

# **Mutation of the Arg191 in FtsZ impairs cytokinetic abscission of *Bacillus subtilis* cells**

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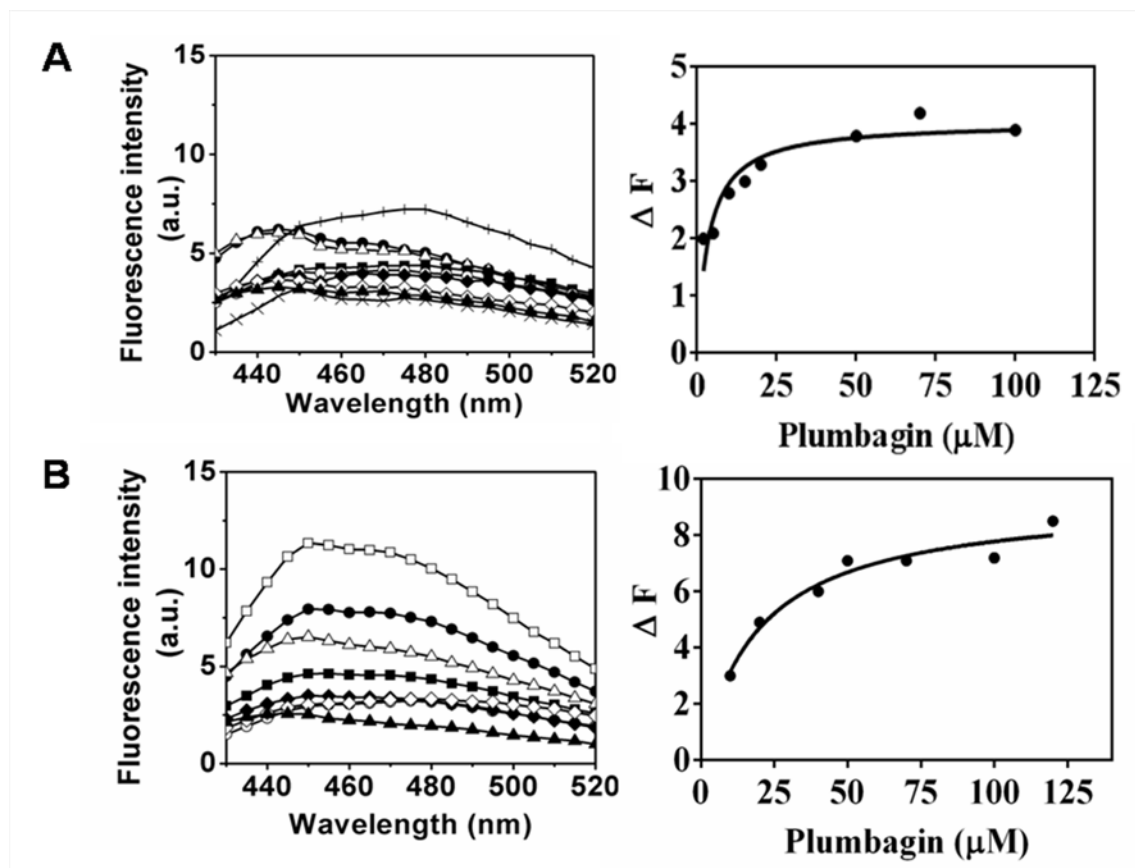
