Supporting Information

Frutescone A–G, Tasmanone-Based Meroterpenoids from the aerial parts of *Baeckea frutescens*

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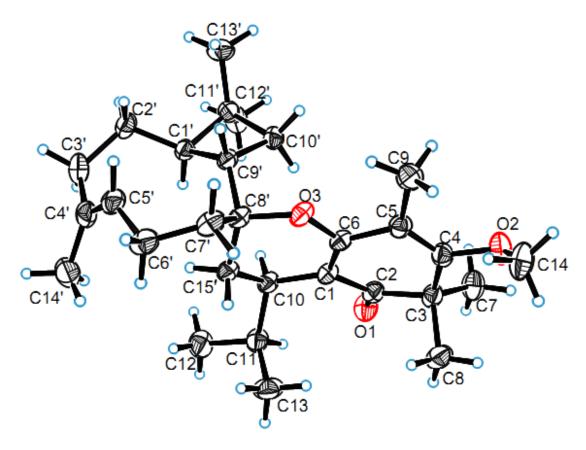


Figure S1. X-ray crystallographic structure of **1** (30% probability level)

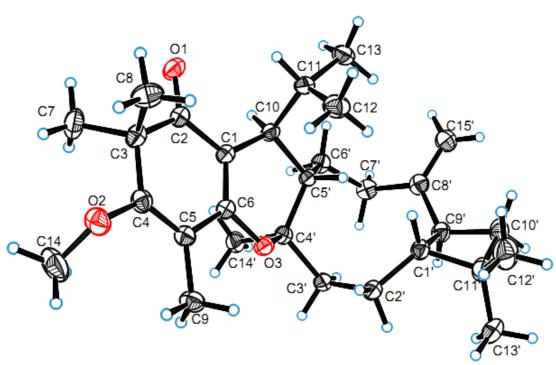


Figure S2. X-ray crystallographic structure of **2** (30% probability level)

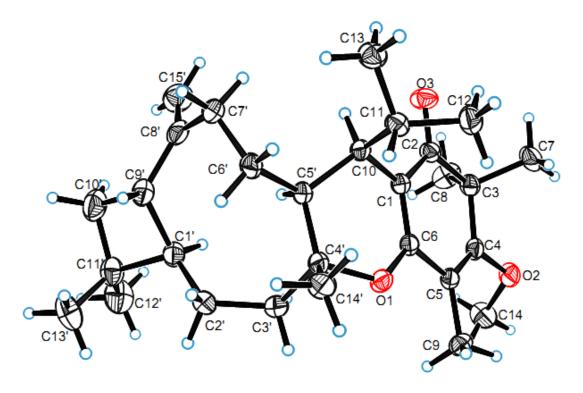


Figure S3. X-ray crystallographic structure of 3 (30% probability level)

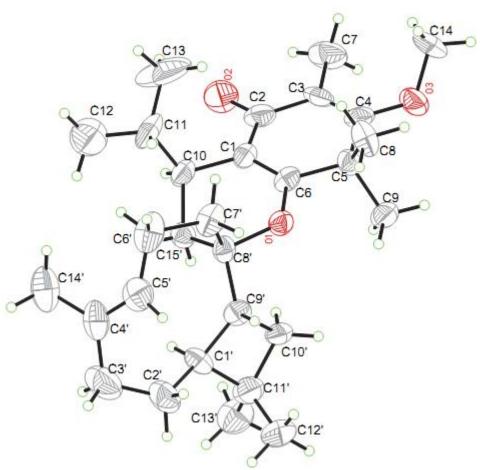


Figure S4. X-ray crystallographic structure of 4 (30% probability level)

Quantum chemical ECD calculation for 5 and (-)-7

Computational data of 5

The systematic random conformational analysis of compound **5** were performed in the SYBYL 8.1 program by using MMFF94s molecular force field, which afforded 12 conformers for **5** with an energy cutoff of 10 kcal mol⁻¹ to the global minima. All of the obtained conformers were further optimized using DFT at the B3LYP/6-31+G(d) level in gas phase by using Gaussian09 software, [1] and 4 conformers of **5** were selected. All of the optimized stable conformers were used for TDDFT computation of the excited stats at the same levels, with the consideration of the first 50 excitations. The overall ECD curves of **5** were weighted by Boltzmann distribution of each conformer (with a half-bandwidth of 0.3eV). The calculated ECD spectra of **5** was subsequently compared with the experimental ones. The ECD spectra were produced by SpecDis 1.6 software. [2]

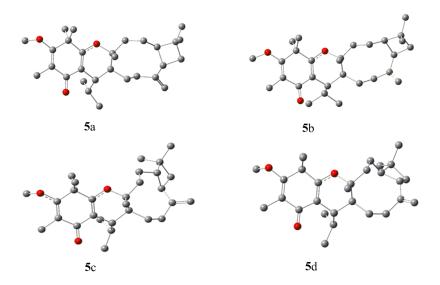


Figure S5-1. Optimized geometries of predominant conformers of **5** in the gas phase at the B3LYP/6-31+G(d) level

For $\mathbf{5a}$ absolute energies = -1357.2675802 atom coordinates:

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random search BF22d000011

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For **5b** absolute energies = -1357.2667287 atom coordinates:

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For 5c absolute energies = -1357.2667286 atom coordinates:

%nprocshared=12

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For **5d** absolute energies = -1357.2681166 atom coordinates:

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C	-1.69371000	-0. 47124300	0. 49431400
C	-2.75043700	-1.55122000	0.39809800
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0	-0.52449200	-0.95537900	0. 95798500
0	-3. 49845300	2. 53222900	-0.05952600
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C	-2.27193800	-2. 61016200	-0. 63262500
0	-5. 05154000	-1.95680100	-0. 15915700
C	-5. 72517400	0.89710700	-0.39370100
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C	3.65682400	-1.20197900	0. 20875300
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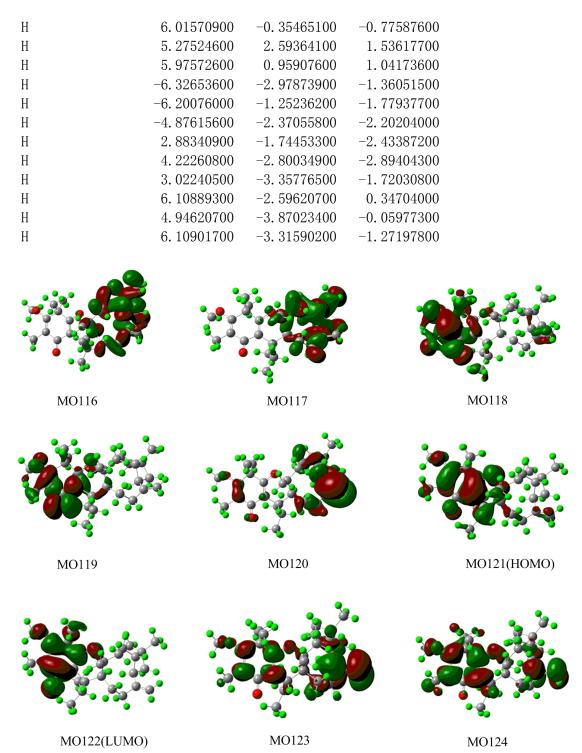


Figure S5-2. Key molecular orbitals involved in important transitions regarding the

ECD spectra of conformer 5c in the gas phase at the B3LYP/6-31+G(d) level

Table S1-1. Selected key transitions and their related rotatory and oscillator strengths of conformer **5**c of **5** at the B3LYP/6-31+G(d) level in the gas phase

	_		_	_	
NO	Energy	Energy Wavelength Osc.	Osc.	R	Major contribtions
	(cm ⁻¹)	(nm)	Strength	(length)	
1	31156.8913	320.95	0.0042	-7.9869	H-2->LUMO (72%)
2	33826.5602	295.62	0.1196	48.8292	HOMO->LUMO(82%)
3	39287.6865	254.53	0.0172	8.9492	H-1->LUMO (94%)
4	40066.0039	249.59	0.1448	47.5732	H-3->LUMO(69%)
5	45076.2707	221.84	0.0008	5.1191	H-6->LUMO (61%)
_	16102 6011	215.12	0.0055	4.0404	H-4->LUMO(29%),
6	46483.6944	215.13	0.0055	4.0404	H-6->LUMO (21%)
7	47126.5120	212.19	0.0633	4.1758	HOMO->L+1(43%)
8	47520.1066	210.44	0.0033	-3.209	HOMO->L+1(38%)
9	47704.8058	209.62	0.0022	2.7044	H-4->LUMO (63%)
10	48342.7841	206.85	0.0002	0.2577	H-5->LUMO (85%)
11	48723.4740	205.24	0.095	28.5924	HOMO->L+1(32%)
12	49288.0566	202.89	0.0295	-28.0208	H-8->LUMO (76%)
12	50027 4204	1067	0.2650	22.2617	H-3->L+1(28%)
13	50837.4324	196.7	0.2659	-33.3617	H-3->L+2(35%)
1.4	51000 4400	102.05	0.025	45 9770	H-1->L+1 (53%)
14	51800.4489	193.05	0.035	-45.8772	H-1->L+2(43%)
15	52216.6269	191.51	0.0213	-17.5747	H-9->LUMO (50%)
16	52250 5772	101.25	0.0027	1 (705	H-2->L+1(56%)
16	52258.5673	191.35	0.0037	1.6795	H-2->L+2 (28%)
17	52002 9045	190.02	0.0257	19.0692	H-3->L+1(47%)
17	52903.8045	189.02	0.0257	18.9683	H-3->L+2 (30%)
18	53292.5599	187.64	0.2461	-49.5004	H-1->L+2 (39%)
19	54379.7846	183.89	0.0006	0.2376	H-9->LUMO (36%)
20	55095.1914	181.5	0.0451	38.9386	H-4->L+1(61%)

Computational data of (-)-7

The systematic random conformational analysis of compound (–)-7 were performed in the SYBYL 8.1 program by using MMFF94s molecular force field, which afforded 8 conformers for (–)-7 with an energy cutoff of 10 kcal mol⁻¹ to the global minima. All of the obtained conformers were further optimized using DFT at the B3LYP/6-31+G(d) level in gas phase by using Gaussian09 software, [1] and 4 conformers of (–)-7 were selected. All of the optimized stable conformers were used for TDDFT computation of the excited stats at the same levels, with the consideration of the first 50 excitations. The overall ECD curves of (–)-7 were weighted by Boltzmann distribution of each conformer (with a half-bandwidth of 0.3eV). The calculated ECD spectra of (–)-7 was subsequently compared with the experimental ones. The ECD spectra were produced by SpecDis 1.6 software. [2]

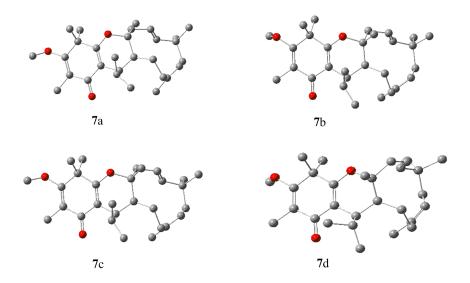


Figure S5-3. Optimized geometries of predominant conformers of (–)-7 in the gas phase at the B3LYP/6-31+G(d) level

For (-)-7a absolute energies = -1357.2716973

atom coordinates:

%nprocshared=12

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guess=read geom=(connectivity, allcheck) test

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For (-)-7b

absolute energies = -1357.2676662

atom coordinates:

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For (-)-7d

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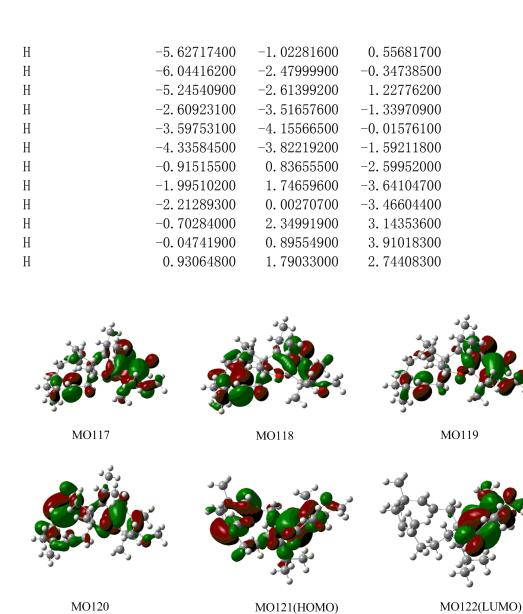
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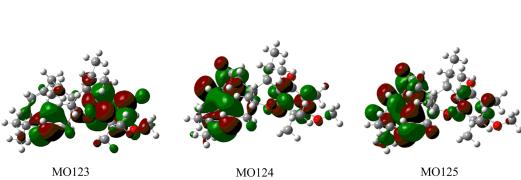


Figure S5-4. Key molecular orbitals involved in important transitions regarding the ECD spectra of conformer **7**a in the gas phase at the B3LYP/6-31+G(d) level

Table S1-2. Selected key transitions and their related rotatory and oscillator strengths of conformer **7**a of (–)-**7** at the B3LYP/6-31+G(d) level in the gas phase

номо	is 121				
NO	Energy	Wavelength	Osc.	R	Material Children
NO	(cm ⁻¹)	(nm)	Strength	(length)	Major contribtions
1	21055 7647	221.55	0.0042	0.7762	H-3->LUMO (32%)
1	31055.7647	321.55	0.0043	-0.7762	H-2->LUMO (48%)
2	33112.0271	301.58	0.1292	-80.2237	H-1->LUMO (13%)
۷	33112.0271	301.36	0.1292	-80.2237	H->LUMO (78%)
3	34983.8462	285.45	0.0048	-0.3988	H-1->LUMO (84%)
3	34703.0402	203.43	0.0040	-0.3700	H->LUMO (15%)
4	38887.7647	256.79	0.0235	29.8081	H-3->LUMO (59%)
7	30007.7047	230.17	0.0233	29.0001	H-2-> LUMO (36%)
5	39789.8462	250.97	0.1328	-58.6780	H-4->LUMO (71%)
6	44700.552	223.40	0.0009	3.7490	H-5->LUMO (80%)
7	45945.7466	217.34	0.0202	1.6724	HUMO->L+136%)
					H-6->LUMO(31%)
8	46484.5792	214.83	0.0335	46.9385	H-1->L+1(18%)
					HUMO->L+1(15%)
9	47271.4842	211.25	0.0510	106.9159	H-6->LUMO (28%)
10	47812.733	208.86	0.0053	-2.4385	H-2->L+1(38%)
10	47012.733	200.00	0.0055	2.4303	H-3->L+1(25%)
11	48014.0905	207.98	0.0041	-2.3365	H-7->LUMO (53%)
11	40014.0903	207.50	0.0041	2.3303	H-6->LUMO (26%)
12	48327.4027	206.63	0.0193	-11.6174	HUMO->L+2(70%)
13	48822.7421	204.54	0.0094	6.6368	HUMO->L+3(40%)
13	40022.7421	204.54	0.0074	0.0300	H-1->L+2(28%)
14	49417.1493	202.08	0.0452	-26.2473	HUMO->L+3(35%)
15	50780.7421	196.65	0.0130	-14.8683	H-8->LUMO (66%)
16	51012.7059	195.76	0.0692	40.8003	H-1->L+3(21%)
17	51205.2036	195.02	0.1173	33.1329	H-4->L+1(49%)
18	52178.1629	191.38	0.2615	80.2018	H-3->L+1(37%)
19	52532.552	190.09	0.1526	75.6914	H-3 ->L+2(18%)
20	53063.3303	188.19	0.0524	-37.7322	H-9-> LUMO (57%)

	m/z 🛆	Ion	Formula	Abundance					
-	441.3365	(M+H)+	C29 H45 O3	408459.7					
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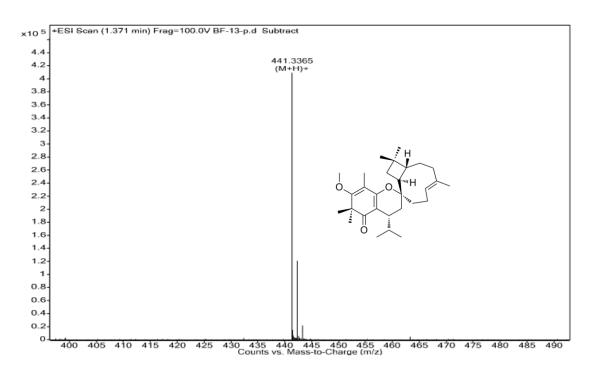


