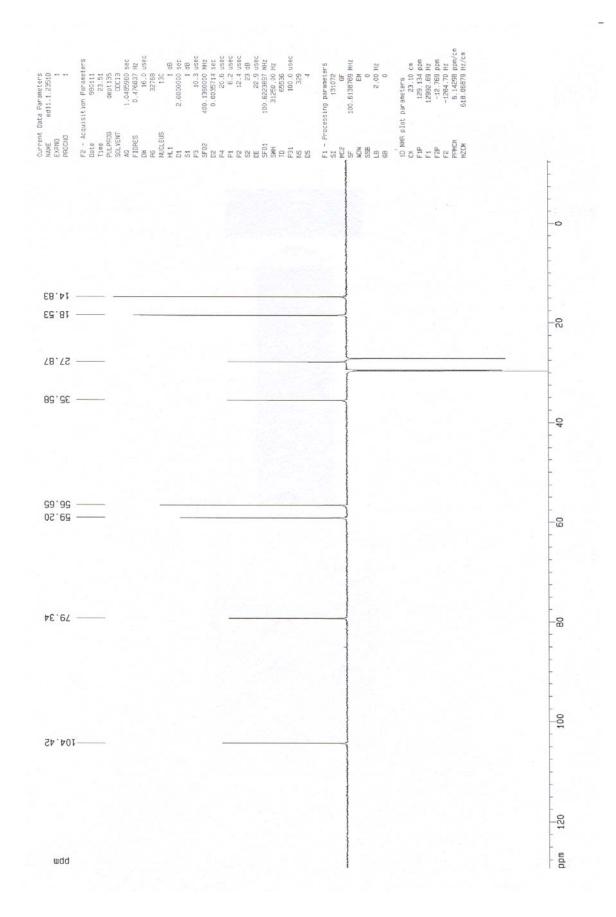
Recorded at 400 MHz(¹H) and 100 MHz (¹³C) in CDCl₃:

¹H NMR spectra

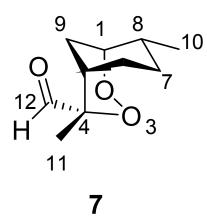
¹³C NMR spectra



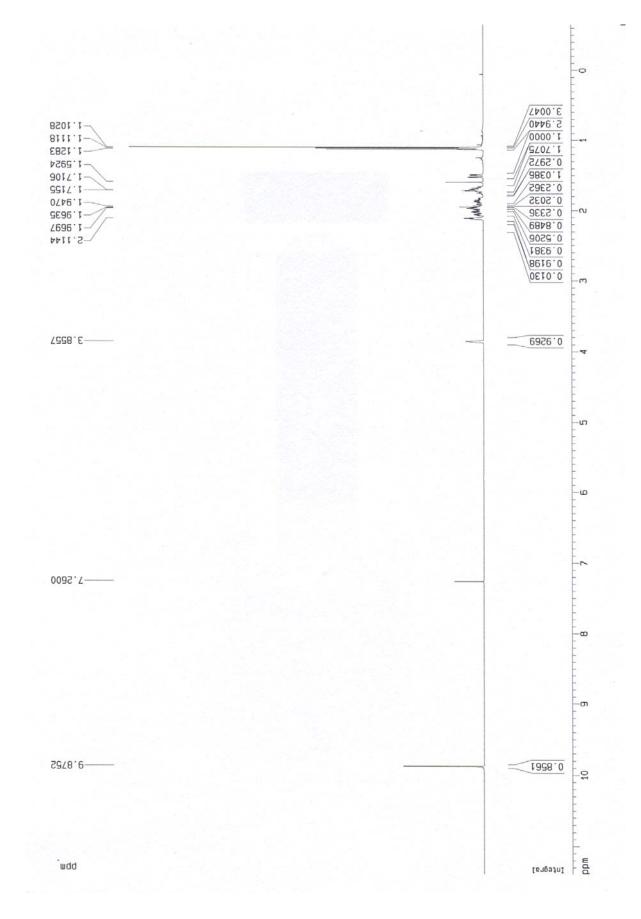


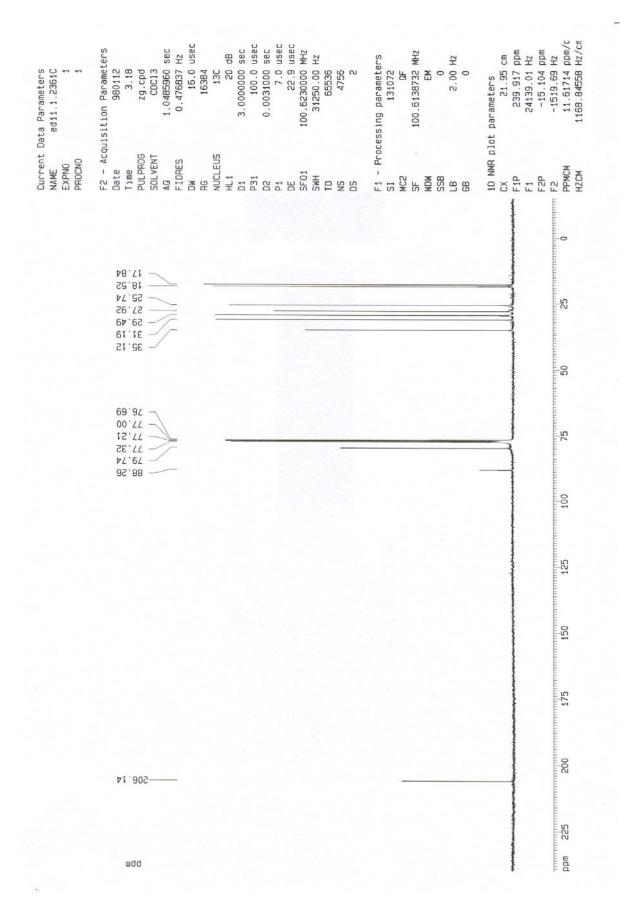


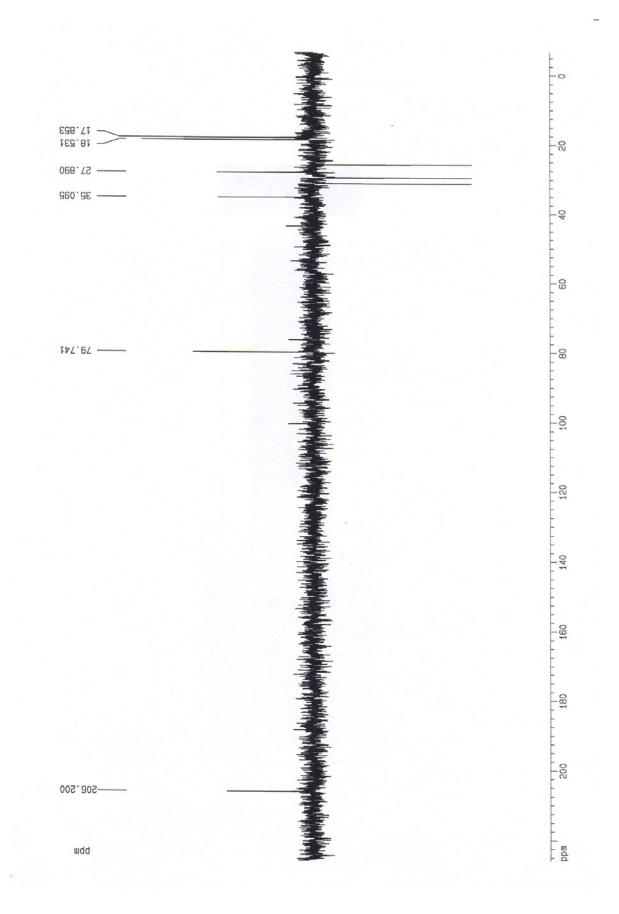
Saturated aldehyde 7:



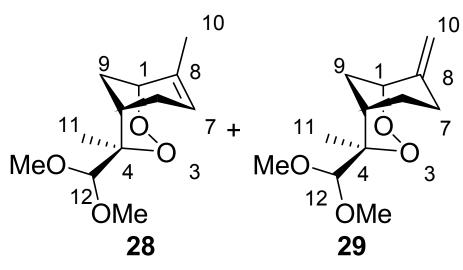
Recorded at 400 MHz(¹H) and 100 MHz (¹³C) in CDCl₃: ¹H NMR spectra ¹³C NMR spectra DEPT spectra







Mixture of unsaturated acetals 28 and 29:



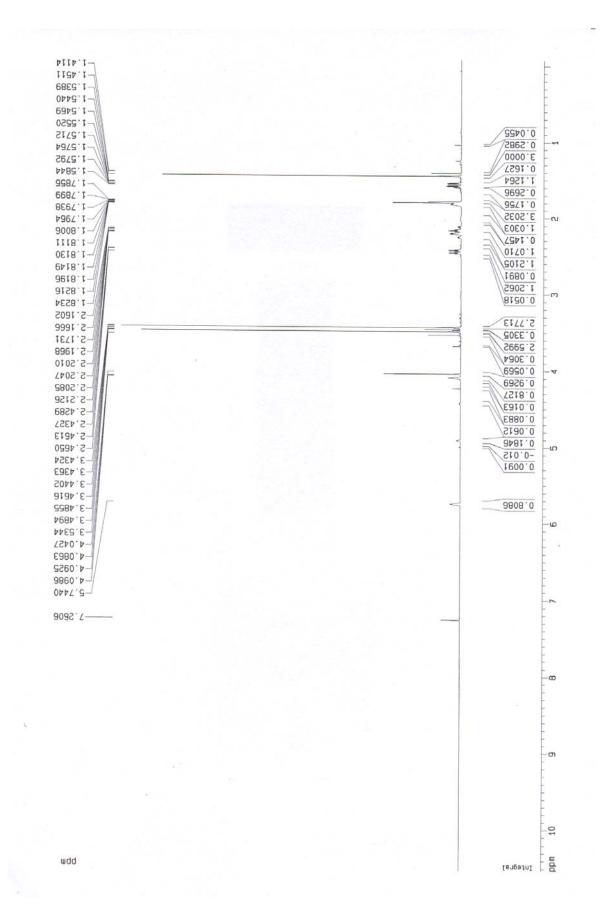
Recorded at 400 MHz(¹H) and 100 MHz (¹³C) in CDCl₃:

¹H NMR spectra

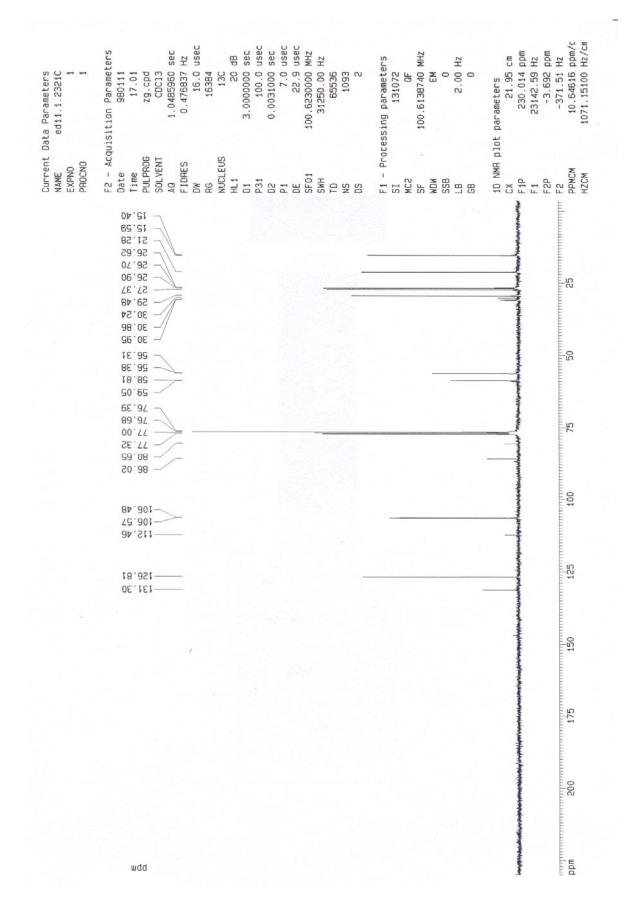
¹³C NMR spectra

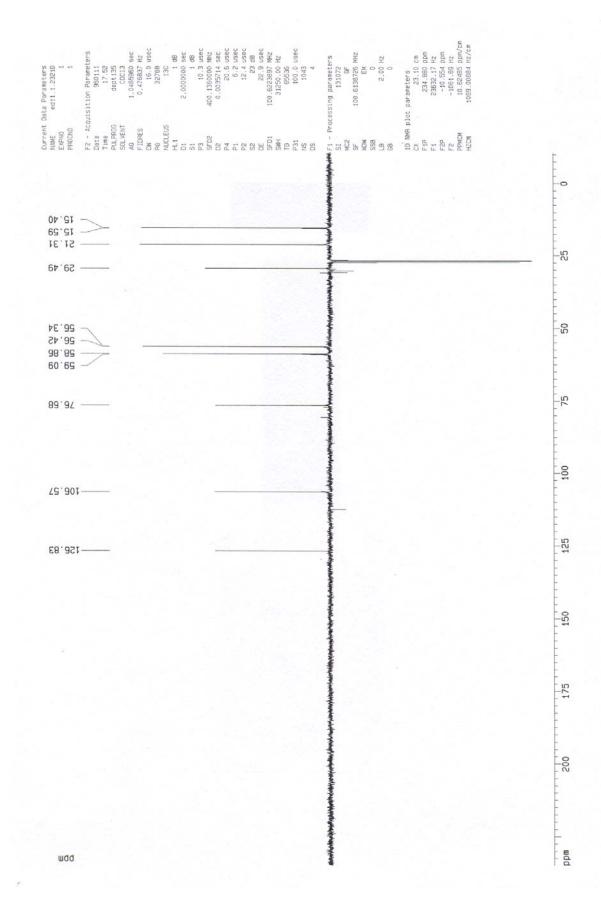
DEPT spectra

COSY spectra

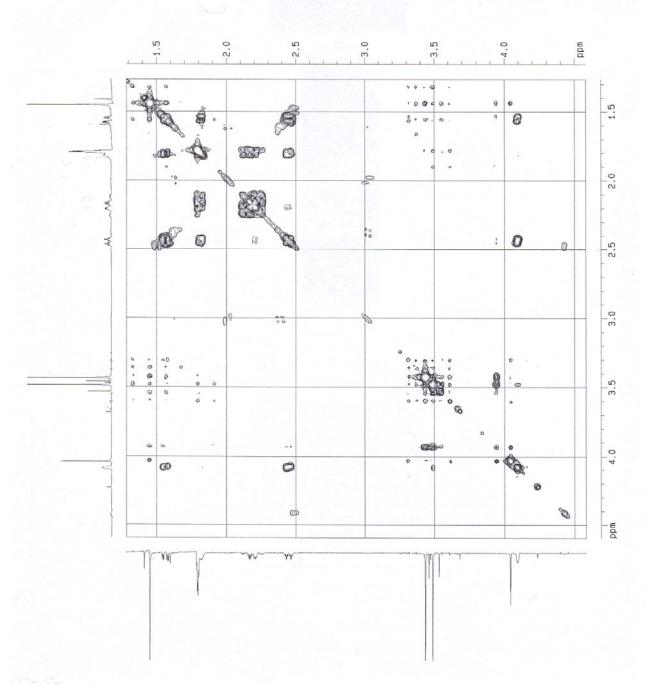


- S49 -

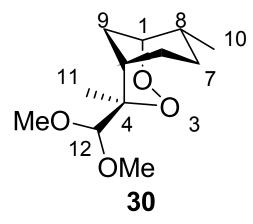




Gurrent Data Parameters edi1.1.2321Co 2 F1 - Acquisition parameter - Processing parane 512 0F 400.1343991 MHz 51NE 0.0 0.0 0 H2 1.500000 sec 12.6 usec 12.6 usec 0.000030 sec 290.0 usec 2463.05 Hz 2463.05 Hz 0.22202 ppm/cm 88.83581 Hz/cm 0.21961 ppm/cm 87.87407 Hz/cm 2078920 sec .405326 Hz 203.0 usec 256 400.135 MHZ 9.621238 Hz 5.155 ppn 0.00 Hz 0 15.00 cm 15.00 cm 4.592 ppm 1837.54 Hz 1.262 ppm 504.98 Hz 4.580 ppm 0.0004060 sec 400.1343943 MHz SINE 1.286 ppm 14.63 Hz 1511100 Par 980111 18.47 cosy45 cosy45 ocessing par 512 - 23 8 Date Time SOLVENT AG FIDRES DN RD NUOLEUS NAME EXPND PROCNO F1 ND0 TD SF01 FIDRES SN CX2 CX1 F2PLD F2PLD F2PLD F2PLD F2PLD F2PLD F2PLD F3PLD F3PL



Saturated acetal 30:



Recorded at 400 MHz(1 H) and 100 MHz (13 C):

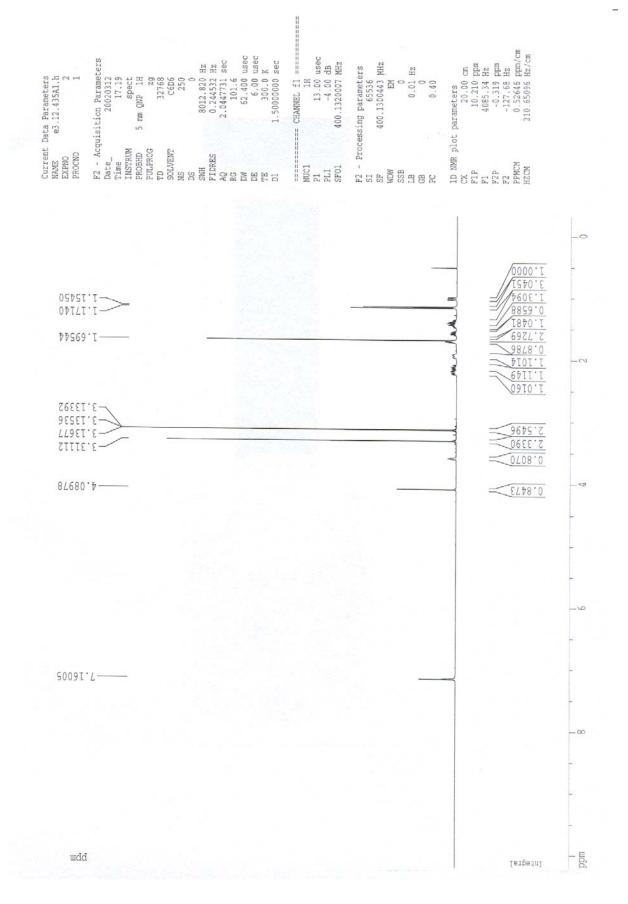
¹H NMR spectra (in C_6D_6)

¹³C NMR spectra (in C₆D₆)

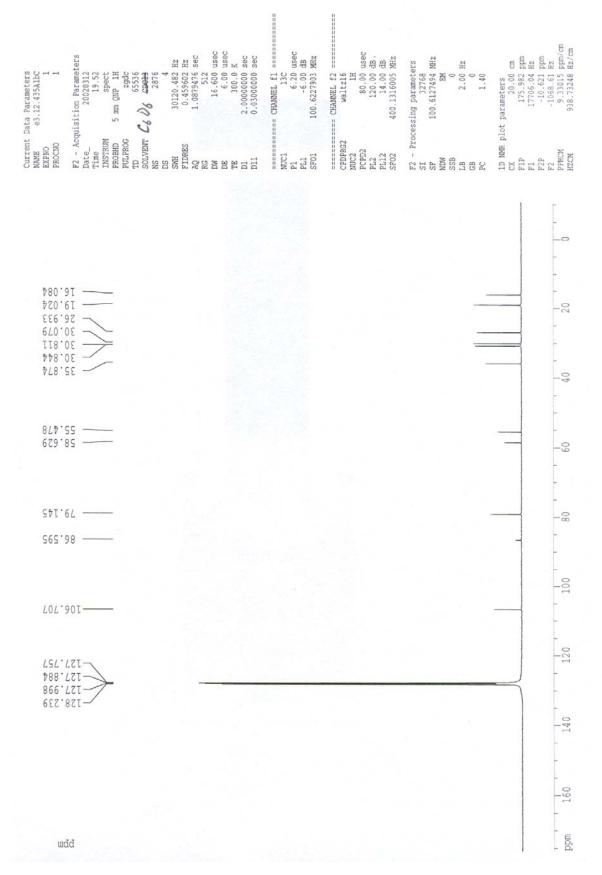
DEPT spectra (in CDCl₃)

COSY spectra (in C_6D_6)

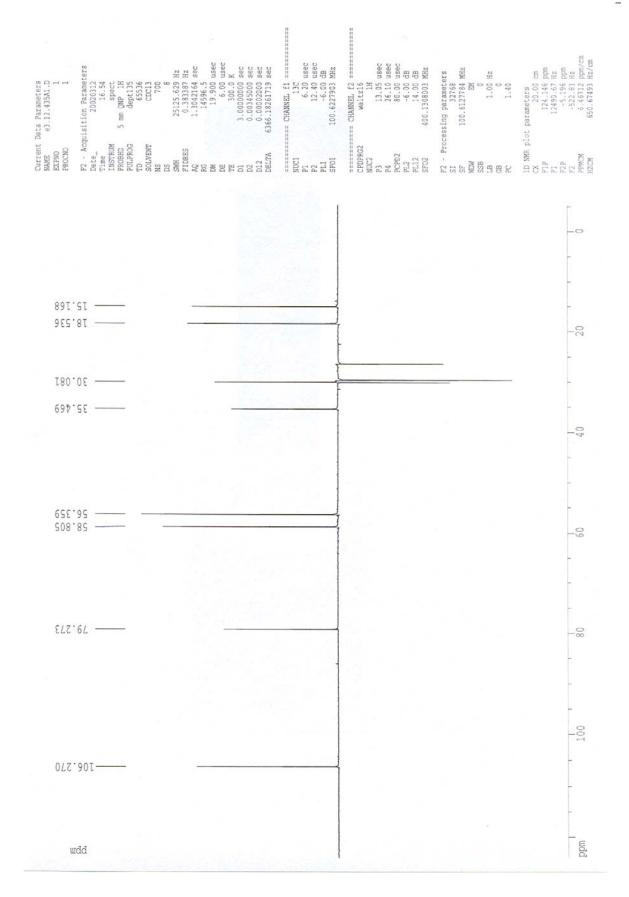
HMQC spectra (in C₆D₆)