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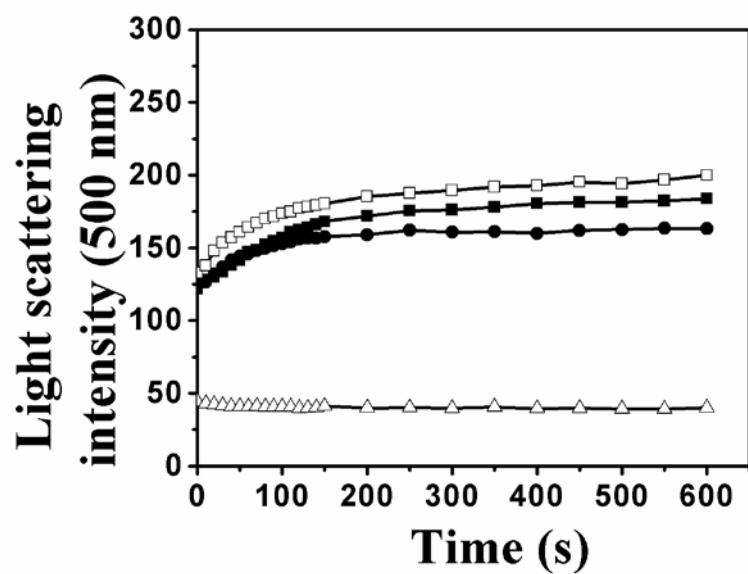
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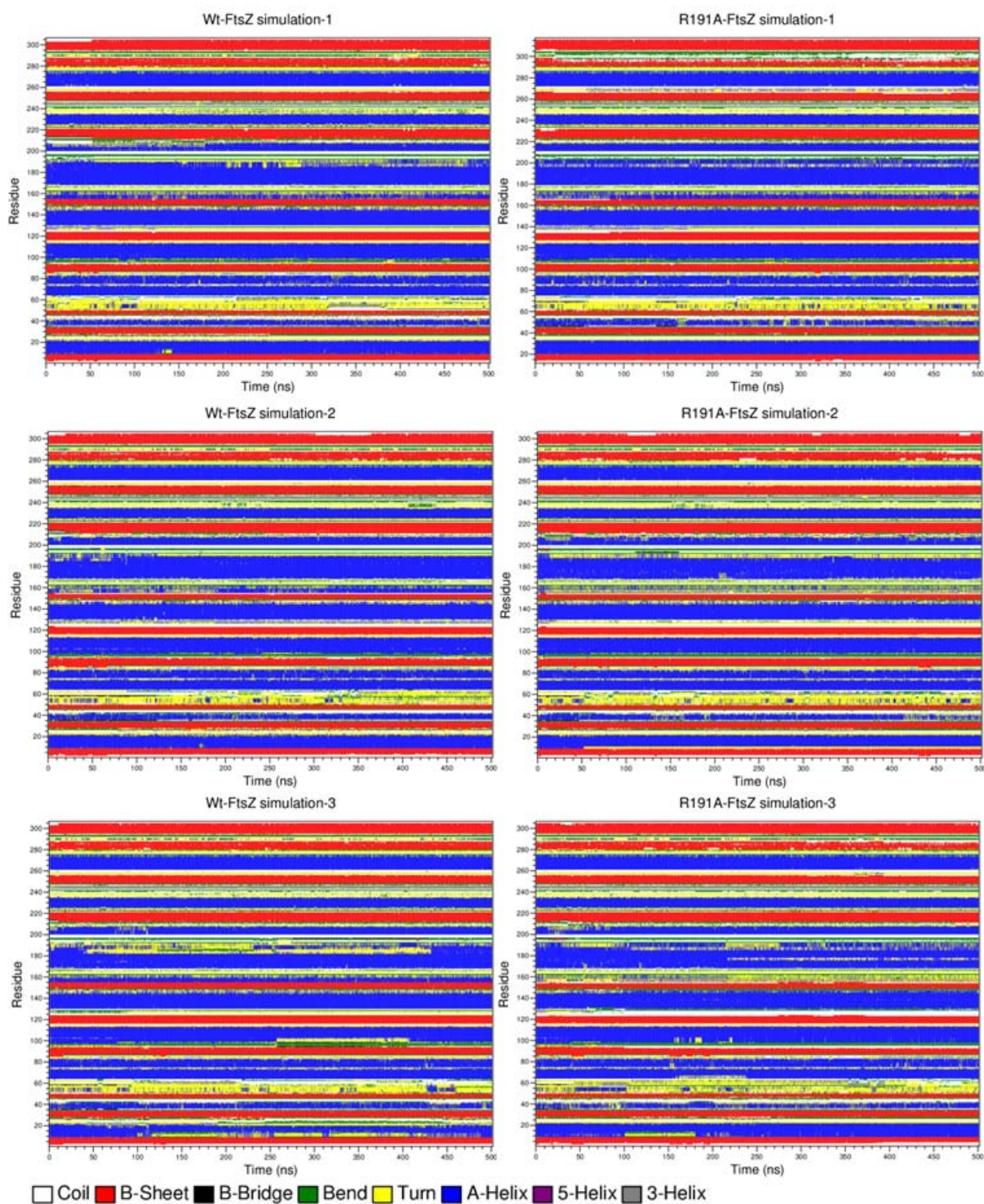
## Supporting Figures



