### **Supporting Information**

# Rapid, mild, and selective ketone and aldehyde hydroboration/reduction mediated by a simple lanthanum catalyst

Victoria L. Weidner, Christopher J. Barger, Massimiliano Delferro,<sup>†</sup> Tracy L. Lohr,\* and Tobin J. Marks\*

Department of Chemistry, Northwestern University, Evanston, Illinois 60208-3113, USA

<sup>†</sup>Current Address: Chemical Sciences & Engineering Division, Argonne National Laboratory, Lemont, Illinois 60439-4803, United States

t-marks@northwestern.edu and tracy.lohr@northwestern.edu

#### **Table of Contents**

Materials and Methods	.S1
Experimental Details	.S2
Kinetic Analysis Details	.S2
Plots for the Determination of Reaction Order with Respect to Aldehyde/Ketone, HBpin, and La <sup>NTMS</sup>	
Spectroscopic Data of Ketone/Aldehyde Hydroboration Products	S6
NMR Spectra of Hydroboration Products and Relevant Alcohols	.S12
References	S78

Materials and Methods. All manipulations of air-sensitive materials were carried out with rigorous exclusion of oxygen and moisture in flame- or oven-dried Schlenk-type glassware on a dualmanifold Schlenk line, interfaced to a high-vacuum line  $(10^{-6} \text{ Torr})$ , or in an argon-filled vacuum atmospheres glovebox with a high capacity recirculator (<1 ppm O<sub>2</sub>). Benzene-d6 (Cambridge Isotope Laboratories; 99+ atom % D) was stored over Na/K alloy and vacuum transferred immediately prior to use. La[N(SiMe<sub>3</sub>)<sub>2</sub>]<sub>3</sub> (La<sup>NTMS</sup>)and hexamethylbenzene were purchased from Sigma-Aldrich Co. and sublimed under high-vacuum (10<sup>-6</sup> Torr). Pinacolborane ("HBpin") was purchased from Sigma-Aldrich Co. and distilled under high-vacuum (10<sup>-6</sup> Torr). Carbonyl-containing substrates were purchased from Sigma-Aldrich Co. and dried over 3Å molecular sieves and distilled off prior to use (for liquid substrates) or dried under vacuum (for solid substrates). Known boryl esters were characterized by <sup>1</sup>H, <sup>13</sup>C, and <sup>11</sup>B NMR and compared to literature values. Unknown boryl esters were also fully characterized by NMR, and then hydrolyzed by refluxing in 1M NaOH/H<sub>2</sub>O and MeOH for 1 hour (for dicyclohexyl methanol and phenyl cyclohexyl methanol) or by refluxing with silica gel and H<sub>2</sub>O for 3 hours (for perflouorodiphenyl methanol and 2-ethynyl benzyl alcohol). The product was extracted with DCM and the organic layer was dried over  $MgSO_4$ , and the solvent was removed by rotary evaporation. If necessary, the crude was purified by column chromatography, using 30% THF in hexanes. The resulting alcohol was characterized by <sup>1</sup>H and <sup>13</sup>C NMR and EI- or ESI-MS.

**Physical and Analytical Measurements**. NMR spectra were recorded on a Bruker Avance III (500 MHz, <sup>1</sup>H ; 125 MHz, <sup>13</sup>C), Varian Inova 500 (500 MHz, <sup>1</sup>H; 125 MHz, <sup>13</sup>C), Agilent DD MR-400

(400 MHz, <sup>1</sup>H; 100 MHz, <sup>13</sup>C; 128 MHz, <sup>11</sup>B), or Agilent DD2 500 (500 MHz, <sup>1</sup>H; 125 MHz, <sup>13</sup>C). Chemical shifts ( $\delta$ ) for <sup>1</sup>H and <sup>13</sup>C are referenced to residual solvent resonances (7.16 and 128.06 ppm, resp., for benzene-d<sub>6</sub>). <sup>11</sup>B shifts are referenced to an external BF<sub>3</sub>·OEt<sub>2</sub> standard. NMR scale reactions were carried out either in Teflon-sealed J. Young tubes or PTFE septum-sealed tubes. Mass spectra were recorded on a Bruker AmaZon SL LC-MS (ESI, Quadrupole ion trap) or Agilent 5973 GC-MS (EI, Quadrupole ion trap).

Typical NMR-Scale Reaction of HBpin with Solid Ketones and Aldehydes and La<sup>NTMS</sup> Catalyst. In a glovebox, the aldehyde/ketone (0.25 mmol) was massed in a vial. 500  $\mu$ L of a stock solution containing HBpin (0.30 mmol, 1.2 equivalents vs. aldehyde/ketone) and the internal standard hexamethylbenzene (50  $\mu$ mol) was added to the vial, and the vial was shaken until all solids were dissolved. This solution was added to a J. Young tap NMR tube, and 100  $\mu$ L of a stock solution containing an appropriate loading of La[N(SiMe<sub>3</sub>)<sub>2</sub>]<sub>3</sub> was added. The tube was capped and shaken, and the reaction was monitored by <sup>1</sup>H NMR.

Typical NMR-Scale Reaction of HBpin with Liquid Ketones and Aldehydes and La<sup>NTMS</sup> Catalyst. In a glovebox, 100  $\mu$ L of a stock solution containing an appropriate loading of La[N(SiMe<sub>3</sub>)<sub>2</sub>]<sub>3</sub> was added to a septum-sealed NMR tube. 500  $\mu$ L of a stock solution containing HBpin (0.30 mmol, 1.2 equivalents vs. aldehyde/ketone) and the internal standard hexamethylbenzene (50  $\mu$ mol) was added to a septum-sealed vial, and both were brought out of the glovebox. The liquid aldehyde/ketone (0.25 mmol) was injected into the vial with HBpin and standard, the vial was shaken, and the contents were injected into the NMR tube with catalyst, all under N<sub>2</sub>. The tube was shaken, and the reaction was monitored by <sup>1</sup>H NMR.

**Scale-Up/Air and Moisture Tolerance Test Reaction.** Benzophenone (1.0 g, 5.5 mmol) and HBpin (0.96 mL, 6.6 mmol) were dissolved in benzene (5 mL) in a vial outside of a glovebox. To this solution was added La<sup>NTMS</sup> (34 mg, 0.055 mmol). After stirring for 5 minutes, volatiles were removed in vacuo, and the resulting white powder was taken up in 10 mL of 10% NaOH in MeOH. The mixture was sonicated and refluxed for 1 hour. The product (diphenylmethanol) was extracted in ethyl acetate and purified by column chromatography (1:5 THF:hexanes). Final yield of diphenylmethanol: 0.87g (86%).

**Typical NMR-Scale Reaction for Kinetic Monitoring by** <sup>1</sup>**H-NMR Arrays**. In a glovebox, 500  $\mu$ L of a stock solution of aldehyde/ketone and 500  $\mu$ L of a stock solution containing HBpin and the internal standard, hexamethylbenzene (50  $\mu$ mol), were mixed in a vial. This solution was then added to a rubber septum-sealed NMR tube, wrapped with parafilm, and removed from the box. At the NMR, the magnet was locked, tuned, and shimmed to the sample, then 100  $\mu$ L of a stock solution containing an appropriate loading of La[N(SiMe\_3)\_2]\_3 was added. The tube was shaken and reinserted into the instrument and scanning was begun. Single (<sup>1</sup>H NMR) scans were collected at regular intervals. Substrate and/or product concentrations were determined relative to the intensity of the internal standard resonance plotted versus time.

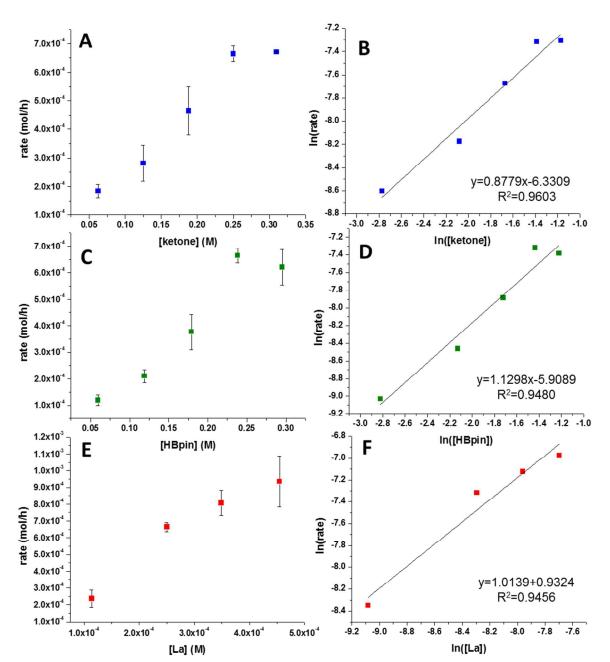
**Kinetic Analysis.** Kinetic analysis of the NMR-scale reactions described above was carried out by collecting multiple (>15) data points early in the reaction (<20% conversion). Under these conditions, the reaction can be approximated as pseudo-zero-order with respect to the substrate concentrations. The product concentration was measured from the area of the R<sub>2</sub>CHOBpin or RCH<sub>2</sub>OBpin peak formed in the product standardized to the methyl peak area of the C<sub>6</sub>Me<sub>6</sub> internal standard. Data were fit by least-squares analysis ( $R^2 > 0.98$ ) according to eq S1, where t is time, [product] is the concentration of product at time t, and m is the rate of reaction.

#### $[product] = mt \tag{Eq. S1}$

Orders for each reactant were determined from the average rates ( $\geq$ 3 trials) at varying concentrations. Ketone/ aldehyde and HBpin concentrations were varied from 25% to 125% (relative to the other reactant) and catalyst concentration was measured at 0.05%, 0.10%, 0.15%, and 0.20% (for dicyclohexylketone) or 0.025%, 0.05%, 0.075%, and 0.1% (for cyclohexylcarboxaldehyde). (Note: in general, ketones react more quickly than aldehydes, except in the case of dicyclohexylketone). These data

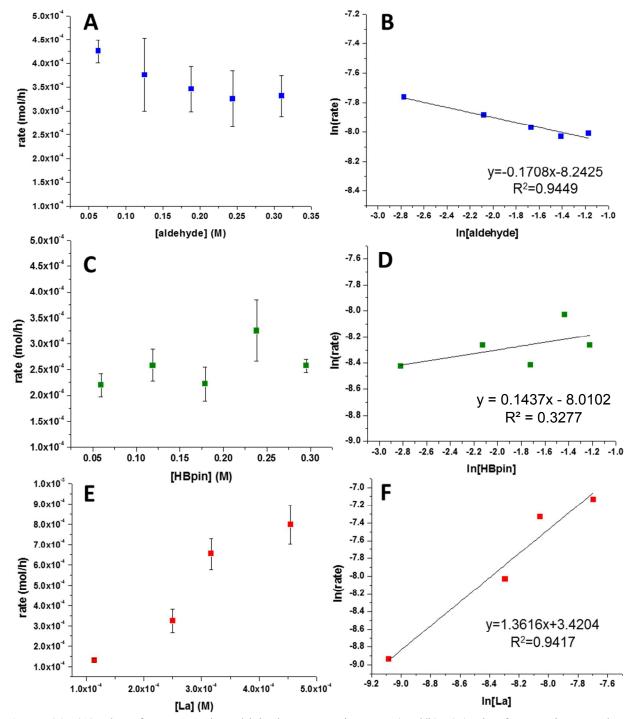
were then plotted as  $\ln(\text{rate})$  vs.  $\ln[\text{ketone}]$ .<sup>1</sup> The negative rate of disappearance of ketone is proportional to the concentration of ketone to the order ( $\alpha$ ) (see eq. S2). Therefore, the order is the slope of a plot of  $\ln(\text{rate})$  vs.  $\ln[\text{ketone}]$  (eq. S3).

$$\frac{-d[ketone]}{dt} = k_{obs}[ketone]^{\alpha}$$
(Eq. S2)



$$\ln(rate) = lnk_{obs} + \propto \ln[ketone]$$
(Eq. S3)

**Figure S1**. (A) Plot of concentration ketone vs. reaction rate (mol/h); (B) Plot for reaction rate law order in [ketone]; (C) Plot of concentration HBpin vs. rate (mol/h); (D) Plot for reaction rate law order in [HBpin]; (E) Plot of concentration  $La^{NTMS}$  vs. rate (mol/h); (F) Plot for reaction rate law order in  $La^{NTMS}$ .



**Figure S2**. (A) Plot of concentration aldehyde vs. reaction rate (mol/h); (B) plot for reaction rate law order in [aldehyde]; (C) Plot of concentration HBpin vs. rate (mol/h); (D) plot for reaction rate law order

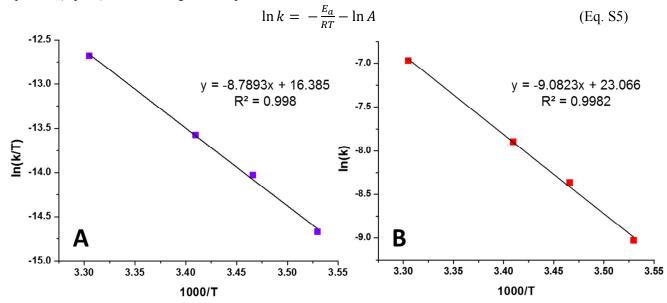
in [HBpin]; (E) Plot of concentration  $La^{NTMS}$  vs. rate (mol/h); (F) plot for reaction rate law order in  $La^{NTMS}$ .

**Temperature Analysis.** Data on the rate dependence on temperature was obtained as shown above. A rate at each temperature were determined from the average rates ( $\geq$ 3 trials) at temperatures set on the NMR and measured using a methanol (<25°C) or ethylene glycol (>25°C) standard.

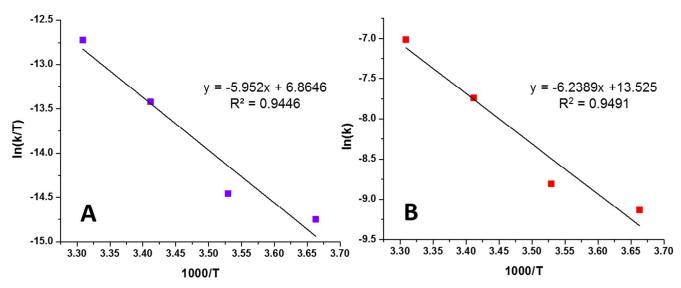
These data were then plotted as 1000/T vs.  $\ln(k/T)^1$  from which the enthalpy and entropy of the transition state could be obtained using the Eyring equation (see eq. S4).  $\Delta H^{\neq}$  is the negative slope times R and  $\Delta S^{\neq}$  is the intercept minus the natural log of  $k_b/h$  times R.

$$\ln\frac{k}{T} = \frac{\Delta H^{\neq}}{RT} \left[ \frac{\Delta S^{\neq}}{R} - \ln\frac{k_b}{h} \right]$$
(Eq. S4)

From a plot of 1000/T vs. ln(k), the activation energy can be obtained using the Arrhenius equation (eq. S5).  $E_a$  is the negative slope times R.



