Supporting Information for

Design of Controllable Bio-Inspired Chiroptic Self-Assemblies

Kai Tao[†], Guy Jacoby[‡], Luba Burlaka[§], Roy Beck[‡] and Ehud Gazit^{,†,}*

[†]Department of Molecular Microbiology and Biotechnology, George S. Wise Faculty of Life Sciences, Tel Aviv University, Tel Aviv 6997801, Israel

[‡]School of Physics and Astronomy, Faculty of Exact Sciences, Tel-Aviv University, Tel Aviv, 6997801, Israel

[§]Institute for Nanotechnology and Advanced Materials, Bar-Ilan University, Ramat Gan, 5290002, Israel

^{II}Department of Materials Science and Engineering, Iby and Aladar Fleischman Faculty of Engineering, Tel Aviv University, Tel Aviv 6997801, Israel

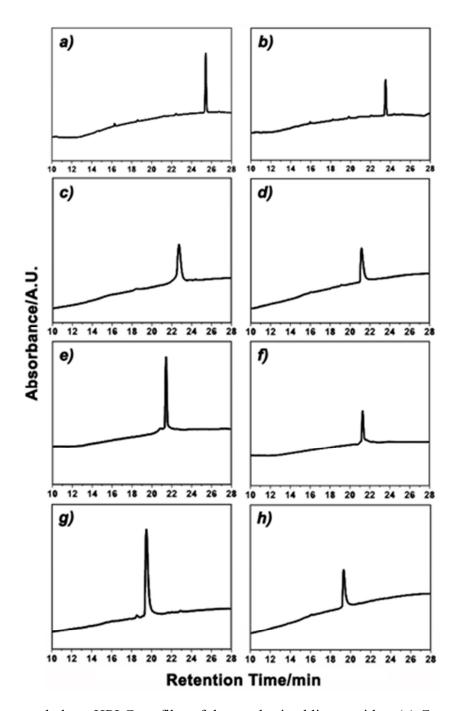
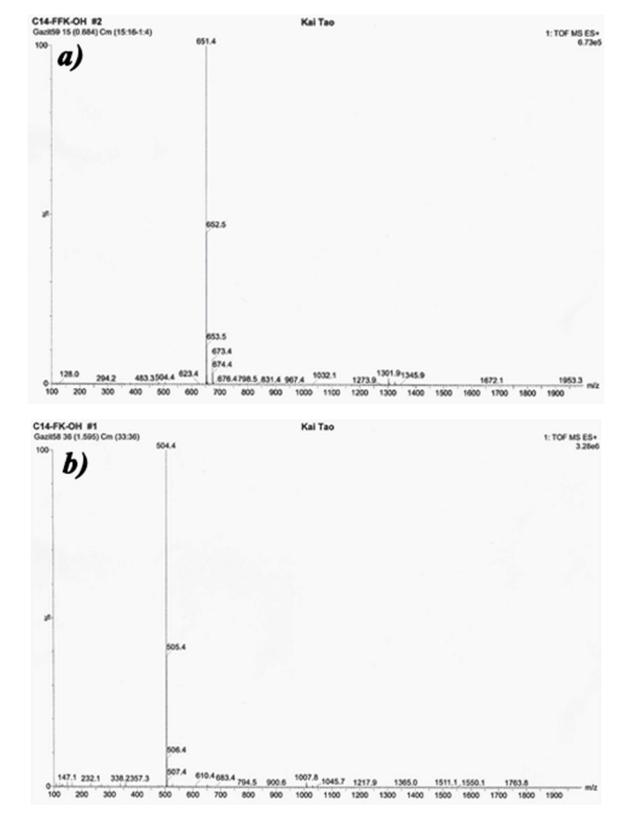


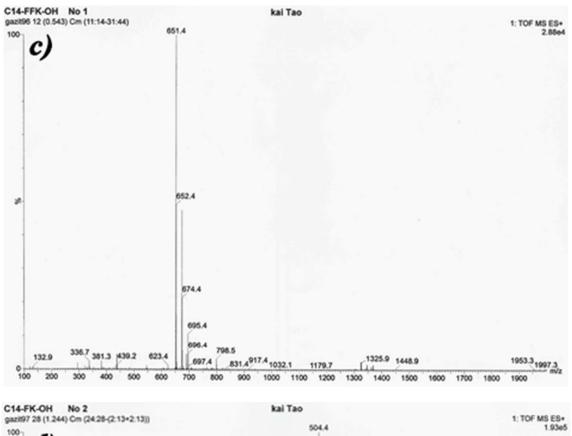
Figure S1. Reversed phase HPLC profiles of the synthesized lipopeptides: (a) C_{14} -L-FFK, (b) C_{14} -L-FK, (c) C_{14} -D-FFK, (d) C_{14} -D-FK, (e) C_{14} -L-YYK, (f) C_{14} -L-YK, (g) C_{14} -D-YYK, (h) C_{14} -D-YK. Note that a gradient elution mode was employed, as shown in Table S1. The monitoring wavelength was set at 214 nm, and the flow rate was 1 mL min⁻¹.