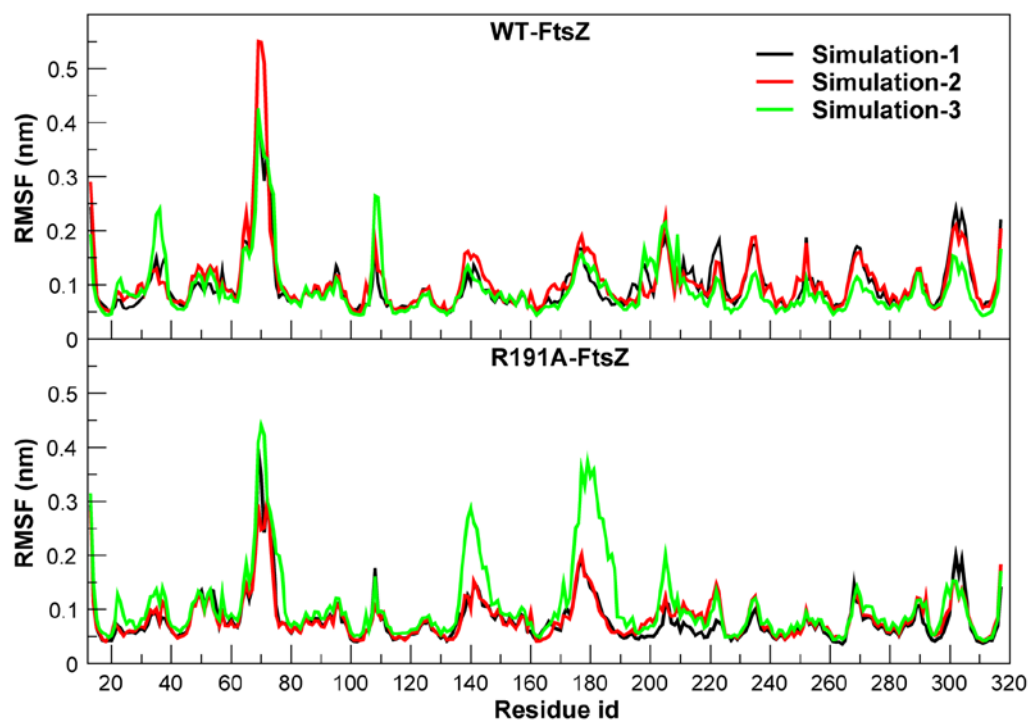
**Fig. S1.** Effects of plumbagin on the fluorescence intensity of ANS in the presence of WT-FtsZ or with R191A-FtsZ. (A) WT-FtsZ (2  $\mu M$ ) (+) was incubated with 2 ( $\bullet$ ), 5 ( $\Delta$ ), 10 ( $\blacksquare$ ), 15 ( $\circ$ ), 20 ( $\blacklozenge$ ), 50 ( $\diamond$ ), 70 ( $\blacktriangle$ ), and 100  $\mu M$  ( $\times$ ) plumbagin at 25  $^{\circ}C$  for 10 min. (B) R191A-FtsZ (2  $\mu M$ ) ( $\square$ ) was incubated with 10 ( $\bullet$ ), 20 ( $\Delta$ ), 40 ( $\blacksquare$ ), 50 ( $\circ$ ), 70 ( $\blacklozenge$ ), 100 ( $\diamond$ ) and 120  $\mu M$  ( $\blacktriangle$ ) plumbagin at 25  $^{\circ}C$  for 10 min. Then, ANS (30  $\mu M$ ) was added in the samples and incubated for 30 min at 25  $^{\circ}C$ . The fluorescence spectra were recorded in the range of 430-520 nm using 350 nm as an excitation wavelength. The fluorescence spectra of plumbagin (0-120  $\mu M$ ) was also recorded as a blank. The change in fluorescence at 475 nm was calculated by subtracting blank from the respective data sets. A dissociation constant of the binding interaction of plumbagin with WT-FtsZ and with R191A-FtsZ was determined from the fluorescence change data as described previously<sup>1</sup>.



