

Enantioselective Syntheses of Furan Atropisomers by an Oxidative Central-to-Axial Chirality Conversion Strategy

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1. General Information

Reactions were run under argon atmosphere in oven-dried glassware. Unless specified, commercial reagents and solvents were used as received. Commercially available catalysts were purchased from Sigma-Aldrich. CHCl₃ was dried using a M-Braun SPS- 800 system.

Analytical thin layer chromatography (TLC) was performed on silica gel 60 F254 aluminum plates (Macherey-Nagel) containing a 254 nm fluorescent indicator. TLC plates were visualized by exposure to short wave ultraviolet light (254 nm) and further visualization was achieved by staining KMnO₄ and heating by a hot air gun. Flash column chromatography was performed using silica gel (35–70 µm, 60 Å, Acros). Organic extracts were dried over anhydrous Na₂SO₄ or MgSO₄.

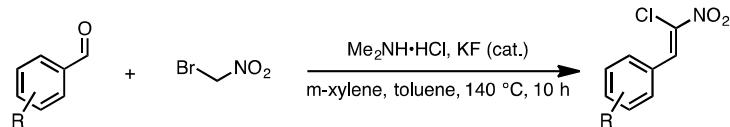
Proton nuclear magnetic resonance (¹H NMR) spectra were recorded with a Bruker AV 300, AV 400, AV 500 or AV 600 MHz spectrometer. Proton chemical shifts are reported in parts per million (δ scale), and are referenced using residual protium in the NMR solvent (CDCl₃: δ 7.26 (CHCl₃)). Data are reported as follows: chemical shift (multiplicity (s = singlet, br s = broad singlet, d = doublet, t = triplet, q = quadruplet, quint = quintuplet, sept = septuplet, m = multiplet), coupling constant(s) (Hz), integration). Carbon-13 nuclear magnetic resonance (¹³C NMR) spectra were recorded with Bruker AV 300, AV 400, AV 500 or AV 600 MHz spectrometers. Carbon chemical shifts are reported in parts per million (δ scale), and are referenced using the carbon resonances of the solvent (δ 77.16 (CHCl₃)). Data are reported as follows: chemical shift (CH_n where n is the number of hydrogen atoms linked to the carbon atom).

HPLC analyses for the determination of enantiomeric excesses were performed on a Merck-Hitachi system equipped with Chiralpak AZ-H, Chiralpak IA, Chiralpak IB, Chiralpak IC, Chiralpak ID, Chiralpak IE, Chiralpak IF, Lux-Cellulose-2 and Lux- Cellulose-4.

Optical Rotations were recorded on an Anton Paar MCP 200 Polarimeter at 589 nm and 25 °C and specific rotations are reported as follows: specific rotation (concentration in grams/100 mL of solution, solvent).

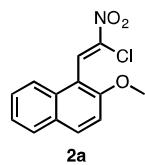
High resolution mass spectra (HRMS) were recorded on a Waters Synapt G2 HDMS apparatus using a positive electrospray (ESI) ionization source.

2. General procedure for the preparation of α -chloronitroalkenes



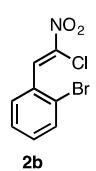
All α -chloronitroalkenes were prepared according to the literature known procedure¹ with slight modification. Substituted benzaldehyde (16.1 mmol, 1.0 equiv), bromonitromethane (4.5 g, 32.2 mmol, 2.0 equiv), dimethylamine hydrochloride (11.82 g, 144.9 mmol, 9.0 equiv), potassium fluoride (0.14 g, 2.4 mmol, 0.15 equiv), toluene (15 mL) and m-xylenes (45 mL) were combined in a 250 mL round bottomed flask and connected to a Dean-Stark trap. The mixture was heated at 140 °C with azeotropic removal of water for 10 hours. The reaction was cool down up to room temperature and saturated sodium bisulfite (NaHSO_3 , 20 mL) was poured in it. The reaction mixture was vigorous stirred before being partitioned between aqueous and organic layer. After separation of organic layer, the aqueous layer was extracted three times with dichloromethane. The combined organics layers were dried over anhydrous Na_2SO_4 and concentrated to afford a brown oil. The oily residue was subsequently purified on a silica column using ethyl acetate/hexane mixture to produce the product. The corresponding bromonitroalkene is often isolated (5-10 %) along with the desired product, which can be used as such in the domino Michael/O-alkylation reaction. Products **2c** ($R = 2\text{-Cl}$),¹ **2e** ($R = \text{CO}_2\text{Me}$),² **2f** ($R = 2\text{-NO}_2$),¹ and **2j** ($R = 2\text{-Me}$)¹ have been synthesized previously using the described procedure.

(Z)-1-(2-chloro-2-nitrovinyl)-2-methoxynaphthalene (2a).



This compound was isolated as a pale yellow solid (3.6 g, 85%); **mp** = 66-67 °C; **Rf** = 0.50 (ethyl acetate/petroleum ether 1:4); **1H NMR** (400 MHz, CDCl_3) δ 8.72 (1H, s, CH), 7.98 (1H, d, J = 9.1 Hz, ArH), 7.85 (1H, d, J = 8.2 Hz, ArH), 7.63 (1H, d, J = 8.7 Hz, ArH), 7.54 (1H, ddd, J = 8.7, 6.7, 1.3 Hz, ArH), 7.42 (1H, ddd, J = 8.2, 6.7, 1.3 Hz, ArH), 7.32 (1H, d, J = 9.1 Hz, ArH), 4.01 (3H, s, CH_3). **13C NMR** (100 MHz, CDCl_3) δ 155.5, 141.7, 132.9, 131.1, 129.3, 128.9, 128.8, 127.9, 124.5, 124.0, 112.8, 112.7, 56.4. **HRMS** (ESI+): $[\text{M}+\text{NH}_4\text{H}]^+$ calcd for $\text{C}_{13}\text{H}_{14}\text{ClN}_2\text{O}_3^+$ 281.0687, found 281.0695.

(Z)-1-bromo-2-(2-chloro-2-nitrovinyl)benzene (2b).



This compound was isolated as a pale yellow solid (3.2 g, 75%); **mp** = 50-51 °C; **Rf** = 0.75 (ethyl acetate/petroleum ether 1:9); **1H NMR** (400 MHz, CDCl_3) δ 8.62 (1H, s, CH), 7.90 (1H, dd, J = 7.9, 1.8 Hz, ArH), 7.71 (1H, dd, J = 7.9, 1.4 Hz, ArH), 7.44 (1H, ddd, J = 7.7, 7.7, 1.4 Hz, ArH), 7.35 (1H, ddd, J = 7.7, 7.7, 1.6 Hz, ArH). **13C NMR** (100 MHz, CDCl_3) δ 139.9, 136.1, 135.6, 132.6, 130.9, 130.7, 127.8, 126.3. **HRMS** (ESI+): $[\text{M}+\text{Ag}]^+$ calcd for $\text{C}_8\text{H}_5\text{BrClNO}_2\text{Ag}^+$ 367.8238, found 367.8239.

(Z)-1-(2-chloro-2-nitrovinyl)-2-iodobenzene (2d).

(Z)-1-(2-chloro-2-nitrovinyl)-2-(trifluoromethyl)benzene (2h).

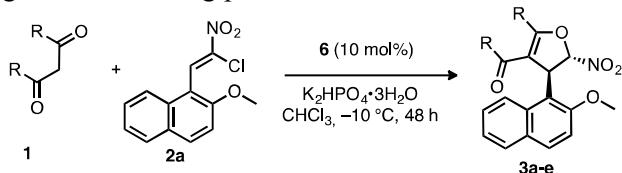
This compound was isolated as a pale yellow oil (3.04 g, 69%); $\text{Rf} = 0.70$ (ethyl acetate/petroleum ether 1:9); $^1\text{H NMR}$ (300 MHz, CDCl_3) δ 8.59 (s, 1H), 7.82 (dd, $J = 13.7, 7.8$ Hz, 2H), 7.69 (t, $J = 7.4$ Hz, 1H), 7.60 (t, $J = 7.6$ Hz, 1H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 141.0, 134.3, 132.3, 130.7, 130.4, 128.7, 128.5, 126.7 (q, $J = 5.5$ Hz), 123.5 (q, $J = 273$ Hz). HRMS (ESI+): $[\text{M}+\text{Ag}]^+$ calcd for $\text{C}_9\text{H}_5\text{NO}_2\text{ClF}_3\text{Ag}^+$ 357.9006, found 357.9006.