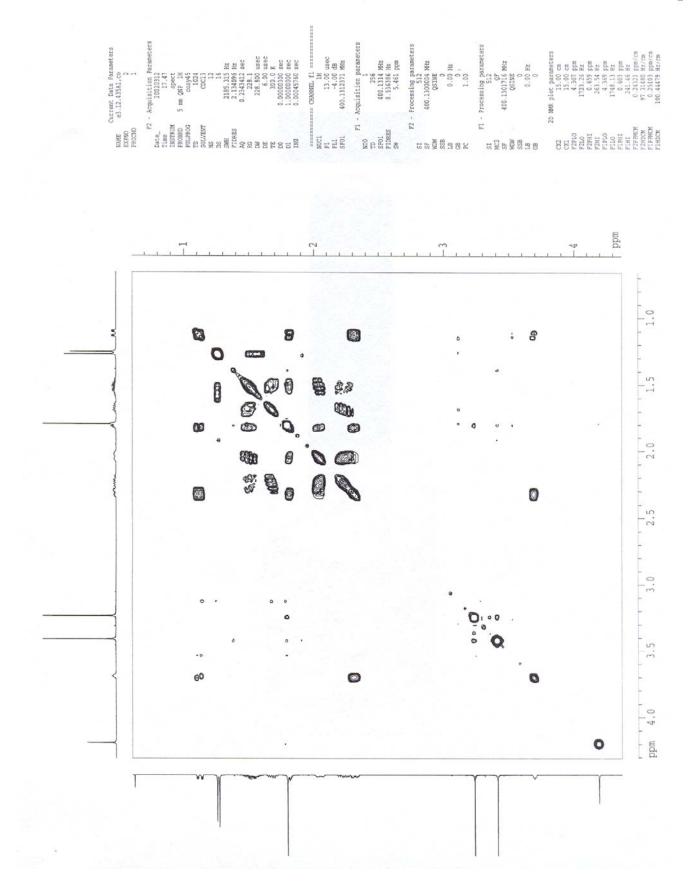


- S54 -

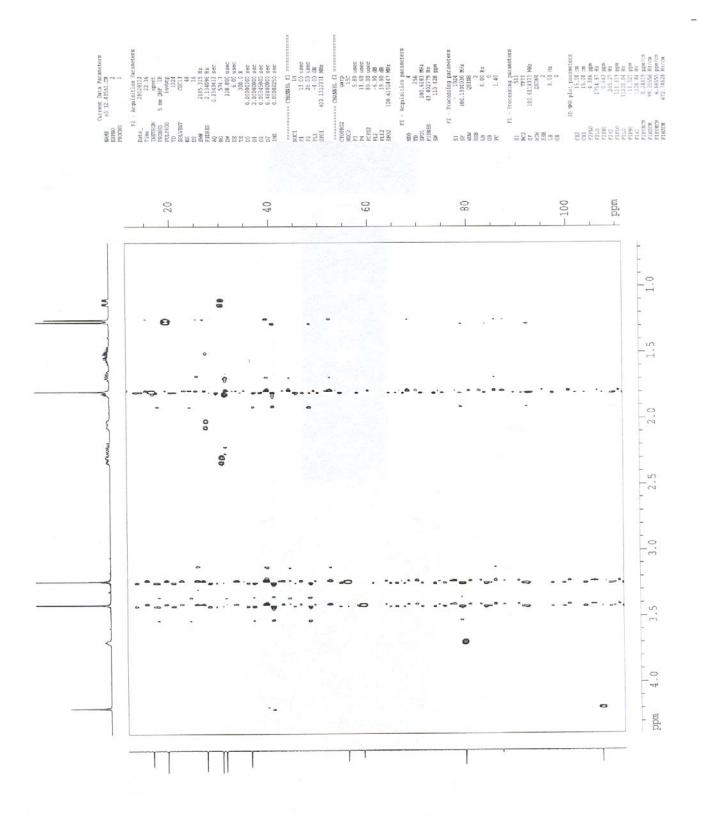


- S55 -



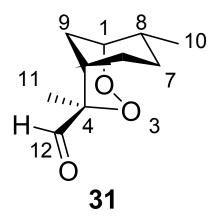


- S57 -



- S58 -

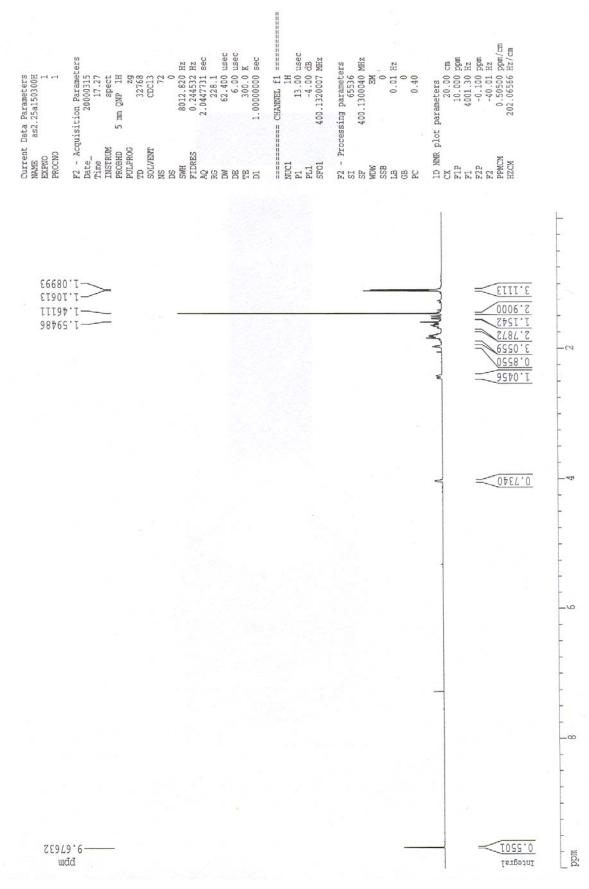
Saturated aldehyde **31**:



Recorded at 400 MHz(¹H) and 100 MHz (¹³C) in CDCl₃:

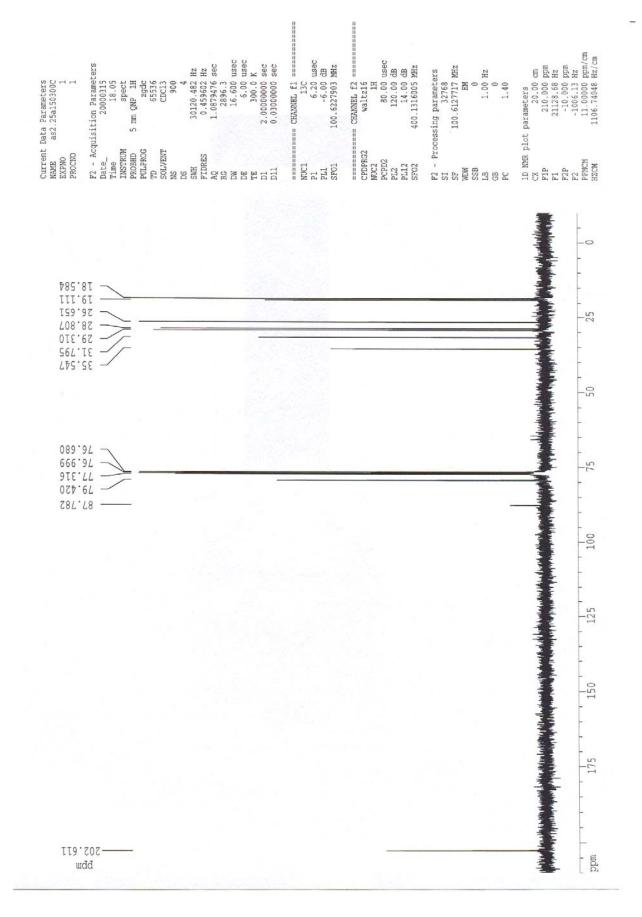
¹H NMR spectra

¹³C NMR spectra

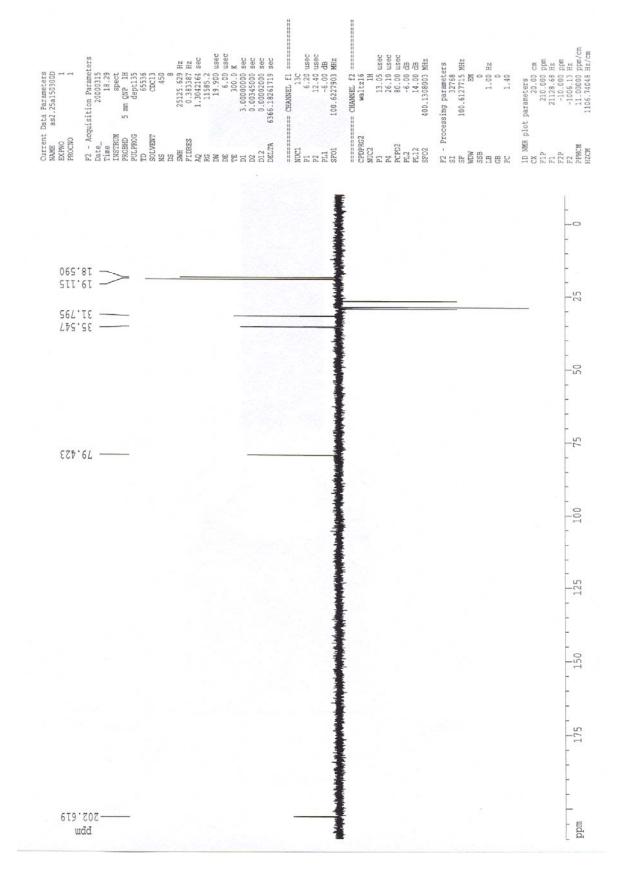


- S60 -

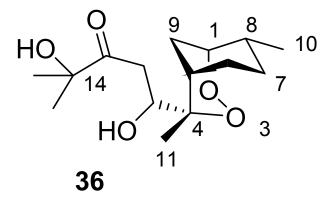
0.7



- S61 -



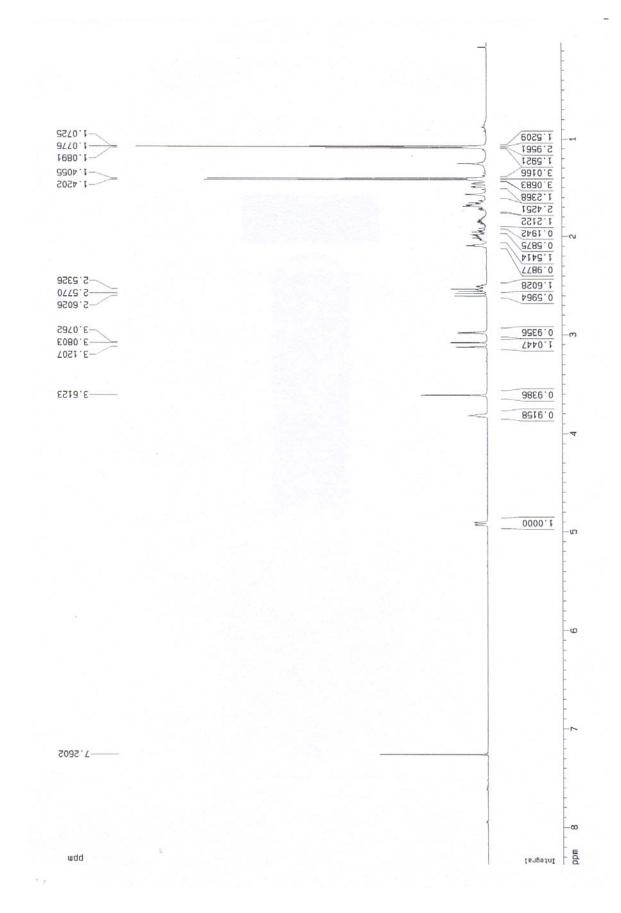
Aldol adduct 36:

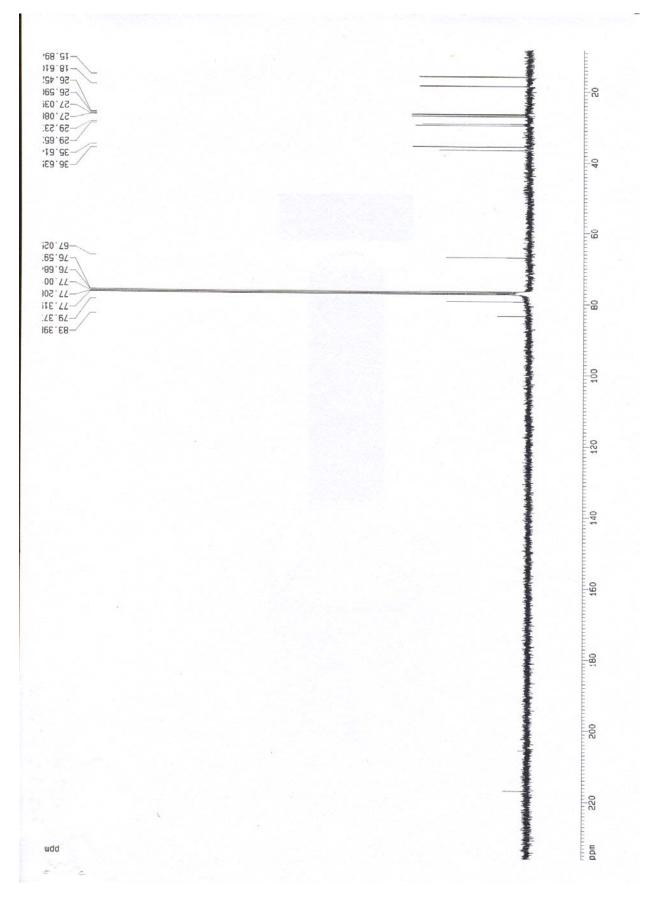


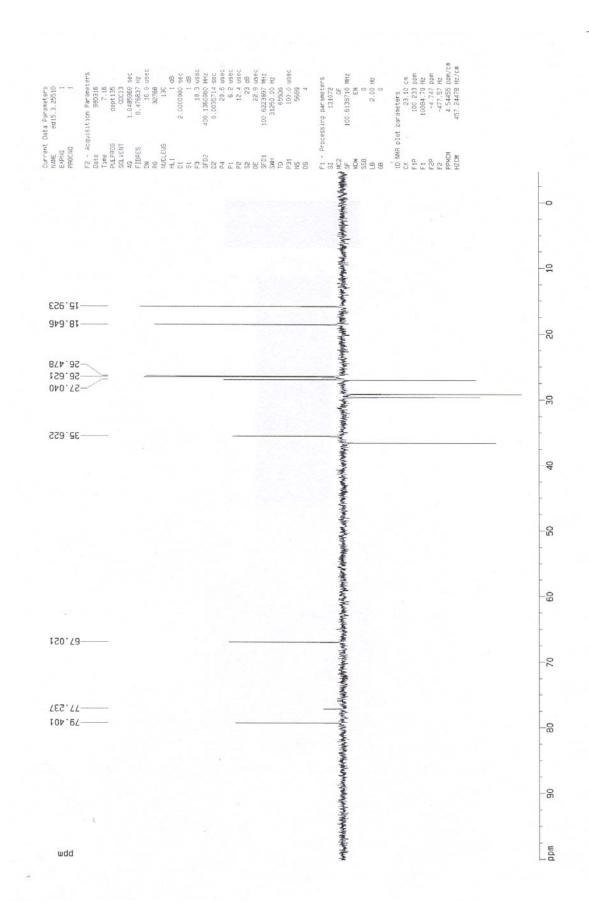
Recorded at 400 MHz(¹H) and 100 MHz (¹³C) in CDCl₃:

¹H NMR spectra

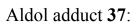
¹³C NMR spectra

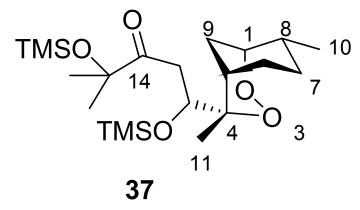






- S66 -

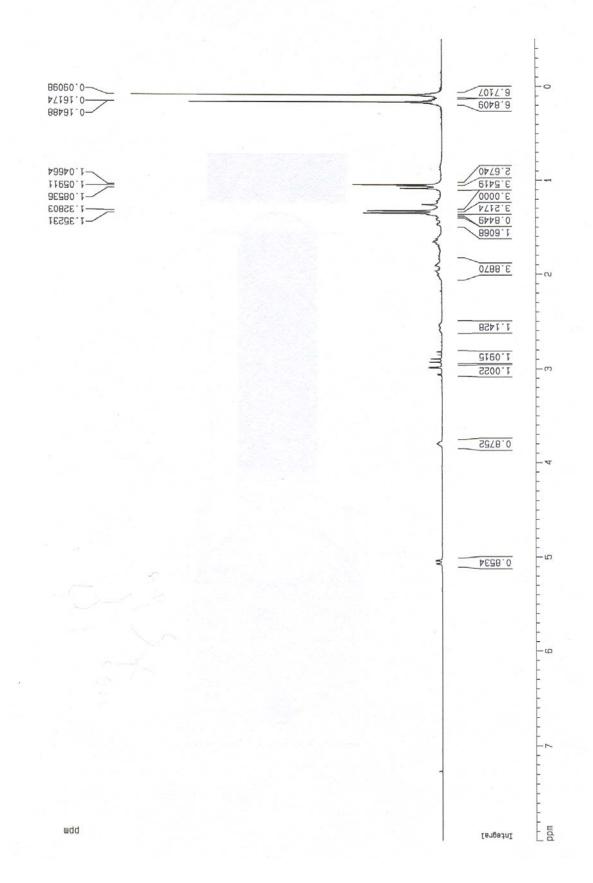


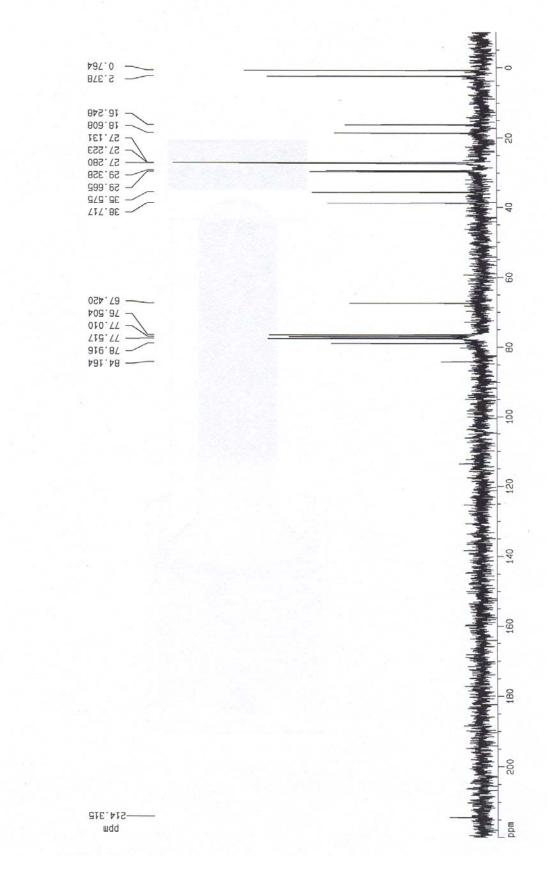


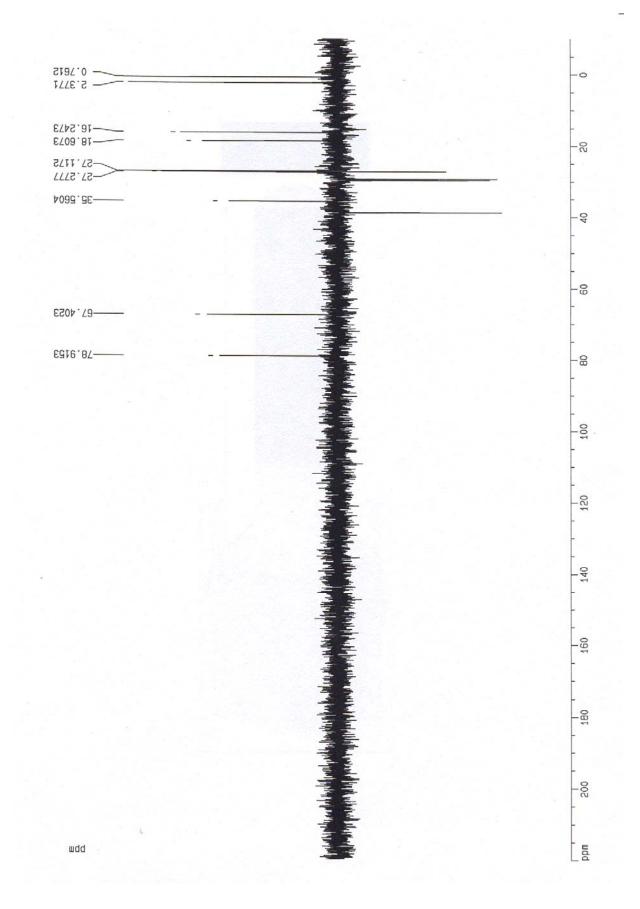
Recorded at 250 MHz(1 H) and 66.7 MHz (13 C) in CDCl₃:

¹H NMR spectra

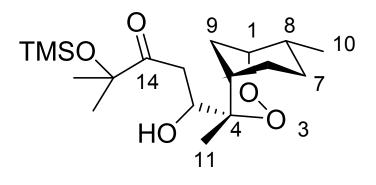
¹³C NMR spectra







Aldol adduct 38:



38

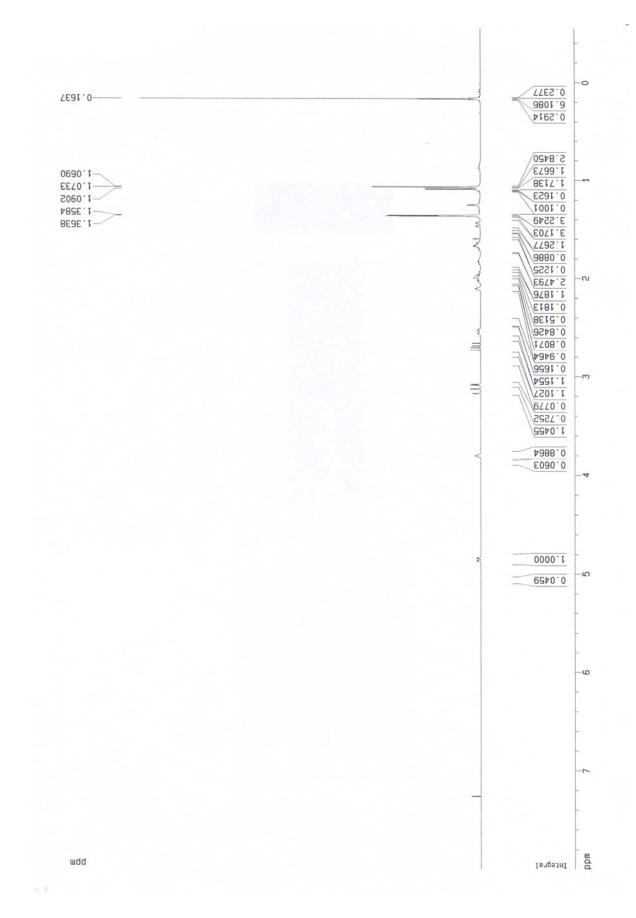
Recorded at 400 MHz(1 H) and 100 MHz (13 C) in CDCl₃:

¹H NMR spectra

¹³C NMR spectra

COSY spectra

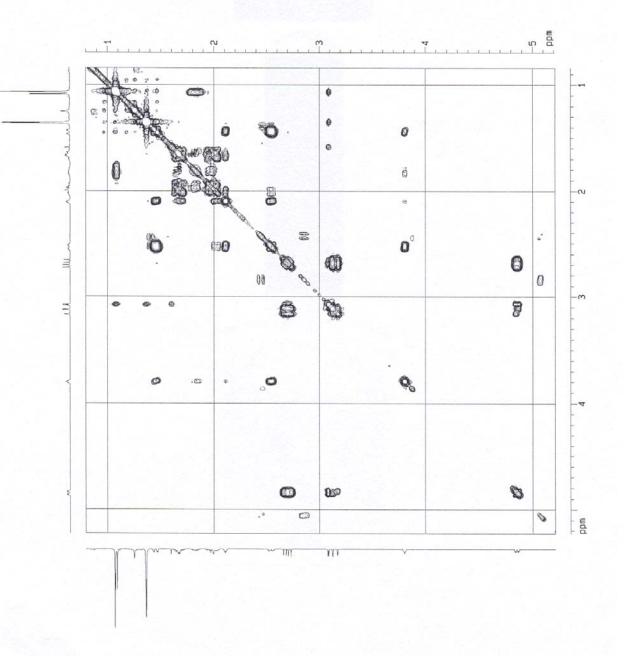
HMQC spectra



MANE ed22.2.250B1C NAME ed22.2.250B1C PROCNO 1	F2 - Acquisition Parameters Date 980223 Time 2.25 PULPADG 29.cpd SQLVENT 0.0485960 sec ATTOPES 0.476437 H7	3.000000 3.0000000 3.0000000	0.0031000 sec 7.0 usec 22.9 usec 31250.00 Mz 65536 544 544	Processing parameters 131072 0F 100.6138701 MHz EM EM 2.00 Hz 0	10 NNR plot parameters CX 21.95 cm C1.95 cm 245.986 ppm F1 24749.62 Hz F2 -64.607 ppm F2 -6500.38 Hz PPMCM 14.14864 ppm/c HZCM 1423.54907 Hz/cm
NAME EXPNO PROCNO	F2 - AC Date Time PULPROG SOLVENT AQ FIDPSC	DW RG NUCLEUS P31 P31	D2 SF01 SWH SS SWH SS SWH	SI SI SF SF SSB SSB SSB SSB SSB SSB SSB SSB S	
					-50
	5.28				
	12.86 57.07 25.98 27.07				
	37.17 35.76 29.70 27.07				
	99.95				
	94, 68 95, 08 95, 67 56, 77 00, 77				
					150
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	78.71S				multiple of the second s
	wdd				T Edd

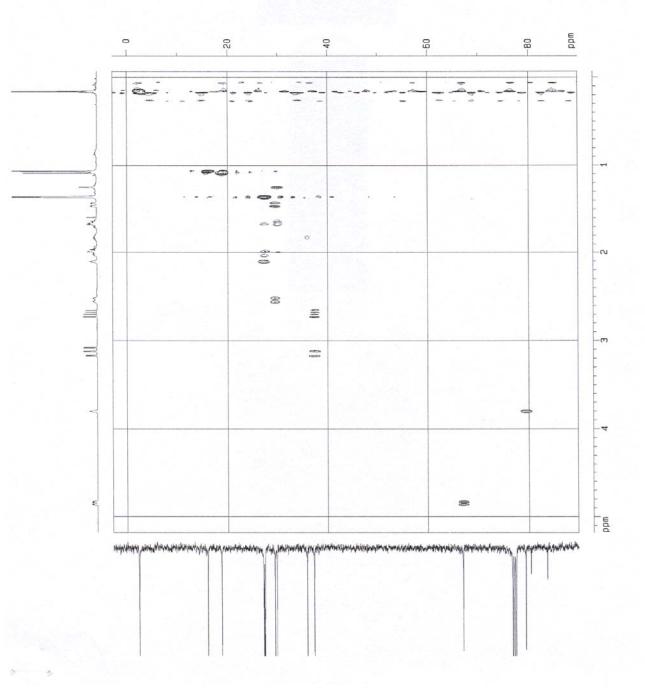
-

1 dB 0.00 sec .2.6 usec .0030 sec 26299 MHz 2824.96 Hz 1924 1024 Current Data Parameters ed22.2 2568100 - Processing parameters 512 0F Acquisition parameter 0F 400.1347353 NHz SINE 0.29232 ppm/cm 116.96681 Hz/cm 0.29416 ppm/cm 117.70334 Hz/cm 256 400.136 MHz 11.034687 Hz 7.050 ppm 0.0003540 sec 15.00 cm 15.00 cm 5.226 ppm 990.99 Hz 0.841 ppm 걒 5.212 ppm Sec Hz use rocessing para 512 400.1343986 N Acquisition P. 980223 00.00 0.00 20 NMR plot par 400.13 F2 - A NAME EXPNO PROCND Date Time PULPPDG SOLVENT A0 DM RG NUCLEUS ND0 TD SFD1 FIDRES SN CX2 CX1 F2PLD F2PLD F2PLD F2PLD F2PLD F1PLD F1PLD F1PLD F1PLD F1PLC F2PPMCN F1PCCN FIDRES P1 DE SSF01 DIS SSF01 DISS SIT NC2 SSF SSB SSB SSB SSB SI MCS SSB MCS SSB MCS H-