**Figure S3.** (A) Plot of 1000/temperature vs.  $\ln(k/T)$  for the lanthanum-catalyzed hydroboration of dicyclohexylketone. (B) Plot of 1000/temperature vs.  $\ln(k)$ .

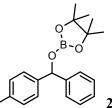


**Figure S4.** (A) Plot of 1000/temperature vs.  $\ln(k/T)$  for the lanthanum-catalyzed hydroboration of cyclohexylcarboxaldehyde. (B) Plot of 1000/temperature vs.  $\ln(k)$ .

#### Characterization Data for Ketone/Aldehyde Hydroboration Products

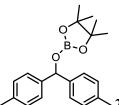


<sup>1</sup>H, <sup>11</sup>B{<sup>1</sup>H} and <sup>13</sup>C{<sup>1</sup>H} spectra are identical to those reported in the literature.<sup>2</sup> <sup>1</sup>H NMR (C<sub>6</sub>D<sub>6</sub>, 500 MHz): 7.45-7.41 (m, 4H), 7.12-7.07 (m, 4H), 7.03-6.98 (tt, 2H, <sup>3</sup>J<sub>HH</sub> = 7.4, <sup>4</sup>J<sub>HH</sub> = 1.2), 6.41 (s, 1H), 0.98 (s, 12H). <sup>11</sup>B{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, 128 MHz): 22.83. <sup>13</sup>C{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, 125 MHz): 143.89, 128.57, 127.54, 126.97, 82.85, 78.53, 24.62.



<sup>2</sup> <u>2-(para-tolylphenylmethoxy)pinacolborane.</u>

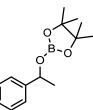
<sup>1</sup>H, <sup>11</sup>B{<sup>1</sup>H} and <sup>13</sup>C{<sup>1</sup>H} spectra are identical to those reported in the literature.<sup>3</sup> <sup>1</sup>H NMR (C<sub>6</sub>D<sub>6</sub>, 500 MHz): 7.46 (d, 2H, <sup>3</sup>J<sub>HH</sub>= 7.7 Hz), 7.37 (d, 2H, <sup>3</sup>J<sub>HH</sub>= 7.7 Hz), 7.11 (t, 2H, <sup>3</sup>J<sub>HH</sub>= 7.5 Hz), 7.01 (t, 1H, 7.5 Hz), 6.93 (d, 2H, <sup>3</sup>J<sub>HH</sub>= 7.7 Hz), 6.43 (s, 1H), 2.05 (s, 3H), 0.99 (s, 12H). <sup>11</sup>B{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, 128 MHz): 22.86. <sup>13</sup>C{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, 125 MHz): 144.15, 141.10, 136.93, 129.28, 128.54, 127.46, 127.01, 126.94, 82.80, 78.43, 24.64, 21.06



#### <u>2-(di-*para*-tolylmethoxy)pinacolborane.</u>

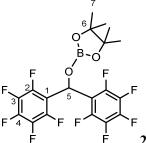
 ${}^{1}H, {}^{11}B{}^{1}H$  and  ${}^{13}C{}^{1}H$  spectra are identical to those reported in the literature.<sup>4</sup>

<sup>1</sup>H NMR ( $C_6D_6$ , 500 MHz): 7.39 (d, 4H, <sup>3</sup>J<sub>HH</sub>= 7.7 Hz), 6.94 (d, 4H, <sup>3</sup>J<sub>HH</sub>= 7.7 Hz), 6.44 (s, 1H), 2.05 (s, 6H), 1.00 (s, 12H). <sup>11</sup>B{<sup>1</sup>H} NMR ( $C_6D_6$ , 128 MHz): 22.84. <sup>13</sup>C{<sup>1</sup>H} NMR ( $C_6D_6$ , 125 MHz): 141.35, 136.82, 129.24, 126.99, 82.76, 78.33, 24.66, 21.06



2-(1-phenylethoxy)pinacolborane.

<sup>1</sup>H, <sup>11</sup>B{<sup>1</sup>H} and <sup>13</sup>C{<sup>1</sup>H} spectra are identical to those reported in the literature.<sup>2</sup> <sup>1</sup>H NMR (C<sub>6</sub>D<sub>6</sub>, 500 MHz): 7.37-7.33 (m, 2H), 7.17-7.11 (m, 2H), 7.08-7.02 (tt, <sup>1</sup>H, <sup>3</sup>J<sub>HH</sub> = 7.4, <sup>4</sup>J<sub>HH</sub> = 2.1 Hz), 5.39 (q, 1H, <sup>3</sup>J<sub>HH</sub> = 6.4 Hz), 1.45 (d, 3H, <sup>3</sup>J<sub>HH</sub> = 6.5 Hz), 1.00 (s, 12H). <sup>11</sup>B{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, 128 MHz): 22.52. <sup>13</sup>C{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, 125 MHz): 145.39, 128.54, 127.35, 125.70, 82.54, 72.94, 25.79, 24.70, 24.62.



2-(di-perfluorophenylmethoxy)pinacolborane.

<sup>1</sup>H NMR ( $C_6D_6$ , 500 MHz): 6.90 (s, 1H, H-5), 1.03 (s, 12H, H-7). <sup>11</sup>B{<sup>1</sup>H} NMR ( $C_6D_6$ , 128 MHz): 22.62. <sup>13</sup>C{<sup>1</sup>H} NMR ( $C_6D_6$ , 125 MHz): 145.13 (dm, C-*Ar*, J=253.6 Hz), 141.60 (dt, C-*Ar*, J=255.4 Hz, J=13.2 Hz), 137.92 (dt, C-*Ar*, J=250.7 Hz, J=14.1 Hz), 131.78 (s, C-*I*), 84.14 (s, C-6), 62.77 (s, C-5), 24.45 (s, C-7).

Perfluorodiphenylmethanol. 62% isolated yield.

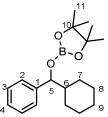
<sup>1</sup>H NMR (C<sub>6</sub>D<sub>6</sub>, 500 MHz): 5.94 (d, 1H, H-5,  ${}^{3}J_{HH} = 5.8$  Hz), 2.20 (d, 1H, OH,  ${}^{3}J_{HH} = 5.8$  Hz).  ${}^{19}F{}^{1}H$ } NMR (C<sub>6</sub>D<sub>6</sub>, 376 MHz): -143.5 - -143.8 (m, 2F), -153.8 (t, 1F, J=22Hz), -161.6 - -162.0 (m, 2F).  ${}^{13}C{}^{1}H$ } NMR (C<sub>6</sub>D<sub>6</sub>, 125 MHz): 144.4 (dm, C-Ar, J=253 Hz), 140.9 (dm, C-Ar, J= 255 Hz), 137.4 (dm, C-Ar, J=251 Hz), 125.5 (s, C-Ar), 113.7 (s, C-OH). LC-MS: [2M-H]<sup>-</sup>: Calc: 726.9813. Found: 726.9818



<sup>1</sup>H NMR (C<sub>6</sub>D<sub>6</sub>, 500 MHz): 3.78 (t, 1H, H-5,  ${}^{3}J_{HH} = 6.0$  Hz), 1.89-1.82 (m, 2H, H-1), 1.75-1.66 (m, 4H, H-Cy), 1.64-1.49 (m, 6H, H-Cy), 1.29-1.07 (m, 10H, H-Cy), 1.10 (s, 12H, H-7). <sup>11</sup>B{}<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, 128 MHz): 22.46. {}^{13}C{}^{1}H NMR (C<sub>6</sub>D<sub>6</sub>, 125 MHz): 82.79 (C-6), 82.14 (C-5), 39.66 (C-1), 30.19 (C-Cy), 27.64 (C-Cy), 26.94 (C-Cy), 26.82 (C-Cy), 26.58 (C-Cy), 24.68 (C-Cy).

Dicyclohexylmethanol. 92% isolated yield.

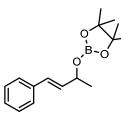
<sup>1</sup>**H** NMR ( $\hat{C}_{6}D_{6}$ , 500 MHz): 2.88 (q, 1H, H-5, <sup>3</sup>J<sub>HH</sub> = 5.7 Hz), 1.85-1.78 (m, 2H, H-*Cy*), 1.77-1.67 (m, 4H, H-*Cy*), 1.66-1.60 (m, 2H, H-*Cy*), 1.51-1.44 (m, 2H, H-*Cy*), 1.40-1.31 (m, 2H, H-*Cy*), 1.25-0.97 (m, 10H, H-*Cy*), 0.81-0.76 (m, 1H, H-O*H*). <sup>13</sup>C{<sup>1</sup>H} NMR ( $C_{6}D_{6}$ , 125 MHz): 80.13 (C-5), 40.30 (C-*I*), 30.33 (C-*Cy*), 27.69 (C-*Cy*), 27.02 (C-*Cy*), 26.96 (C-*Cy*), 26.65 (C-*Cy*). GC-MS [M-H<sub>2</sub>O]<sup>+</sup>: Calc: 178.17; Found: 178.25.



# <sup>9</sup> <u>2-(cyclohexylphenylmethoxy)pinacolborane.</u>

<sup>1</sup>H NMR (C<sub>6</sub>D<sub>6</sub>, 500 MHz): 7.38-7.33 (m, 2H, H-2), 7.20-7.12 (m, 2H, H-3), 7.10-7.04 (m, <sup>1</sup>H, H-4), 5.03 (d, 1H, H-5, <sup>3</sup>J<sub>HH</sub> = 6.4 Hz), 2.03-1.94 (m, <sup>1</sup>H, H-6), 1.75-1.45 (m, 6H, H-*Cy*), 1.25-1.05 (m, 4H, H-*Cy*), 1.03 (s, 6H, H-*I1*), 0.99 (s, 6H, H-*I1*). <sup>11</sup>B{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, 128 MHz): 22.60. <sup>13</sup>C{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, 125 MHz): 143.13 (C-*I*), 128.25 (C-*Ar*), 127.38 (C-*Ar*), 127.05 (C-*Ar*), 82.47 (C-*I0*), 81.48 (C-5), 45.45 (C-6), 29.65 (C-*Cy*), 28.63 (C-*Cy*), 26.78 (C-*Cy*), 26.40 (C-*Cy*), 24.64 (C-*Cy*). Cyclohexyl(phenyl)methanol. 83% isolated yield.

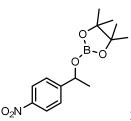
<sup>1</sup>**H** NMR (C<sub>6</sub>D<sub>6</sub>, 500 MHz): 7.22-7.17 (m, 4H, H-*Ar*), 7.12-7.08 (m, 1H, H-*Ar*), 4.10 (dd, 1H, H-5,  ${}^{3}J_{HH} = 6.8 \text{ Hz}, {}^{4}J_{HH} = 3.3 \text{ Hz}$ ), 2.04-1.98 (m, 1H, H-6), 1.72-1.65 (m, 1H, H-*Cy*), 1.61-1.48 (m, 3H, H-*Cy*), 143-1.36 (m, 1H, H-*Cy*), 1.19 (d, 1H, H-*Cy*,  ${}^{3}J_{HH} = 3.4 \text{ Hz}$ ), 1.18-0.98 (m, 4H, H-*Cy*), 0.94-0.85 (m, 1H, H-*Cy*), 1<sup>3</sup>C{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, 125 MHz): 144.61 (C-*1*), 127.40 (C-*Ar*), 126.99 (C-*Ar*), 79.15 (C-5), 45.54 (C-6), 29.71 (C-*Cy*), 28.92 (C-*Cy*), 26.86 (C-*Cy*), 26.54 (C-*Cy*), 26.47 (C-*Cy*). GC-MS [M]<sup>+</sup>: Calc: 190.14; Found: 190.20.



2-(1-cinnamylethoxy)pinacolborane.

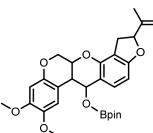
<sup>1</sup>H, <sup>11</sup>B{<sup>1</sup>H} and <sup>13</sup>C{<sup>1</sup>H} spectra are identical to those reported in the literature.<sup>2</sup>

<sup>1</sup>H NMR ( $C_6D_6$ , 500 MHz): 7.22-7.17 (m, 2H), 7.12-7.06 (m, 2H), 7.05-6.99 (m, 1H), 6.64 (dd, 1H,  ${}^{3}J_{HH}$ = 16 Hz,  ${}^{4}J_{HH}$  = 0.95 Hz), 6.19 (dd, 1H,  ${}^{3}J_{HH}$  = 16 Hz,  ${}^{3}J_{HH}$  = 5.9 Hz), 4.98 (ddq, 1H,  ${}^{3}J_{HH}$  = 6.2 Hz,  ${}^{3}J_{HH}$  = 6.4 Hz,  ${}^{4}J_{HH}$  = 1.2 Hz), 1.33 (d, 3H,  ${}^{3}J_{HH}$  = 6.4 Hz), 1.06 (d, 12H,  ${}^{3}J_{HH}$  = 2.3 Hz)  ${}^{11}B{}^{1}H{}$  NMR ( $C_6D_6$ , 128 MHz): 22.50.  ${}^{13}C{}^{1}H{}$  NMR ( $C_6D_6$ , 125 MHz): 137.43, 132.72, 129.49, 128.76, 127.65, 126.88, 82.51, 71.47, 24.95, 24.80, 24.67, 23.43.



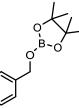
### 2-(1-(4-nitrophenyl)ethoxy)pinacolborane.

<sup>1</sup>H, <sup>11</sup>B{<sup>1</sup>H} and <sup>13</sup>C{<sup>1</sup>H} spectra are identical to those reported in the literature.<sup>2</sup> <sup>1</sup>H NMR (C<sub>6</sub>D<sub>6</sub>, 500 MHz): 7.80-7.72 (m, 2H), 7.00-6.92 (m, 2H), 5.13 (q, <sup>1</sup>H, <sup>3</sup>J<sub>HH</sub> = 6.5 Hz), 1.20 (d, 3H, <sup>3</sup>J<sub>HH</sub> = 6.5 Hz), 0.94 (s, 12H). <sup>11</sup>B{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, 128 MHz): 22.43. <sup>13</sup>C{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, 125 MHz): 151.82, 147.47, 126.09, 123.64, 82.94, 71.96, 25.33, 24.62.