(Z)-(2-(2-chloro-2-nitrovinyl)phenyl)diphenylphosphine oxide (2i).

This compound was isolated as a pale yellow solid (2.2 g, 58%); $\text{Rf} = 0.34$ (ethyl acetate/petroleum ether 1:1); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 8.9 (1H, s, CH), 7.79 (1H, dd, $J = 7.5, 4.1$ Hz, ArH), 7.73-7.60 (5H, m, ArH), 7.59-7.53 (1H, m, ArH), 7.53-7.40 (5H, m, ArH), 7.32 (1H, ddd, $J = 13.6, 7.78, 1.4$ Hz, ArH). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 139.5, 134.8 (d, $J = 6.5$ Hz), 133.0 (d, $J = 99.0$ Hz), 133.7 (d, $J = 10.8$ Hz), 133.5, 132.6 (d, $J = 2.7$ Hz), 132.3 (d, $J = 1.9$ Hz), 132.2, 132.1 (d, $J = 9.8$ Hz), 131.4 (d, $J = 5.4$ Hz), 131.2, 130.6 (d, $J = 8.9$ Hz), 129.9 (d, $J = 12.1$ Hz), 129.0 (d, $J = 12.1$ Hz). HRMS (ESI+): $[\text{M}+\text{Ag}]^+$ calcd for $\text{C}_{20}\text{H}_{15}\text{NO}_3\text{PClAg}^+$ 489.9524, found 489.9528.

3. General procedure for the enantioselective synthesis of dihydrofurans 3a-e

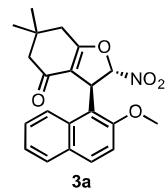
Catalyst **6** was synthetized according to literature procedure.³ Racemic samples were synthesized according to the following procedure.⁴



A solution of the corresponding α -chloronitroalkenes (0.28 mmol, 1.0 equiv) and catalyst **6** (10 mol %) in CHCl_3 (0.03 M, 9 mL) was cooled at 0°C . Then, diketone derivate (0.36 mmol, 1.3 equiv) and K_2HPO_4 (0.42 mmol, 1.5 equiv) were added sequentially. The reaction mixture was stirred at same temperature for 48 hours. When the reaction was complete as

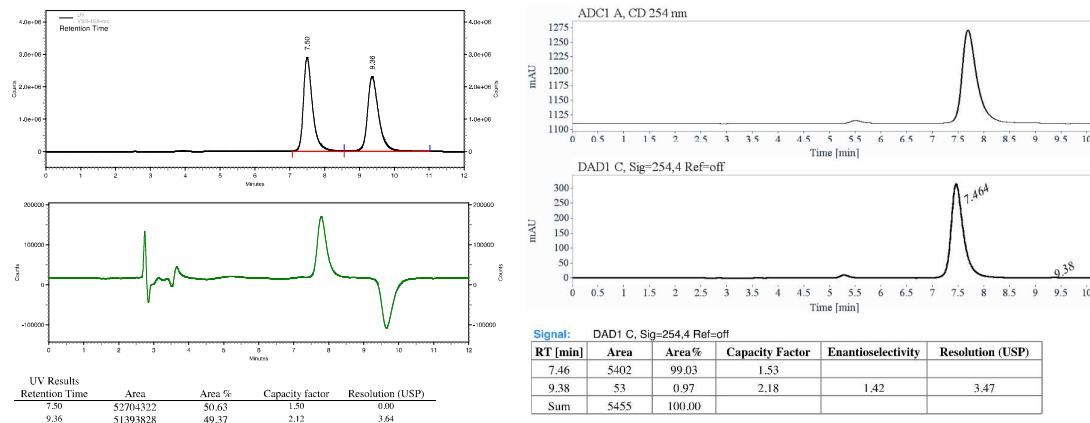
indicated by TLC analysis, the crude product was directly purified by means of flash silica gel column chromatography.

(2*S*,3*S*)-3-(2-methoxynaphthalen-1-yl)-6,6-dimethyl-2-nitro-3,5,6,7-tetrahydro-benzo-furan-4(2*H*)-one (3a).

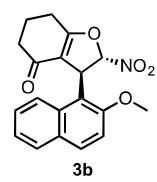


This compound was isolated as a pale yellow solid (94 mg, 92%); **mp** = 154-155 °C; **R_f** = 0.35 (ethyl acetate/petroleum ether 3:7); **dr** (*trans/cis*) > 20:1; **HPLC** (Ulmo (S,S), Heptane/ethanol = 80/20, flow rate = 1.0 mL/min, λ = 254 nm): t_{major} = 7.46 min, t_{minor} = 9.38 min, ee = 99%; $[\alpha]_D^{25}$ (CHCl₃, *c* = 1.0) = +465; **1H NMR** (400 MHz, CDCl₃) δ 8.17 (1H, brs, ArH), 7.85 (1H, d, *J* = 9.0 Hz, ArH), 7.80 (1H, d, *J* = 8.1 Hz, ArH), 7.56 (1H, brs, ArH), 7.39 (1H, dd, *J* = 9.0, 7.6 Hz, ArH), 7.24 (1H, brs, ArH), 6.29 (1H, d, *J* = 2.9 Hz, CH), 5.46 (1H, brs, CH), 3.90 (3H, s, CH₃), 2.72-2.47 (2H, m), 2.25 (1H, d, *J* = 16.4 Hz, CH), 2.13 (1H, d, *J* = 16.4 Hz, CH), 1.19 (3H, s, CH₃), 1.11 (3H, s, CH₃). **13C NMR** (100 MHz, CDCl₃) δ 193.0, 174.3, 155.3, 132.8, 130.8, 129.5, 128.8, 127.6, 123.9, 122.6, 117.6, 113.8, 112.6, 111.7, 56.0, 51.4, 45.9, 37.4, 34.7, 29.2, 28.0. **HRMS** (ESI+): [M+H]⁺ calcd for C₂₁H₂₂NO₅⁺ 368.1492, found 368.1492.

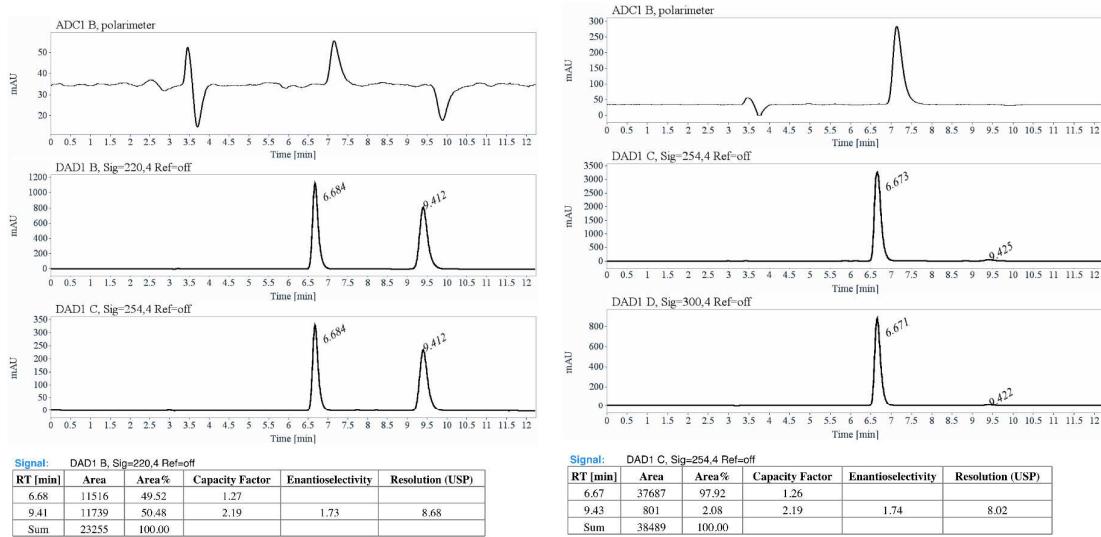
Method description : Ulmo (S,S), Heptane/Ethanol 80/20, 1 ml/min, UV 254 nm et polarimetre



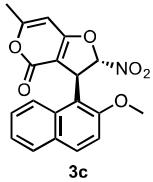
(2*S*,3*S*)-3-(2-methoxynaphthalen-1-yl)-2-nitro-3,5,6,7-tetrahydrobenzofuran-4(2*H*)-one (3b).