Figure S6. HR-ESI-MS spectrum of 1

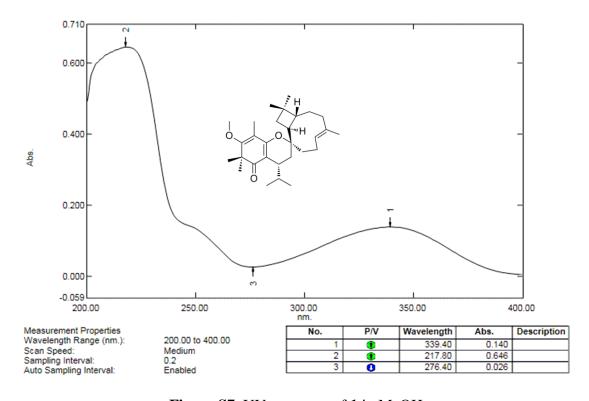


Figure S7. UV spectrum of 1 in MeOH

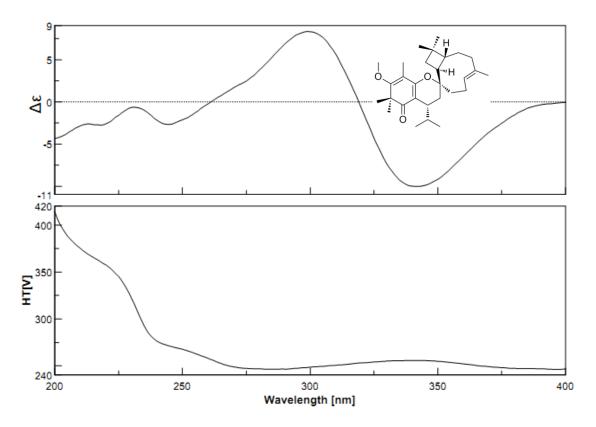


Figure S8. CD spectrum of 1 in MeOH

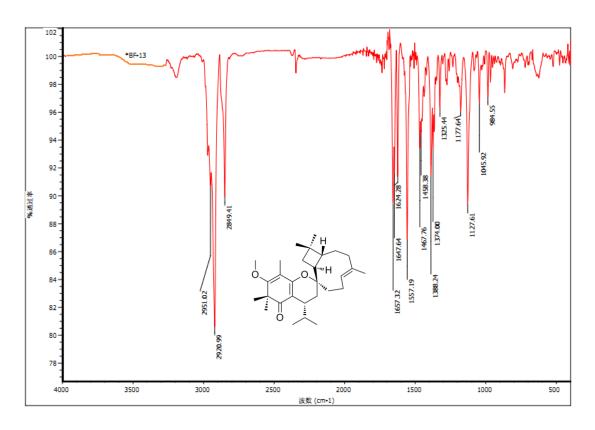


Figure S9. IR spectrum of 1

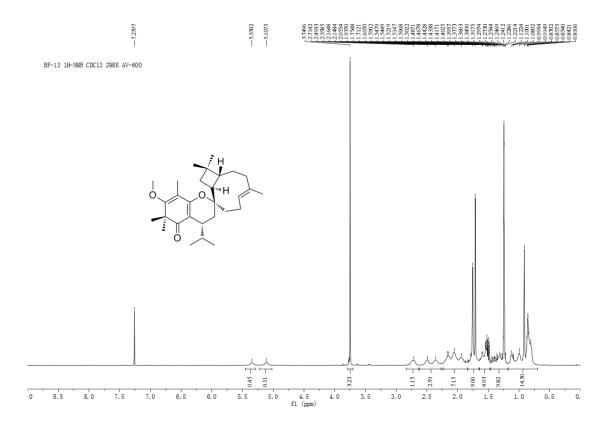


Figure S10-1. ¹H NMR spectrum of 1 in CDCl₃ (600 MHz, 298K)

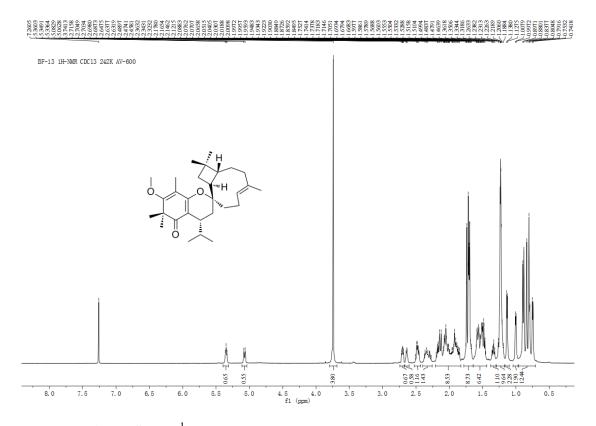


Figure S10-2. ¹H NMR spectrum of 1 in CDCl₃ (600 MHz, 242K)

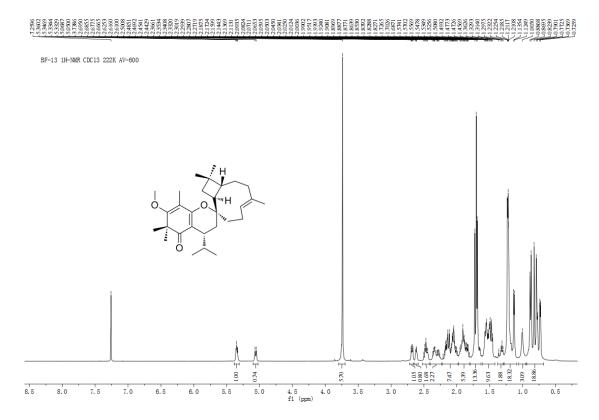


Figure S10-3. ¹H NMR spectrum of 1 in CDCl₃ (600 MHz, 222K)

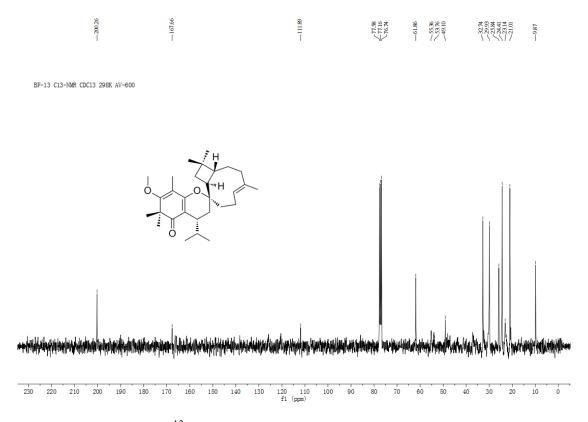


Figure S11-1. ¹³C NMR spectrum of **1** in CDCl₃ (150 MHz, 298K)

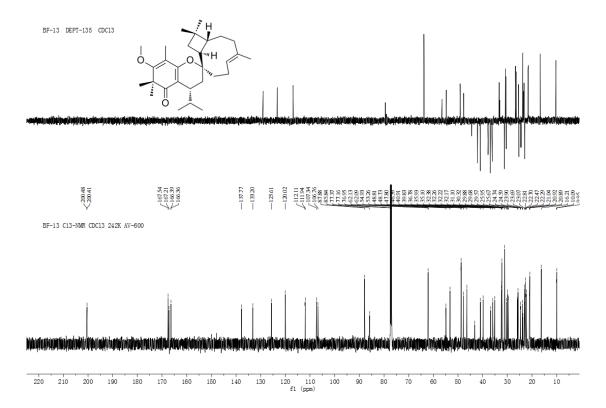


Figure S11-2. ¹³C NMR spectrum of **1** in CDCl₃ (150 MHz, 242K)

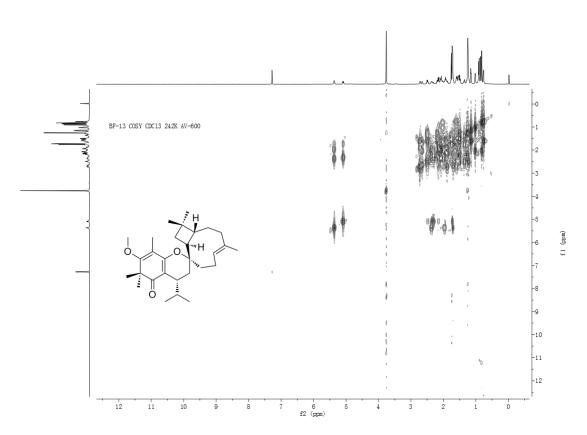


Figure S12. ¹H-¹H COSY spectrum of **1** in CDCl₃ (600 MHz, 242K)

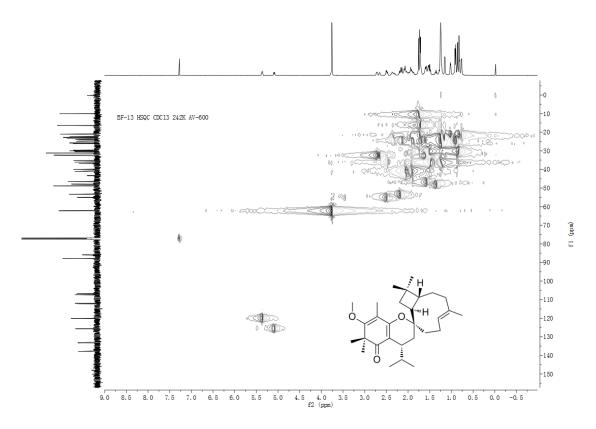


Figure S13. HSQC spectrum of 1 in CDCl₃ (600 MHz, 242K)

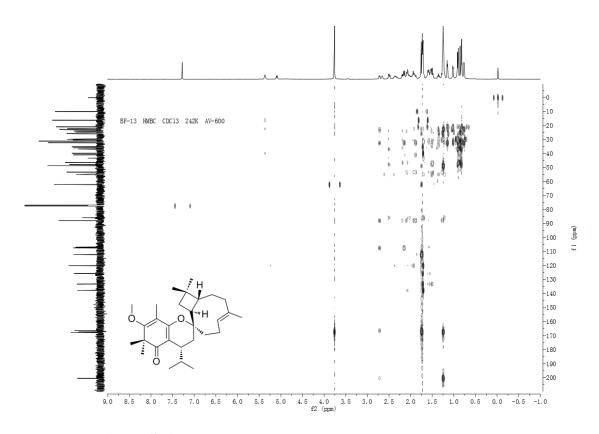


Figure S14. HMBC spectrum of 1 in CDCl₃ (600 MHz, 242K)

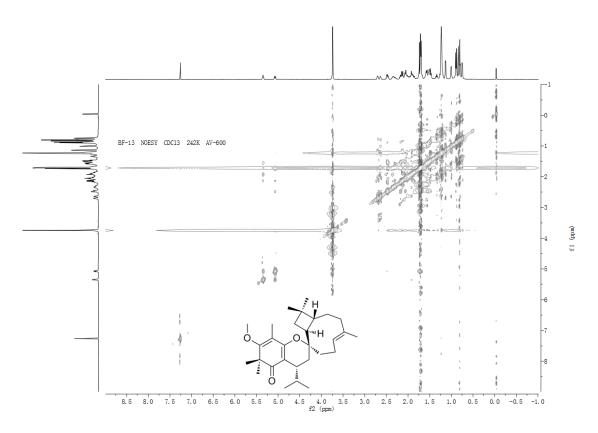


Figure S15. NOESY spectrum of 1 in CDCl₃ (600 MHz, 242K)

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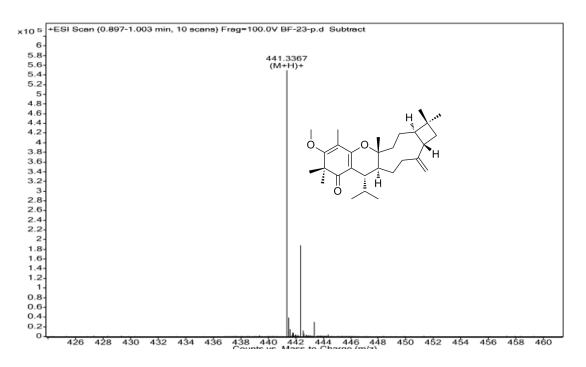


Figure S16. HR-ESI-MS spectrum of 2

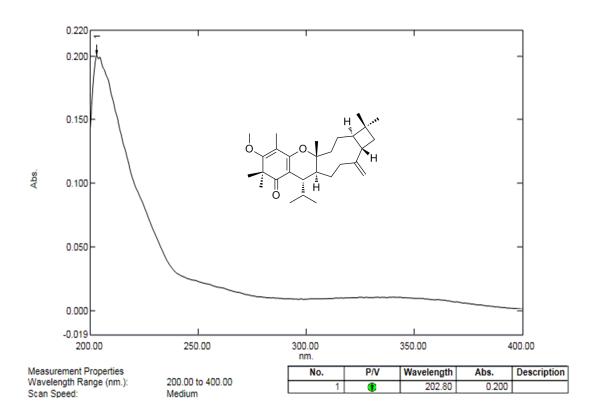


Figure S17. UV spectrum of 2 in MeOH

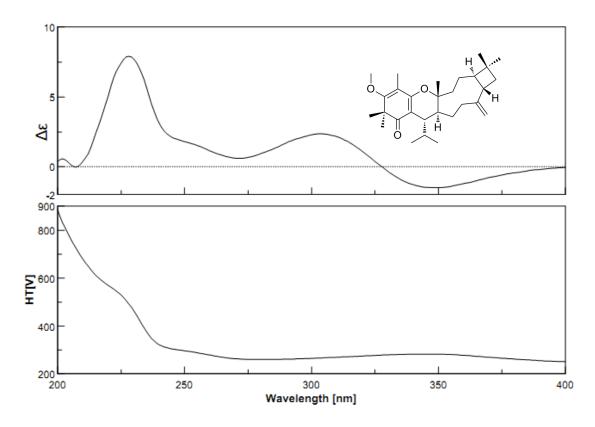


Figure S18. CD spectrum of 2 in MeOH

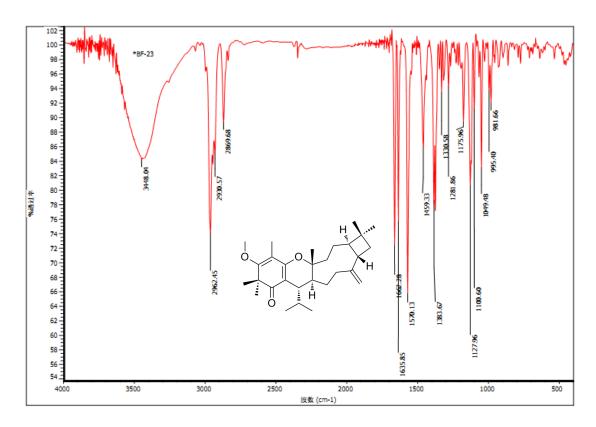


Figure S19. IR spectrum of 2

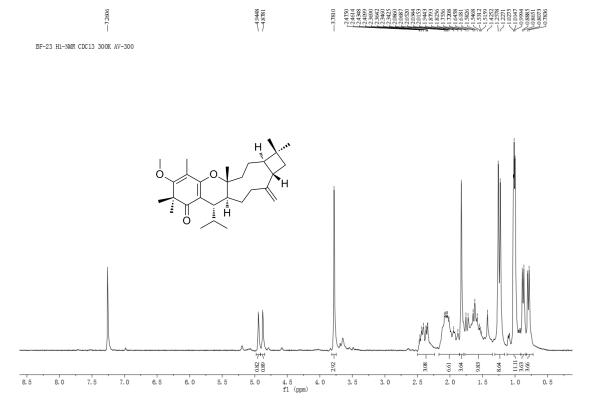


Figure S20. ¹H NMR spectrum of 2 in CDCl₃ (300 MHz)

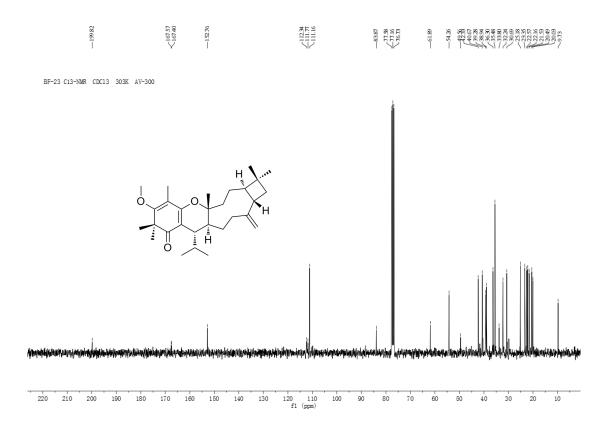


Figure S21. ¹³C NMR spectrum of **2** in CDCl₃ (75 MHz)