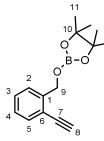
2-(Rotenoxy)pinacolborane.

<sup>1</sup>**H** NMR ( $C_6D_6$ , 500 MHz): 7.26 (d, 1H, J=8.2 Hz), 6.71 (s, 1H), 6.60 (d, 1H, J=8.2 Hz), 6.51 (s, 1H), 5.48 (d, 1H, J=4.0), 5.08-5.05 (m, 1H), 4.96 (t, 1H, J=8.8 Hz), 4.89 (t, 1H, J=10 Hz), 4.76-4.73 (m, 1H), 4.67-4.61 (m, 1H), 4.17 (ddd, 1H, J=1.2, 4.8, 9.8 Hz), 3.59 (s, 3H), 3.31 (s, 3H), 3.12-3.01 (m, 2H), 2.93-2.85 (m, 1H), 1.58 (s, 3H), 0.95 (s, 6H), 0.92 (s, 6H). <sup>11</sup>B{<sup>1</sup>H} NMR ( $C_6D_6$ , 128 MHz): 22.29. <sup>13</sup>C{<sup>1</sup>H} NMR ( $C_6D_6$ , 125 MHz): 162.60, 150.67, 150.19, 150.12, 144.54, 144.40, 130.38, 114.64, 113.96, 113.20, 111.15, 109.91, 102.88, 101.54, 86.70, 82.67, 70.17, 69.41, 65.45, 56.63, 55.36, 38.03, 32.57, 24.80, 24.31, 17.32. LC-MS [M+Na]<sup>+</sup> Calc.: 545.232, Found: 545.233



## 2-(benzyloxy)pinacolborane.

<sup>1</sup>H, <sup>11</sup>B{<sup>1</sup>H} and <sup>13</sup>C{<sup>1</sup>H} spectra are identical to those reported in the literature.<sup>2</sup> <sup>1</sup>H NMR (C<sub>6</sub>D<sub>6</sub>, 500 MHz): 7.32-7.28 (m, 2H), 7.16-7.10 (m, 2H), 7.08-7.02 (m, 1H), 4.94 (s, 2H), 1.04 (s, 12H). <sup>11</sup>B{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, 128 MHz): 22.79. <sup>13</sup>C{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, 125 MHz): 140.1, 128.59, 127.57, 127.05, 82.75, 66.96, 24.70.



2-(2-ethynylbenzyloxy)pinacolborane.

<sup>1</sup>H NMR (C<sub>6</sub>D<sub>6</sub>, 500 MHz): 7.61 (m, 1H), 7.35 (dd, 1H,  ${}^{3}J_{HH}$ = 7.7 Hz), 7.04 (dt, 1H,  ${}^{3}J_{HH}$ = 7.7 Hz,  ${}^{4}J_{HH}$ = 0.96 Hz), 6.86 (m, 1H), 5.34 (s, 2H) 2.89 (s, 1H), 1.03 (s, 12H). <sup>11</sup>B{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, 128 MHz): 22.76.

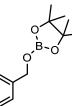
<sup>13</sup>C{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, 125 MHz): 142.45, 132.67, 129.22, 127.10, 126.24, 119.94, 82.83, 81.11, 65.26, 24.68

#### **<u>2-ethynylbenzyl alcohol</u>.** 77% isolated yield.

<sup>1</sup>**H** NMR (C<sub>6</sub>D<sub>6</sub>, 500 MHz): 7.38 (dd, 1H, H-2,  ${}^{3}J_{HH} = 7.7$  Hz,  ${}^{4}J_{HH} = 0.9$  Hz), 7.32 (d, 1H, H-5,  ${}^{3}J_{HH} = 7.7$  Hz), 7.01 (dt, 1H, H-3,  ${}^{3}J_{HH} = 7.7$  Hz,  ${}^{4}J_{HH} = 1.1$  Hz), 6.85 (t, 1H, H-4,  ${}^{3}J_{HH} = 7.7$  Hz), 4.67 (s, 2H, H-9), 2.85 (s, 1H, H-8), 1.42 (br s, 1H, OH).  ${}^{13}C{^{1}H}$  NMR (C<sub>6</sub>D<sub>6</sub>, 125 MHz): 144.34 (C-1), 132.82 (C-Ar), 129.23 (C-Ar), 127.15 (C-Ar), 127.00 (C-Ar), 120.22 (C-6), 82.37 (C-9), 81.53 (C-7), 63.43 (C-8). GC-MS [M]<sup>+</sup>: Calc: 132.06; Found: 132.10.

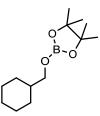
2-(cinnamylmethoxy)pinacolborane.

<sup>1</sup>H, <sup>11</sup>B{<sup>1</sup>H} and <sup>13</sup>C{<sup>1</sup>H} spectra are identical to those reported in the literature.<sup>5</sup> <sup>1</sup>H NMR (C<sub>6</sub>D<sub>6</sub>, 500 MHz): 7.22-7.16 (m, 2H), 7.13-7.07 (m, 2H), 7.06-7.00 (m, 1H), 6.62 (dt, 1H, <sup>3</sup>J<sub>HH</sub> = 15.9 Hz, <sup>4</sup>J<sub>HH</sub> = 1.7 Hz), 6.19 (dt, 1H, <sup>3</sup>J<sub>HH</sub> = 15.9 Hz, <sup>3</sup>J<sub>HH</sub> = 5.3 Hz), 4.55 (dd, 2H, <sup>3</sup>J<sub>HH</sub> = 5.3 Hz, <sup>4</sup>J<sub>HH</sub> = 1.7 Hz), 1.08 (s, 12H). <sup>11</sup>B{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, 128 MHz): 22.70. <sup>13</sup>C{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, 125 MHz): 137.36, 130.91, 128.76, 127.69, 127.52, 126.85, 82.70, 65.54, 24.74.



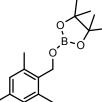
2-(4-chlorobenzyloxy)pinacolborane.

<sup>1</sup>H, <sup>11</sup>B{<sup>1</sup>H} and <sup>13</sup>C{<sup>1</sup>H} spectra are identical to those reported in the literature.<sup>2</sup> <sup>1</sup>H NMR (C<sub>6</sub>D<sub>6</sub>, 500 MHz): 7.09-7.04 (m, 2H), 7.02-6.96 (m, 2H), 4.76 (s, 2H), 1.03 (s, 12H). <sup>11</sup>B{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, 128 MHz): 22.64. <sup>13</sup>C{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, 125 MHz): 138.46, 133.35, 128.71, 128.40, 82.87, 66.09, 24.69.



## 2-(cyclohexylmethoxy)pinacolborane.

<sup>1</sup>H, <sup>11</sup>B{<sup>1</sup>H} and <sup>13</sup>C{<sup>1</sup>H} spectra are identical to those reported in the literature.<sup>2</sup> <sup>1</sup>H NMR (C<sub>6</sub>D<sub>6</sub>, 500 MHz): 3.76 (d, 2H, <sup>3</sup>J<sub>HH</sub> = 64 Hz), 1.75-1.67 (m, 2H), 1.65-1.57 (m, 2H), 1.56-1.45 (m, 2H), 1.18-1.04 (m, 3H), 1.07 (s, 12H), 0.97-0.84 (m, 2H). <sup>11</sup>B{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, 128 MHz): 22.29. <sup>13</sup>C{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, 125 MHz): 82.37, 70.60, 39.86, 29.74, 26.90, 26.18, 24.77.



2-(mesitylmethoxy)pinacolborane.

<sup>1</sup>H, <sup>11</sup>B{<sup>1</sup>H} and <sup>13</sup>C{<sup>1</sup>H} spectra are identical to those reported in the literature.<sup>6</sup> <sup>1</sup>H NMR (C<sub>6</sub>D<sub>6</sub>, **500** MHz): 6.73 (s, 2H), 5.03 (s, 2H), 2.37 (s, 6H), 2.12 (s, 3H), 1.04 (s, 12H). <sup>11</sup>B{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, **128** MHz): 22.58. <sup>13</sup>C{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, **125** MHz): 137.78, 137.39, 132.98, 129.35, 82.53, 61.53, 24.70, 21.07, 14.64.

N//

2-(4-cyanobenzyloxy)pinacolborane.

<sup>1</sup>H, <sup>11</sup>B{<sup>1</sup>H} and <sup>13</sup>C{<sup>1</sup>H} spectra are identical to those reported in the literature.<sup>4</sup> <sup>1</sup>H NMR (C<sub>6</sub>D<sub>6</sub>, 500 MHz): 7.03-6.99 (m, 2H), 6.92-6.87 (m, 2H), 4.667 (s, 2H), 1.04 (s, 12H). <sup>11</sup>B{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, 128 MHz): 22.66. <sup>13</sup>C{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, 125 MHz): 144.57, 132.10, 126.78, 118.85, 111.61, 83.10, 65.84, 24.67.

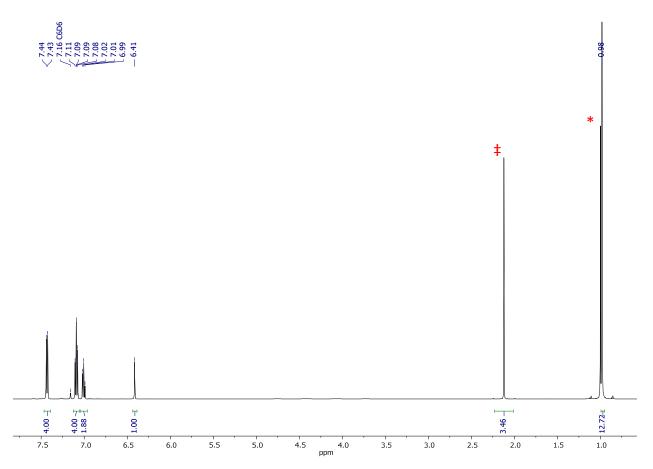


#### 2-(ferrocenylmethoxy)pinacolborane.

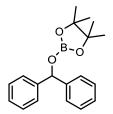
<sup>1</sup>H, <sup>11</sup>B{<sup>1</sup>H} and <sup>13</sup>C{<sup>1</sup>H} spectra are identical to those reported in the literature.<sup>2</sup> <sup>1</sup>H NMR (C<sub>6</sub>D<sub>6</sub>, 500 MHz): 4.75 (s, 2H), 4.21 (dd, 2H, <sup>3</sup>J<sub>HH</sub> = 1.9 Hz, <sup>4</sup>J<sub>HH</sub> = 1.9 Hz), 3.98 (s, 5H), 3.95 (dd, 2H, <sup>3</sup>J<sub>HH</sub> = 1.9 Hz, <sup>4</sup>J<sub>HH</sub> = 1.9 Hz), 1.07 (s, 12H). <sup>11</sup>B{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, 128 MHz): 22.67. <sup>13</sup>C{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, 125 MHz): 86.08, 82.61, 69.02, 68.80, 68.52, 63.44, 24.82.

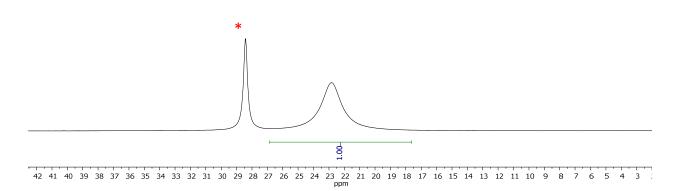
# 2-(4-N,N-dimethylaminobenzyloxy)pinacolborane

<sup>1</sup>H, <sup>11</sup>B{<sup>1</sup>H} and <sup>13</sup>C{<sup>1</sup>H} spectra are identical to those reported in the literature.<sup>4</sup> <sup>1</sup>H NMR (C<sub>6</sub>D<sub>6</sub>, 500 MHz): 7.36-7.32 (m, 2H), 6.60-6.53 (m, 2H), 5.00 (s, 2H), 2.50 (s, 6H), 1.07 (s, 12H). <sup>11</sup>B{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, 128 MHz): 22.80. <sup>13</sup>C{<sup>1</sup>H} NMR (C<sub>6</sub>D<sub>6</sub>, 125 MHz): 150.16, 128.57, 112.42, 82.15, 66.85, 39.92, 24.39.

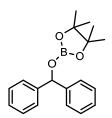


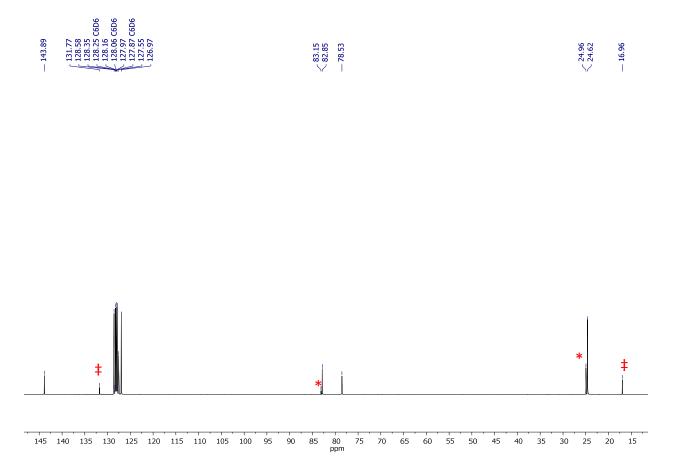
**Figure S5**. <sup>1</sup>H NMR spectrum of 2-(diphenylmethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.



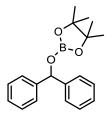


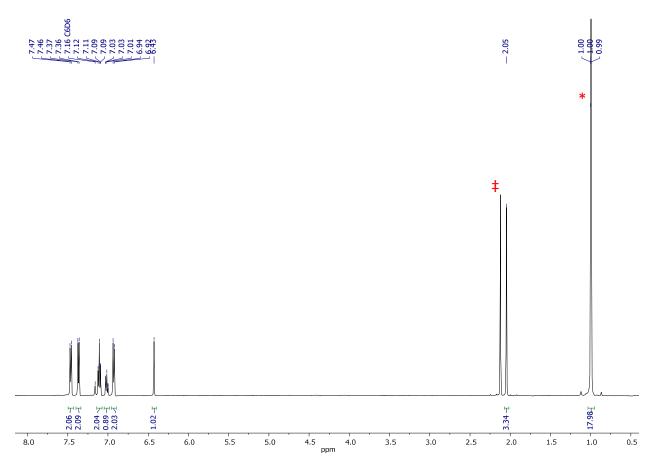
**Figure S6**. <sup>11</sup>B NMR spectrum of 2-(diphenylmethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin.