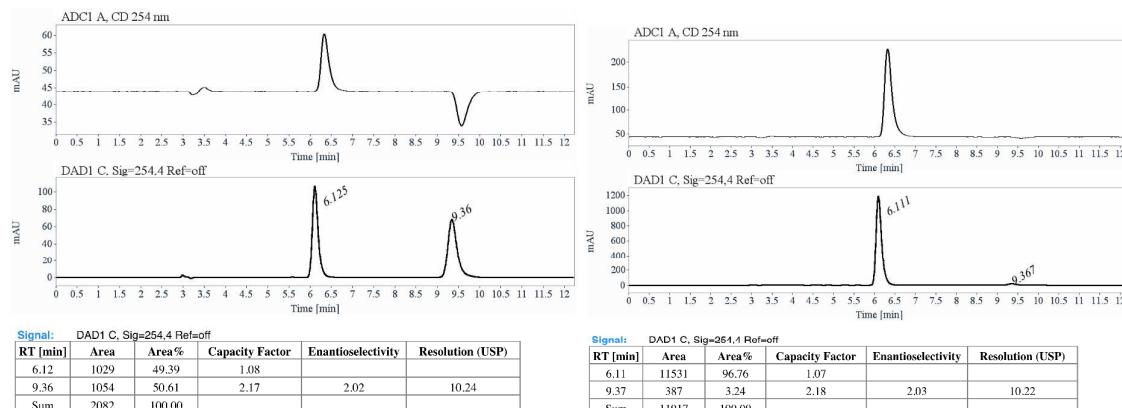
This compound was isolated as a white solid (88 mg, 92%); **mp** = 163-165 °C; **R_f** = 0.35 (ethyl acetate/petroleum ether 3:7); **dr** (*trans/cis*) > 20:1; **HPLC** (Lux-Cellulose-4, Heptane/ethanol = 50/50, flow rate = 1.0 mL/min, λ = 254 nm): t_{major} = 6.68 min, t_{minor} = 9.41 min, ee = 96%; $[\alpha]_D^{25}$ (CHCl₃, *c* = 1.0) = +489; **1H NMR** (400 MHz, CDCl₃) δ 8.17 (1H, brs, ArH), 7.84 (1H, d, *J* = 9.1 Hz, ArH), 7.80 (1H, d, *J* = 8.2 Hz, ArH), 7.56 (1H, brs, ArH), 7.39 (1H, dd, *J* = 7.6, 7.6 Hz, ArH), 7.28-7.22 (1H, m, ArH), 6.27 (1H, d, *J* = 3.5 Hz, CH), 5.49 (1H, brs, CH), 3.90 (3H, s, CH₃), 2.83-2.65 (2H, m), 2.37-2.03 (4H, m). **13C NMR** (100 MHz, CDCl₃) δ 193.5, 175.1, 155.6, 132.7, 130.8, 129.6, 128.9, 127.6, 124.0, 122.6, 117.9, 115.3, 113.3, 111.4, 56.5, 45.9, 37.0, 23.6, 21.8. **HRMS** (ESI+): [M+H]⁺ calcd for C₁₉H₁₈NO₅⁺ 340.1179, found 340.1179.



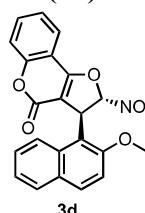
(2S,3S)-3-(2-methoxynaphthalen-1-yl)-6-methyl-2-nitro-2,3-dihydro-4H-furo[3,2-c]pyran-4-one (3c).



This compound was isolated as a white solid (92 mg, 93%); **mp** = 156–157 °C; **R_f** = 0.37 (ethyl acetate/petroleum ether 1:1); **dr** (*trans/cis*) > 20:1; **HPLC** (Lux-Cellulose-4, Heptane/ethanol = 50/50, flow rate = 1.0 mL/min, λ = 254 nm): t_{major} = 6.13 min, t_{minor} = 9.36 min, ee = 94%; $[\alpha]_D^{25}$ (CHCl₃, *c* = 1.0) = +381; **1H NMR** (400 MHz, CDCl₃) δ 8.16 (1H, brs, ArH), 7.88 (1H, d, *J* = 9.0 Hz, ArH), 7.82 (1H, d, *J* = 8.3 Hz, ArH), 7.58 (1H, brs, ArH), 7.40 (1H, dd, *J* = 7.8, 6.6 Hz, ArH), 7.24 (1H, d, *J* = 9.0 Hz, ArH), 6.38 (1H, d, *J* = 3.3 Hz, CH), 6.23 (1H, s, CH), 5.62 (1H, brs, CH), 3.83 (3H, s, CH₃), 2.31 (3H, s, CH₃). **13C NMR** (100 MHz, CDCl₃) δ 169.9, 167.2, 159.6, 155.6, 132.8, 131.3, 129.5, 129.0, 127.9, 124.1, 122.2, 116.6, 113.0, 111.9, 101.3, 95.0, 56.3, 45.4, 20.7. **HRMS** (ESI+): [M+H]⁺ calcd for C₁₉H₁₆NO₆⁺ 354.0972, found 354.0977.

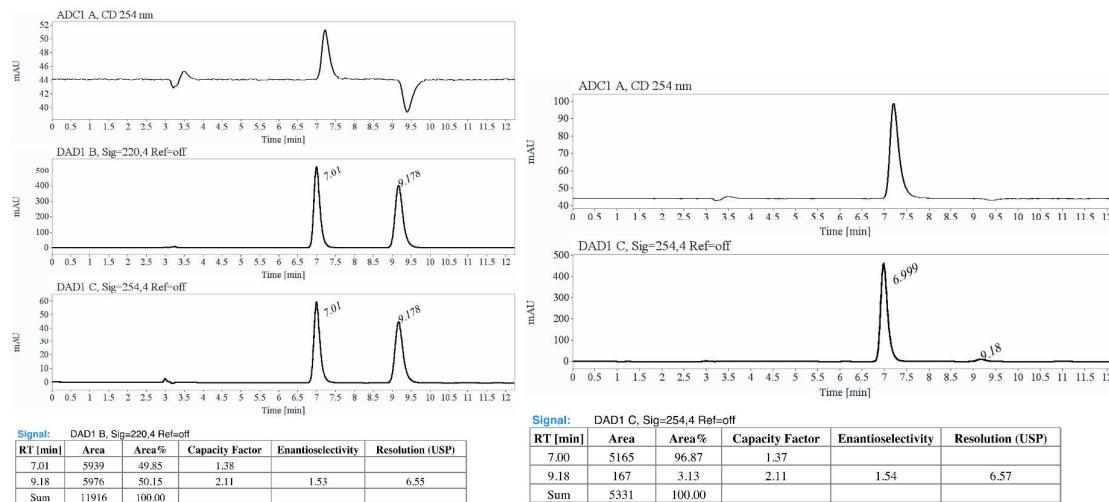


(2S,3S)-3-(2-methoxynaphthalen-1-yl)-2-nitro-2,3-dihydro-4H-furo[3,2-c]chromen-4-one (3d).



This compound was isolated as a white solid (102 mg, 94%); **mp** = 210–215 °C; **R_f** = 0.65 (ethyl acetate/petroleum ether 3:7); **dr** (*trans/cis*) > 20:1; **HPLC** (Lux-Cellulose-4, Heptane/ethanol = 50/50, flow rate = 1.0 mL/min, λ