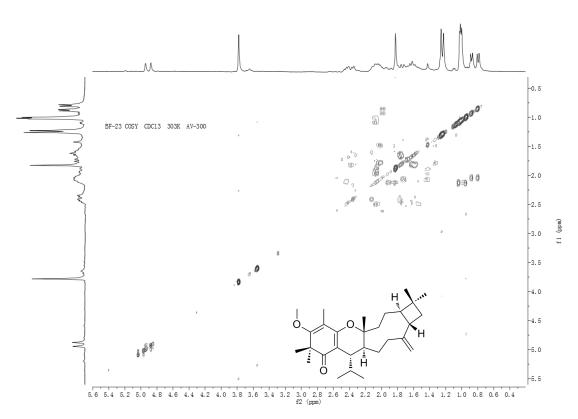


Figure S22. ¹H-¹H COSY spectrum of **2** in CDCl₃ (300 MHz)

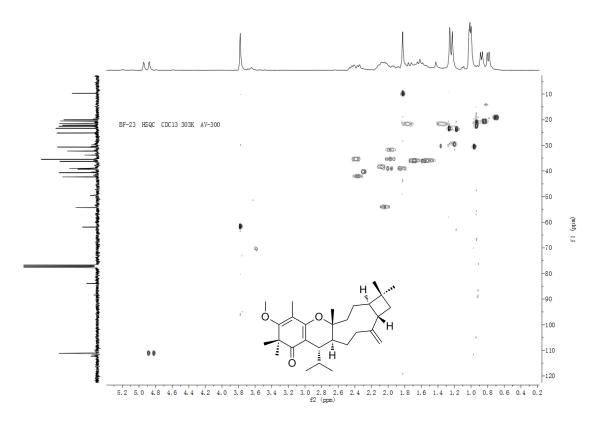


Figure S23. HSQC spectrum of 2 in $CDCl_3$ (300 MHz)

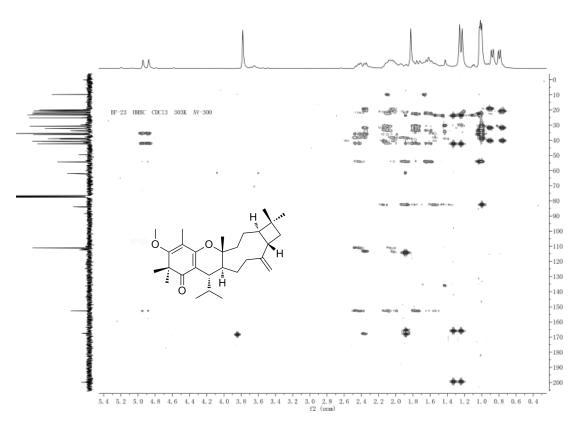


Figure S24. HMBC spectrum of 2 in CDCl₃ (300 MHz)

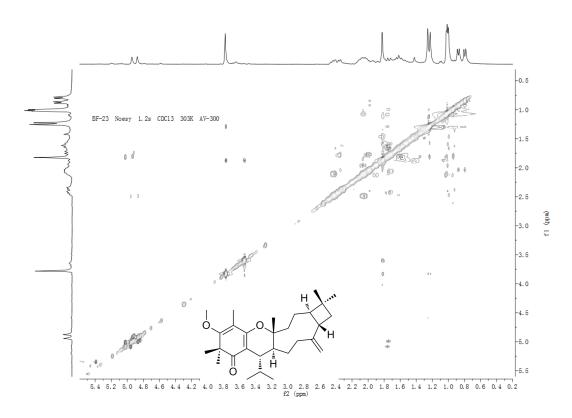


Figure S25. NOESY spectrum of 2 in CDCl₃ (300 MHz)

	m/z	lon	Formula /	Abundance					
	441.3359	(M+H)+	C29 H45 O3	814759					
	Best	Formula (M)	Ion Formula	Calc m/z	Score ∇	Cross Score	Mass	Calc Mass	Diff (ppm)
ŧ	 <u> </u>	C29 H44 O3	C29 H45 O3	441.3363	97.64		440.3287	440.329	0.79

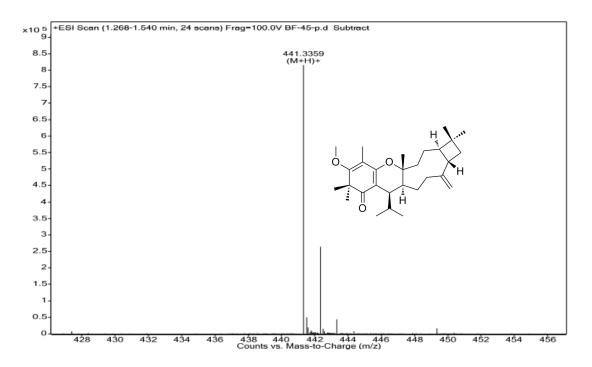


Figure S26. HR-ESI-MS spectrum of 3

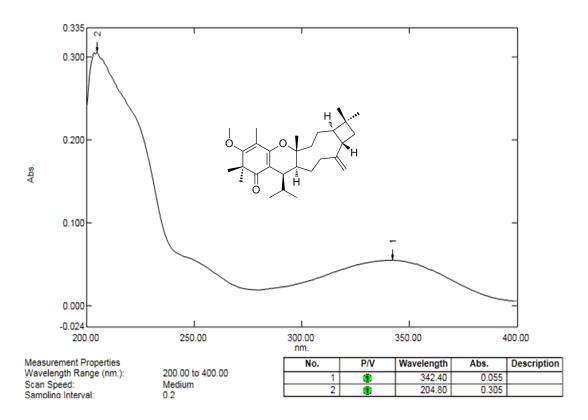


Figure S27. UV spectrum of 3 in MeOH

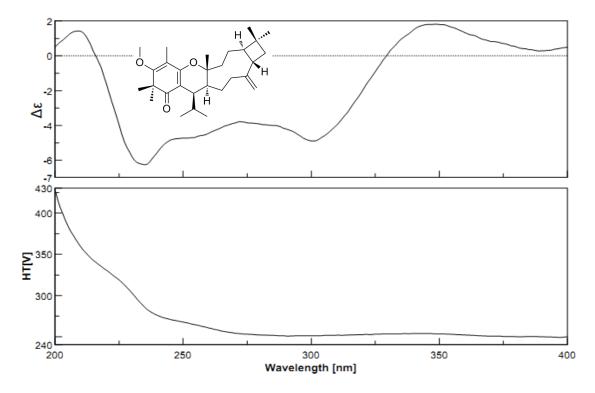


Figure S28. CD spectrum of 3 in MeOH

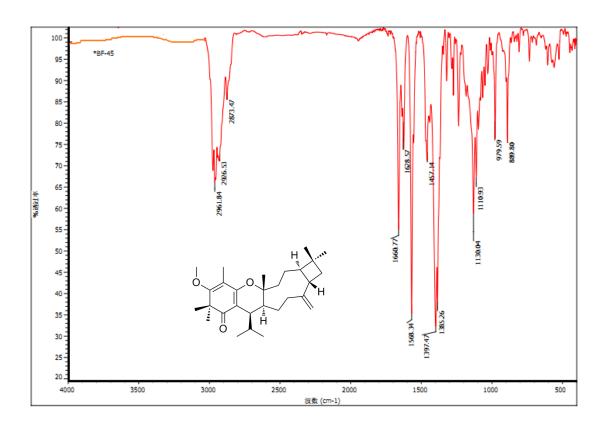


Figure S29. IR spectrum of 3

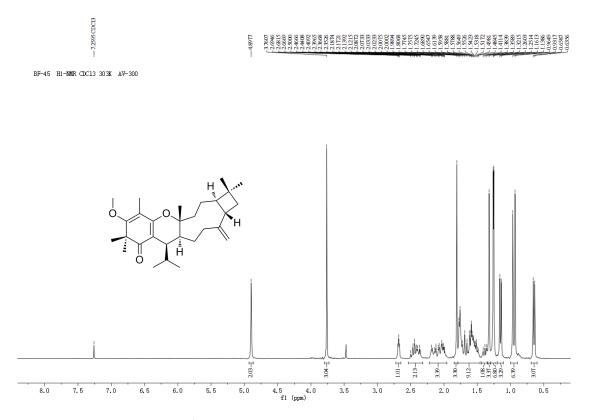


Figure S30. ¹H NMR spectrum of 3 in CDCl₃ (300 MHz)

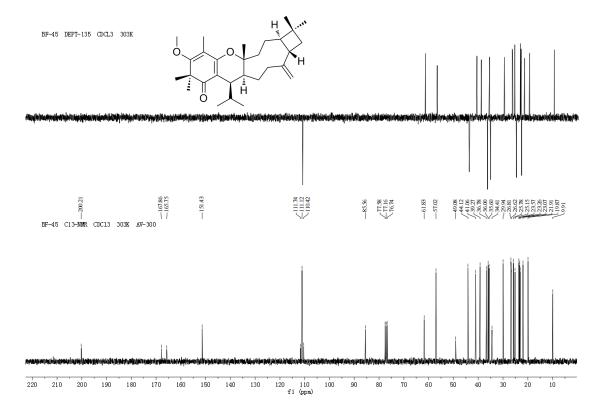


Figure S31. ¹³C NMR spectrum of 3 in CDCl₃ (75 MHz)

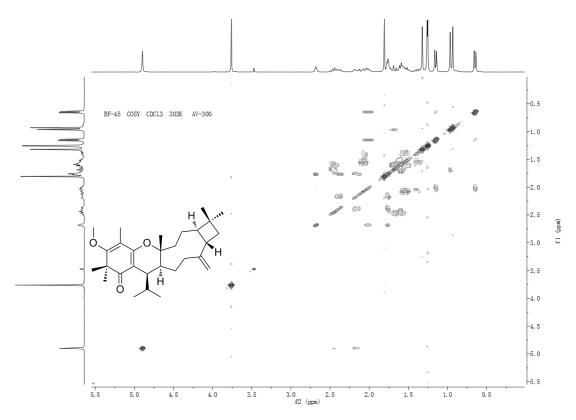


Figure S32. ¹H-¹H COSY spectrum of **3** in CDCl₃ (300 MHz)

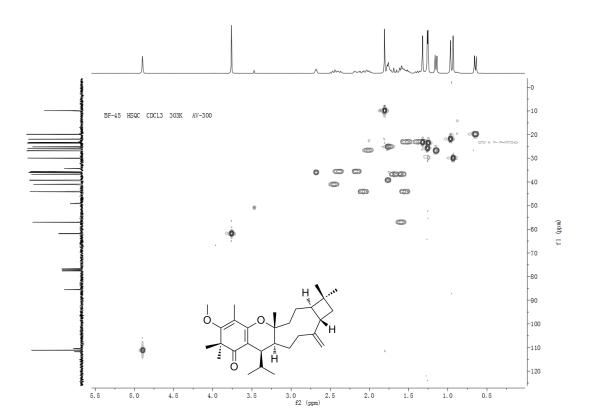


Figure S33. HSQC spectrum of 3 in CDCl₃ (300 MHz)

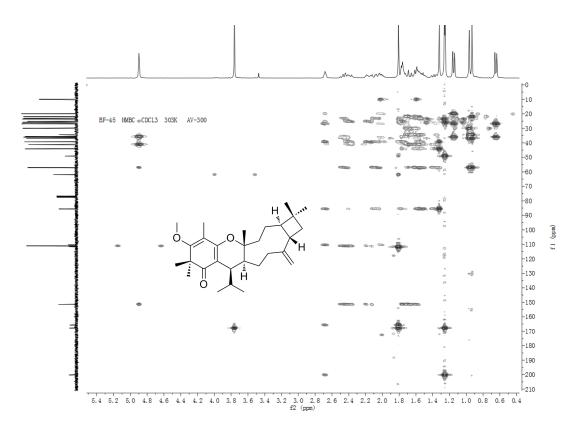


Figure S34. HMBC spectrum of 3 in CDCl₃ (300 MHz)

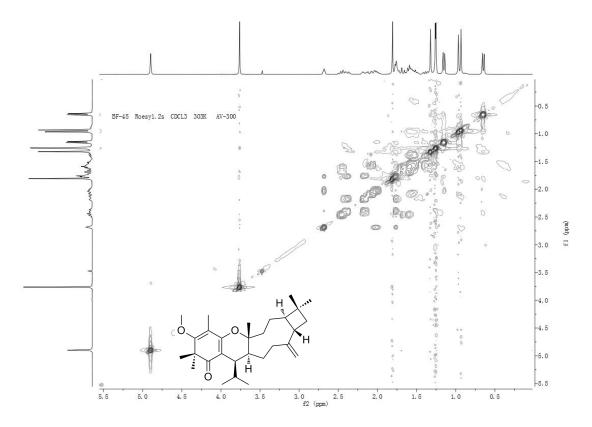


Figure S35. ROESY spectrum of 3 in CDCl₃ (300 MHz)

	m/z	Ion	Formula 🛆	Abundance					
	441.3357	(M+H)+	C29 H45 O3	780117.9					
	Best	Formula (M)	Ion Formula	Calc m/z	Score ∇	Cross Score	Mass	Calc Mass	Diff (ppm)
+	V	C29 H44 O3	C29 H45 O3	441.3363	96.55		440.3285	440.329	1.24

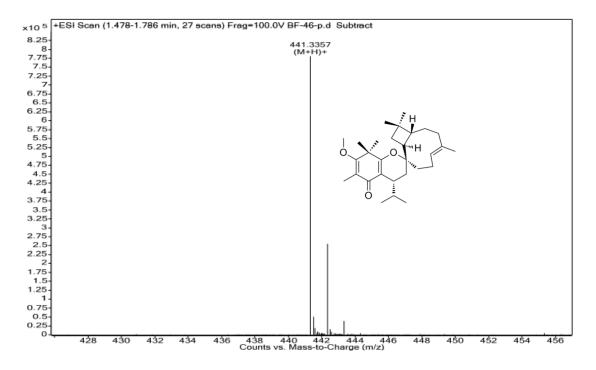


Figure S36. HR-ESI-MS spectrum of 4

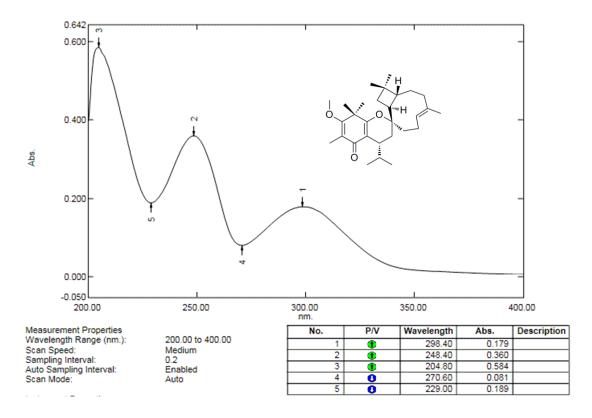


Figure S37. UV spectrum of 4 in MeOH

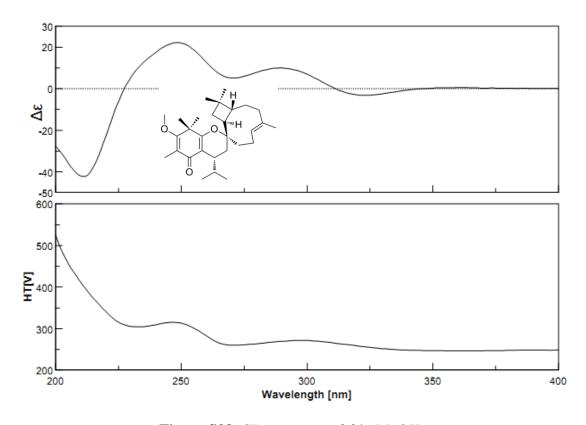


Figure S38. CD spectrum of 4 in MeOH

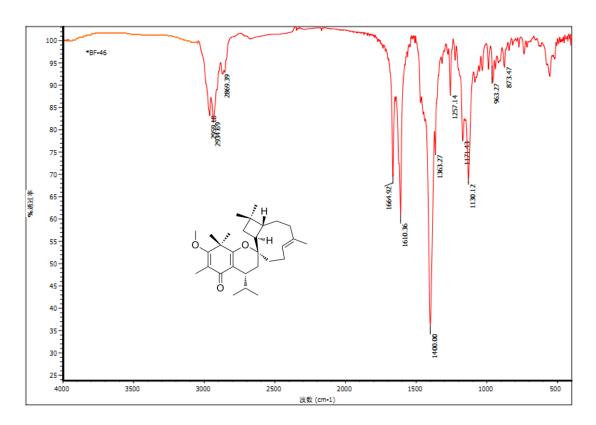


Figure S39. IR spectrum of 4

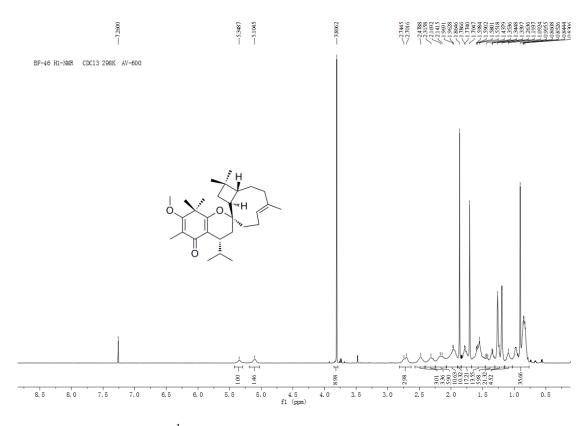


Figure S40-1. ¹H NMR spectrum of **4** in CDCl₃ (600 MHz, 298K)