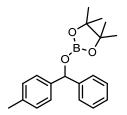


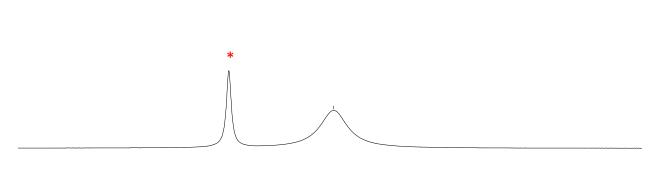
**Figure S7**. <sup>13</sup>C NMR spectrum of 2-(diphenylmethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.





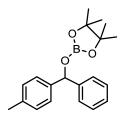
**Figure S8**. <sup>1</sup>H NMR spectrum of 2-(para-tolylphenylmethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.

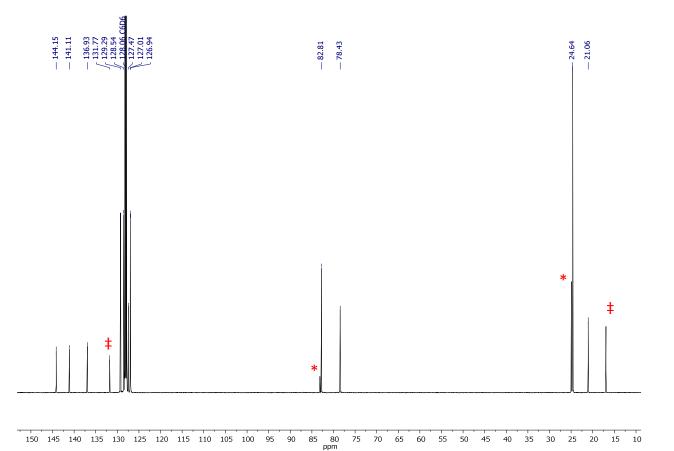




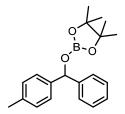
39 38 37 36 35 34 33 32 31 30 29 28 27 26 25 24 23 22 21 20 19 18 17 16 15 14 13 12 11 10 9 8 7 ppm

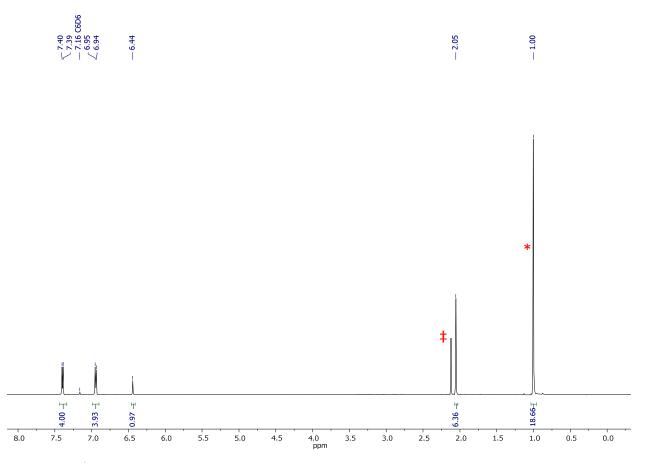
**Figure S9**. <sup>11</sup>B NMR spectrum of 2-(para-tolylphenylmethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin.



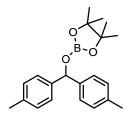


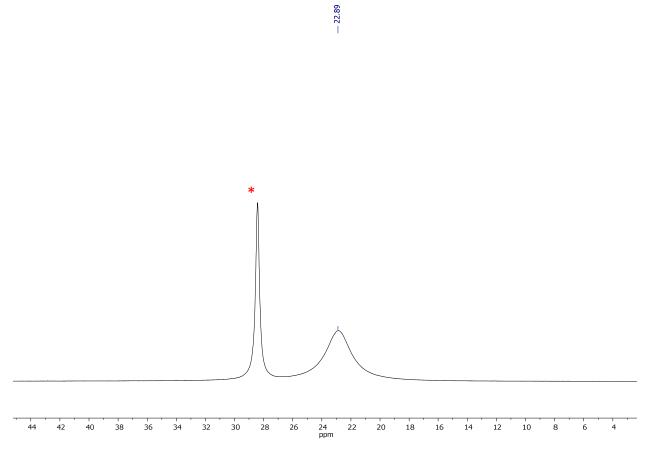
**Figure S10.** <sup>13</sup>C NMR spectrum of 2-(para-tolylphenylmethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.



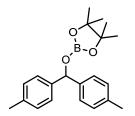


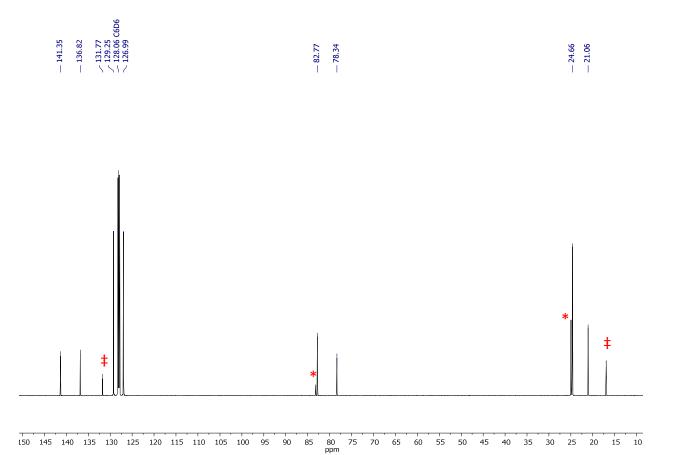
**Figure S11.** <sup>1</sup>H NMR spectrum of 2-(di-para-tolylmethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.



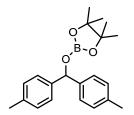


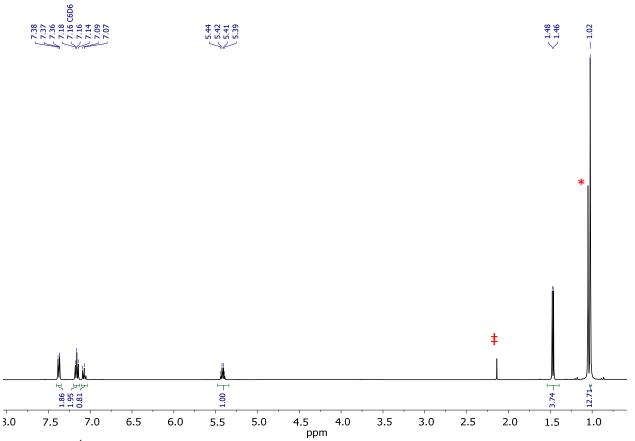
**Figure S12.** <sup>11</sup>B NMR spectrum of 2-(di-para-tolylmethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin.





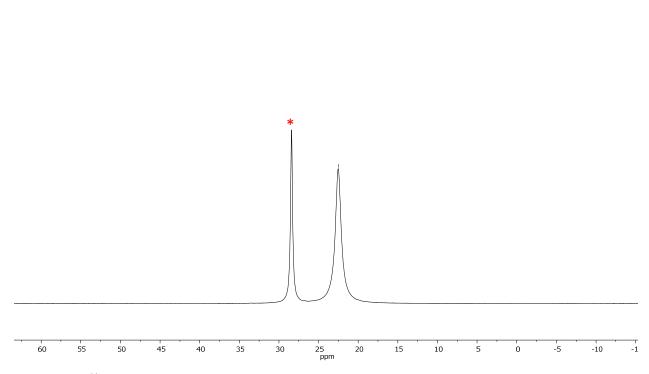
**Figure S13**. <sup>13</sup>C NMR spectrum of 2-(di-para-tolylmethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.



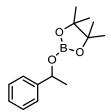


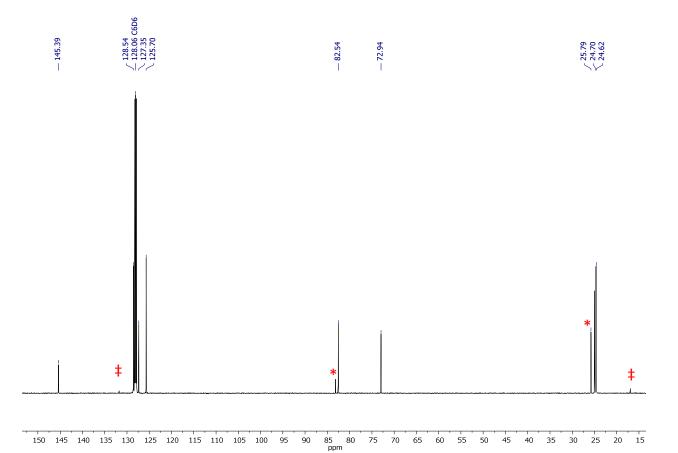
**Figure S14.** <sup>1</sup>H NMR spectrum of 2-(1-phenylethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.

or or B-o

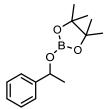


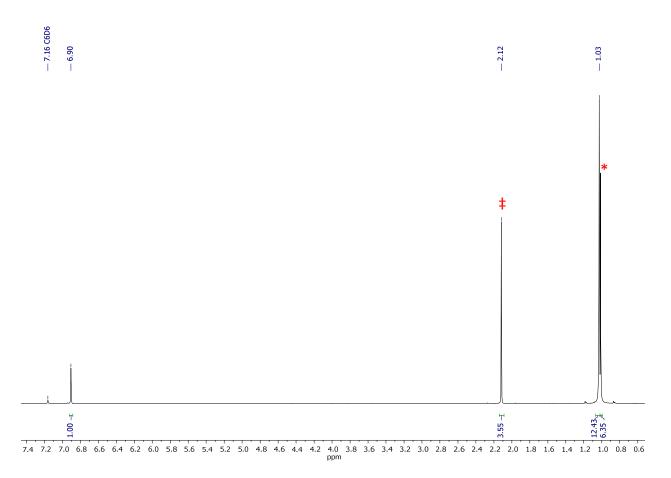
**Figure S15.** <sup>11</sup>B NMR spectrum of 2-(1-phenylethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin.



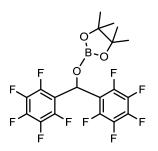


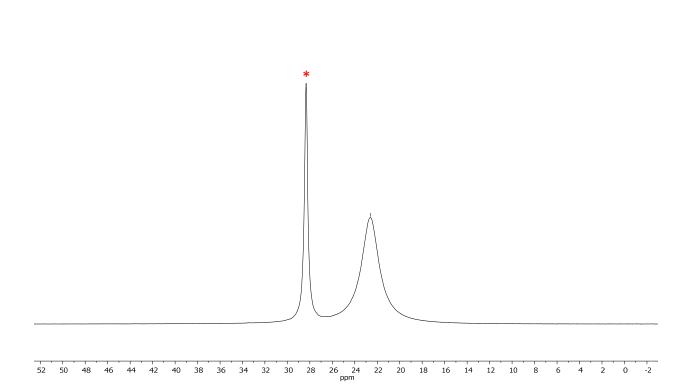
**Figure S16.** <sup>13</sup>C NMR spectrum of 2-(1-phenylethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.



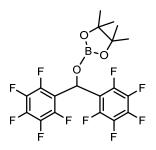


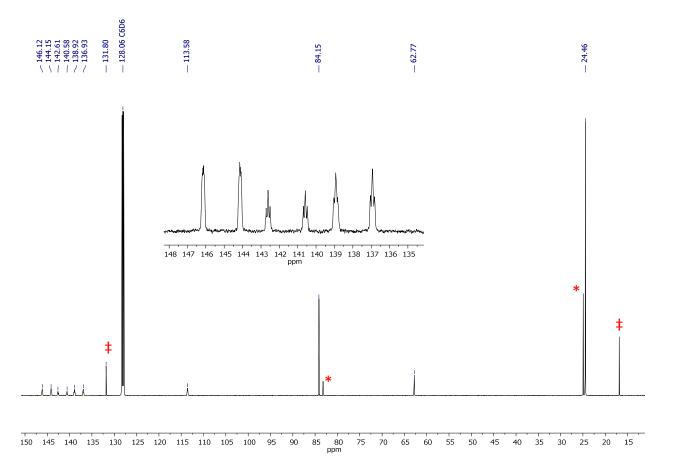
**Figure S17.** <sup>1</sup>H NMR spectrum of 2-(di-perfluorophenylmethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.



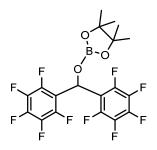


**Figure S18.** <sup>11</sup>B NMR spectrum of 2-(di-perfluorophenylmethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin.





**Figure S19.** <sup>13</sup>C NMR spectrum of 2-(di-perfluorophenylmethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.



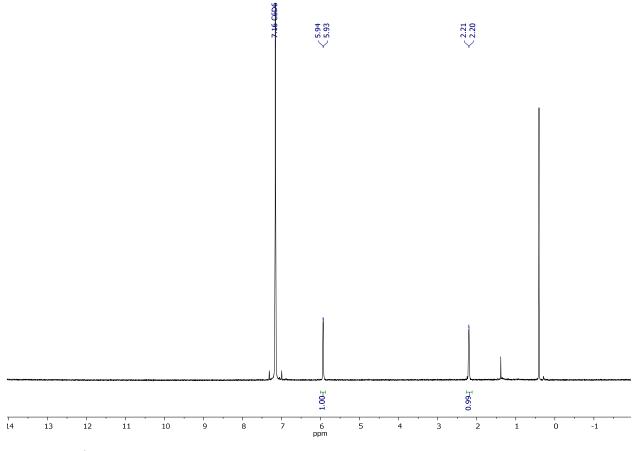
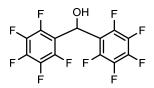


Figure S20. <sup>1</sup>H NMR spectrum of perfluorobenzhydrol acquired in benzene-d<sub>6</sub>.



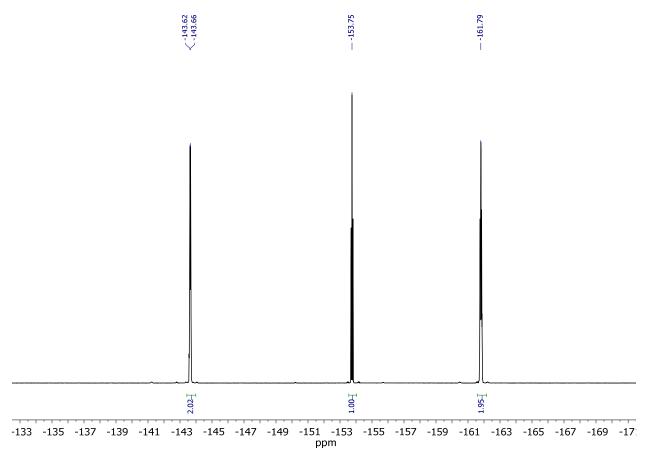
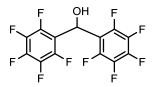


Figure S21. <sup>19</sup>F NMR spectrum of perfluorobenzhydrol acquired in benzene-d<sub>6</sub>.