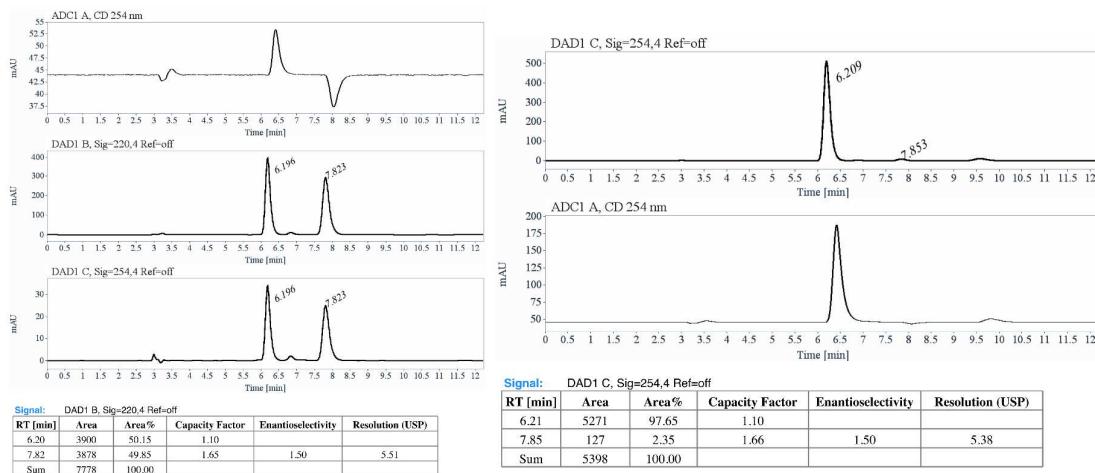
$\lambda = 254$ nm): $t_{\text{major}} = 7.01$ min, $t_{\text{minor}} = 9.18$ min, ee = 94%; $[\alpha]_D^{25}$ (CHCl_3 , $c = 1.0$) = +499; **1H NMR** (400 MHz, DMSO-*d*₆) δ 8.40 (1H, d, $J = 9.1$ Hz, ArH), 8.02 (1H, d, $J = 9.1$ Hz, ArH), 7.99-7.93 (2H, m, ArH), 7.81-7.76 (1H, m, ArH), 7.62 (1H, dd, $J = 9.1, 7.1$ Hz, ArH), 7.56-7.51 (2H, m, ArH), 7.49-7.43 (2H, m, ArH), 7.09 (1H, d, $J = 3.5$ Hz, CH), 6.10 (1H, d, $J = 3.5$ Hz, CH), 3.72 (3H, s, CH_3). **13C NMR** (100 MHz, DMSO-*d*₆) δ 164.2, 157.2, 155.9, 154.4, 133.5, 132.6, 130.6, 128.8, 128.5, 127.2, 125.0, 123.7, 122.9, 122.5, 117.0, 114.0, 111.6, 111.0, 104.5, 56.5, 44.8. **HRMS** (ESI+): $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{22}\text{H}_{16}\text{NO}_6$ ⁺ 390.0972, found 390.0969.



(2*S*,3*S*)-3-(2-methoxynaphthalen-1-yl)-5-methyl-2-nitro-3,5-dihydrofuro[3,2-*c*]quinolin-4(2*H*)-one (3e).

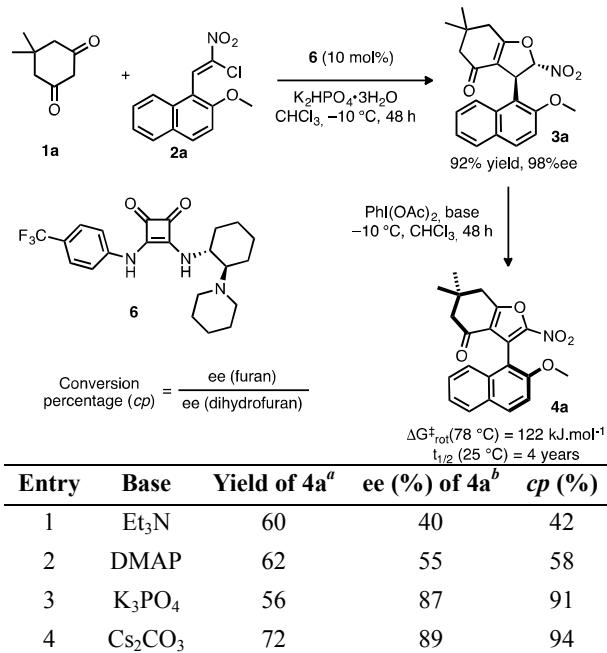


This compound was isolated as a white solid (100 mg, 89%); **mp** = 210-215 °C; **R_f** = 0.50 (ethyl acetate/petroleum ether 1:1); **dr** (*trans/cis*) > 20:1; **HPLC** (Lux-Cellulose-4, Heptane/ethanol = 50/50, flow rate = 1.0 mL/min, $\lambda = 254$ nm): $t_{\text{major}} = 6.20$ min, $t_{\text{minor}} = 7.82$ min, ee = 95%; $[\alpha]_D^{25}$ (CHCl_3 , $c = 1.0$) = +520; **1H NMR** (400 MHz, DMSO-*d*₆) δ 8.43 (1H, d, $J = 9.0$ Hz, ArH), 8.01-7.91 (3H, m, ArH), 7.79-7.73 (1H, m, ArH), 7.64-7.57 (2H, m, ArH), 7.48-7.40 (3H, m, ArH), 7.56-7.51 (2H, m, ArH), 7.49-7.43 (2H, m, ArH), 6.95 (1H, d, $J = 3.1$ Hz, CH), 5.97 (1H, d, $J = 3.1$ Hz, CH), 3.66 (3H, s, CH_3), 3.48 (3H, s, CH_3). **13C NMR** (100 MHz, DMSO-*d*₆) δ 160.3, 158.2, 155.7, 140.4, 132.7, 131.9, 130.2, 128.8, 128.3, 126.9, 123.5, 123.4, 122.3, 118.2, 115.5, 114.1, 111.7, 110.6, 109.3, 56.4, 46.2, 30.7, 28.6. **HRMS** (ESI+): $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{23}\text{H}_{19}\text{N}_2\text{O}_5$ ⁺ 403.1288, found 403.1288.



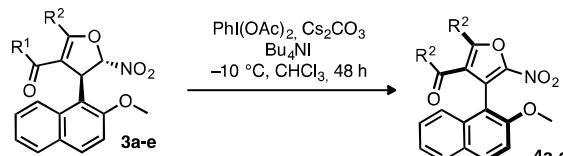
4. General procedure for the oxidation of dihydrofuran 3a-e to furan atropisomers 4a-e

Optimization of the reaction conditions



^aIsolated yield after column chromatography. ^bDetermined by HPLC analysis on chiral stationary phase.

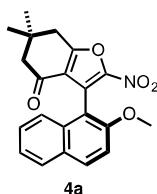
General procedure



The mixture of enantioenriched 2,3-dihydrofuran **3a-3e** (0.27 mmol, 1.0 equiv), PhI(OAc)₂ (210 mg, 0.65 mmol, 2.4 equiv), and Cs₂CO₃ (132 mg, 0.41 mmol, 1.6 equiv) in CHCl₃ (6

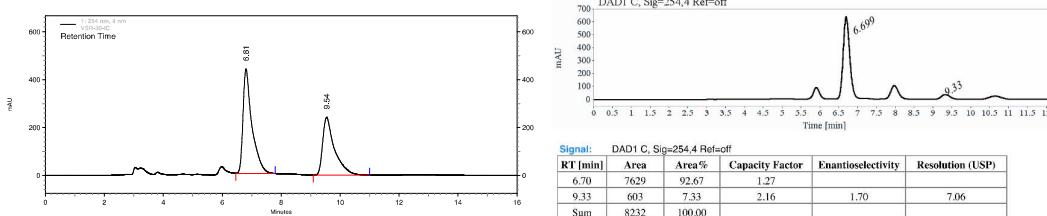
mL) was treated with Bu_4NI (120 mg, 0.32 mmol, 1.2 equiv) at -5°C . The reaction was allowed to stir at same temperature for 48 h. Upon completion as shown by TLC, the reaction mixture was washed with saturated $\text{Na}_2\text{S}_2\text{O}_3$ (20 mL) and extracted using dichloromethane (3×5 mL). The organic layer was dried over Na_2SO_4 and concentrated *in vacuo*. The residue was purified by column chromatography on silica gel ($\text{MeOH}/\text{CH}_2\text{Cl}_2$) to provide corresponding furan **4a-e**.