BF-46 H1-NMR CDC13 242K AV-600

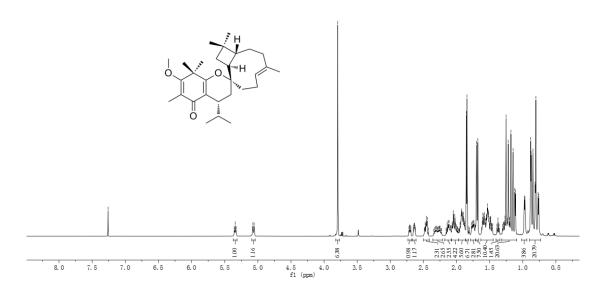


Figure S40-2. ¹H NMR spectrum of 4 in CDCl₃ (600 MHz, 242K)

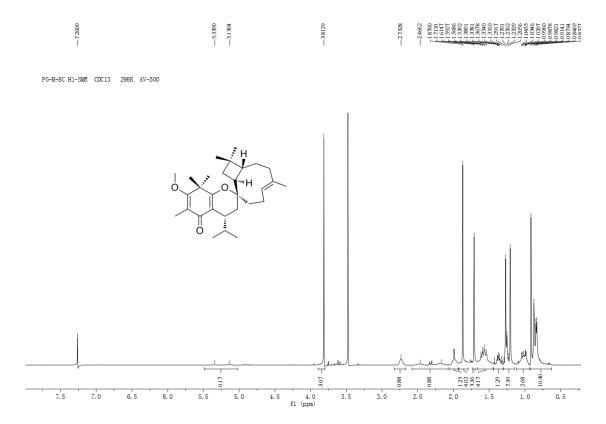


Figure S41. ¹H NMR spectrum of 4 (synthetic) in CDCl₃ (500 MHz, 298K)

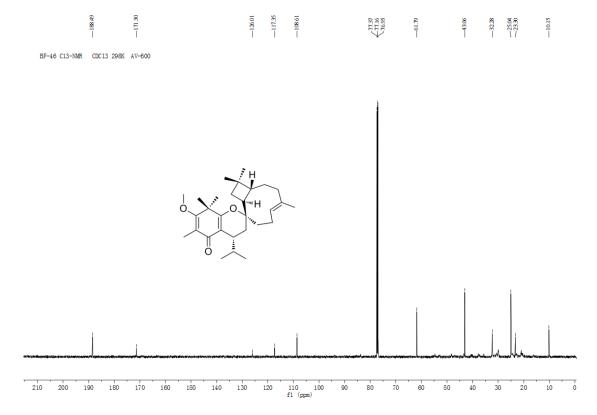


Figure S42-1. ¹³C NMR spectrum of **4** in CDCl₃ (150 MHz, 298K)

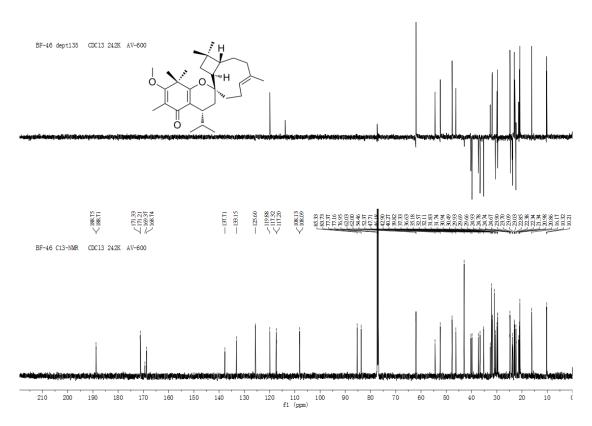


Figure S42-2. ¹³C NMR spectrum of **4** in CDCl₃ (150 MHz, 242K)

PG-M-8C C13-NMR CDC13 298K AV-500

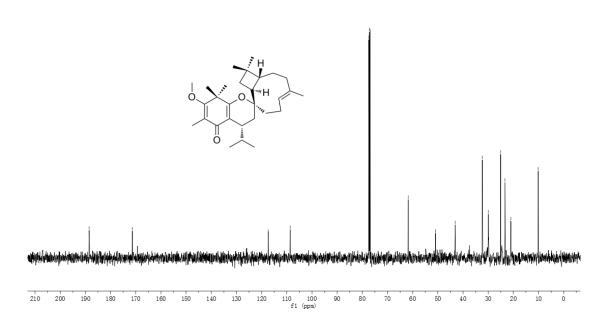


Figure S43. ¹³C NMR spectrum of 4 (synthetic) in CDCl₃ (125 MHz, 298K)

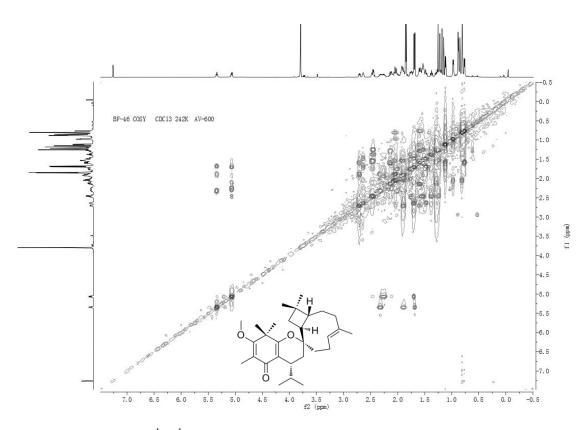


Figure S44. $^{1}\text{H-}^{1}\text{H COSY}$ spectrum of **4** in CDCl₃ (600 MHz, 242K)

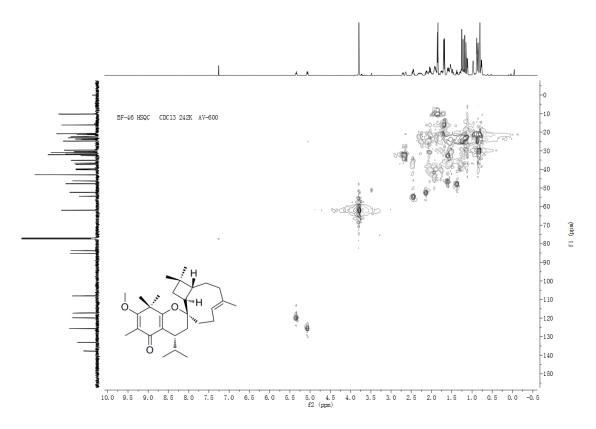


Figure S45. HSQC spectrum of 4 in CDCl₃ (600 MHz, 242K)

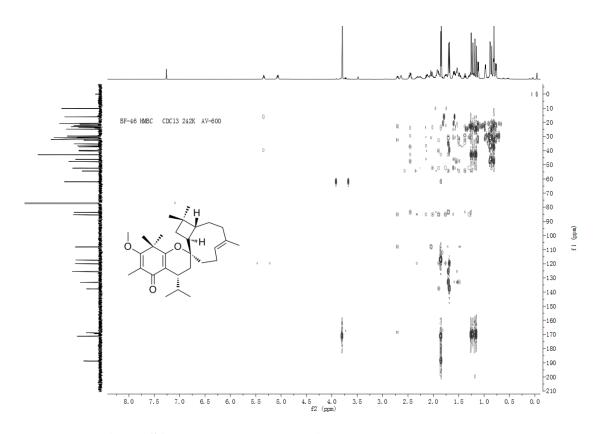


Figure S46. HMBC spectrum of 4 in CDCl₃ (600 MHz, 242K)

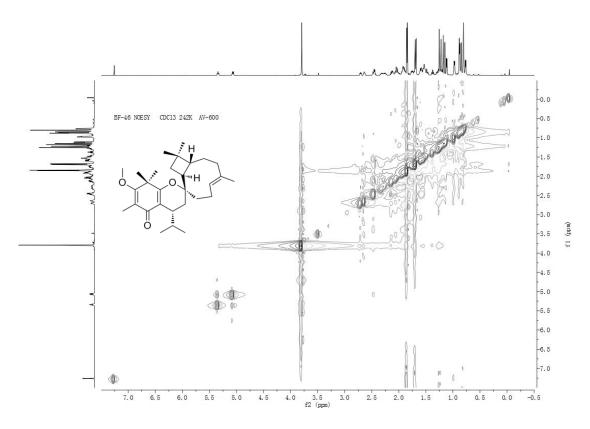


Figure S47. NOESY spectrum of 4 in CDCl₃ (600 MHz, 242K)

		m/z	lon	Formula	Abundance					
		441.3363	(M+H)+	C29 H45 O3	1285718					
	Γ	Best	Formula (M)	Ion Formula	Calc m/z	Score ∇	Cross Score	Mass	Calc Mass	Diff (ppm)
[+	V	C29 H44 O3	C29 H45 O3	441.3363	98.94		440.329	440.329	0.05

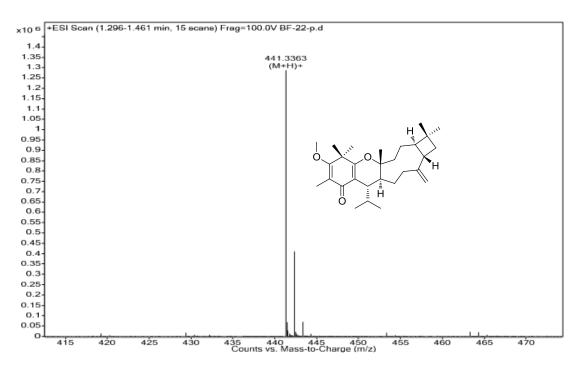


Figure S48. HR-ESI-MS spectrum of **5**

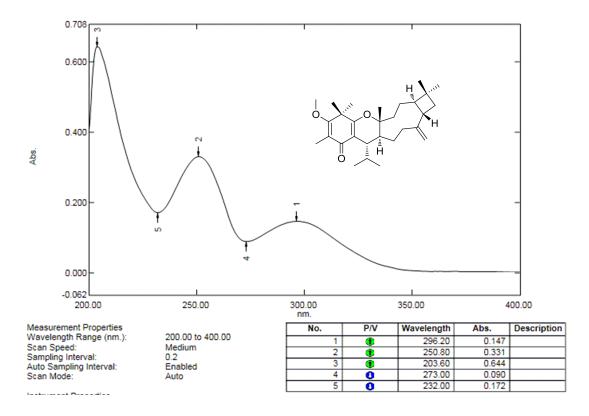


Figure S49. UV spectrum of 5 in MeOH

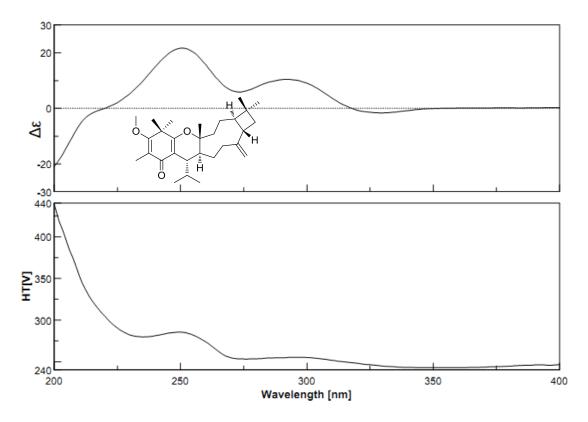


Figure S50. CD spectrum of 5 in MeOH

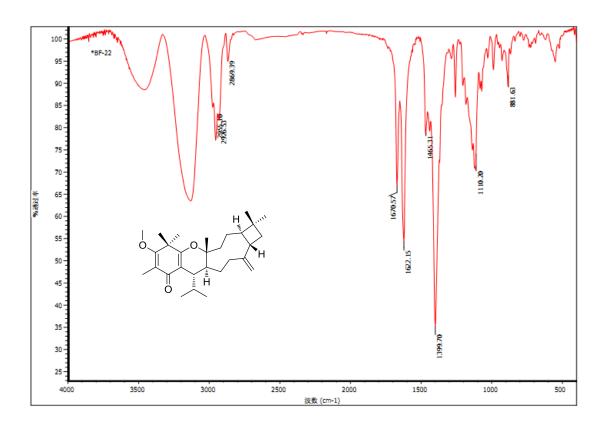


Figure S51. IR spectrum of 5

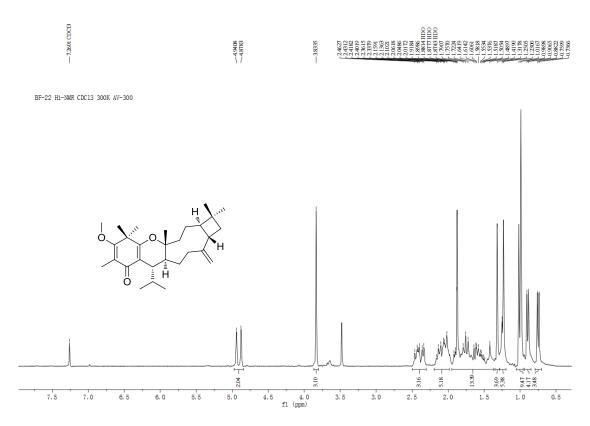


Figure S52. ¹H NMR spectrum of **5** in CDCl₃ (300 MHz)

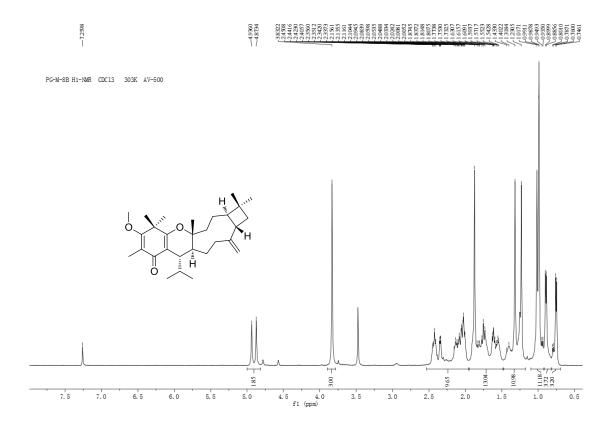


Figure S53. ¹H NMR spectrum of **5** (synthetic) in CDCl₃ (500 MHz)

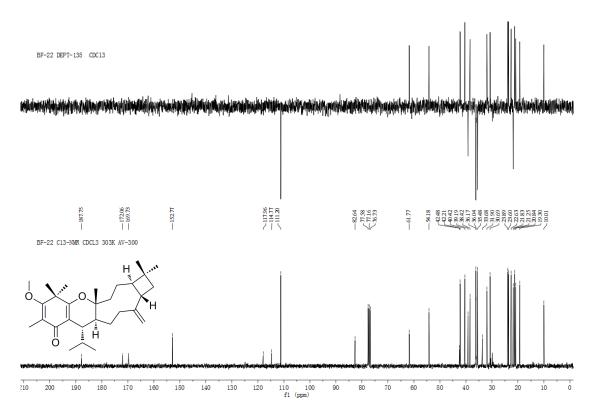


Figure S54. ¹³C NMR spectrum of 5 in CDCl₃ (75 MHz)