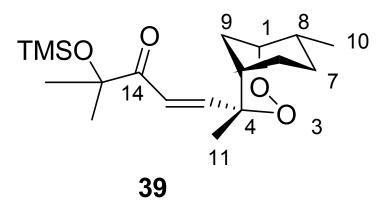


- S74 -





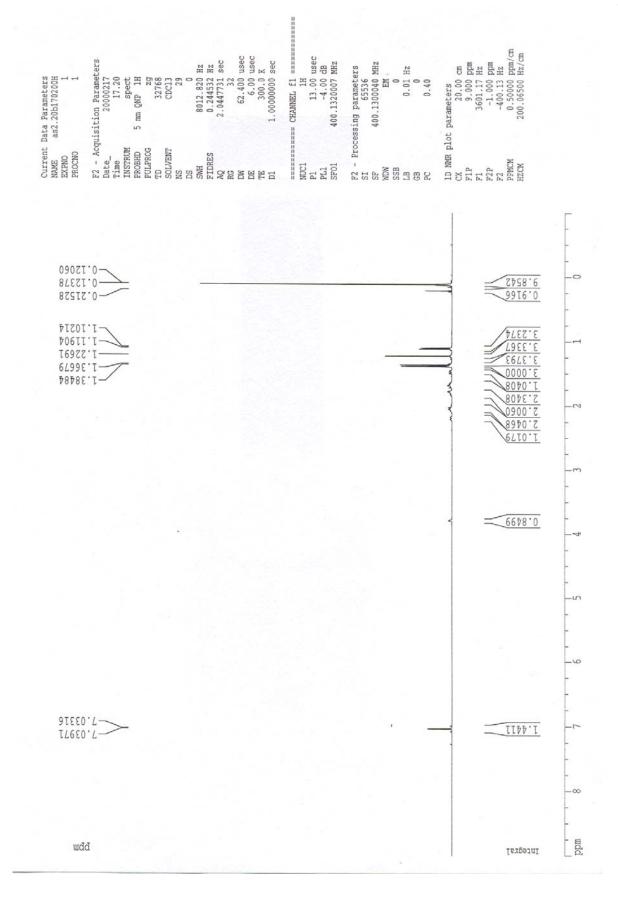
Unsaturated ketone **39**:



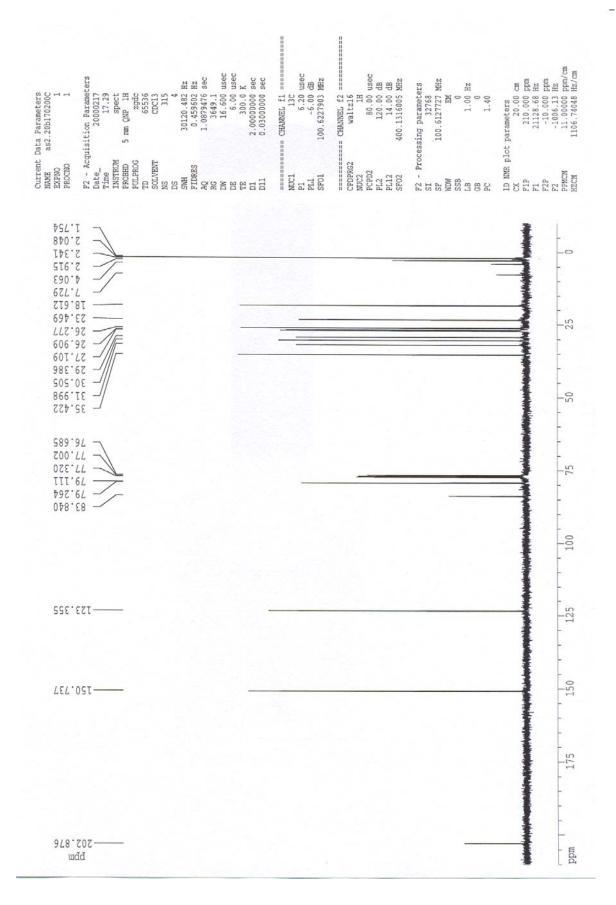
Recorded at 400 MHz(¹H) and 100 MHz (¹³C) in CDCl₃:

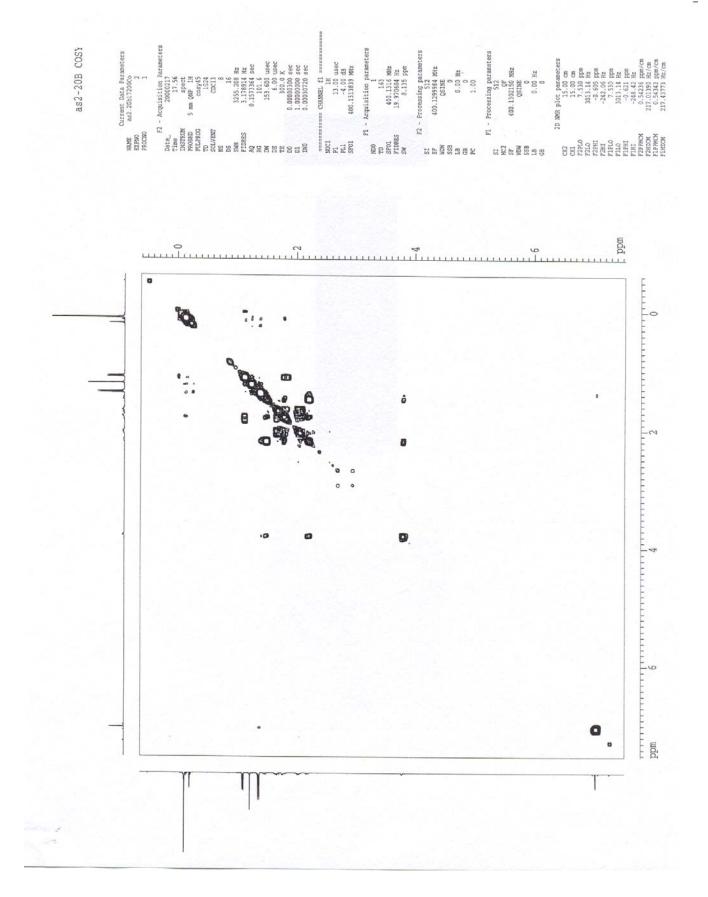
¹H NMR spectra

¹³C NMR spectra

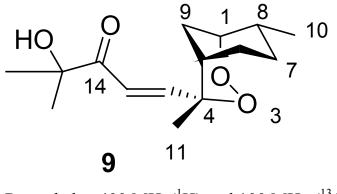


- S77 -





Unsaturated ketone 9:



Recorded at 400 MHz(¹H) and 100 MHz (¹³C) in CDCl₃:

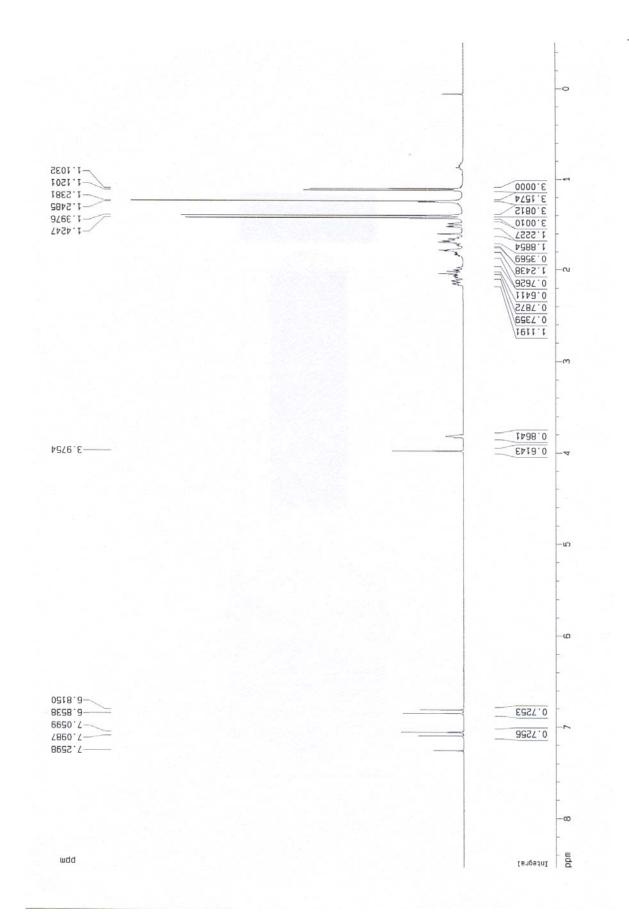
¹H NMR spectra

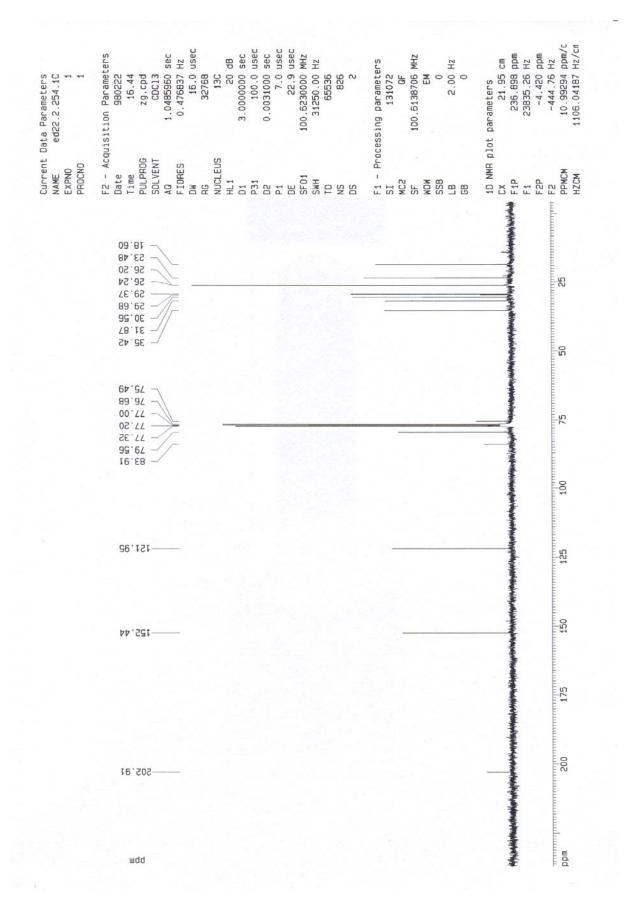
¹³C NMR spectra

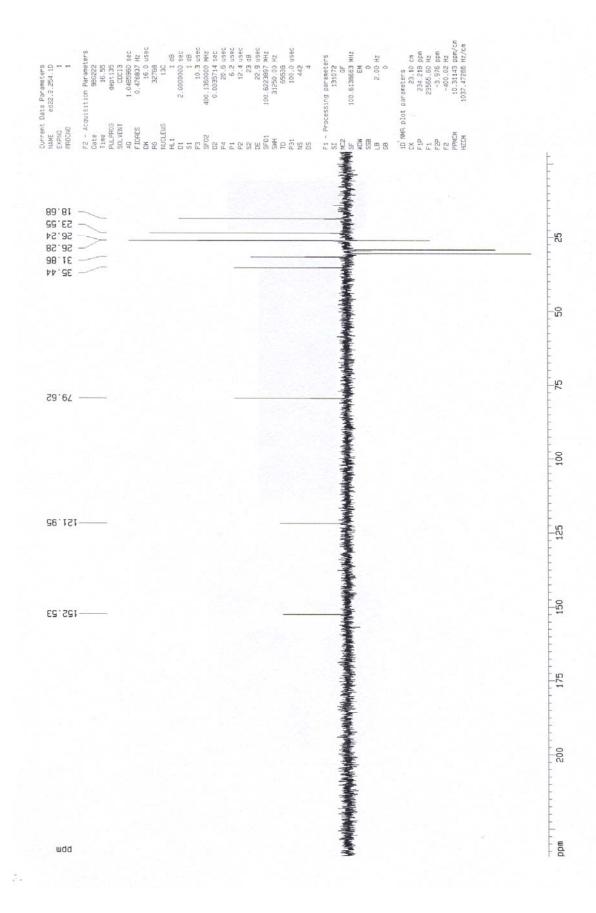
DEPT spectra

COSY spectra

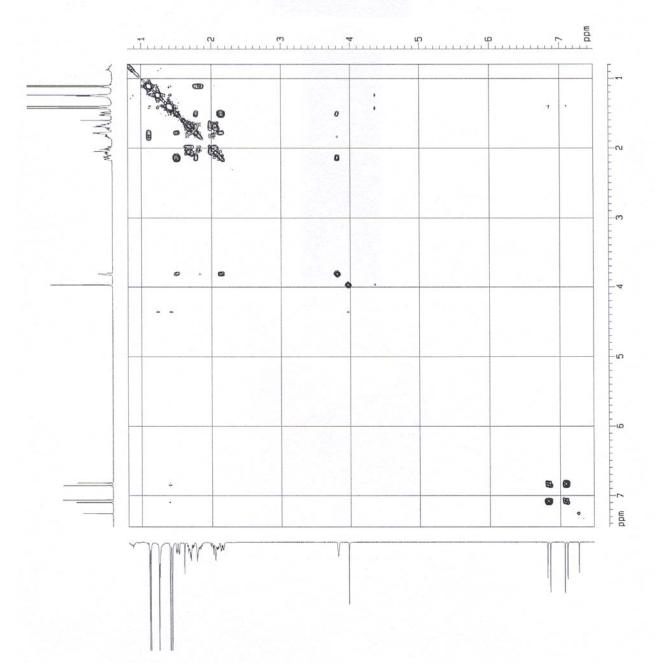
HMQC spectra

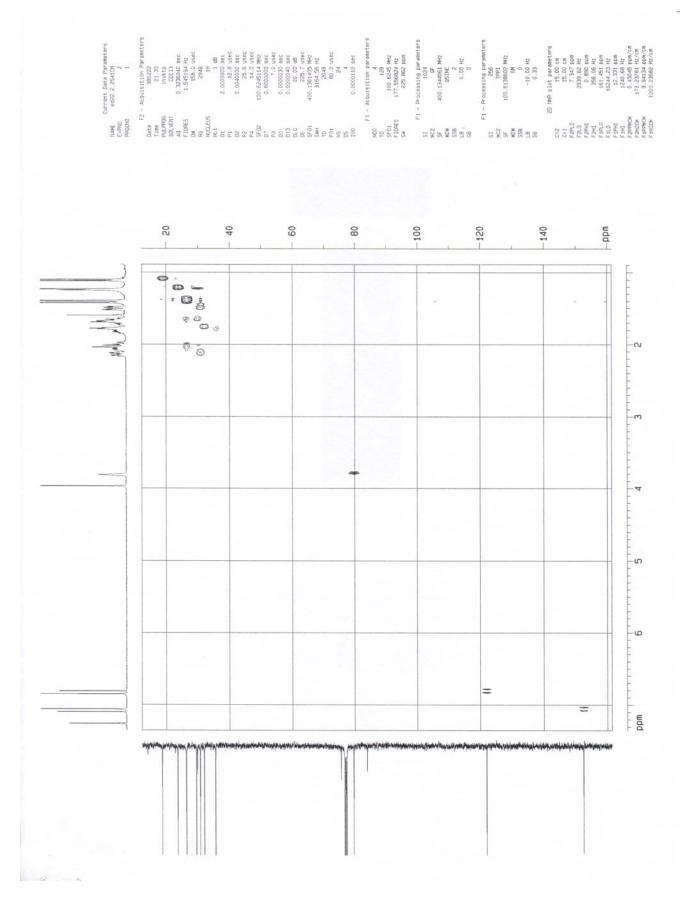




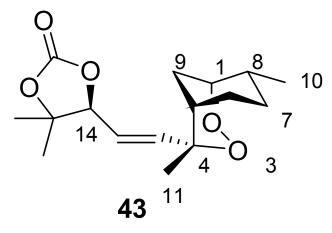


Parameters .Co 1	cm Parameters 20222 0.01 0.01 0.01 0.01 0.01 0.01 0.0	4 (316) sec 1 256 256 11365 Hz 11365 Hz 11365 Hz 11365 Hz 11367 Hz 512 512 6 512 6 7 1367 Hz 512 0 0 0 0 0 0 0 0 0	00 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	450 p 99 b 99 b 99 b 99 b 90 b 90 b 90 b 90 b
urrent Data Bd22.2.2541	- Acoulstine F 900225 900225 900209 900209 900308 9195.0 195.0 195.0 195.0 195.0 195.0 10000000 0 10000000 12.6 0 00000000 12.6 10000000 1354.455 100012.6 10000000	0.0003160 - Acquisition 255 400.135 7.909 7.909 7.909 7.909 12.351545 7.909 7.909 7.909 7.909 7.909 7.909 7.909 7.9000 7.90000 7.90000 7.90000 7.90000 7.90000 7.9000000000000000000000000000000000000	- Processing 5 400.13422 5 19 0.0 0.0 20 MMR plot pu	7. 29802 0. 317 7. 2933 203 203 203 203 177,559 0.44
C NAME EXPND PROCND	F2 Date Time Sourent A0 Sourent A0 NUD-EUS F1DPES F	DS 1000 1000 1000 1000 1000 1000 1000 10	SSI	F2PL0 F2PL0 F2PHI F2PHI F2PHI F1PL0 F1PL0 F1PL0 F1PL0 F1PL0 F1PL0 F1PL0 F1PL0 F1PL0 F1PL0 F1PL0





Carbonate **43**:

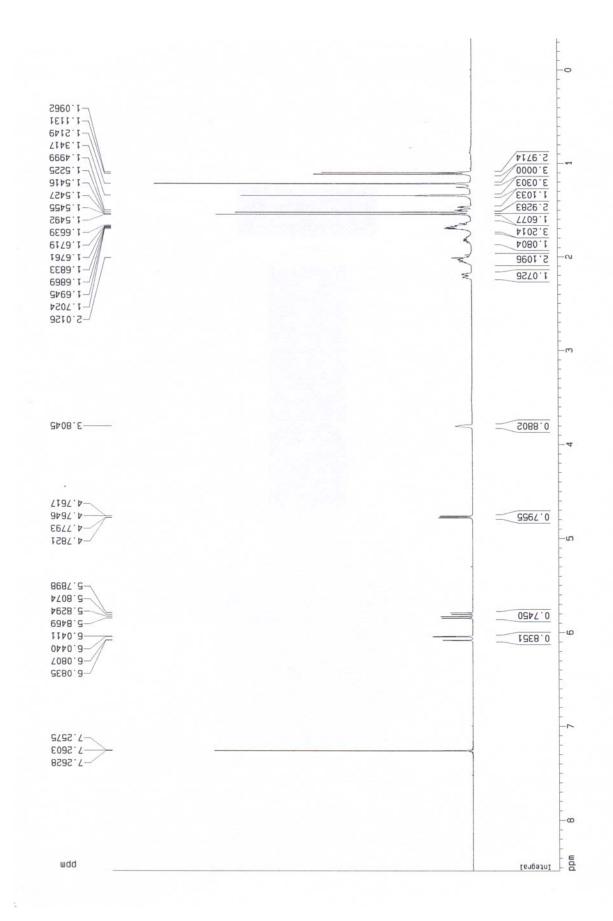


Recorded at 400 MHz(¹H) and 100 MHz (¹³C) in CDCl₃:

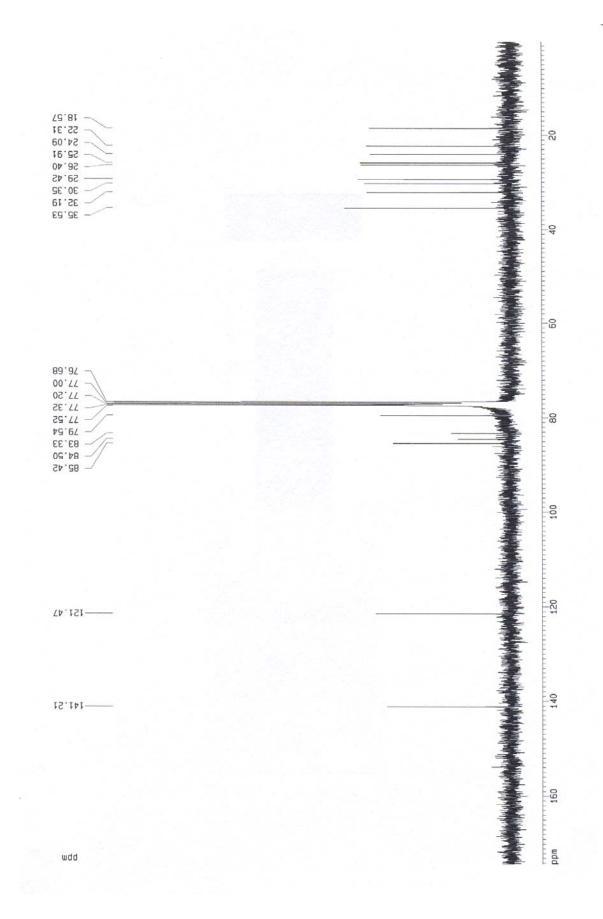
¹H NMR spectra

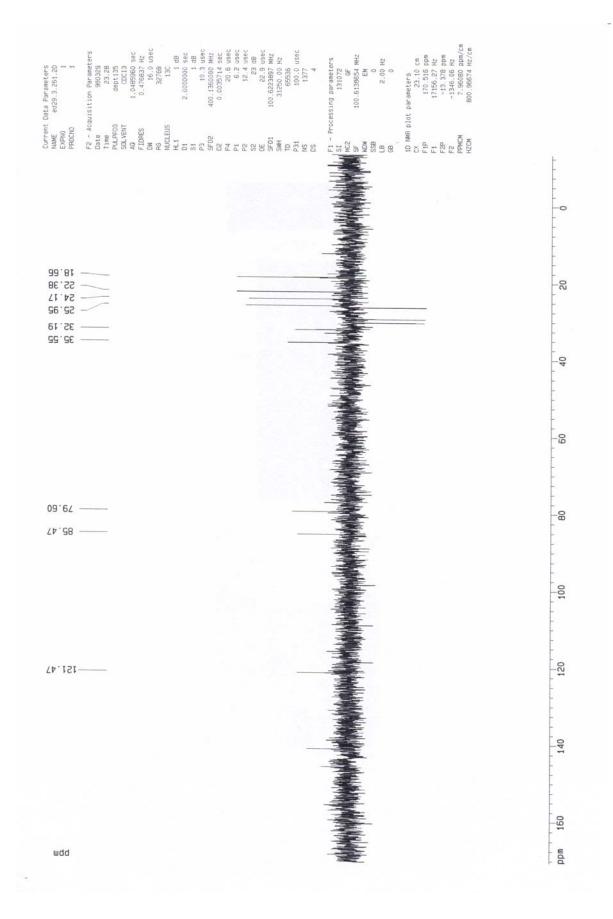
¹³C NMR spectra

DEPT spectra



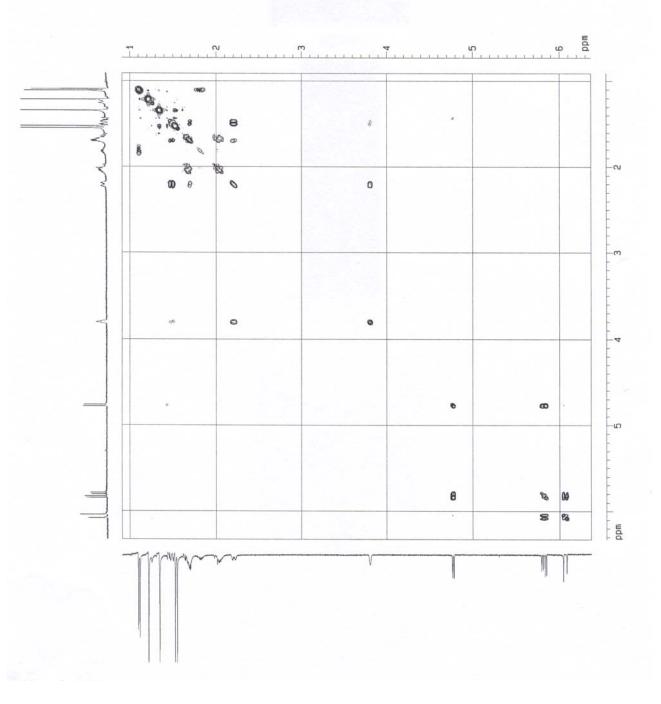
- S87 -



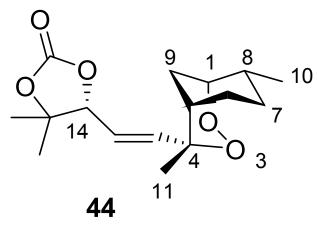


Current Data Parameters NAME ed29.3.261.2Co EXPND 2 PROCND 1

1 200 2 1	ition Parameters 980330	1.14 CDSy45	66280 sec	43136 HZ 192.0 usec	2048 1H		1.5000000 sec			4.17		ष ग	0.0003840 sec	tion parameters	1 256	359	6.508 ppm	ing parameters	512	43939 MHZ	SINE	0.00 Hz	ing parameters		5	SINE PRIZ	0	0.00 Hz 0	para	8	15.00 cm	85	912	ZH 10.00E	B0.74	912			36355 ppm/cm	46700 Hz/cm
ed29,3,26	- Acquisit	C	0.19	6 1			1.50	0.00	S ADA	28			0.00	- Acquisit		400.1	1.01	- Process		400.13			- Process			400,13			DIN PIO			8		τ, T	52		ē	144	0	145.
NAME EXPND PROCND	F2 Date	PULPP06	AQ Fronce	DW	RG NUCI FLIS	HLI	5 5	18	DE	SNH	2 5	2 2	ONI	F1	100	SF01	SW	Ĩ.	IS	5	NDM	8 8	ε.	Is	WC2	MON	SSB	8		CX5	CX1 Foot n	F2LD	F2PHI	F2HI	FILD	F 1PHI	F1HI	F 2H7CM	F 1PPMCM	F1HZCM