(*aS*)-3-(2-methoxynaphthalen-1-yl)-6,6-dimethyl-2-nitro-6,7-dihydrobenzofuran-4(5*H*)-one (4a)

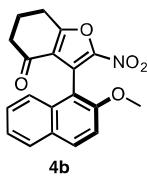


This compound was isolated as an orange solid (71 mg, 72%); **mp** = 165-166 $^\circ\text{C}$; **R_f** = 0.34 (ethyl acetate/petroleum ether 3:7); **HPLC** (Chiralpak IC, Heptane/ethanol = 70/30, flow rate = 1.0 mL/min, λ = 254 nm): t_{major} = 6.81 min, t_{minor} = 9.54 min, ee = 85%; **Chirality conversion percentage** = 94%; $[\alpha]_D^{25}$ (CHCl_3 , c = 1.0) = -60; **¹H NMR** (400 MHz, CDCl_3) δ 7.97 (1H, d, J = 9.1 Hz, ArH), 7.84 (1H, dd, J = 7.6, 1.8 Hz, ArH), 7.40-7.28 (4H, m, ArH), 3.84 (3H, s, OCH₃), 2.95 (2H, d, J = 1.0 Hz, CH₂), 2.40 (2H, s, CH₂), 1.22 (3H, s, CH₃), 1.21 (3H, s, CH₃). **¹³C NMR** (100 MHz, CDCl_3) δ 191.5, 165.9, 155.0, 149.2, 132.1, 131.6, 128.8, 128.7, 127.3, 123.9, 123.5, 122.0, 119.8, 112.8, 110.7, 56.5, 52.6, 37.6, 35.0, 28.7, 28.6. **HRMS** (ESI+): $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{21}\text{H}_{19}\text{NO}_5^+$ 366.1336, found 366.1338.

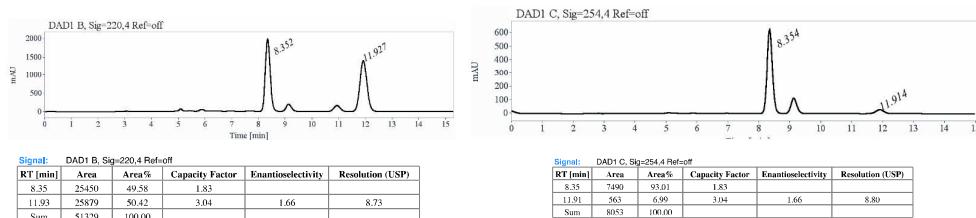
Method description : Chiralpak IC, Heptane/Ethanol 70/30, 1 ml/min, DAD



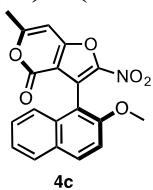
(*aS*)-3-(2-methoxynaphthalen-1-yl)-2-nitro-6,7-dihydrobenzofuran-4(5*H*)-one (4b)



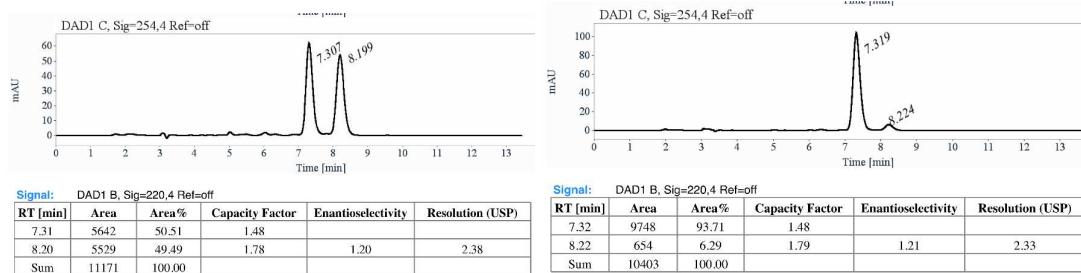
This compound was isolated as a yellow solid (46 mg, 51%); **mp** = 203-204 $^\circ\text{C}$; **R_f** = 0.35 (ethyl acetate/petroleum ether 3:7); **HPLC** (Lux-Cellulose-2, Heptane/ethanol = 50/50, flow rate = 1.0 mL/min, λ = 254 nm): t_{major} = 8.35 min, t_{minor} = 11.93 min, ee = 86%; **Chirality conversion percentage** = 90%; $[\alpha]_D^{25}$ (CHCl_3 , c = 1.0) = -20; **¹H NMR** (400 MHz, CDCl_3) δ 7.97 (1H, d, J = 9.1 Hz, ArH), 7.85-7.82 (1H, m, ArH), 7.38-7.28 (4H, m, ArH), 3.85 (3H, s, OCH₃), 3.11 (2H, t, J = 6.3 Hz, CH₂), 2.52 (2H, dd, J = 7.4, 5.1 Hz, CH₂), 2.30 (2H, pent, J = 6.3, CH₂). **¹³C NMR** (100 MHz, CDCl_3) δ 191.8, 166.8, 155.0, 132.2, 131.7, 128.9, 128.7, 127.3, 123.9, 123.5, 123.0, 119.9, 112.9, 110.8, 56.6, 38.4, 23.9, 21.9. **HRMS** (ESI+): $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{19}\text{H}_{16}\text{NO}_5^+$ 338.1023, found 338.1023.



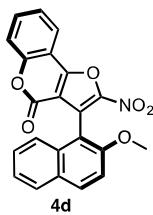
(aS)-3-(2-methoxynaphthalen-1-yl)-6-methyl-2-nitro-4H-furo[3,2-c]pyran-4-one (4c)



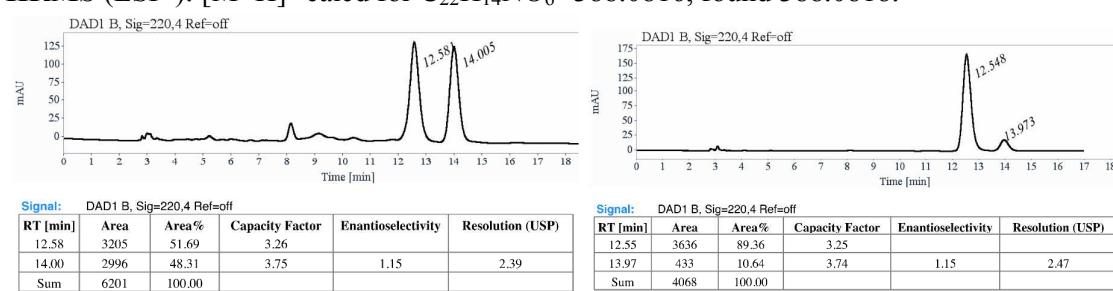
This compound was isolated as a yellow solid (76 mg, 80%); **mp** = 227-229 °C; **R_f** = 0.45 (ethyl acetate/petroleum ether 3:7); **HPLC** (Chiralpak AZ-H, Heptane/ethanol = 50/50, flow rate = 1.0 mL/min, λ = 254 nm): *t*_{major} = 7.31 min, *t*_{minor} = 8.20 min, ee = 87%; **Chirality conversion percentage** = 92%; $[\alpha]_D^{25}$ (CHCl₃, *c* = 1.0) = -158; **¹H NMR** (400 MHz, CDCl₃) δ 8.01 (1H, d, *J* = 9.1 Hz, ArH), 7.86 (1H, dd, *J* = 7.72, 1.76 Hz, ArH), 7.43-7.31 (4H, m, ArH), 6.54 (1H, s, CH), 3.90 (3H, s, OCH₃), 2.39 (3H, s, CH₃). **¹³C NMR** (100 MHz, CDCl₃) δ 165.9, 160.6, 156.7, 155.4, 148.5, 132.4, 132.0, 128.9, 127.7, 124.2, 123.3, 121.2, 112.8, 110.3, 109.3, 95.1, 56.6, 29.9, 21.0. **HRMS** (ESI+): [M+H]⁺ calcd for C₁₉H₁₄NO₆⁺ 352.0816, found 352.0815.



(aS)-3-(2-methoxynaphthalen-1-yl)-2-nitro-4H-furo[3,2-c]chromen-4-one (4d)



This compound was isolated as an orange solid (91 mg, 87%); **mp** = 227-229 °C; **R_f** = 0.72 (ethyl acetate/petroleum ether 3:7); **HPLC** (Lux-Cellulose-2, Heptane/ethanol = 50/50, flow rate = 1.0 mL/min, λ = 254 nm): *t*_{major} = 12.54 min, *t*_{minor} = 13.97 min, ee = 79%; **Chirality conversion percentage** = 81%; $[\alpha]_D^{25}$ (CHCl₃, *c* = 1.0) = -174; **¹H NMR** (400 MHz, CD₂Cl₂) δ 8.15 (1H, td, *J* = 7.3, 1.6 Hz, ArH), 8.10 (1H, dd, *J* = 8.8, 1.8 Hz, ArH), 7.96-7.91 (1H, m, ArH), 7.76-7.70 (1H, m, ArH), 7.55-7.39 (6H, m, ArH), 3.90 (3H, s, OCH₃), 2.39 (3H, s, CH₃). **¹³C NMR** (100 MHz, CD₂Cl₂) δ 142.7, 141.4, 141.3, 140.4, 135.5, 119.7, 118.1, 117.9, 114.7, 114.6, 113.6, 111.4, 110.1, 109.2, 108.2, 107.5, 103.7, 98.8, 98.4, 97.2, 95.2, 42.5. **HRMS** (ESI+): [M+H]⁺ calcd for C₂₂H₁₄NO₆⁺ 388.0816, found 388.0818.