PG-M-8B C13-NMR CDC13 303K AV-500

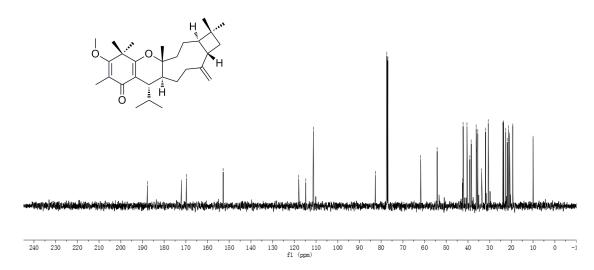


Figure S55. ¹³C NMR spectrum of 5 (synthetic) in CDCl₃ (125 MHz)

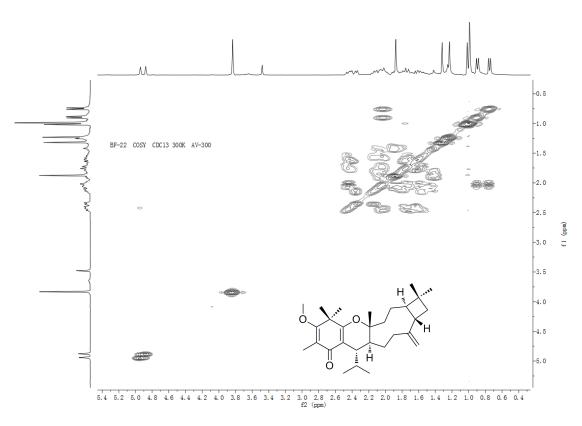


Figure S56. ¹H-¹H COSY spectrum of **5** in CDCl₃ (300 MHz)

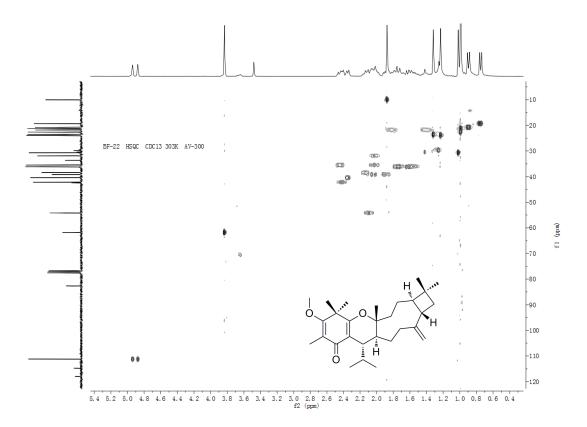


Figure S57. HSQC spectrum of $\bf 5$ in CDCl₃ (300 MHz)

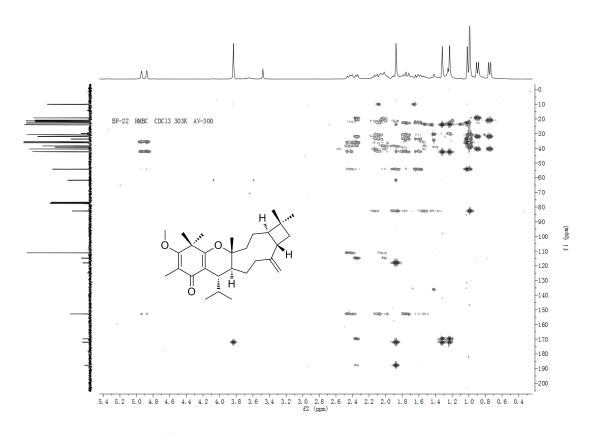


Figure S58. HMBC spectrum of 5 in CDCl₃ (300 MHz)

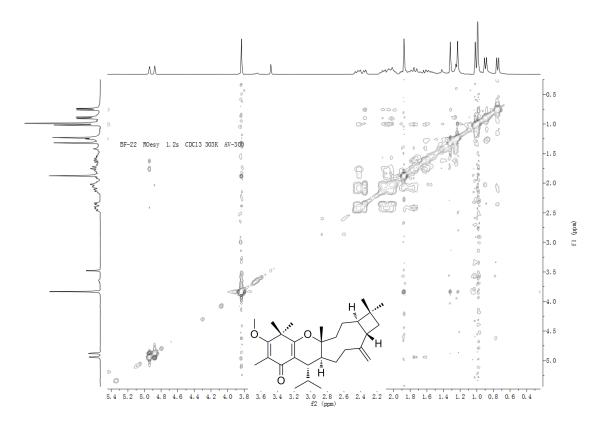


Figure S59. ROESY spectrum of 5 in CDCl₃ (300 MHz)

	m/z $ riangle$	lon	Formula	Abundance					
-[441.3363	(M+H)+	C29 H45 O3	1249737.6					
	Best	Formula (M)	Ion Formula	Calc m/z	Score ∇	Cross Score	Mass	Calc Mass	Diff (ppm)
[V	C29 H44 O3	C29 H45 O3	441.3363	98.06		440.329	440.329	0

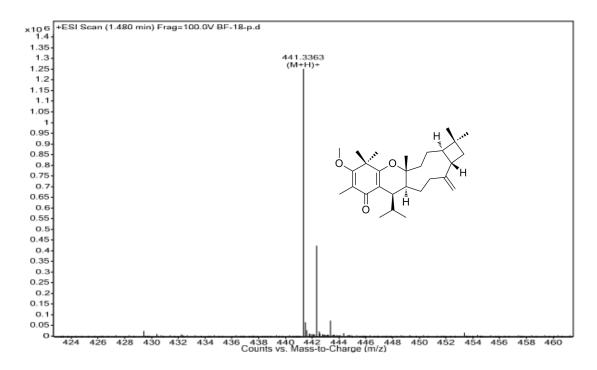


Figure S60. HR-ESI-MS spectrum of 6

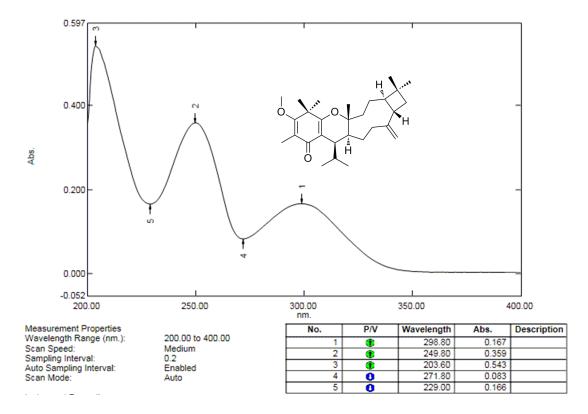


Figure S61. UV spectrum of 6 in MeOH

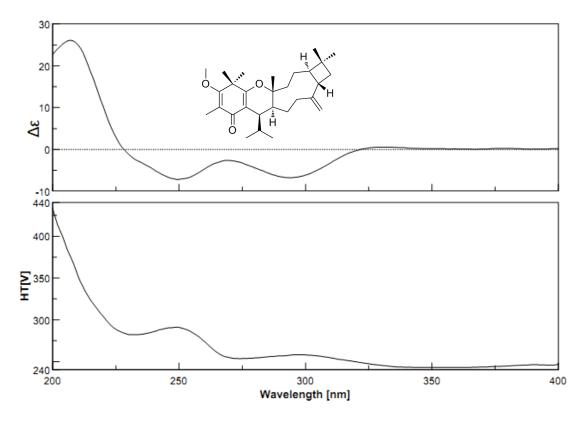


Figure S62. CD spectrum of 6 in MeOH

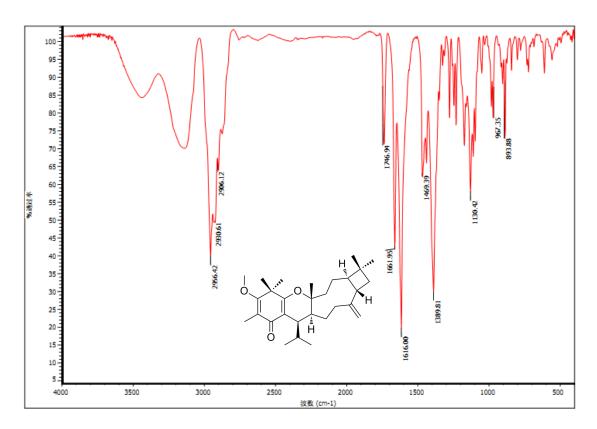


Figure S63. IR spectrum of 6

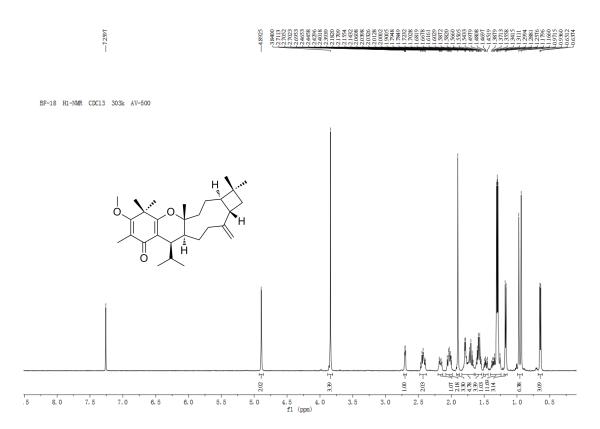


Figure S64. ¹H NMR spectrum of 6 in CDCl₃ (500 MHz)

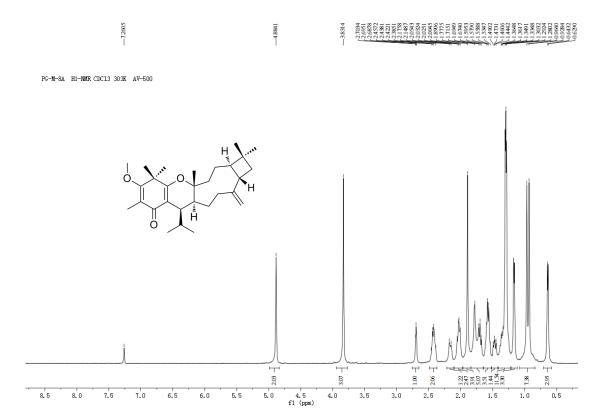


Figure S65. ¹H NMR spectrum of **6** (synthetic) in CDCl₃ (500 MHz)

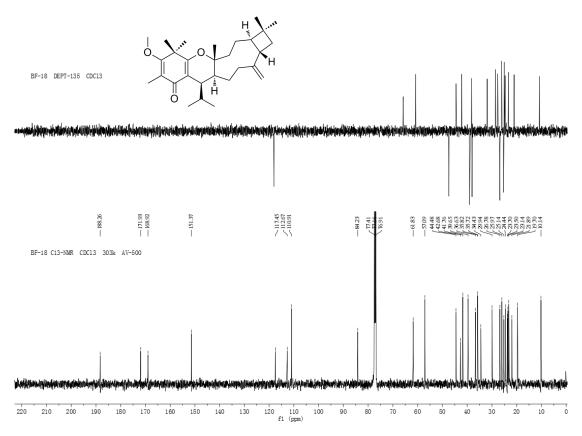


Figure S66. ¹³C NMR spectrum of 6 in CDCl₃ (125 MHz)



PG-M-8A C13-NNMR CDC13 303K AV-500

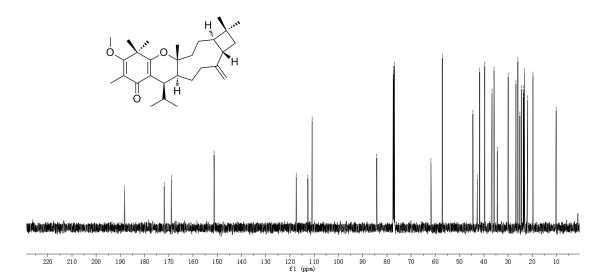


Figure S67. ¹³C NMR spectrum of 6 (synthetic) in CDCl₃ (125 MHz)

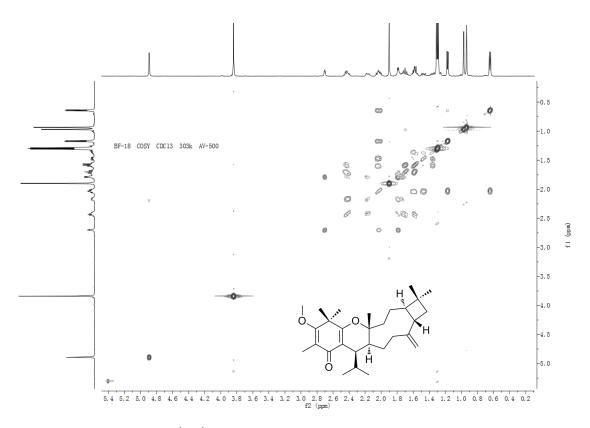


Figure S68. ¹H-¹H COSY spectrum of **6** in CDCl₃ (500 MHz)

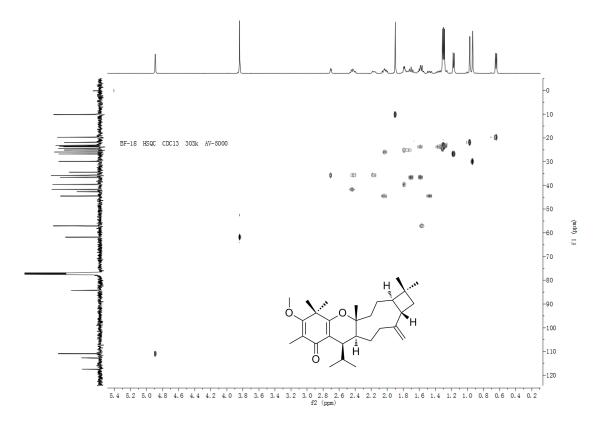


Figure S69. HSQC spectrum of 6 in CDCl₃ (500 MHz)

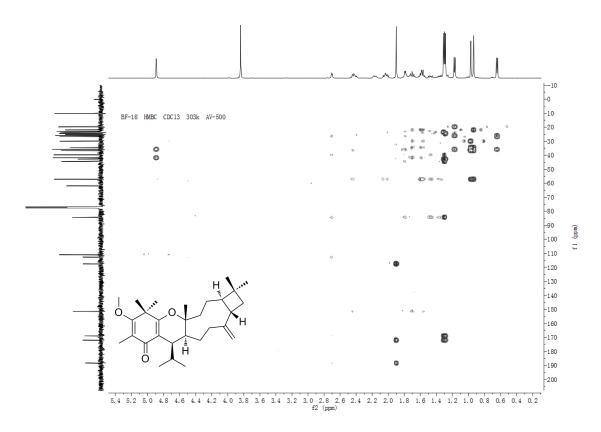


Figure S70. HMBC spectrum of 6 in CDCl₃ (500 MHz)

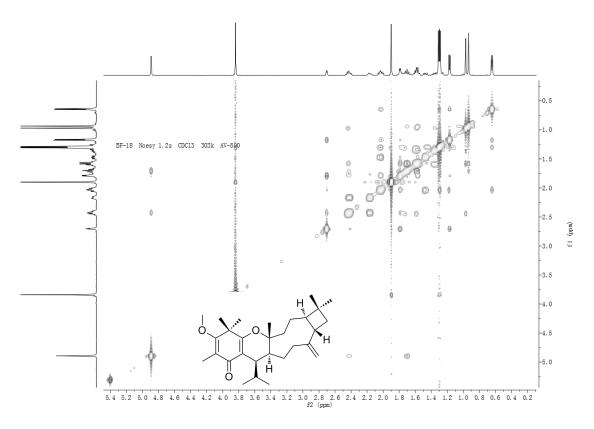


Figure S71. NOESY spectrum of 6 in CDCl₃ (500 MHz)

	m/z 👃	lon	Formula	Abundance					
	441.3366	(M+H)+	C29 H45 O3	1192332.3					
	Best	Formula (M)	Ion Formula	Calc m/z	Score ∇	Cross Score	Mass	Calc Mass	Diff (ppm)