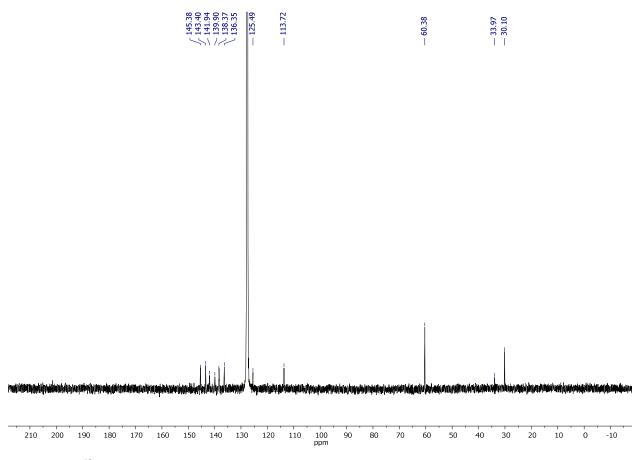
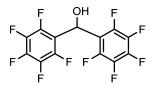
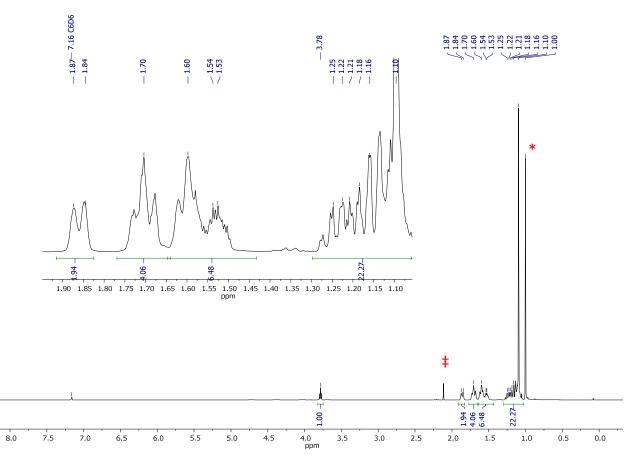
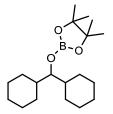


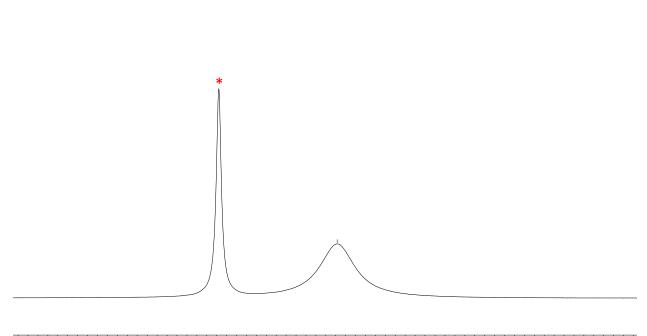
Figure S22. <sup>13</sup>C NMR spectrum of perfluorobenzhydrol acquired in benzene-d<sub>6</sub>.





**Figure S23.** <sup>1</sup>H NMR spectrum of 2-(dicyclohexylmethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.

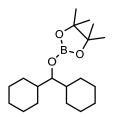


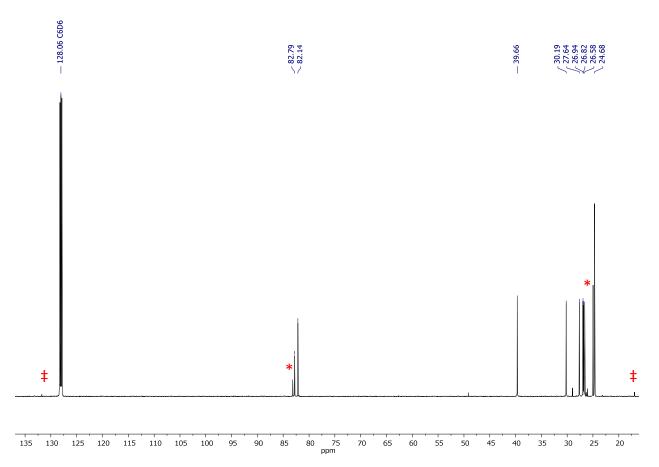


— 22.43

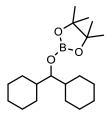
38 37 36 35 34 33 32 31 30 29 28 27 26 25 24 23 22 21 20 19 18 17 16 15 14 13 12 11 10 9 8 ppm

**Figure S24.** <sup>11</sup>B NMR spectrum of 2-(dicyclohexylmethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin.





**Figure S25.** <sup>13</sup>C NMR spectrum of 2-(dicyclohexylmethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.



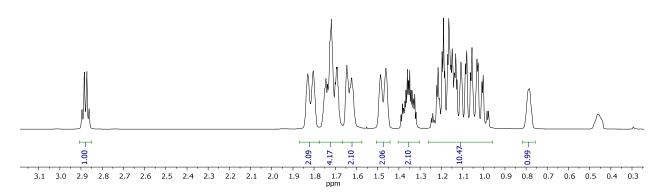
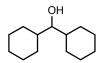
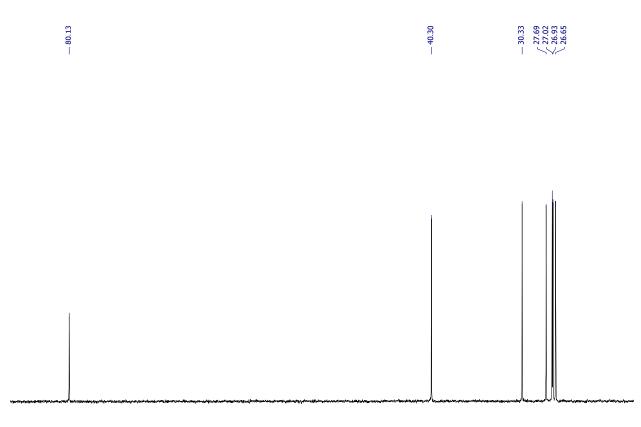
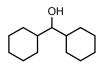


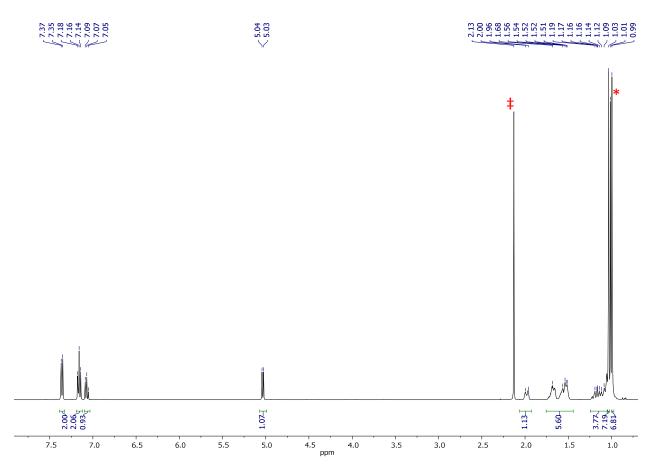
Figure S26. <sup>1</sup>H NMR spectrum of dicyclohexyl methanol acquired in benzene-d<sub>6</sub>.



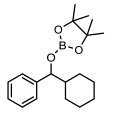


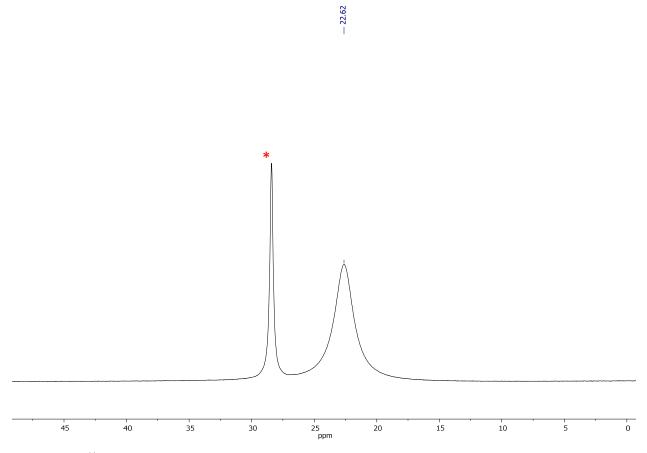
 $\frac{1}{86} = 84 = 82 = 80 = 78 = 76 = 74 = 72 = 70 = 68 = 66 = 64 = 62 = 60 = 58 = 56 = 54 = 52 = 50 = 48 = 46 = 44 = 42 = 40 = 38 = 36 = 34 = 32 = 30 = 28 = 26 = 24 = 22 = 20 = 10$ Figure S27. <sup>13</sup>C NMR spectrum of dicyclohexyl methanol acquired in benzene-d<sub>6</sub>.



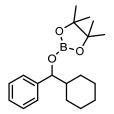


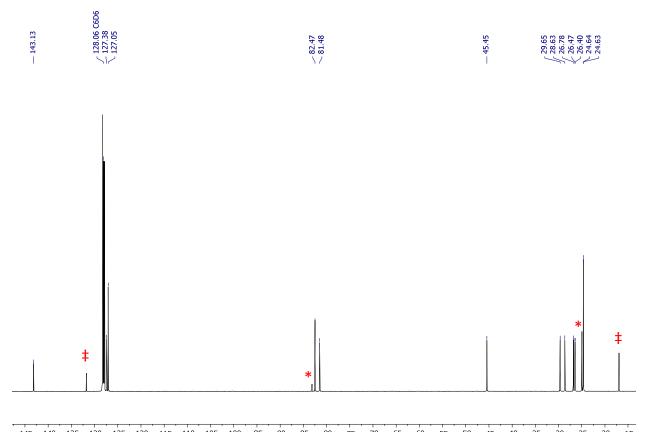
**Figure S28.** <sup>1</sup>H NMR spectrum of 2-(phenylcyclohexylmethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.



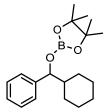


**Figure S29.** <sup>11</sup>B NMR spectrum of 2-(phenylcyclohexylmethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin.





<sup>145</sup> <sup>140</sup> <sup>135</sup> <sup>130</sup> <sup>125</sup> <sup>120</sup> <sup>115</sup> <sup>110</sup> <sup>105</sup> <sup>100</sup> <sup>95</sup> <sup>90</sup> <sup>85</sup> <sup>80</sup> <sup>75</sup> <sup>70</sup> <sup>65</sup> <sup>60</sup> <sup>55</sup> <sup>50</sup> <sup>45</sup> <sup>40</sup> <sup>35</sup> <sup>30</sup> <sup>25</sup> <sup>20</sup> <sup>15</sup> **Figure S30.** <sup>13</sup>C NMR spectrum of 2-(phenylcyclohexylmethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.



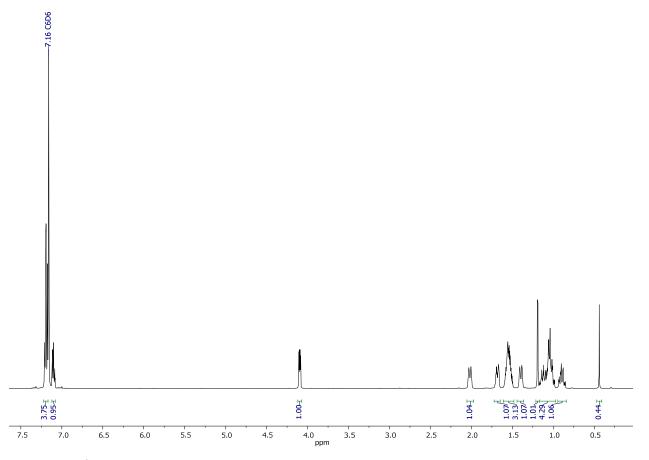
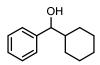


Figure S31. <sup>1</sup>H NMR spectrum of cyclohexyl(benzyl)alcohol acquired in benzene-d<sub>6</sub>.



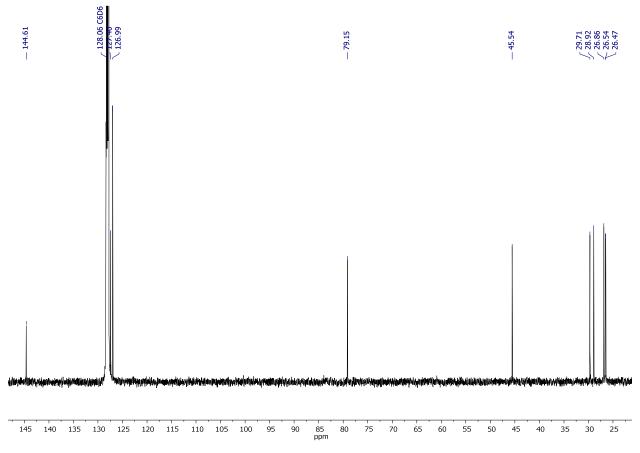
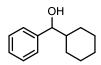
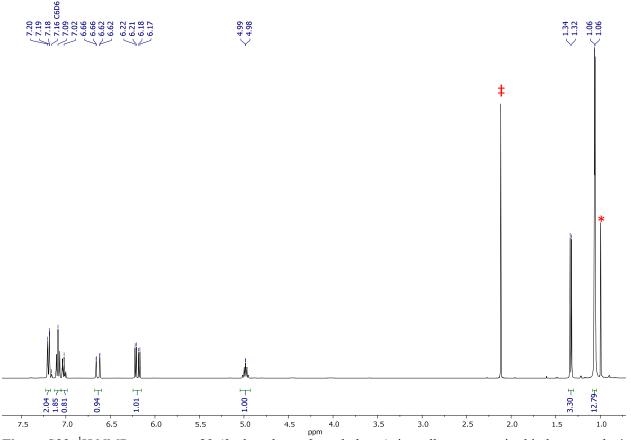
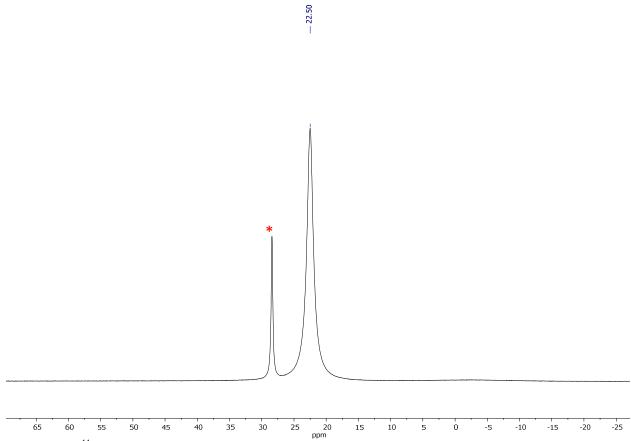


Figure S32. <sup>13</sup>C NMR spectrum of cyclohexyl(benzyl)alcohol acquired in benzene-d<sub>6</sub>.