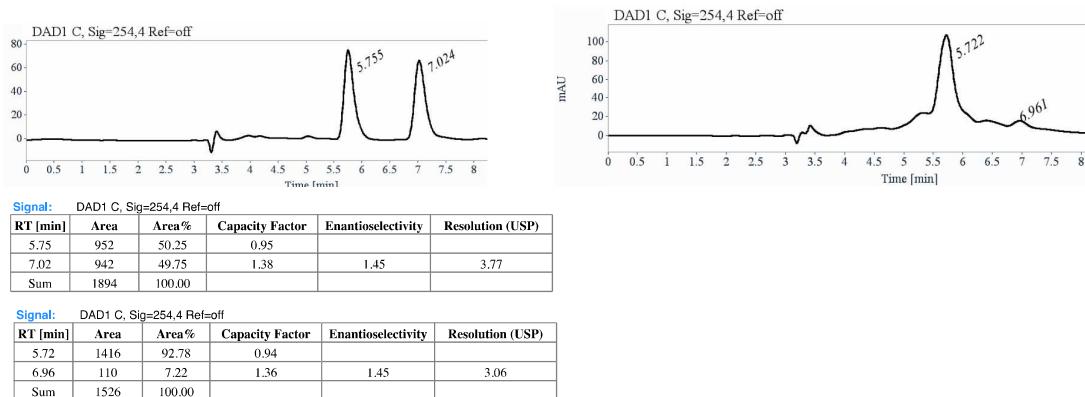


(aS)-3-(2-methoxynaphthalen-1-yl)-5-methyl-2-nitrofuro[3,2-c]quinolin-4(5H)-one (4e)



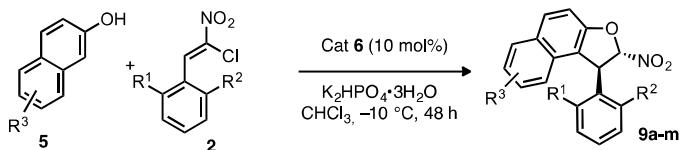
This compound was isolated as an orange solid (75 mg, 69%). This product proved to be very insoluble in most organic solvents; **mp** = 312-314 °C; **R_f** = 0.50 (ethyl acetate/petroleum ether 1:1); **HPLC** (Chiralpak IB, Heptane/ethanol/chloroform = 50/40/10, flow rate = 1.0 mL/min, λ = 254

nm): $t_{\text{major}} = 5.72$ min, $t_{\text{minor}} = 6.96$ min, ee = 86%; **Chirality conversion percentage** = 90%; $[\alpha]_D^{25}$ (CHCl_3 , $c = 1.0$) = +9; **$^1\text{H NMR}$** (400 MHz, $\text{DMSO}-d_6$) δ 8.21 (1H, d, $J = 7.9$, ArH), 8.12 (1H, d, $J = 9.0$ Hz, ArH), 7.96 (1H, d, $J = 7.8$ Hz, ArH), 7.84 (1H, ddd, $J = 8.7, 7.8, 1.1$ Hz, ArH), 7.72 (1H, d, $J = 8.6$ Hz, ArH), 7.58 (1H, d, $J = 9.2$ Hz, ArH), 7.55-7.49 (2H, m, ArH), 7.41-7.32 (2H, m, ArH), 3.81 (3H, s, OCH_3), 3.56 (3H, s, CH_3). **HRMS** (ESI $^+$): $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{23}\text{H}_{17}\text{N}_2\text{O}_5^+$ 401.1132, found 401.1132.



5. General procedure for the enantioselective synthesis of dihydrofurans 10a-10m

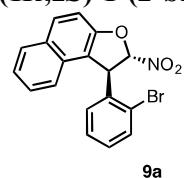
The synthesis of racemic dihydrofuran was accomplished using the following procedure: Triethylamine (2.0 equiv) was added to a solution of chloronitroalkene **2** (1.0 equiv) and 2-naphthol derivative **5** (1.2 equiv) in chloroform ($C = 0.1 \text{ mol.L}^{-1}$). The reaction was stirred at room temperature until TLC showed consumption of starting materials. Purification by flash column chromatography affords the desired product.



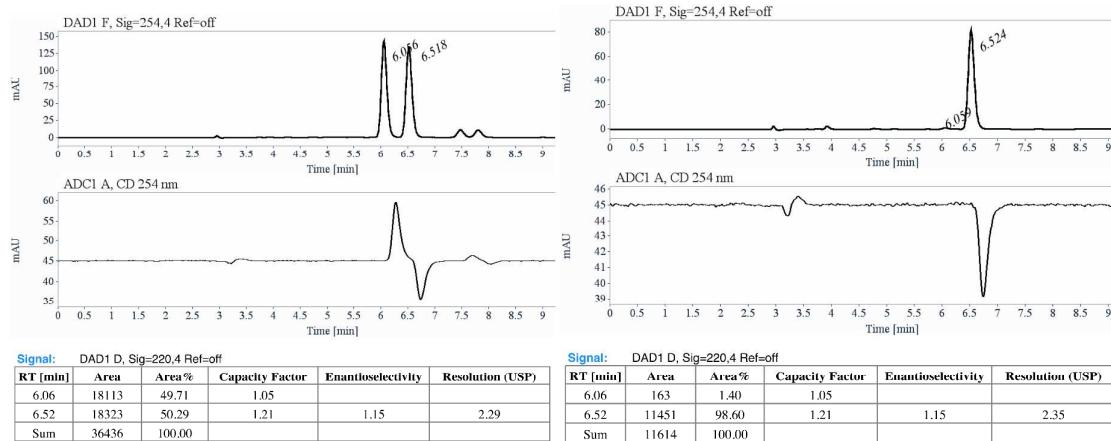
A solution of the corresponding α -chloronitroalkenes (0.38 mmol, 1.0 equiv) and catalyst **6** (10 mol %) in CHCl_3 (0.1 M, 4 mL) was cooled at 0°C . Then, 2-naphthol derivative (0.49 mmol, 1.3 equiv) and K_2HPO_4 (0.95 mmol, 2.5 equiv) were added sequentially. The reaction mixture was stirred at same temperature for 48-72 hours. When the reaction was complete as indicated by TLC analysis, the crude product was directly purified by flash column chromatography on silica gel.

(1*R*,2*S*)-1-(2-bromophenyl)-2-nitro-1,2-dihydronaphtho[2,1-*b*]furan (**9a**)

This compound was isolated as a white solid (105 mg, 75%); **mp** = 115-117 $^\circ\text{C}$; **Rf** = 0.42 (ethyl acetate/petroleum ether 1:9); **dr** (*trans/cis*) > 20:1; **HPLC** (Lux-Cellulose 2, Heptane/ethanol = 90/10, flow rate = 1.0 mL/min, $\lambda = 254$ nm): $t_{\text{major}} = 6.52$ min, $t_{\text{minor}} = 6.06$ min, ee = 97%; $[\alpha]_D^{25}$ (CHCl_3 , $c = 1.0$) = +201; **$^1\text{H NMR}$** (400 MHz, CDCl_3) δ 7.94 (1H, d, $J = 8.8$ Hz,