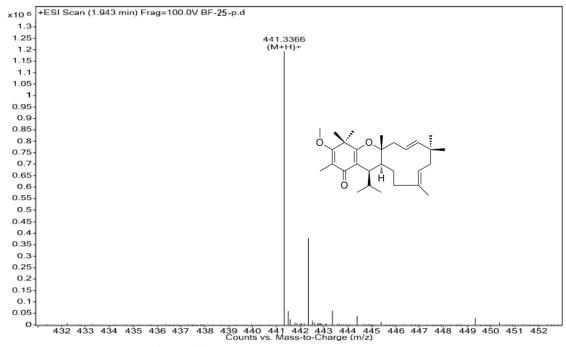


Figure S72. HR-ESI-MS spectrum of (±)-7

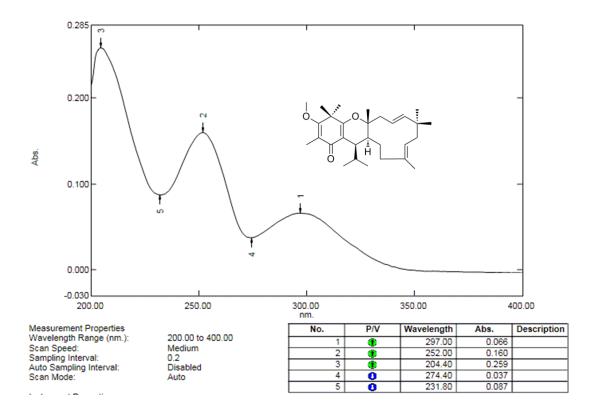


Figure S73. UV spectrum of (\pm) -7 in MeOH

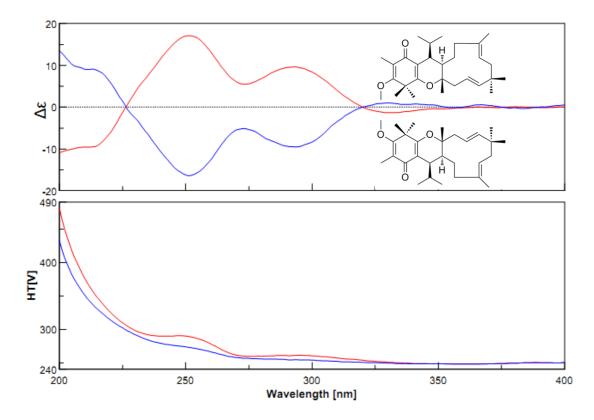


Figure S74. CD spectrum of (+)-7 and (-)-7 in MeOH

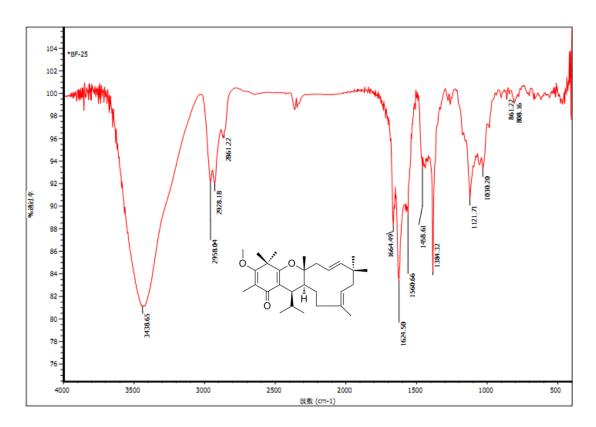


Figure S75. IR spectrum of (\pm) -7

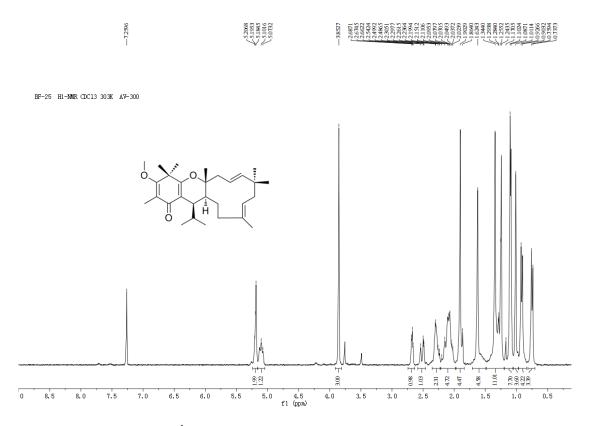


Figure S76. 1 H NMR spectrum of (\pm)-7 in CDCl₃ (300 MHz)

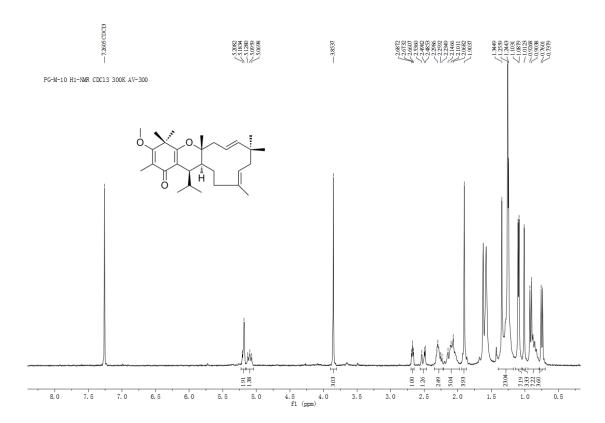


Figure S77. ¹H NMR spectrum of (±)-**7** (synthetic) in CDCl₃ (300 MHz)

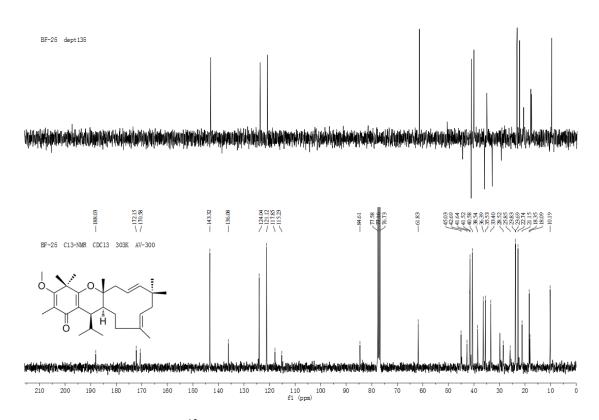


Figure S78. ¹³C NMR spectrum of (±)-**7** in CDCl₃ (75 MHz)

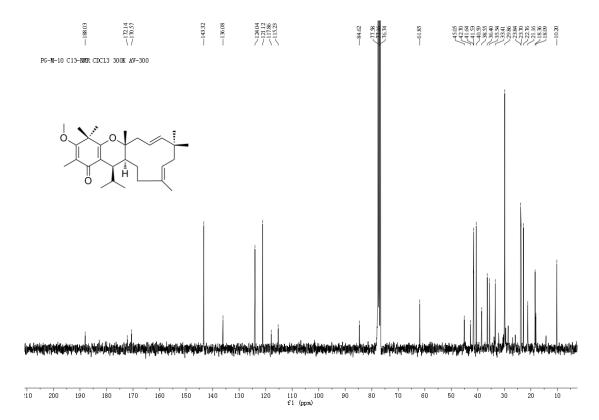


Figure S79. 13 C NMR spectrum of (±)-7 (synthetic) in CDCl₃ (75 MHz)

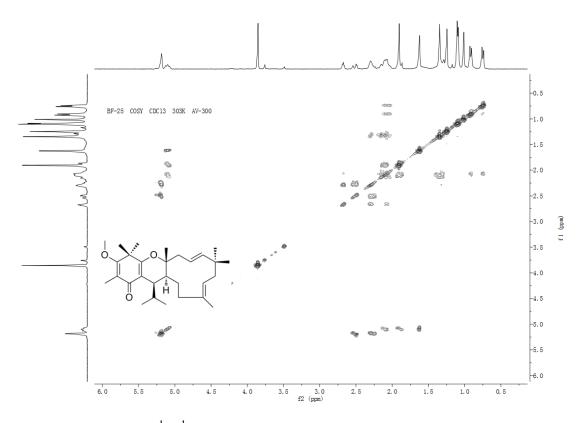


Figure S80. ¹H-¹H COSY spectrum of (±)-**7** in CDCl₃ (300 MHz)

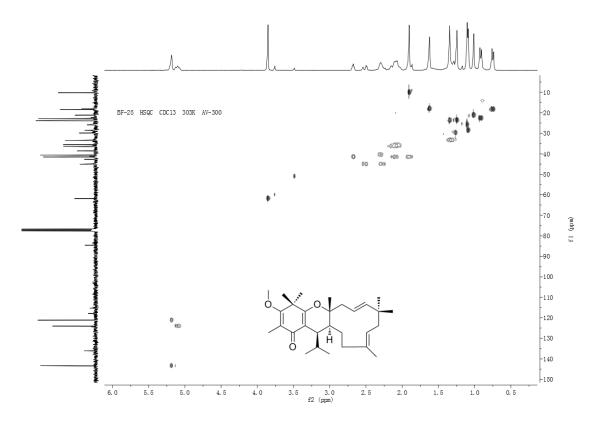


Figure S81. HSQC spectrum of (±)-7 in CDCl₃ (300 MHz)

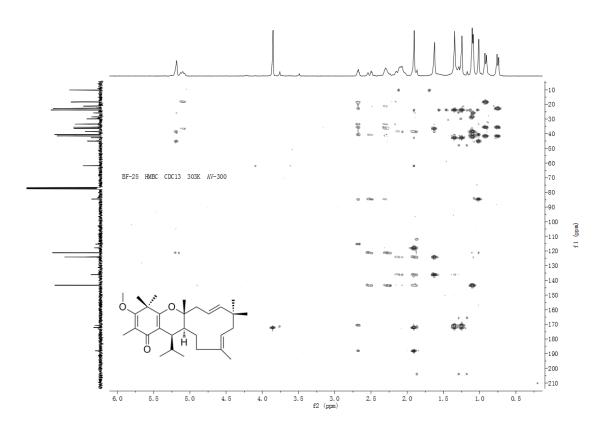


Figure S82. HMBC spectrum of (\pm) -7 in CDCl₃ (300 MHz)

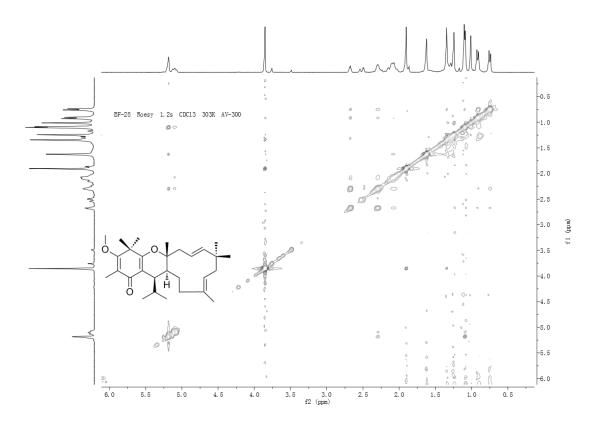


Figure S83. ROESY spectrum of (±)-7 in CDCl₃ (300 MHz)

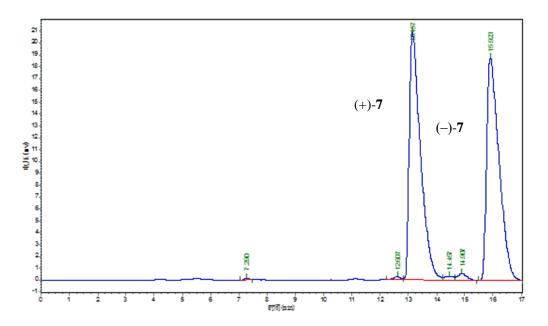


Figure 84. Chiral HPLC chromatogram of (\pm)-7

[Chiral HPLC separation of (\pm)-7 was carried out on a Daicel Chiralcel OD-RH (250 \times 10 mm, 5 μ m), using MeCN-H₂O (85:15, v/v) as mobile phase at a flow rate of 4 mL/min at room temperature with UV detection at 254 nm.]

Display Report - Selected Window Selected Analysis

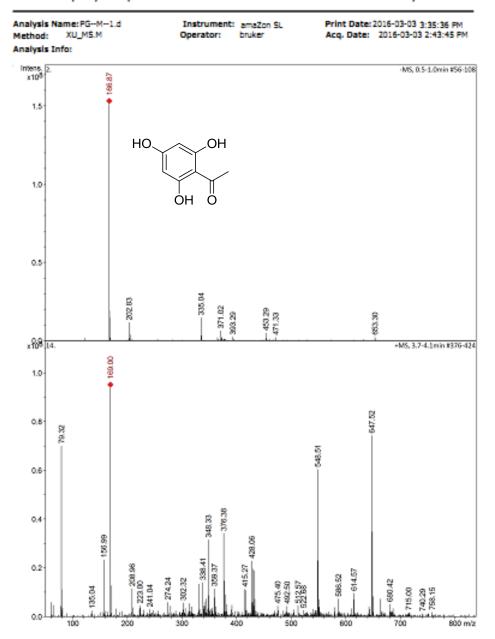


Figure S85. ESI-MS spectrum of 12

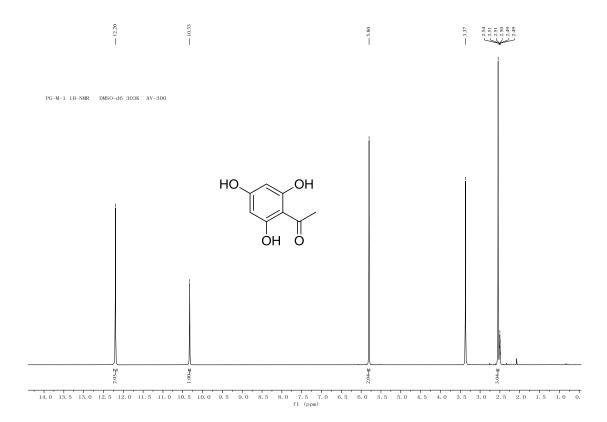


Figure S86. ¹H NMR spectrum of 12 in CDCl₃ (300 MHz)

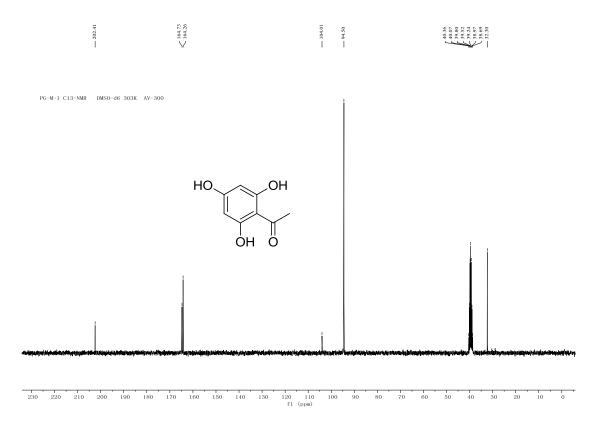


Figure S87. ¹³C NMR spectrum of **12** in CDCl₃ (75 MHz)

Display Report - Selected Window Selected Analysis

Print Date: 2016-03-24 5:27:54 PM Analysis Name: PG--M--28.d Instrument: amaZon SL Method: XU_MS.M Acq. Date: 2016-03-24 5:17:41 PM Operator: bruker Analysis Info: Intens. 4. x10⁸ -M5, 1.6-2.1min#163-214 18002 2.0 HO 1.5 1.0 0.5 166.01 -M52[208.97], 1.6-2.1min#164-215 164,99 1.25 1.00 0.75 0.50 0.25 198.03 459.07 123.08 268.96 2189 la +M5, 3.1-3.5min #312-354 5-647.50 149.03 300.18 318.49 338.51 107.05

Figure S88. ESI-MS spectrum of 13

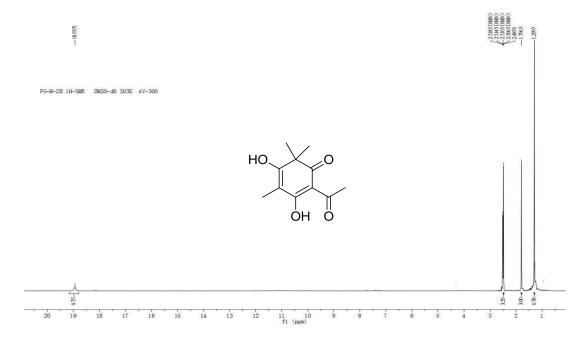


Figure S89. ¹H NMR spectrum of 13 in CDCl₃ (300 MHz)

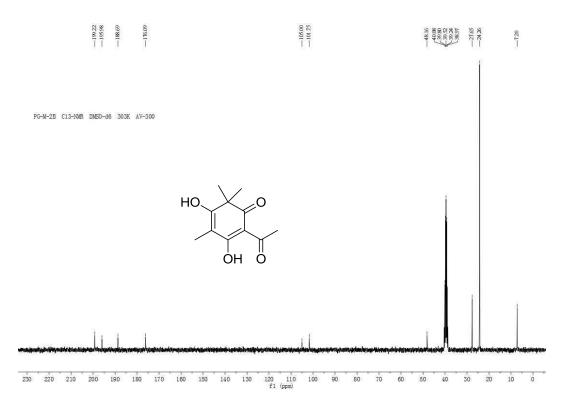


Figure S90. ¹³C NMR spectrum of 13 in CDCl₃ (75 MHz)

References

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- [2] T. Bruhn, A. Schaumlöffel, Y. Hemberger, G. Bringmann, SpecDis version 1.60, University of Wuerzburg, Germany, 2012.