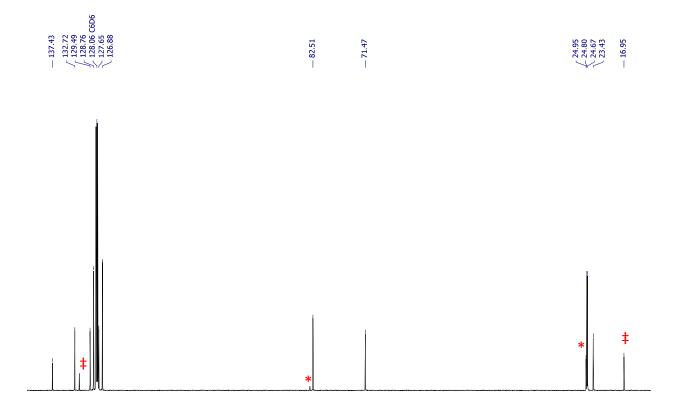


**Figure S33.** <sup>1</sup>H NMR spectrum of 2-(3-phenylprop-3-enylethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.

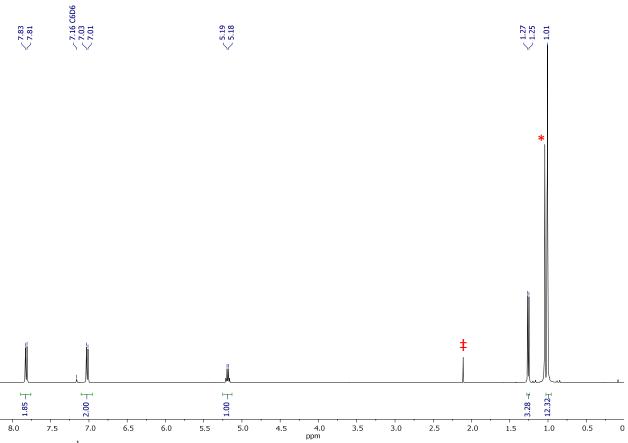


**Figure S34.** <sup>11</sup>B NMR spectrum of 2-(3-phenylprop-3-enylethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin.

0

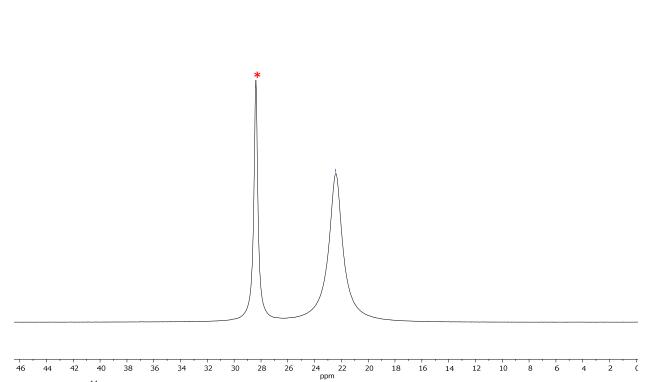


140 135 130 125 120 115 110 105 100 80 75 70 65 ppm **Figure S35.** <sup>13</sup>C NMR spectrum of 2-(3-phenylprop-3-enylethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.



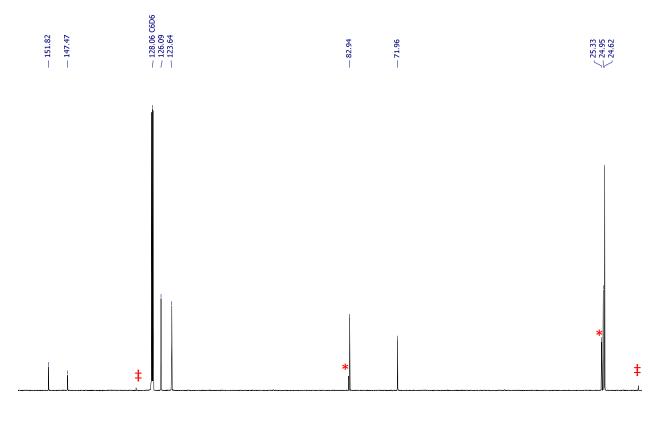
**Figure S36.** <sup>1</sup>H NMR spectrum of 2-(4-nitrophenylmethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.

ó O<sub>2</sub>N



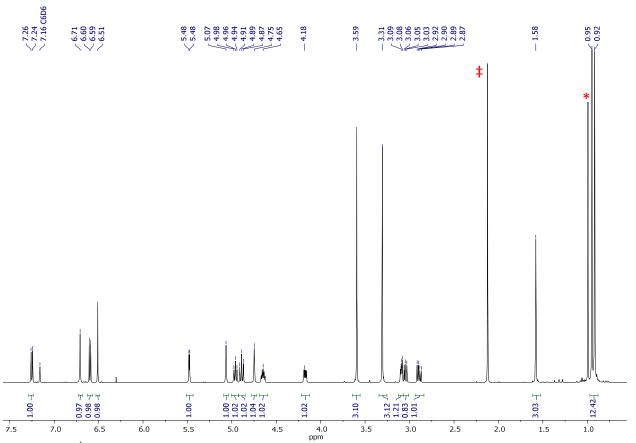
**Figure S37.** <sup>11</sup>B NMR spectrum of 2-(4-nitrophenylmethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin.

O O<sub>2</sub>N

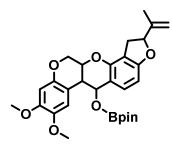


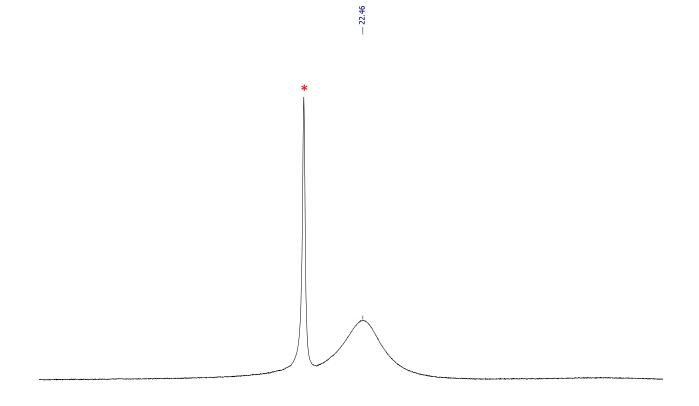
**Figure S38.** <sup>13</sup>C NMR spectrum of 2-(4-nitrophenylmethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.

۰, 0<sub>2</sub>N

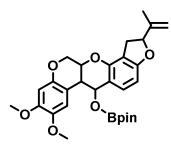


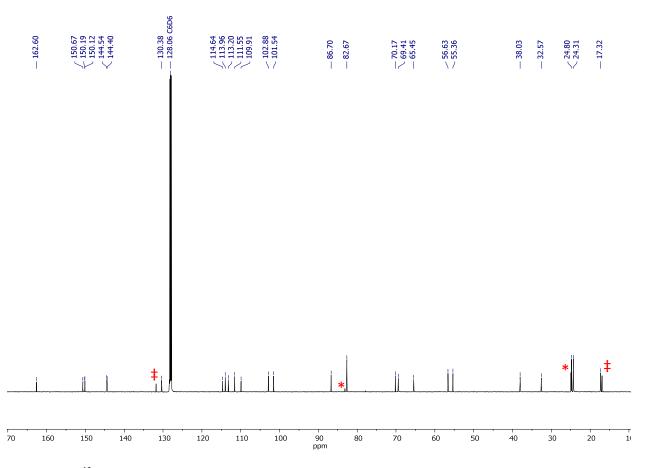
**Figure S39.** <sup>1</sup>H NMR spectrum of 2-(rotenoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.



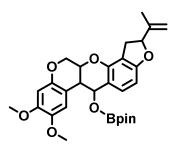


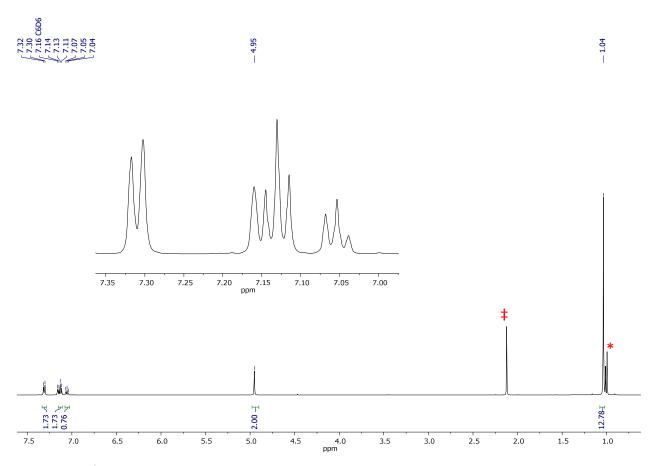
 $_{54}$   $_{52}$   $_{50}$   $_{48}$   $_{46}$   $_{44}$   $_{42}$   $_{40}$   $_{38}$   $_{36}$   $_{34}$   $_{32}$   $_{30}$   $_{28}$   $_{26}$   $_{24}$   $_{22}$   $_{20}$   $_{18}$   $_{16}$   $_{14}$   $_{12}$   $_{10}$   $_{8}$   $_{6}$   $_{4}$   $_{2}$   $_{0}$   $_{-2}$   $_{-4}$   $_{-6}$  -**Figure S40.**  $^{11}$ B NMR spectrum of 2-(rotenoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin.



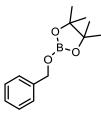


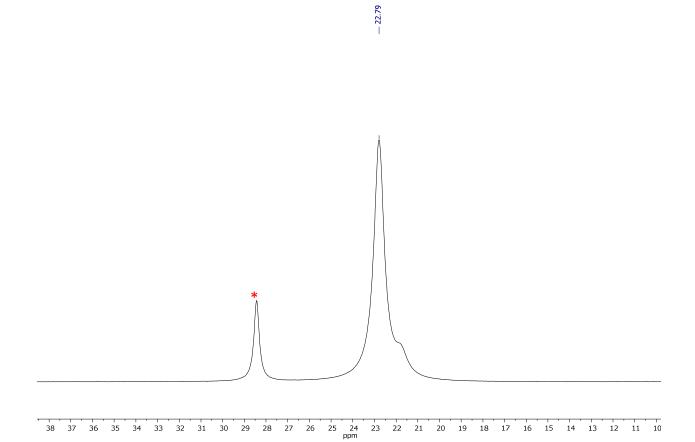
**Figure S41.** <sup>13</sup>C NMR spectrum of 2-(rotenoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.



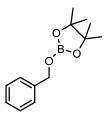


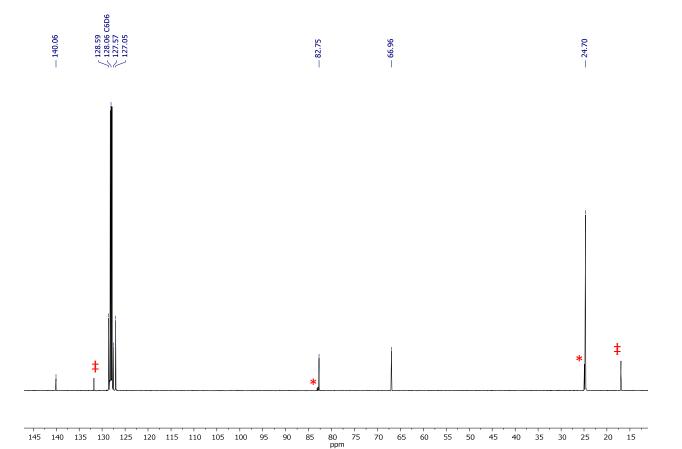
**Figure S42.** <sup>1</sup>H NMR spectrum of 2-(benzyloxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.





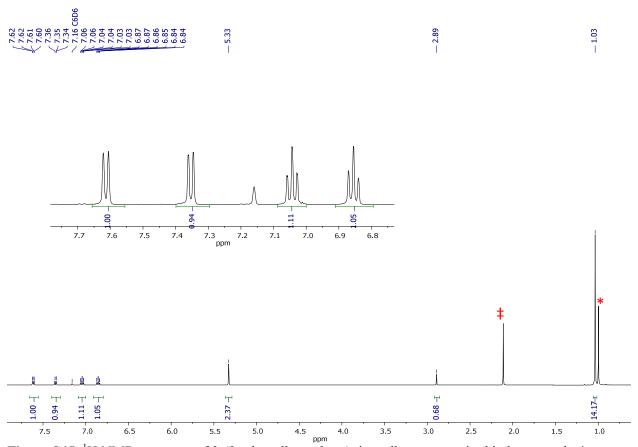
**Figure S43.** <sup>11</sup>B NMR spectrum of 2-(benzyloxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin.