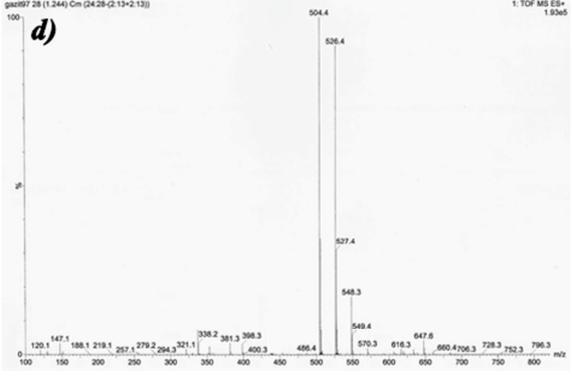
Time/min	A% 0.1% (v/v) TFA in water	B% 0.1% (v/v) TFA in acetonitrile
4	90	10
25	10	90
32	90	10

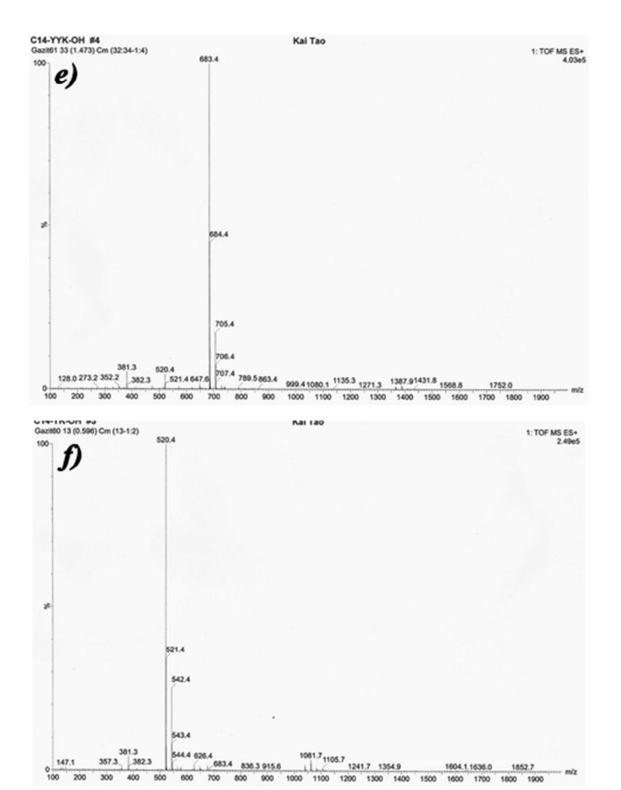
 Table S1. HPLC gradient elution conditions

It is evident that the lipopeptide peaks highly dominate the profiles and their relative area is more than 95%, indicating the high purities of the synthesized compounds.









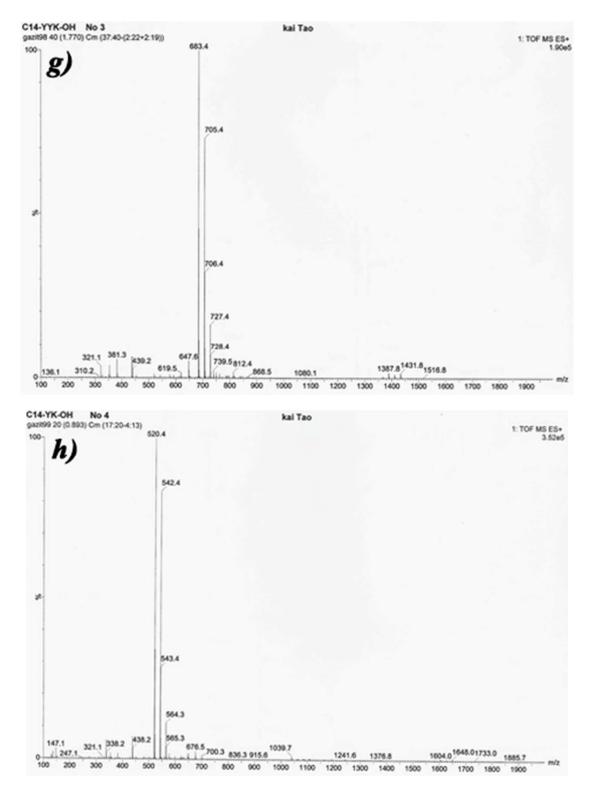


Figure S2. MS spectra of the synthesized lipopeptides: (a) C_{14} -*L*-FFK, (b) C_{14} -*L*-FK, (c) C_{14} -*D*-FK, (d) C_{14} -*D*-FK, (e) C_{14} -*L*-YYK, (f) C_{14} -*L*-YK, (g) C_{14} -*D*-YYK, (h) C_{14} -*D*-YK.