**Fig. S2.** The effect of plumbagin on the assembly kinetics of R191A-FtsZ. R191A-FtsZ (12 μM) was incubated without (□) or with 20 (■) and 40 μM (●) plumbagin for 15 min on ice and then, the assembly kinetics was monitored by adding 1 mM GTP at 37 °C. The light scattering intensity of buffer (Δ) [25 mM PIPES (pH 6.8), 50 mM KCl and 10 mM MgCl<sub>2</sub>] was also monitored as a blank.

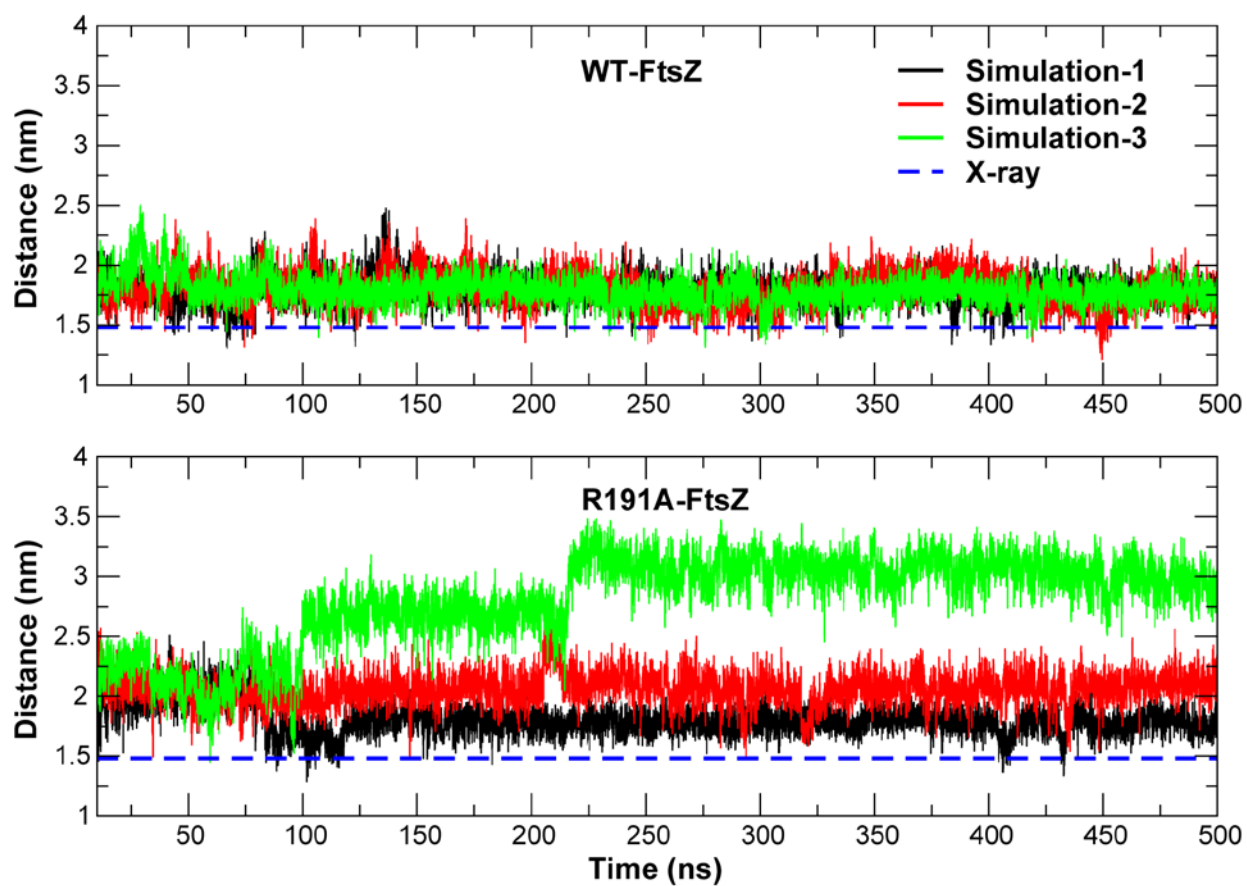


**Fig. S3:** Secondary structures of WT-FtsZ and R191A-FtsZ. Molecular dynamics simulations of WT-FtsZ and R191A-FtsZ generated 20,000 structures in one trajectory. For each structure, secondary structure was calculated using DSSP tool from GROMACS package. On the y-axis secondary structure of each residue is shown and x-axis shows how the secondary structure of each residue evolved during the course of the simulation. H5-helix: residues 179-203 and T7-loop: residues 204 to 210.



**Fig. S4: Root mean square fluctuation analysis of WT-FtsZ and R191A-FtsZ**

**simulations.** Root mean square fluctuation analysis was performed to identify regions of structural change. Three simulations each for the WT-FtsZ and R191A-FtsZ that were performed are shown in black, red, and green.



**Fig. S5:** Distance plot between the residue F138 and N176 C-alpha atoms in WT-FtsZ and R191A-FtsZ simulations.



**Table S1. The role of different domains of FtsZ.**

| Domian or residues of FtsZ  | Function  |
|---|---|
| 1. N-terminal domain and C-terminal domain of <i>T. maritima</i> FtsZ   | 1. Both domains can fold independently into functional tertiary structure. <sup>2</sup>   |
| 2. Poorly conserved last 6 residues (NRNKRG) of <i>B. subtilis</i> FtsZ   | 2. These residues are essentially required to promote the high degree of lateral interactions between FtsZ polymers. The change in this region of FtsZ produces significant defect in cell division <i>in vivo</i> . <sup>3</sup> |
