- Pag S2. Experimental part: general Pag S3. <sup>1</sup>H and <sup>13</sup>C NMR spectra of compound **6** Pag S4. <sup>1</sup>H and <sup>13</sup>C NMR spectra of compound **7** Pag S5. <sup>1</sup>H and <sup>13</sup>C NMR spectra of compound **8** Pag S6. <sup>1</sup>H and <sup>13</sup>C NMR spectra of compound **9** Pag S7. <sup>1</sup>H and <sup>13</sup>C NMR spectra of compound **10**

**General**: Column chromatography: Silica Gel 60. Analytical grade solvents were used.  $Na_2SO_4$  was used to dry solutions before the evaporation. Reactions were monitored by TLC on Si 60  $F_{254}$  (0.25 mm) plates, which were visualized by UV inspection and/or staining with  $(NH_4)_2MoO_4$  and heating.

**Spectroscopy**: <sup>1</sup>H NMR (500 MHz) and <sup>13</sup>C NMR (125 MHz) were recorded at room temperature on a spectrometer endowed of an inverse multinuclear 5 mm probehead equipped with a shielded gradient coil. The spectra were recorded in CDCl<sub>3</sub>, and the solvent signals (7.26 and 77.0 ppm, respectively) were used as reference. The chemical shifts ( $\delta$ ) are given in ppm, and the coupling constants (J) in Hz. COSY, HMQC and HMBC experiments were recorded with gradient enhancements using sine shaped gradient pulses. For the 2D heteronuclear correlation spectroscopy the refocusing delays were optimised for  ${}^{1}J_{\text{CH}}$ =145 Hz and  ${}^{n}J_{\text{CH}}$ =10 Hz. The raw data were transformed and the spectra were evaluated with the standard XWIN-NMR software (rev. 010101).

ESIMS<sup>n</sup> spectra were recorded on a LCQ ion trap mass spectrometer equipped with a NT<sup>TM</sup> data system and an Electrospray interface (ESI). Mass spectrometer conditions were optimized in order to achieve maximum sensitivity. ESI conditions: source voltage 4.5 kV, sheath gas flow rate 60 au, source current 80 mA, capillary voltage -38 V and capillary temperature 200°C. Full scan spectra from 150 to 2000 u in the negative ion mode were obtained (scan time 1 s). Ion trap conditions: acquisition in automatic gain control with a max-inject time of 200 msec. For the MS<sup>n</sup> analyses the [M-H] molecular ions were isolated with an isolation width of 3 m/z units and fragmented using an activation amplitude of 30% for MS<sup>2</sup> experiments and 30% for MS<sup>n</sup> experiments. HREIMS spectra were taken at 70 eV. FAB mass spectra were recorded in positive mode.

Horse radish peroxidase (HRP) was purchased from Roche as lyophilized powder (# 127 361, M<sub>r</sub> ~44000 g/mol, 220 U/mg); catalase from bovine liver was purchased from Fluka as crystalline suspension in water (# 60630, M<sub>r</sub> ~ 240000 g/mol, 65000 U/mg, 20 mg/mL); albumin from eggs was purchased from Merck as dry fine powder (# 967, M<sub>r</sub> ~45000 g/mol); human serum albumin (HSA) was purchased from Fluka as dry powder (# 05430, M<sub>r</sub> ~68000 g/mol).