HPLC-Q/TOF-MS analysis of the petroleum ether portion of the aerial parts of *B. frutescens*

Preparation of sample solution

The fresh aerial parts of *B. frutescens* L. were ground into powders. Sample powders (5 g) were percolated with 100 mL methanol at room temperature for 48 h. The crude methanol extract was concentrated *in vacuo*, suspended in H_2O and partitioned with petroleum ether (PE, 60–90 °C). The petroleum ether extract of *B. frutescens* was subjected on a silica flash column (25-40 μ m, 25 g) with PE-EtOAc (100:0, 95:5, each 100 mL) as eluent. Then, the PE: EtOAc (95:5) fraction was concentrated and analyzed on LC-MS.

Preparation of mixed standards

Mixed standards (0.02~0.1 mg/mL for 1–7) were prepared in methanol. 2 μ L was injected into LC-MS for analysis.

Instrument and chromatographic conditions

The HPLC analysis was performed on an Agilent series 1200 HPLC system equipped with a quaternary pump, a degasser, an autosampler, a thermostated column compartment and a diode array detector. Chromatographic separation was carried out at 25 °C on an Agilent Poroshell 120 EC-C18 column (4.6×50 mm, 2.7 µm) with the gradient program of mobile phase MeCN/H₂O ($85:15 \rightarrow 95:5$). The flow rate was 1 mL/min.

All MS experiments were conducted on an Agilent 6210 Q/TOF mass spectrometer equipped with an ESI source. The MS conditions were as follows: The mass range was set at m/z 100–1200; drying gas temperature, 350 °C; drying gas flow, 8 L/min; nebulizer pressure, 35 psi; capillary voltage, 4000 V. Both MS and MS/MS data were performed in positive mode. Collision energy was set at 25 V. Data acquisition was performed with MassHunter Workstation.

Result and Conclusion

To confirm that compounds **1–7** are naturally occurring in the plant, the petroleum ether portion extract of the aerial parts of *B. frutescens* was analyzed by LC-Q/TOF-MS (**Figure S91**). Compounds **1–7** were detected in the crude petroleum ether portion extract by comparsion of the HPLC retention times, HRMS spectra, MS² spectra, and UV absorptions with those of isolates (**Figure S92–S98**). The above results indicated that compounds **1–7** are naturally occurring products.

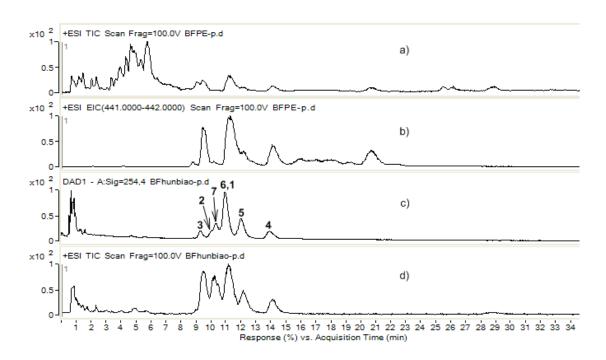


Figure S91. The HPLC-Q/TOF-MS analysis of the petroleum ether portion from *B*. *frutescens* and the mixed standards. a) The total ion chromatogram (TIC) for the petroleum ether portion extract of *B. frutescens*. b) The extracted ion chromatogram (EIC) for m/z 441.0000 to 442.0000 from the petroleum ether portion extract. c) The HPLC-UV chromatogram for the mixed standards. d) The total ion chromatogram (TIC) for the mixed standards.

Figure S92-1. The extracted ion chromatogram (EIC) for m/z 441.0000 to 442.0000, retention time: 9.44 min; m/z 441.3384 [M+H]⁺ (calcd for C₂₉H₄₅O₃, m/z 441.3333)

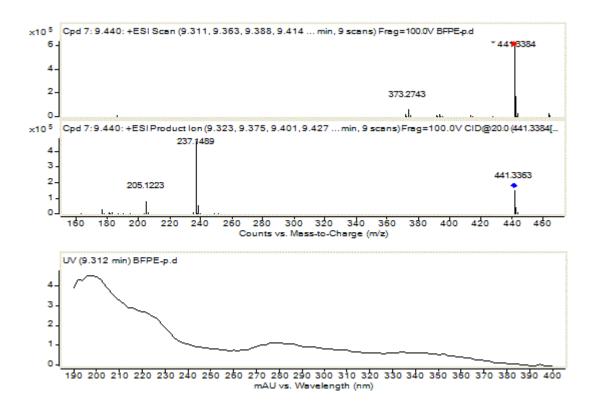


Figure S92-2. The total ion chromatogram (TIC) for compound **3**, retention time: 9.20 min; m/z 441.3420 [M+H]⁺

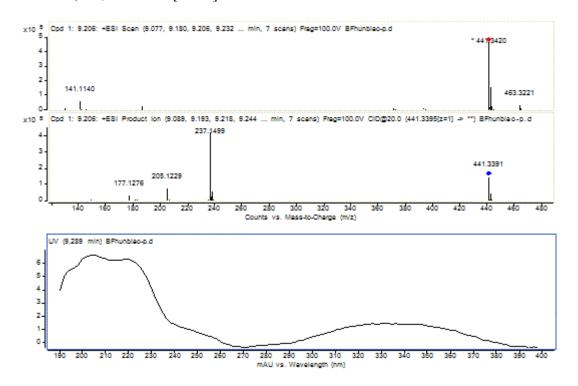


Figure S93-1. The extracted ion chromatogram (EIC) for m/z 441.0000 to 442.0000, retention time: 9.97 min; m/z 441.3390 [M+H]⁺ (calcd for C₂₉H₄₅O₃, m/z 441.3316)

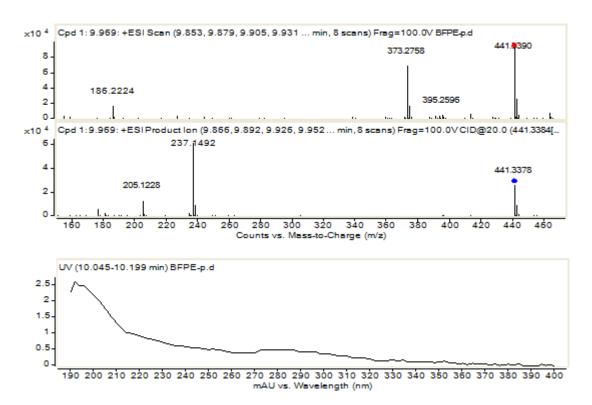


Figure S93-2. The total ion chromatogram (TIC) for compound **2**, retention time: 9.75 min; m/z 441. 3416 [M+H]⁺

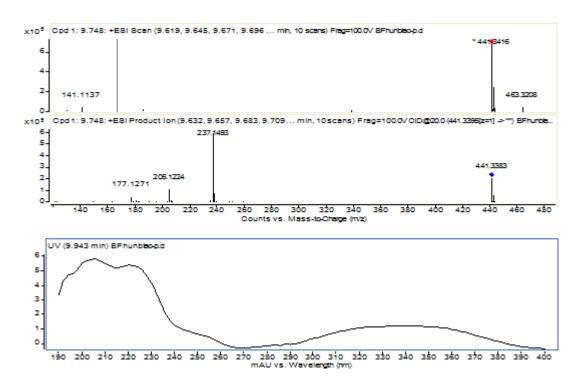


Figure S94-1. The extracted ion chromatogram (EIC) for m/z 441.0000 to 442.0000, retention time: 10.25 min; m/z 441.3417 [M+H]⁺ (calcd for C₂₉H₄₅O₃, m/z 441.3428)

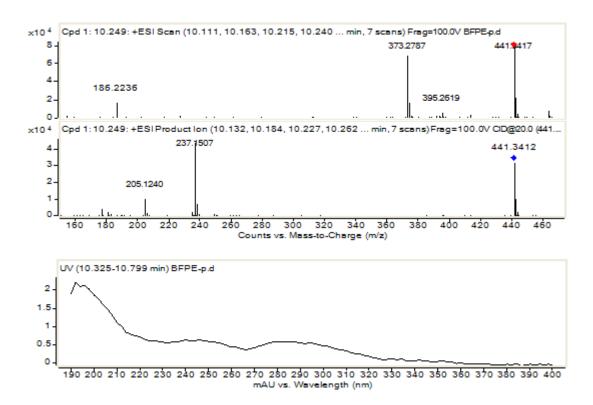


Figure S94-2. The total ion chromatogram (TIC) for compound **7**, retention time: 10.26 min; $m/z 441.3501 [M+H]^+$

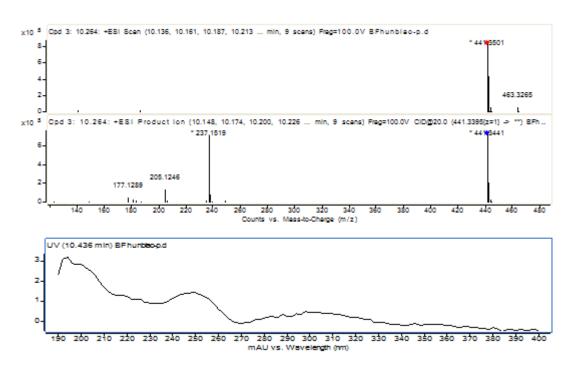


Figure S95-1. The extracted ion chromatogram (EIC) for m/z 441.0000 to 442.0000, retention time: 10.82 min; m/z 441. 3390 [M+H]⁺ (calcd for C₂₉H₄₅O₃, m/z 441.3354)

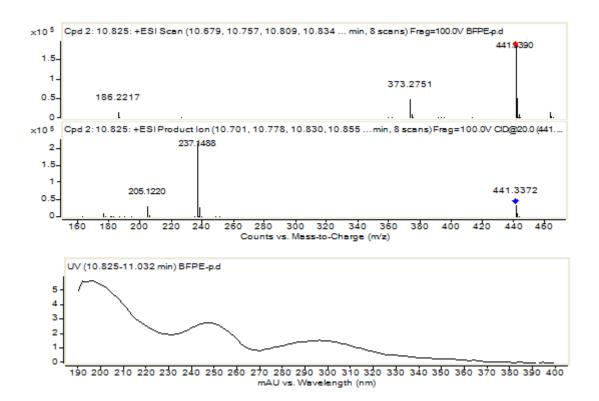


Figure S95-2. The total ion chromatogram (TIC) for compound **6**, retention time: 10.78 min; m/z 441. 3373 [M+H]⁺

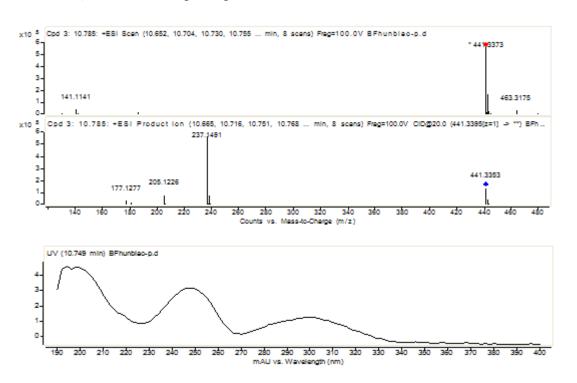


Figure S96-1. The extracted ion chromatogram (EIC) for m/z 441.0000 to 442.0000, retention time: 11.61 min; m/z 441. 3394 [M+H]⁺ (calcd for C₂₉H₄₅O₃, m/z 441.3317)

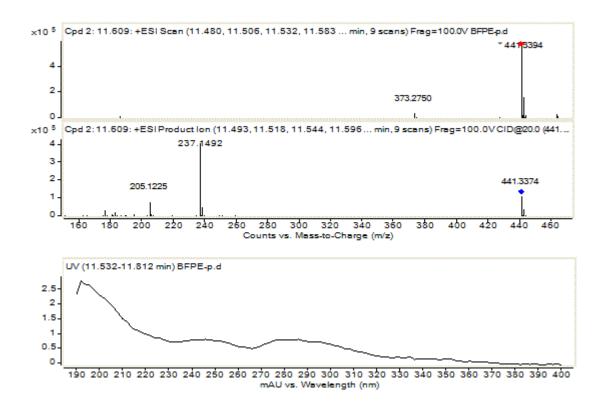


Figure S96-2. The total ion chromatogram (TIC) for compound **1**, retention time: 11.50 min; $m/z 441.3482 [M+H]^+$

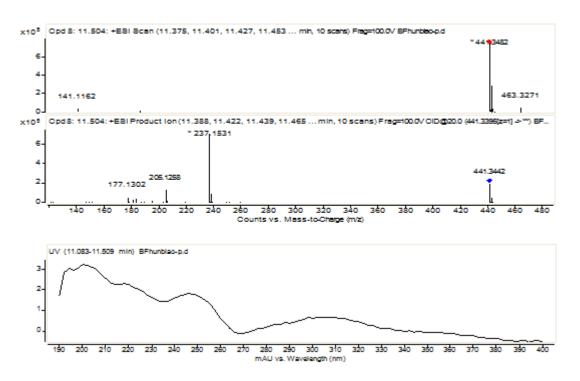


Figure S97-1. The extracted ion chromatogram (EIC) for m/z 441.0000 to 442.0000, retention time: 12.14 min; m/z 441. 3392 [M+H]⁺ (calcd for C₂₉H₄₅O₃, m/z 441.3306)

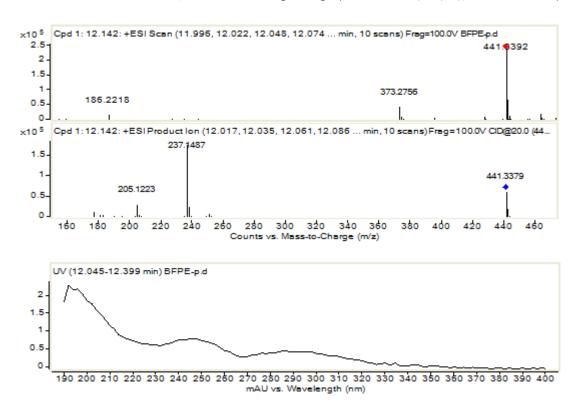


Figure S97-2. The total ion chromatogram (TIC) for compound **5**, retention time: 11.76 min; $m/z 441.3375 [M+H]^+$

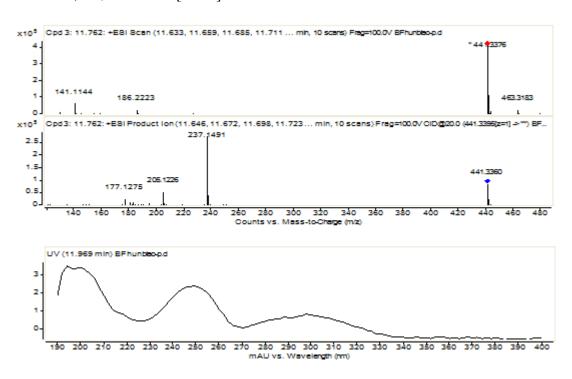


Figure S98-1. The extracted ion chromatogram (EIC) for m/z 441.0000 to 442.0000, retention time: 13.97 min; m/z 441. 3439 [M+H]⁺ (calcd for C₂₉H₄₅O₃, m/z 441.3397)

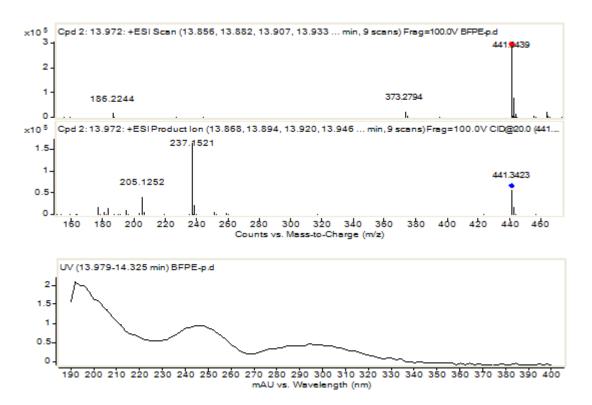
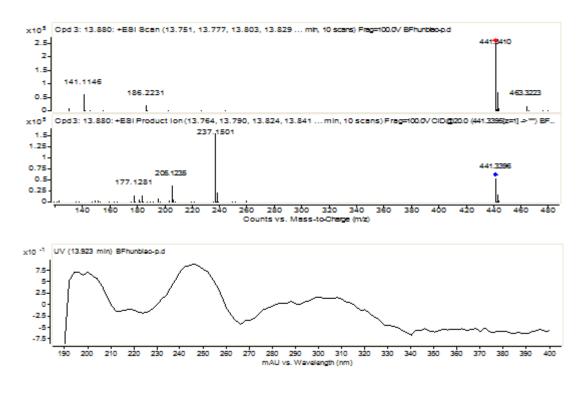
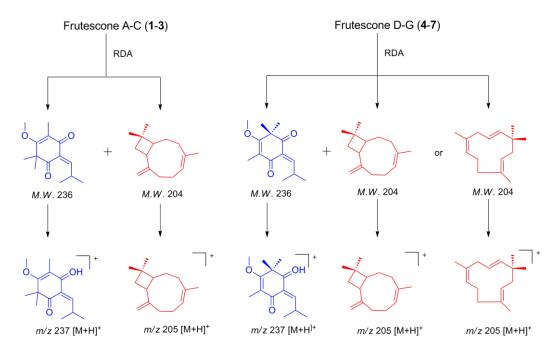


Figure S98-2. The total ion chromatogram (TIC) for compound **4**, retention time: 13.88 min; $m/z 441.3410 [M+H]^+$



Scheme S1. Possible Formation for Major Fragments of Frutescone A–G (1–7) in (+) ESI-MS² Spectrum



HPLC-MSD Trap analysis of the CH_2Cl_2 extract of the aerial parts of B. frutescens

Preparation of sample solution

The fresh aerial parts of *B. frutescens* were ground into powders and passed through a 60-mesh (0.3 mm) sieve. Sample powders (5 g) were extracted by ultrasonator with 100 mL CH₂Cl₂ at room temperature with for 40 min. The crude CH₂Cl₂ extract was concentrated *in vacuo*, and was subjected to a silica flash column (25-40 µm, 25 g) with PE-EtOAc (100:0, 95:5, each 100 mL) as eluent. Then, the PE: EtOAc (95:5) fraction was concentrated and analyzed on LC-MS.