The observed molecular masses are consistent with the calculated ones, demonstrating the high purities of the lipopeptide samples, which are well in accordance with the HPLC analysis described above:

(a) C₁₄-*L*-FFK:

Expected masses $[M+H]^+=651.9$, $[M+Na]^+=673.9$;

Observed masses $[M+H]^+=651.4$, $[M+Na]^+=673.4$.

(b) C₁₄-*L*-FK:

Expected masses $[M+H]^+=504.7;$

Observed masses $[M+H]^+=504.4$.

(c) C₁₄-*D*-FFK:

Expected masses [M+H]⁺=651.9, [M+Na]⁺=654.9;

Observed masses $[M+H]^+=651.4$, $[M+Na]^+=674.4$.

(d) C₁₄-*D*-FK:

Expected masses $[M+H]^+=504.7$, $[M+Na]^+=526.7$;

Observed masses $[M+H]^+=504.4$, $[M+Na]^+=526.4$.

(e) C₁₄-*L*-YYK:

Expected masses [M+H]⁺=683.9, [M+Na]⁺=705.9;

Observed masses $[M+H]^+=683.4$, $[M+Na]^+=705.4$.

Expected masses [M+H]⁺=520.7, [M+Na]⁺=542.7;

Observed masses $[M+H]^+=520.4$, $[M+Na]^+=542.4$.

(g) C₁₄-*D*-YYK:

Expected masses [M+H]⁺=683.9, [M+Na]⁺=705.9;

Observed masses $[M+H]^+=683.4$, $[M+Na]^+=705.4$.

(h) C₁₄-*D*-YK:

Expected masses [M+H]⁺=520.7, [M+Na]⁺=542.7;

Observed masses $[M+H]^+=520.4$, $[M+Na]^+=542.4$.

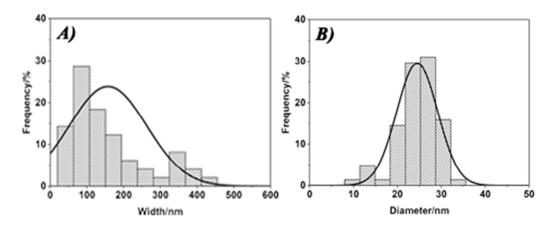


Figure S3. Statistical width distribution of nanoribbons self-assembled by C_{14} -FFK (A) and diameter distribution of nanofibers self-assembled by C_{14} -FK (B).

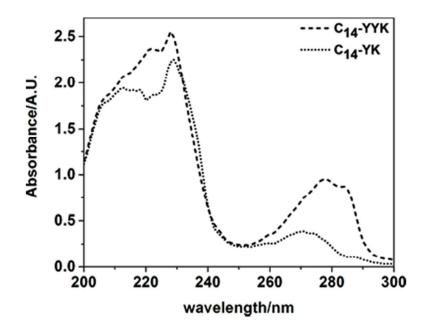


Figure S4. UV-Vis absorption spectra of 1.0 mg mL⁻¹ C_{14} -YYK (dashed lines) and C_{14} -YK (dotted lines) in HFIP/water (1/49, v/v) at pH 7.0. It can be clearly seen that the phenolic groups present an intensive absorption at around 280 nm.

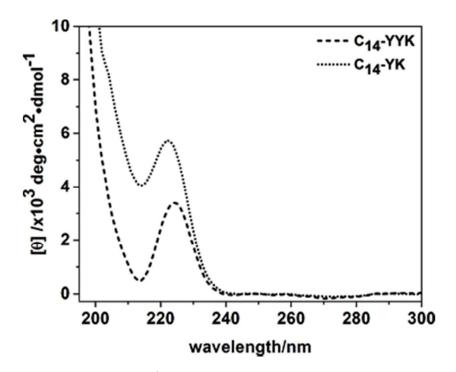


Figure S5. CD spectra of 1.0 mg mL⁻¹ C_{14} -YYK (dashed lines) and C_{14} -YK (dotted lines) in HFIP. Note that there is no obvious CD signal for free phenolic moieties.

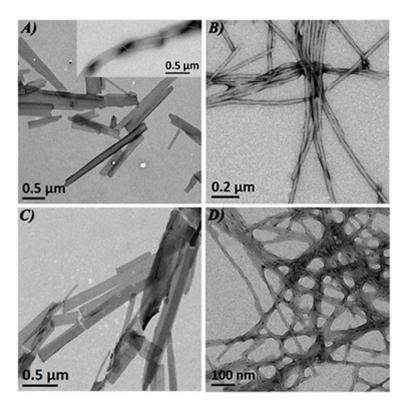


Figure S6. TEM micrographs representing the nanoribbons self-assembled by C_{14} -D-FFK (A) and C_{14} -D-YYK (C), and the nanofibers self-assembled by C_{14} -D-FK (B) and C_{14} -D-YK (D). The inset in (A) shows the right-handed twisting of the nanoribbons.