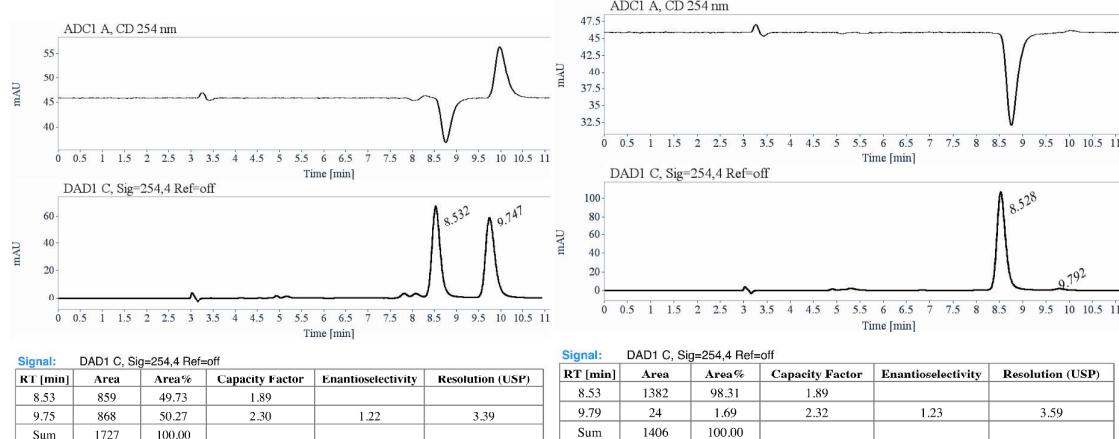


ArH), 7.91-7.87 (1H, m, ArH), 7.73 (1H, dd, J = 7.9, 1.5 Hz, ArH), 7.46 (1H, d, J = 8.8 Hz, ArH), 7.42-7.34 (3H, m, ArH), 7.17 (1H, ddd, J = 7.6, 7.6, 2.2 Hz, ArH), 7.14-7.08 (1H, m, ArH), 6.63 (1H, brd, J = 6.5 Hz, ArH), 6.11 (1H, s, CH), 5.91 (1H, s, CH). ^{13}C NMR (100 MHz, CDCl_3) δ 156.7, 137.0, 133.7, 131.8, 131.1, 130.1, 129.5, 129.4, 129.2, 128.5, 128.1, 124.8, 124.1, 123.2, 118.9, 112.1, 112.0, 53.9. HRMS (ESI $^+$): $[\text{M}+\text{Ag}]^+$ calcd for $\text{C}_{18}\text{H}_{12}\text{BrNO}_3\text{Ag}^+$ 475.9046, found 475.9046.



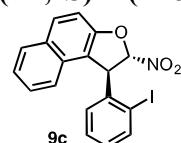
(1*R*,2*S*)-1-(2-chlorophenyl)-2-nitro-1,2-dihydronaphtho[2,1-*b*]furan (9b)

This compound was isolated as a white solid (101 mg, 82%); **mp** = 112-114 °C; **R_f** = 0.57 (ethyl acetate/petroleum ether 1:9); **dr** (*trans/cis*) > 20:1; HPLC (Lux-Amylose 2, Heptane/ethanol = 90/10, flow rate = 1.0 mL/min, λ = 254 nm): t_{major} = 8.53 min, t_{minor} = 9.79 min, ee = 97%; $[\alpha]_D^{25}$ (CHCl_3 , *c* = 1.0) = +185; ^1H NMR (400 MHz, CDCl_3) δ 7.94 (1H, d, J = 9.0 Hz, ArH), 7.91-7.87 (1H, m, ArH), 7.54 (1H, d, J = 7.7, ArH), 7.45 (1H, d, J = 9.0 Hz, ArH), 7.42-7.33 (3H, m, ArH), 7.28-7.23 (1H, m, ArH), 7.07 (1H, dd, J = 7.7, 7.7 Hz, ArH), 6.64 (1H, brd, J = 7.7 Hz, ArH), 6.11 (1H, d, J = 1.2 Hz, CH), 5.88 (1H, s, CH). ^{13}C NMR (100 MHz, CDCl_3) δ 156.7, 135.2, 133.8, 131.8, 131.1, 130.4, 129.9, 129.6, 129.3, 129.2, 128.1, 127.9, 124.8, 123.1, 118.9, 112.1, 112.0, 51.4. HRMS (ESI $^+$): $[\text{M}+\text{Ag}]^+$ calcd for $\text{C}_{18}\text{H}_{12}\text{ClNO}_3\text{Ag}^+$ 431.9551, found 431.9558.

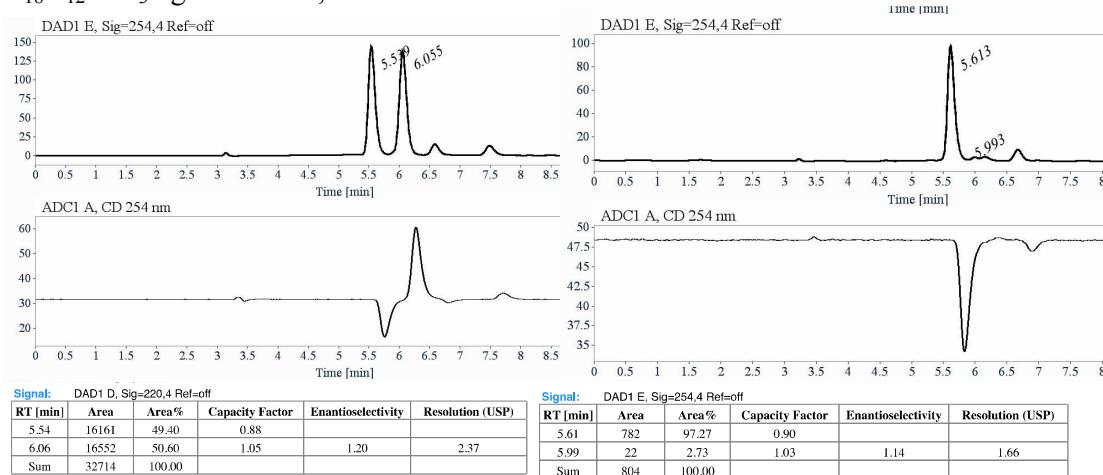


(1*R*,2*S*)-1-(2-iodophenyl)-2-nitro-1,2-dihydronaphtho[2,1-*b*]furan (9c)

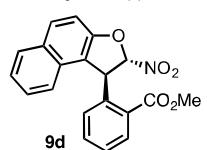
This compound was synthesized on 0.59 mmol scale and was isolated as a light green solid (196 mg, 80%); **mp** = 104-105 °C; **R_f** = 0.40 (ethyl acetate/petroleum ether 1:9); **dr** (*trans/cis*) > 20:1; HPLC (Lux-Amylose 2, Heptane/ethanol = 90/10, flow rate = 1.0 mL/min, λ = 254 nm): t_{major} = 8.53 min, t_{minor} = 9.79 min, ee = 97%; $[\alpha]_D^{25}$ (CHCl_3 , *c* = 1.0) = +185; ^1H NMR (400 MHz, CDCl_3) δ 7.94 (1H, d, J = 9.0 Hz, ArH), 7.91-7.87 (1H, m, ArH), 7.54 (1H, d, J = 7.7, ArH), 7.45 (1H, d, J = 9.0 Hz, ArH), 7.42-7.33 (3H, m, ArH), 7.28-7.23 (1H, m, ArH), 7.07 (1H, dd, J = 7.7, 7.7 Hz, ArH), 6.64 (1H, brd, J = 7.7 Hz, ArH), 6.11 (1H, d, J = 1.2 Hz, CH), 5.88 (1H, s, CH). ^{13}C NMR (100 MHz, CDCl_3) δ 156.7, 135.2, 133.8, 131.8, 131.1, 130.4, 129.9, 129.6, 129.3, 129.2, 128.1, 127.9, 124.8, 123.1, 118.9, 112.1, 112.0, 51.4. HRMS (ESI $^+$): $[\text{M}+\text{Ag}]^+$ calcd for $\text{C}_{18}\text{H}_{12}\text{INO}_3\text{Ag}^+$ 431.9551, found 431.9558.