Preparation of mixed standards

Mixed standards (0.02~0.1 mg/mL for 1–7) were prepared in methanol. 2 μ L was injected into LC-MS for analysis.

Instrument and chromatographic conditions

The HPLC analysis was performed on an Agilent series 1100 HPLC system \$80

equipped with a quaternary pump, a degasser, an autosampler, a thermostated column compartment and a diode array detector. Chromatographic separation was carried out at 25 $^{\circ}$ C on an Agilent Poroshell 120 EC-C18 column (4.6 \times 50 mm, 2.7 μ m) with the gradient program of mobile phase MeCN/H₂O (85:15 \rightarrow 95:5). The flow rate was 1 mL/min.

All MS experiments were conducted on an Agilent 1100 series LC-MSD Trap mass spectra with an electrospray interface (ESI). The MS conditions were as follows: The mass range was set at m/z 50–1200; drying gas temperature, 190 °C; drying gas flow, 9 L/min; nebulizer pressure, 16 psi; capillary voltage, 140 V.; Trap drive, 63.4; rolling averages, 1 cts. Both MS and MS/MS data were performed in positive mode. Data acquisition was performed with Bruker Compass DataAnalysis 4.1.

Result and Conclusion

The fresh aerial parts of *B. frutescens* L. was extracted by ultrasonator at room temperature with CH₂Cl₂, more efficient than petroleum ether. Then the CH₂Cl₂ extract was analyzed by HPLC-MSD Trap (**Figure S99**). Compounds **1–7** were also detected in the CH₂Cl₂ extract by comparison of the HPLC retention time and MSⁿ spectra, with those of isolates (see the Supporting Information, **Figure S99-S106**). Thus, compounds **1–7** are proved to be natural occurring products in *B. frutescens*, not artifacts produced during the extraction and isolation procedure.

Figure S99. The LC-MSD Trap analysis of the CH_2Cl_2 extract of *B. frutescens* and the mixed standards. a) The HPLC-UV chromatogram (254 nm) for the CH_2Cl_2 extract of *B. frutescens*. b) The base peak chromatogram (BPC) for the CH_2Cl_2 extract. c) The extracted ion chromatogram (EIC) for m/z 441 from CH_2Cl_2 extract. d) The HPLC-UV chromatogram (254 nm) for the mixed standards. e) The base peak chromatogram (BPC) for the mixed standards. f) The extracted ion chromatogram (EIC) for m/z 441 from the mixed standards.

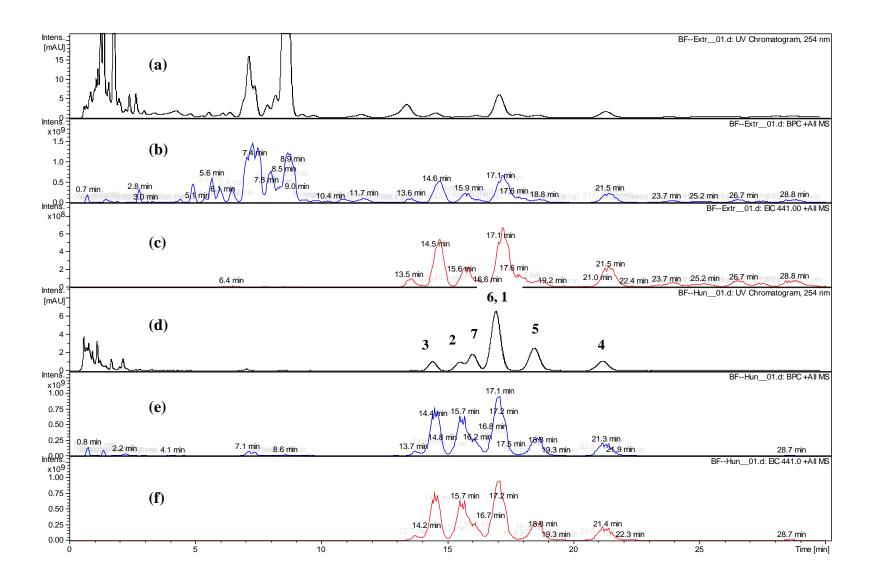


Figure S100-1. The extracted ion chromatogram (EIC) for m/z 441, retention time:

14.5 min; *m/z* 441.29 [M+H]⁺

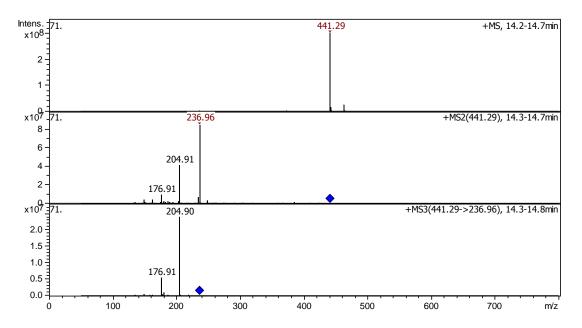


Figure S100-2. The base peak chromatogram (BPC) for compound 3, retention time:

14.5 min; *m/z* 441.29 [M+H]⁺

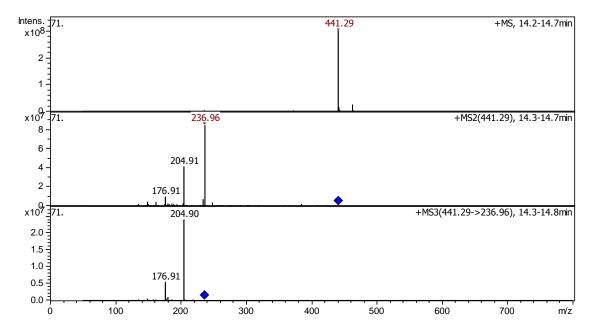


Figure S101-1. The extracted ion chromatogram (EIC) for m/z 441, retention time: 15.6 min; m/z 441.29 [M+H]⁺

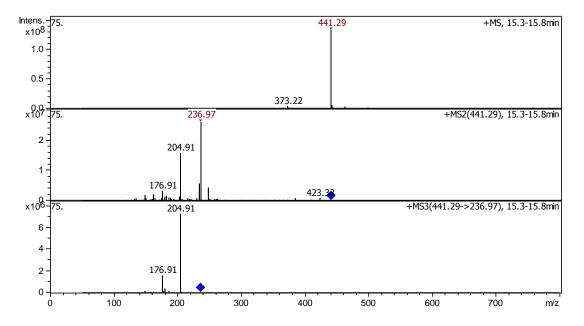


Figure S101-2. The base peak chromatogram (BPC) for compound **2**, retention time: 15.6 min; m/z 441. 29 [M+H]⁺

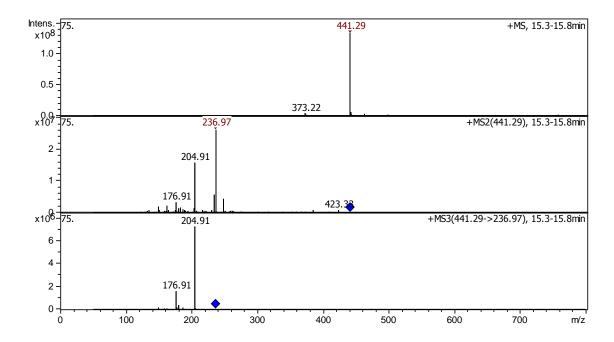


Figure S102-1. The extracted ion chromatogram (EIC) for m/z 441, retention time: 16.1 min; m/z 441.29 [M+H]⁺

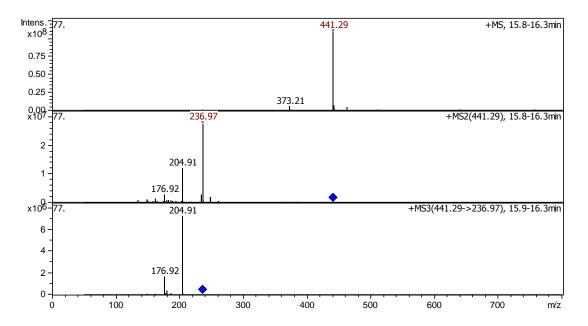


Figure S102-2. The base peak chromatogram (BPC) for compound **7**, retention time: 16.2 min; $m/z 441.29 [M+H]^+$

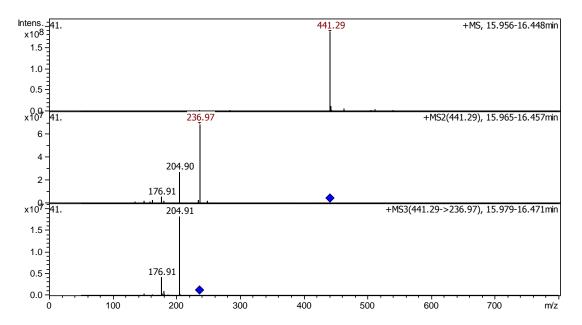


Figure S103-1. The extracted ion chromatogram (EIC) for m/z 441, retention time: 17.1 min; m/z 441. 29 [M+H]⁺

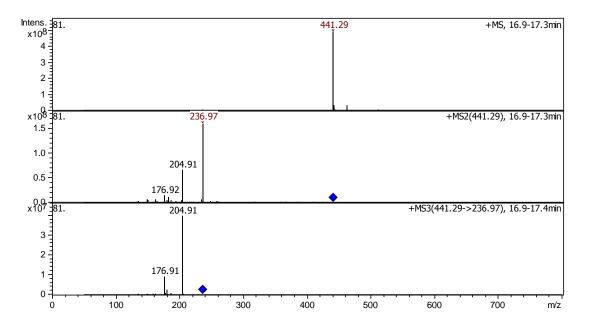


Figure S103-2. The base peak chromatogram (BPC) for compound **6**, retention time: 17.2 min; $m/z 441. 29 [M+H]^+$

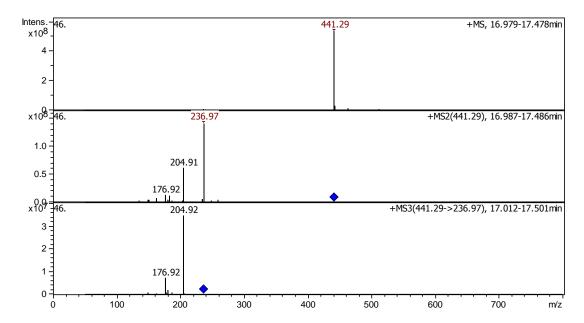


Figure S104-1. The extracted ion chromatogram (EIC) for m/z 441, retention time: 17.6 min; m/z 441. 28 [M+H]⁺

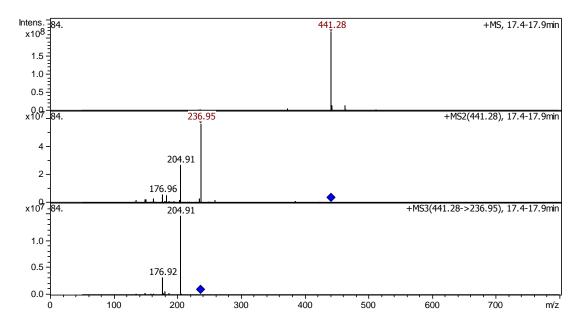


Figure S104-2. The base peak chromatogram (BPC) for compound **1**, retention time: 17.6 min; $m/z 441. 28 [M+H]^+$

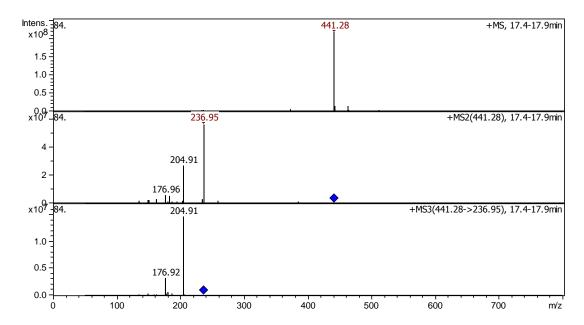


Figure S105-1. The extracted ion chromatogram (EIC) for m/z 441, retention time: 18.6 min; m/z 441. 28 [M+H]⁺

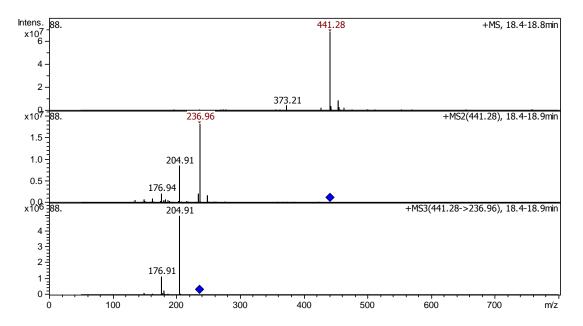


Figure S105-2. The base peak chromatogram (BPC) for compound **5**, retention time: 18.7 min; $m/z 441. 28 [M+H]^+$

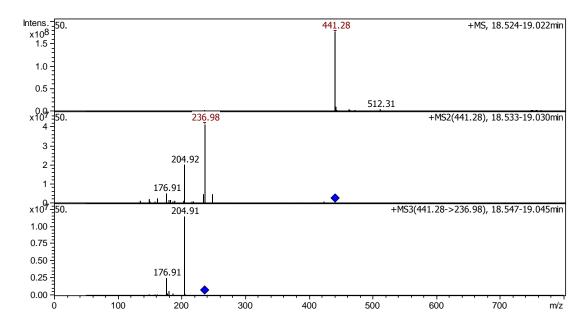


Figure S106-1. The extracted ion chromatogram (EIC) for m/z 441, retention time: 21.5 min; m/z 441. 30 [M+H]⁺

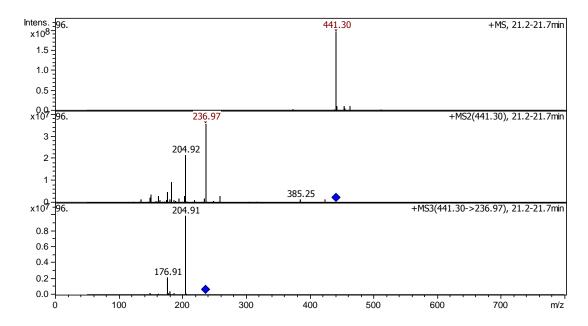


Figure S106-2. The base peak chromatogram (BPC) for compound **4**, retention time: 21.3 min; $m/z 441. 29 [M+H]^+$