Carbonate 44:

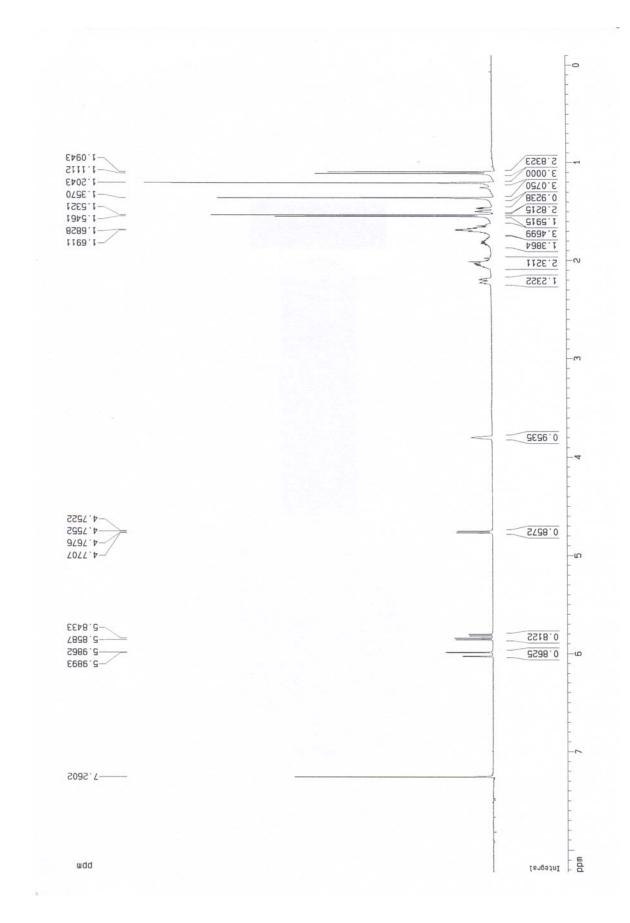


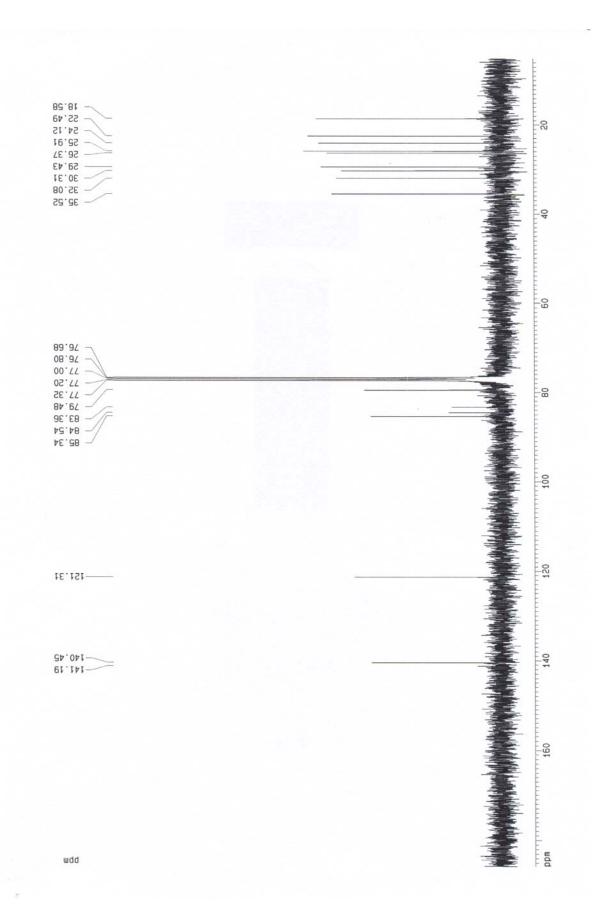
Recorded at 400 MHz(¹H) and 100 MHz (¹³C) in CDCl₃:

¹H NMR spectra

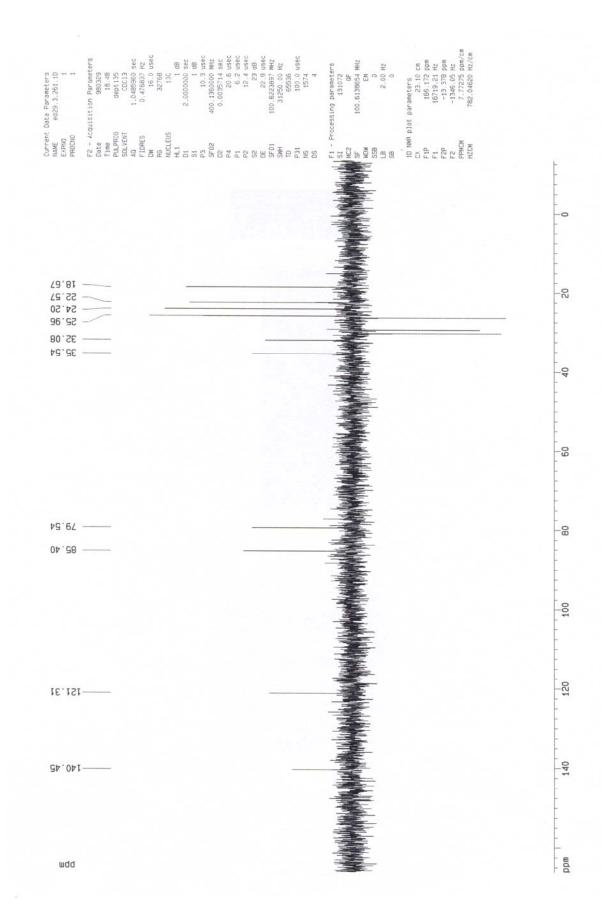
¹³C NMR spectra

DEPT spectra

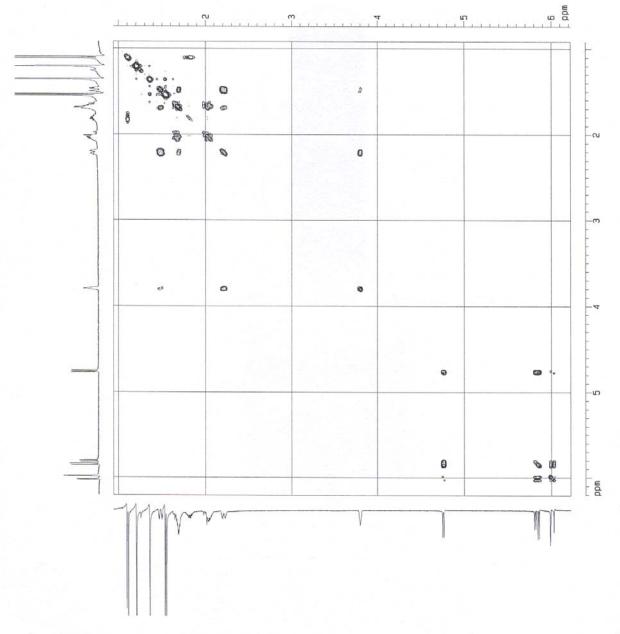




- S93 -

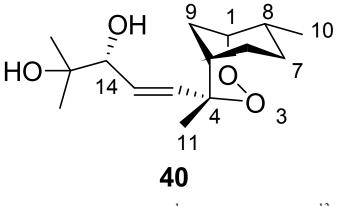








Epimer of yingzhaosu A **40**:

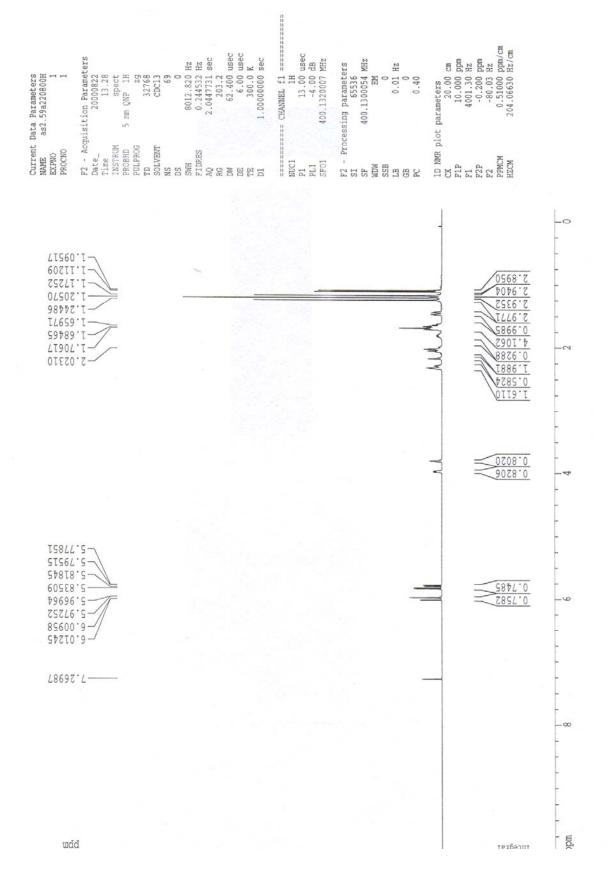


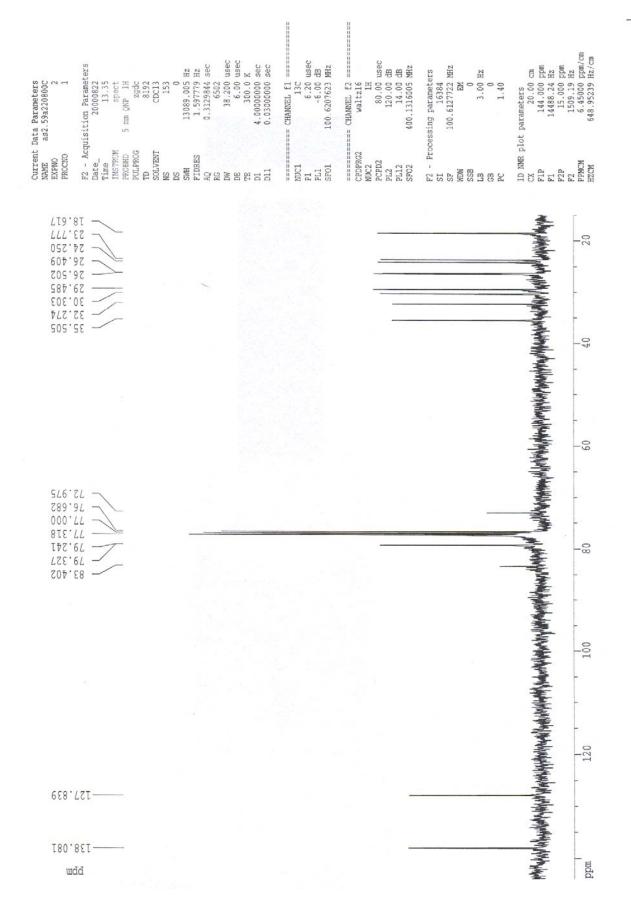
Recorded at 400 MHz(1 H) and 100 MHz (13 C) in CDCl₃:

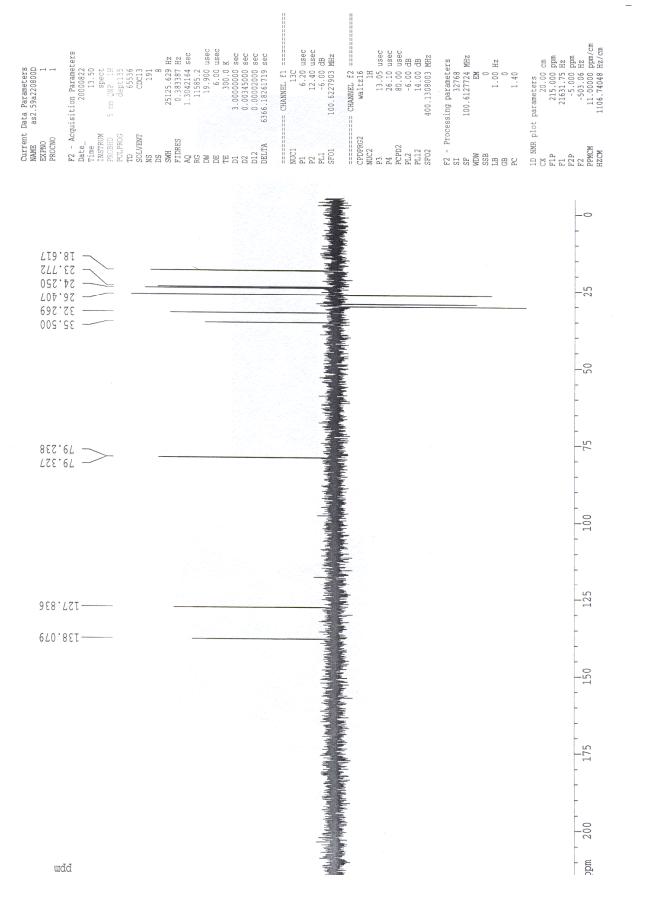
¹H NMR spectra

¹³C NMR spectra

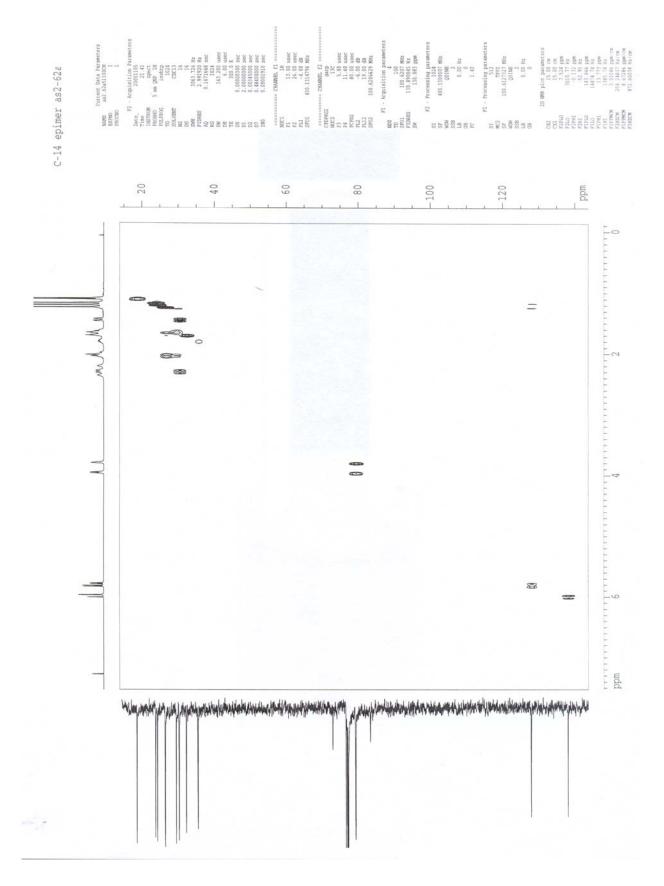
DEPT spectra



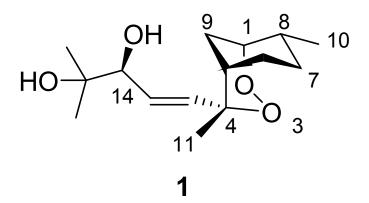




- S99 -



Yingzhaosu A (1):



Recorded at 400 MHz(¹H) and 100 MHz (¹³C) in CDCl₃:

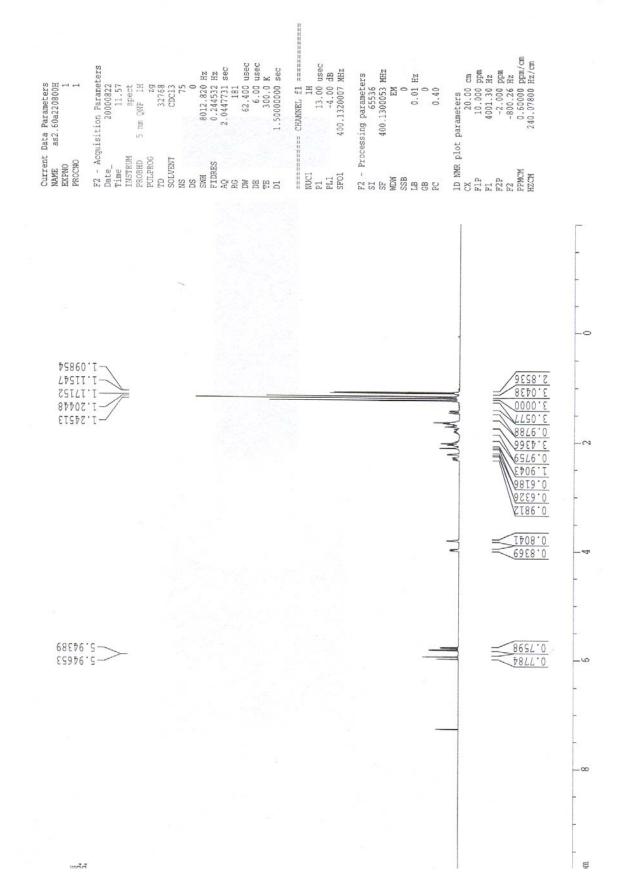
¹H NMR spectra

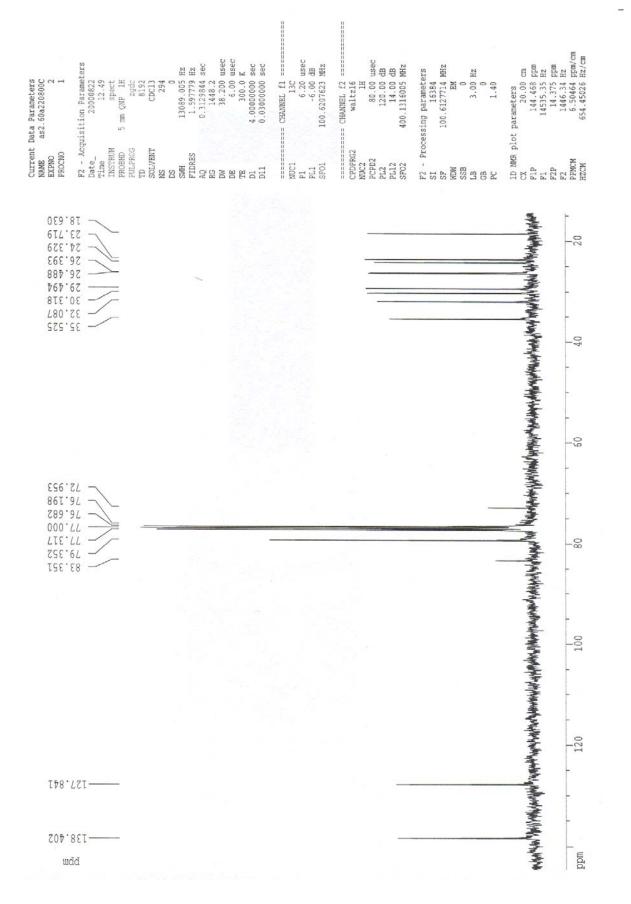
¹³C NMR spectra

DEPT spectra

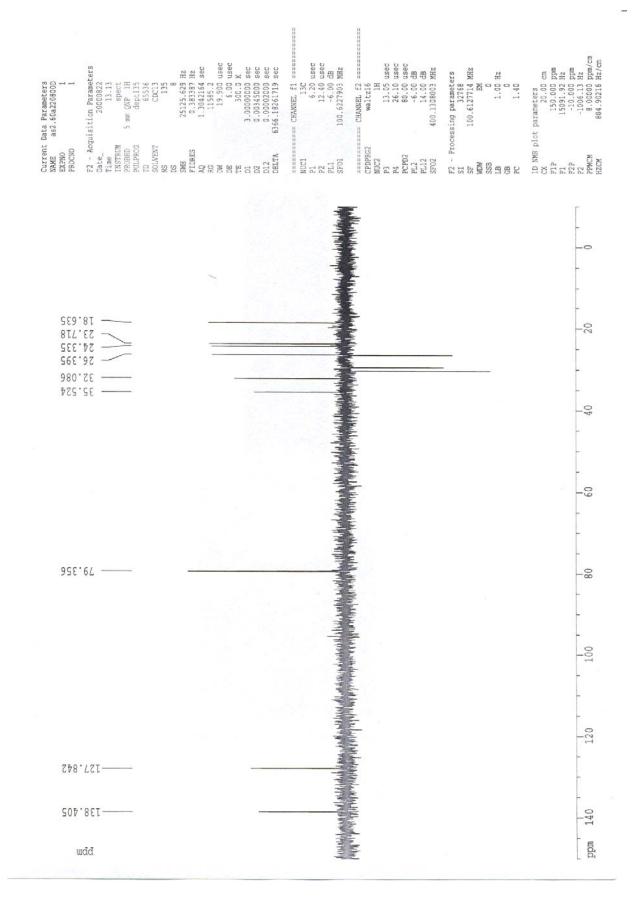
COSY spectra

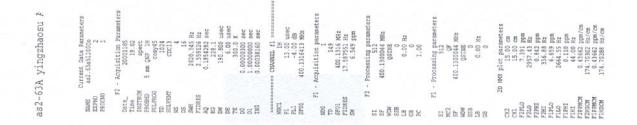
HMQC spectra

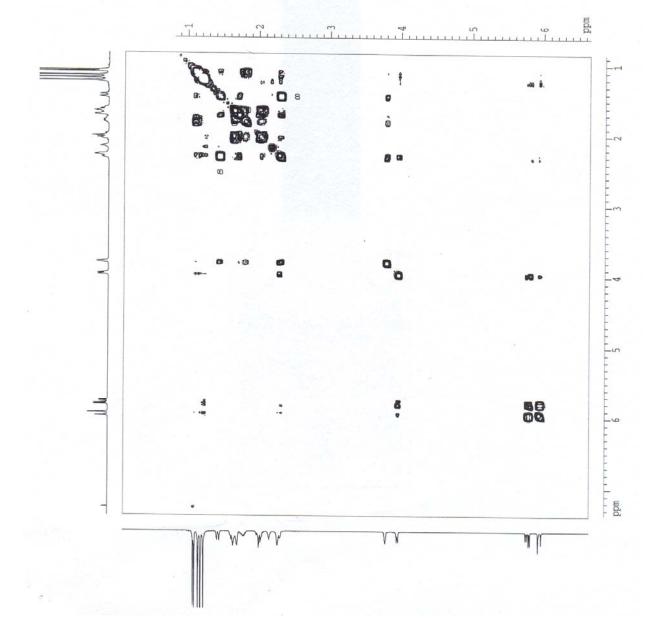


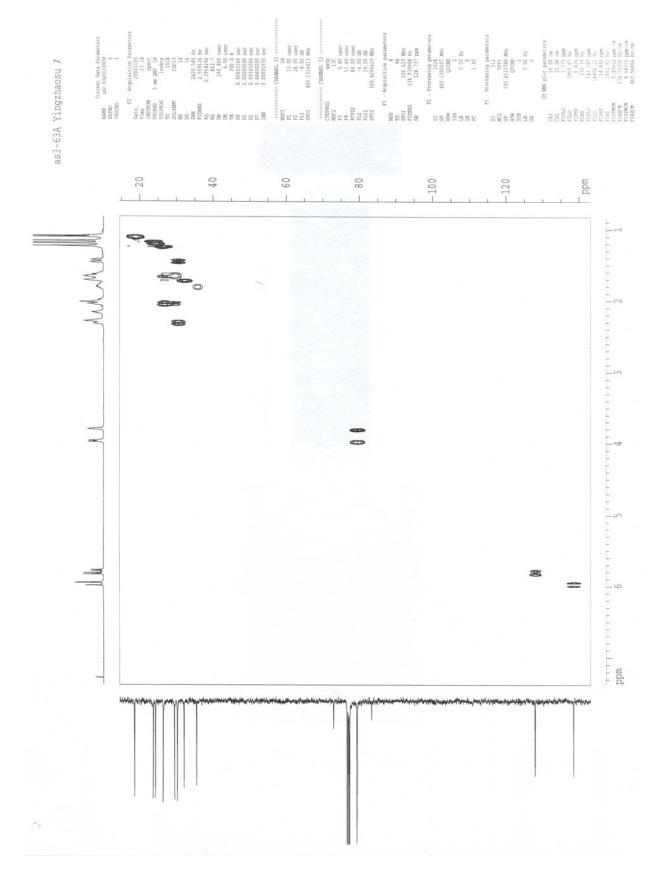


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X-ray structural data for yingzhaosu A (1)

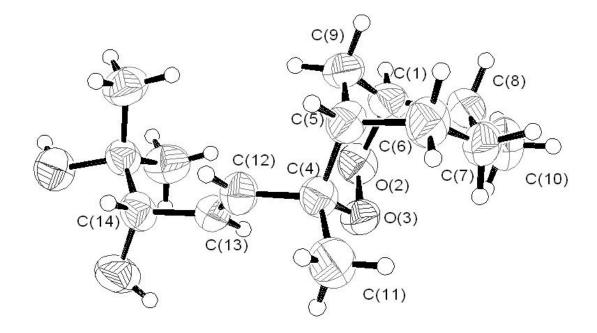


Figure S1. Perspective view (ORTEP) of the X-ray structure of yingzhaosu A (1). Thermal ellipsoids are drawn at the 50% probability level. For more details see the attached CIF file.