| 3. a) N1-FtsZ (1–178 residues) and N2-FtsZ (1–204 residues) of <i>B. subtilis</i> FtsZ<br>b) C1-FtsZ (205–366 residues), C2-FtsZ (176-366 residues) and C3-FtsZ (176-382 residues) of <i>B. subtilis</i> FtsZ | 3. a) Both N-domains have ability to polymerize and form filamentous polymers independently. <sup>4</sup><br>b) These C-domains cannot form polymers and also inhibited the Polymerization of FL-FtsZ. <sup>4</sup>               |
| 4) Mutations of Asn207, Asp209, and Asp212 in the T7 loop of EcFtsZ   | 4) Severely affected GTP hydrolysis. <sup>5</sup>   |
| 5) H7 helix of <i>Methanococcus jannaschi</i> FtsZ  | 5) H7-helix maintains communication between N-and C-terminal domain, and bending of it can regulate assembly/disassembly of FtsZ. <sup>6</sup>  |

**Table S2. Comparison of root mean square deviation of backbone atoms of entire protein and only helices H4 and H7.**

|            | Simulation-1        |             | Simulation-2       |             | Simulation-3       |             |
|------------|---------------------|-------------|--------------------|-------------|--------------------|-------------|
|            | Entire protein (nm) | H5 & H7(nm) | Entire protein(nm) | H5 & H7(nm) | Entire protein(nm) | H5 & H7(nm) |
| WT-FtsZ    | 0.16 ± 0.02         | 0.19 ± 0.02 | 0.19 ± 0.04        | 0.2 ± 0.03  | 0.17 ± 0.02        | 0.18 ± 0.03 |
| R191A-FtsZ | 0.17 ± 0.02         | 0.2 ± 0.03  | 0.17 ± 0.02        | 0.19 ± 0.02 | 0.19 ± 0.02        | 0.27 ± 0.02 |

Average ± Standard deviation

## **Movie S1: The MD simulation of R191A-FtsZ and WT-FtsZ.**

Movie is attached as FtsZ\_HelixTilt.vlc file.

### **References:**

- (1) Bhattacharya, A., Jindal, B., Singh, P., Datta, A., and Panda, D. (2013) Plumbagin inhibits cytokinesis in *Bacillus subtilis* by inhibiting FtsZ assembly--a mechanistic study of its antibacterial activity. *FEBS J.* 280, 4585–4599.
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