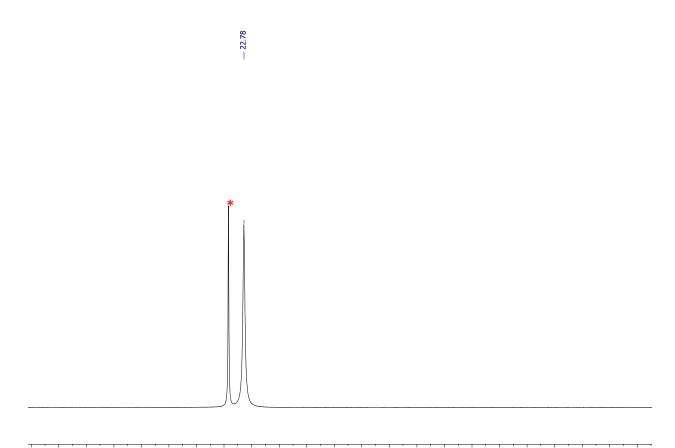


**Figure S44.** <sup>13</sup>C NMR spectrum of 2-(benzyloxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.

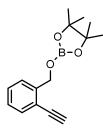
, P . В-

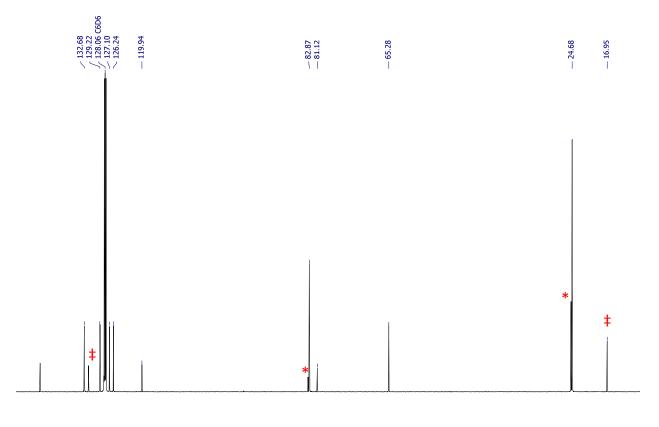


**Figure S45.** <sup>1</sup>H NMR spectrum of 2-(2-ethynylbenzyloxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.



**Figure S46.** <sup>11</sup>B NMR spectrum of 2-(2-ethynylbenzyloxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin.





<sup>145</sup> <sup>140</sup> <sup>135</sup> <sup>130</sup> <sup>125</sup> <sup>120</sup> <sup>115</sup> <sup>110</sup> <sup>105</sup> <sup>100</sup> <sup>95</sup> <sup>90</sup> <sup>85</sup> <sup>80</sup> <sup>75</sup> <sup>70</sup> <sup>65</sup> <sup>60</sup> <sup>55</sup> <sup>50</sup> <sup>45</sup> <sup>40</sup> <sup>35</sup> <sup>30</sup> <sup>25</sup> <sup>20</sup> <sup>15</sup> <sup>1</sup> **Figure S47.** <sup>13</sup>C NMR spectrum of 2-(2-ethynylbenzyloxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.

B-C

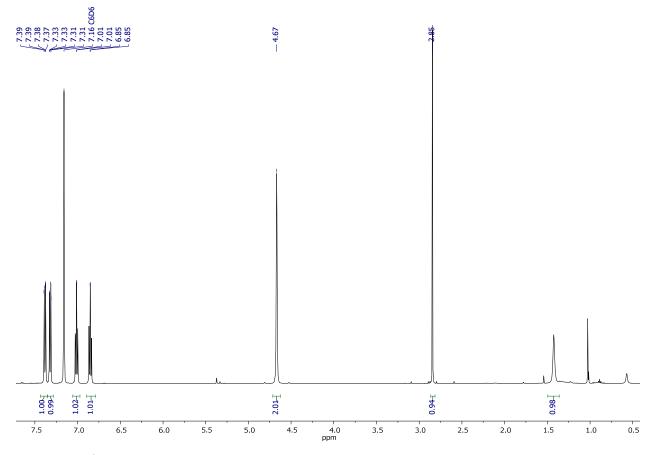


Figure S48. <sup>1</sup>H NMR spectrum of 2-ethynylbenzyl alcohol acquired in benzene-d<sub>6</sub>.



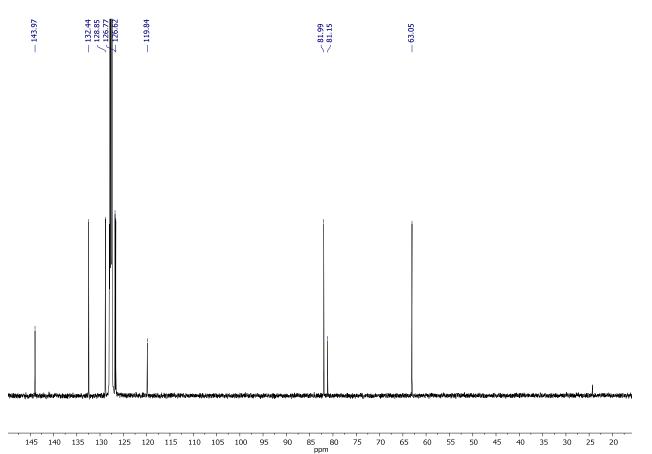
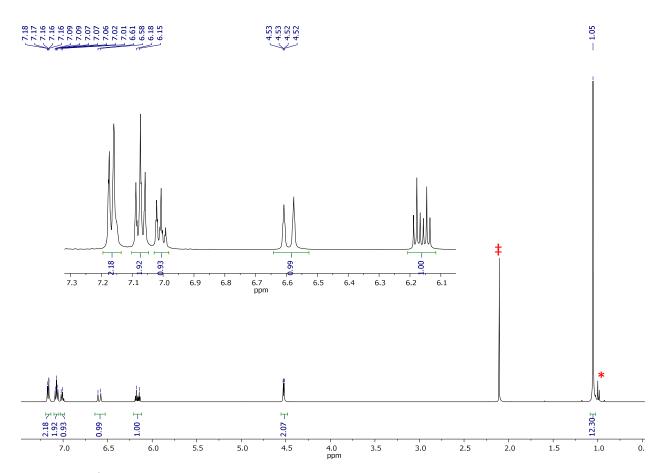




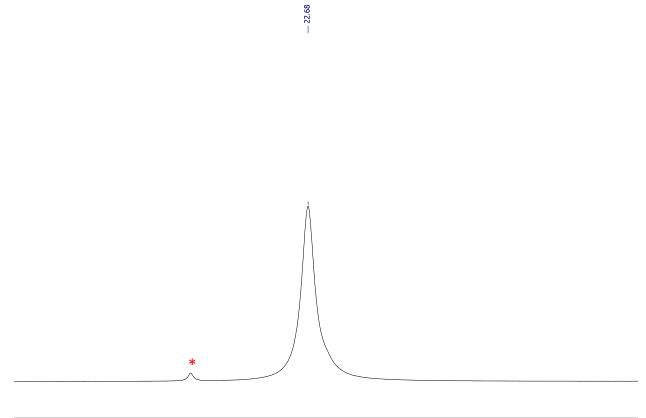
Figure S49. <sup>13</sup>C NMR spectrum of 2-ethynylbenzyl alcohol acquired in benzene-d<sub>6</sub>.





**Figure S50.** <sup>1</sup>H NMR spectrum of 2-(3-phenylprop-3-enylmethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.

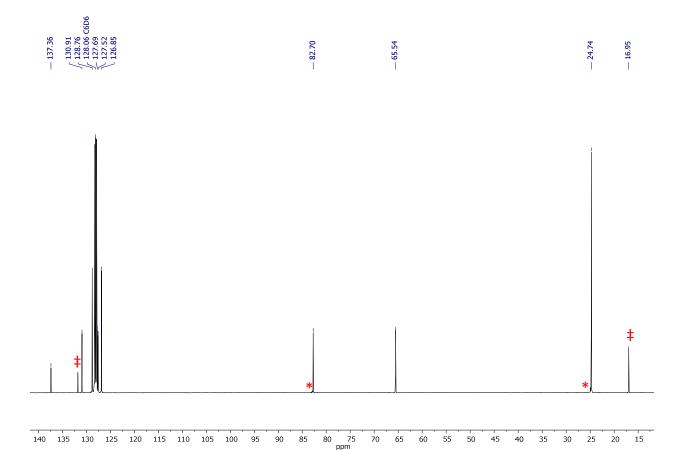
Ý



## 37 36 35 34 33 32 31 30 29 28 27 26 25 24 23 22 21 20 19 18 17 16 15 14 13 12 11 10 9 8 7 ppm <

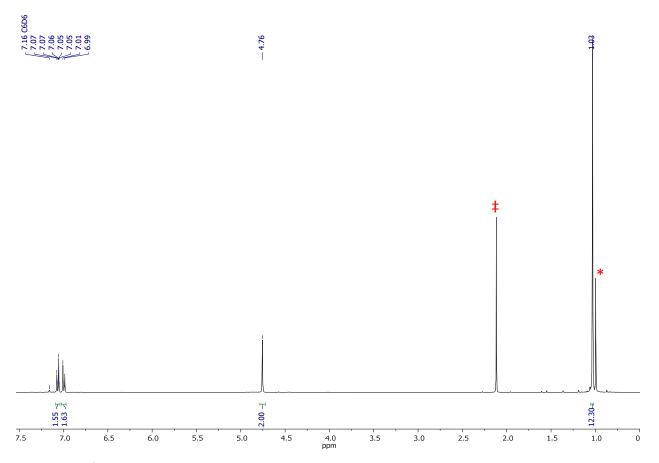
**Figure S51.** <sup>11</sup>B NMR spectrum of 2-(3-phenylprop-3-enylmethoxy)pinacolborane acquired in benzene- $d_6$ . \* indicates excess HBpin.

°, °, B-0

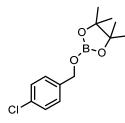


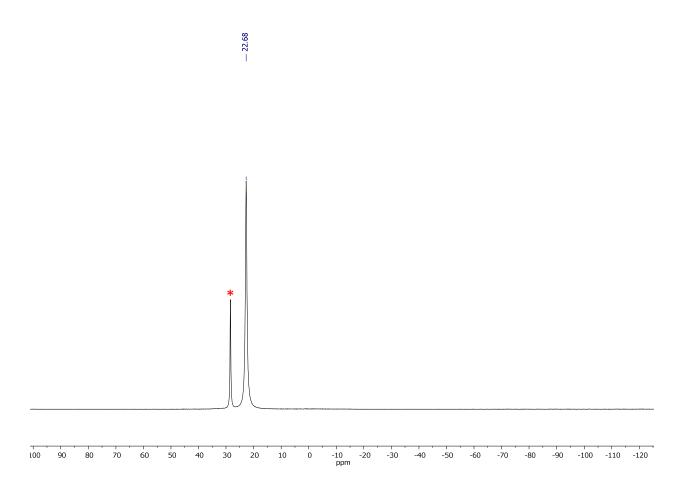
**Figure S52.** <sup>13</sup>C NMR spectrum of 2-(3-phenylprop-3-enylmethoxy)pinacolborane acquired in benzened<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.

о́,в. О́,В.

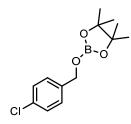


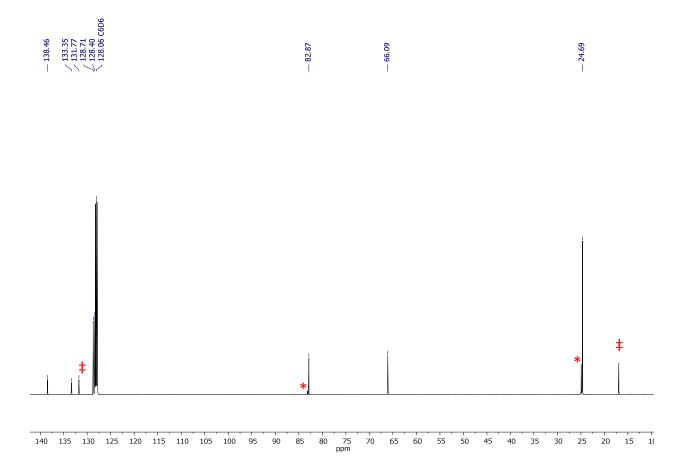
**Figure S53.** <sup>1</sup>H NMR spectrum of 2-(4-chlorobenzyloxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.





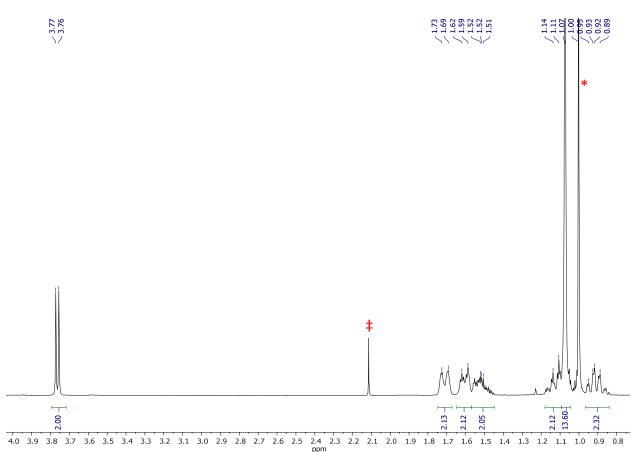
**Figure S54.** <sup>11</sup>B NMR spectrum of 2-(4-chlorobenzyloxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin.





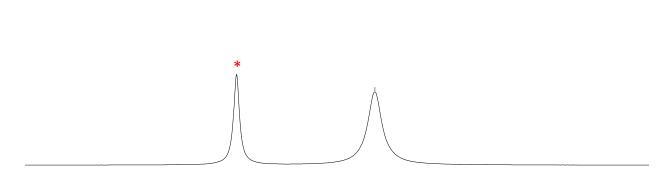
**Figure S55.** <sup>13</sup>C NMR spectrum of 2-(4-chlorobenzyloxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.

о<sup>В-0</sup>

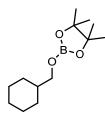


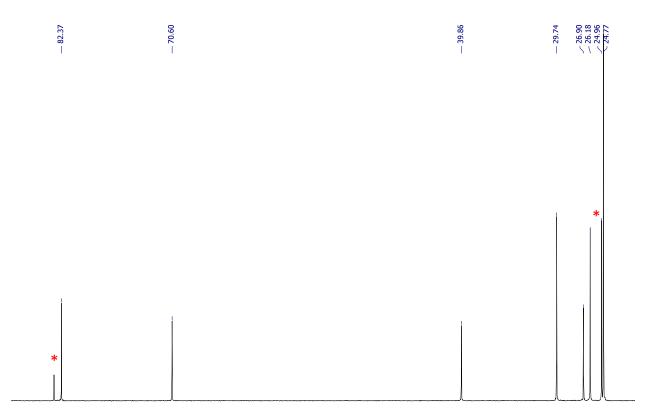
**Figure S56.** <sup>1</sup>H NMR spectrum of 2-(cyclohexylmethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.

or B-o



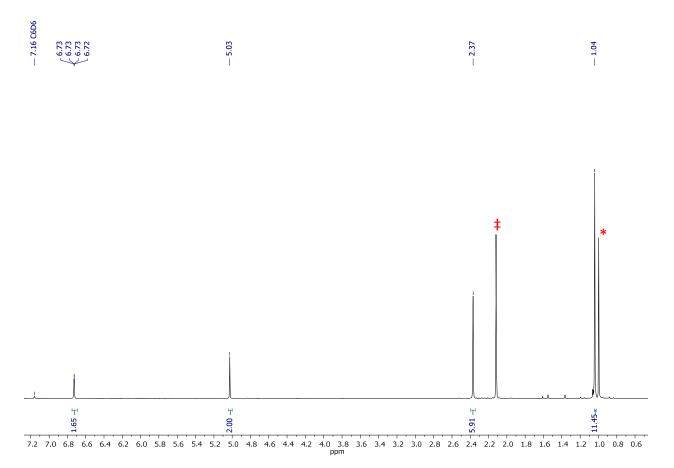
 $_{37}$   $_{36}$   $_{35}$   $_{34}$   $_{33}$   $_{32}$   $_{31}$   $_{30}$   $_{29}$   $_{28}$   $_{27}$   $_{26}$   $_{25}$   $_{24}$   $_{23}$   $_{22}$   $_{21}$   $_{20}$   $_{19}$   $_{18}$   $_{17}$   $_{16}$   $_{15}$   $_{14}$   $_{13}$   $_{12}$   $_{11}$ **Figure S57.**  $^{11}$ B NMR spectrum of 2-(cyclohexylmethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin.



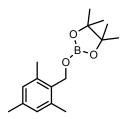


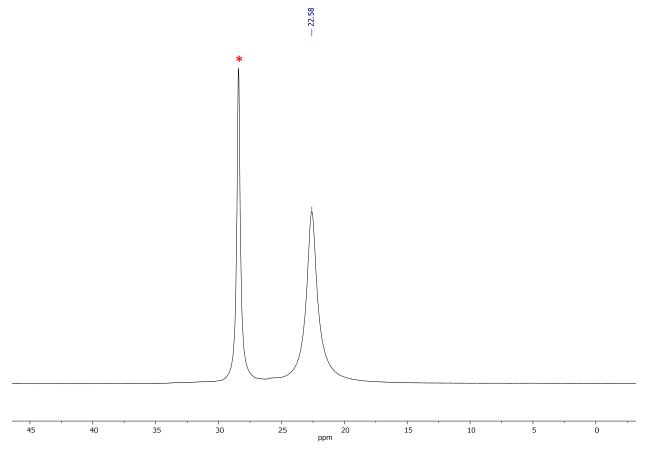
**Figure S58.** <sup>13</sup>C NMR spectrum of 2-(cyclohexylmethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin.

о , В-0

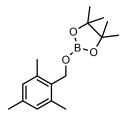


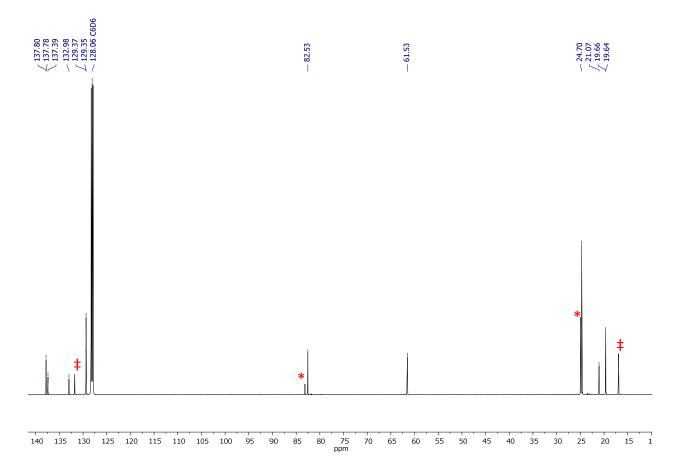
**Figure S59.** <sup>1</sup>H NMR spectrum of 2-(2,4,6-trimethylbenzyloxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.



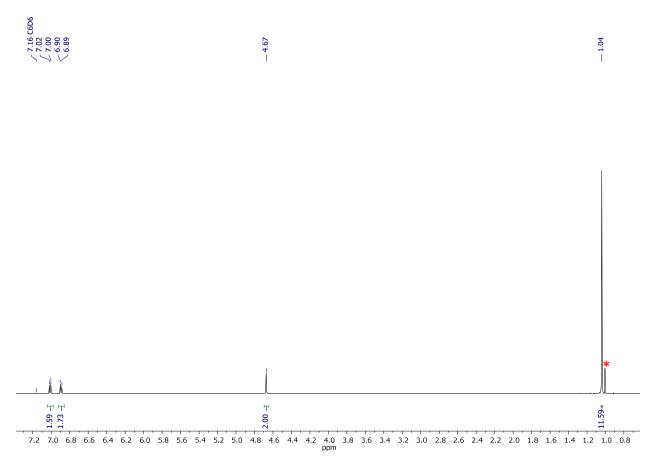


**Figure S60.** <sup>11</sup>B NMR spectrum of 2-(2,4,6-trimethylbenzyloxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin.



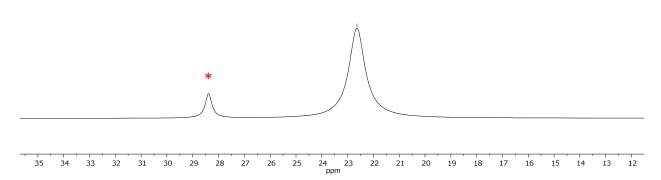


**Figure S61.** <sup>13</sup>C NMR spectrum of 2-(2,4,6-trimethylbenzyloxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.



**Figure S62.** <sup>1</sup>H NMR spectrum of 2-(4-cyanobenzyloxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin.

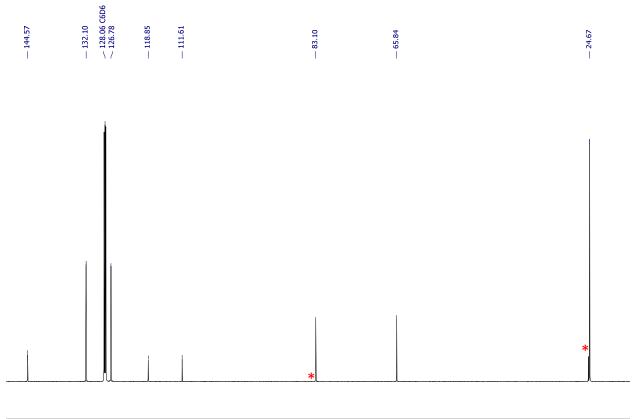
о́,<sup>в</sup>-о́ N∥



**Figure S63.** <sup>11</sup>B NMR spectrum of 2-(4-cyanobenzyloxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin.

о~ ,<sup>в</sup>-о́ N∭

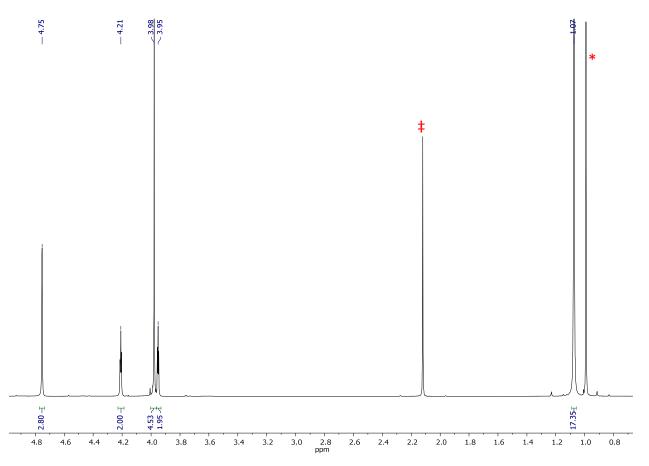
— 22.64



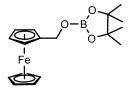
145 140 135 130 125 120 115 110 105 100 95 90 85 80 75 70 65 60 55 50 45 40 35 30 25 20 ppm

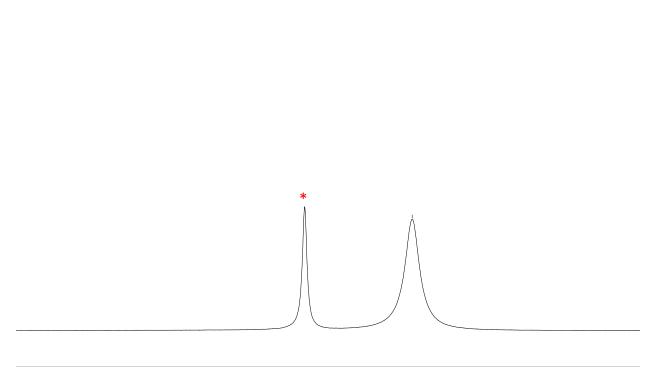
**Figure S64.** <sup>13</sup>C NMR spectrum of 2-(4-cyanobenzyloxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin.

0,<sup>4</sup>-0 N∭



**Figure S65.** <sup>1</sup>H NMR spectrum of 2-(ferrocenylmethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin, <sup>‡</sup> indicates hexamethylbenzene (internal standard) resonance.

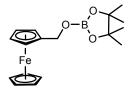


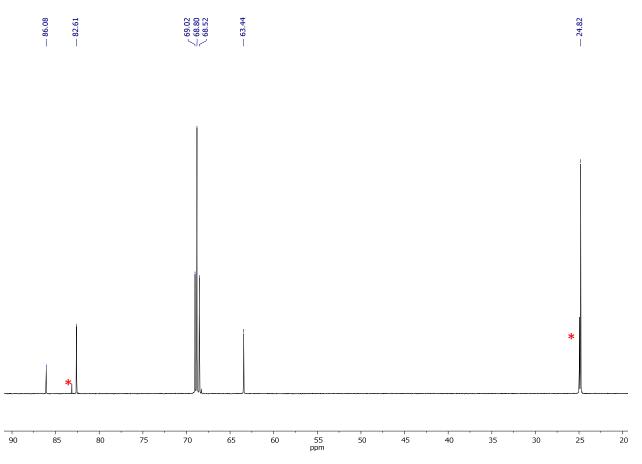


- 22.64

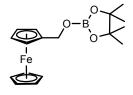
43 42 41 40 39 38 37 36 35 34 33 32 31 30 29 28 27 26 25 24 23 22 21 20 19 18 17 16 15 14 13 12 11 ppm

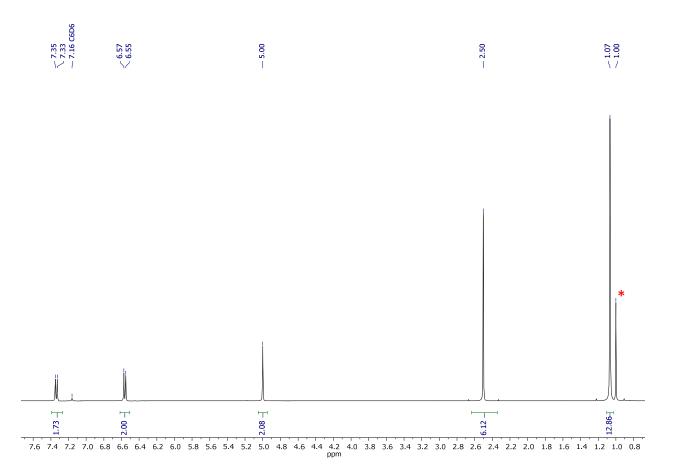
**Figure S66.** <sup>11</sup>B NMR spectrum of 2-(ferrocenylmethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin.



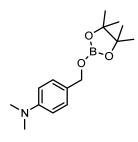


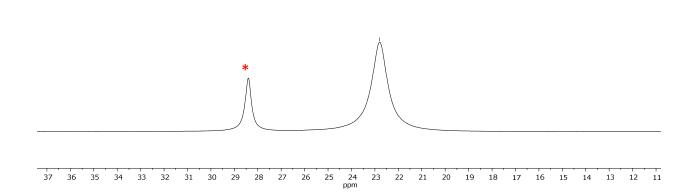
**Figure S67.** <sup>13</sup>C NMR spectrum of 2-(ferrocenylmethoxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin.



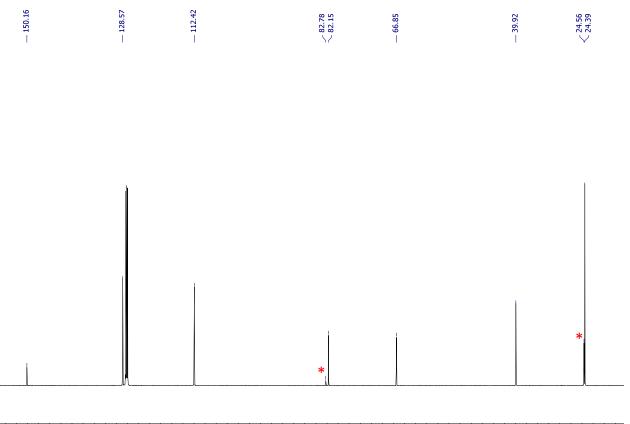


**Figure S68.** <sup>1</sup>H NMR spectrum of 2-(4-N,N-dimethylaminobenzyloxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin.





**Figure S69.** <sup>11</sup>B NMR spectrum of 2-(4-N,N-dimethylaminobenzyloxy)pinacolborane acquired in benzene-d<sub>6</sub>. \* indicates excess HBpin.



155 150 145 140 135 130 125 120 115 110 105 100 95 90 85 80 75 70 65 60 55 50 45 40 35 30 25 20 ppm

**Figure S70.** <sup>13</sup>C NMR spectrum of 2-(4-N,N-dimethylaminobenzyloxy)pinacolborane acquired in benzene- $d_6$ . \* indicates excess HBpin.

°,<sup>B</sup>~0′

## References

- (1) Dudnik, A.S.; Weidner, V. L.; Motta, A.; Delferro, M.; Marks, T. J. Nat. Chem. 2014, 6, 1100-1107.
- (2) Eedugurala, N.; Wang, Z.; Chaudhary, U.; Nelson, N.; Kandel, K.; Kobayashi, T.; Slowing, I. I.; Pruski, M; Sadow, A. D. ACS Catal. 2015, 5, 7399-7414.
- (3) Oluyadi, A. A.; Ma, S.; Muhoro, C. N. Organometallics 2013, 32, 70-78.
- (4) Chong, C. C.; Hirao, H.; Kinjo, R. Angew. Chem., Int. Ed. 2015, 54, 190-194.
- (5) Yang, Z.; Zhong, M.; Ma, X.; De, S.; Anusha, C.; Parameswaran, P.; Roesky, H. W. *Angew. Chem., Int. Ed.* **2015**, 54, 10225-10229.
- (6) Arrowsmith, M.; Hadlington, T. J.; Hill, M. S.; Kociok-Koehn, G. Chem. Commun. 2012, 48, 4567-4569.