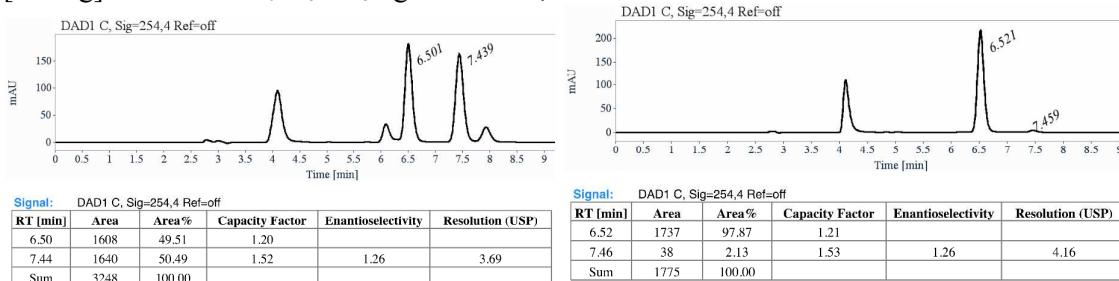
acetate/petroleum ether 1:9); **dr** (*trans/cis*) > 20:1; **HPLC** (Chiralpak IB, Heptane/ethanol = 90/10, flow rate = 1.0 mL/min, λ = 254 nm): t_{major} = 5.61 min, t_{minor} = 5.99 min, ee = 97%; $[\alpha]_D^{25}$ (CHCl_3 , c = 1.0) = +221; **$^1\text{H NMR}$** (400 MHz, CDCl_3) δ 8.01 (1H, dd, J = 7.9, 1.1 Hz, ArH), 7.93 (1H, d, J = 9.0 Hz, ArH), 7.88 (1H, dd, J = 7.3, 1.6 Hz, ArH), 7.45 (1H, d, J = 8.9 Hz, ArH), 7.44-7.32 (3H, m, ArH), 7.18-7.11 (1H, m, ArH), 6.99 (1H, ddd, J = 7.8, 7.8, 1.6 Hz, ArH), 6.59 (1H, d, J = 7.2 Hz, ArH), 6.06 (1H, s, CH), 5.82 (1H, s, CH). **$^{13}\text{C NMR}$** (100 MHz, CDCl_3) δ 156.6, 140.6, 140.5, 131.9, 131.1, 130.3, 129.5, 129.4, 129.2, 128.8, 128.1, 127.5, 124.9, 123.5, 119.7, 112.3, 112.0, 58.8. **HRMS** (ESI+): $[\text{M}+\text{Ag}]^+$ calcd for $\text{C}_{18}\text{H}_{12}\text{INO}_3\text{Ag}^+$ 523.8907, found 523.8912.



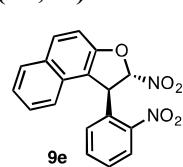
Methyl 2-((1*S*,2*S*)-2-nitro-1,2-dihydropyran-1-yl)benzoate (9d)



This compound was isolated as a white solid (91 mg, 69%); **mp** = 165-167 °C; **R_f** = 0.47 (ethyl acetate/petroleum ether 1:9); **dr** (*trans/cis*) > 20:1; **HPLC** (Lux-Cellulose 2, Heptane/ethanol = 80/20, flow rate = 1.0 mL/min, λ = 254 nm): t_{major} = 6.52 min, t_{minor} = 7.46 min, ee = 96%; $[\alpha]_D^{25}$ (CHCl_3 , c = 1.0) = +194; **$^1\text{H NMR}$** (400 MHz, CDCl_3) δ 8.07 (1H, d, J = 7.8 Hz, ArH), 7.92 (1H, d, J = 8.9 Hz, ArH), 7.92-7.87 (1H, m, ArH), 7.44-7.26 (6H, m, ArH), 6.73 (1H, d, J = 6.1 Hz, ArH), 6.42 (1H, s, CH), 6.14 (1H, s, CH), 4.02 (3H, s, OCH₃). **$^{13}\text{C NMR}$** (100 MHz, CDCl_3) δ 168.1, 156.9, 138.6, 133.1, 131.5, 131.4, 131.0, 129.8, 129.7, 129.2, 129.1, 128.5, 127.8, 124.6, 123.4, 119.1, 113.7, 111.9, 52.9, 50.3. **HRMS** (ESI+): $[\text{M}+\text{Ag}]^+$ calcd for $\text{C}_{20}\text{H}_{15}\text{NO}_5\text{Ag}^+$ 455.9996, found 456.0002.

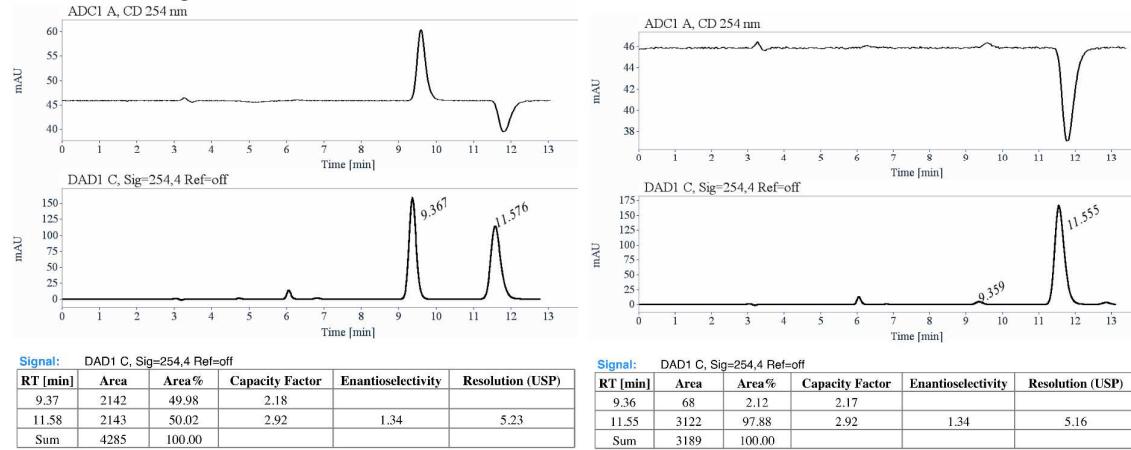


(1*S*,2*S*)-2-nitro-1-(2-nitrophenyl)-1,2-dihydropyran-1-ylbenzoate (9e)

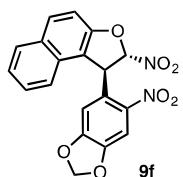


This compound was synthesized on 0.62 mmol and isolated as a white solid (193 mg, 93%); **mp** = 95-97 °C; **R_f** = 0.42 (ethyl acetate/petroleum ether 2:8); **dr** (*trans/cis*) > 20:1; **HPLC** (Chiralpak AZ-H, Heptane/ethanol = 80/20, flow rate = 1.0 mL/min, λ = 254 nm): t_{major} = 11.56 min, t_{minor} = 9.36 min, ee = 96%; $[\alpha]_D^{25}$ (CHCl_3 , c = 1.0) = +230; **$^1\text{H NMR}$** (400 MHz,

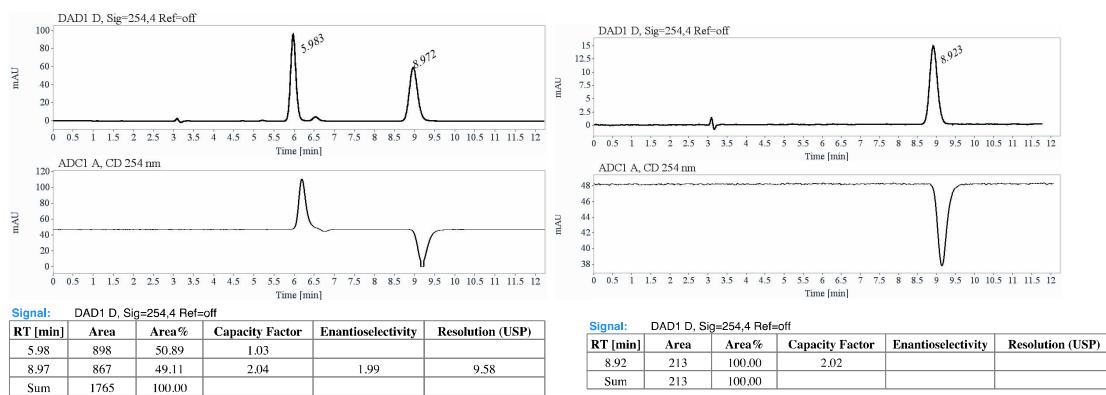
CDCl_3) δ 8.08 (1H, d, J = 8.1 Hz, ArH), 7.96 (1H, d, J = 8.9 Hz, ArH), 7.93-7.88 (1H, m, ArH), 7.50 (1H, ddd, J = 8.1, 7.1, 1.6 Hz, ArH), 7.46-7.37 (4H, m, ArH), 7.33-7.28 (1H, m, ArH), 6.80 (1H, d, J = 6.3 Hz, ArH), 6.20 (1H, s, CH), 6.06 (1H, s, CH). ^{13}C NMR (100 MHz, CDCl_3) δ 157.1, 149.0, 134.2, 132.3, 132.0, 131.1, 130.3, 129.8, 129.4, 129.3, 128.3, 125.5, 125.0, 122.9, 117.5, 112.4, 111.9, 49.2. HRMS (ESI+): $[\text{M}+\text{Ag}]^+$ calcd for $\text{C}_{18}\text{H}_{12}\text{N}_2\text{O}_5\text{Ag}^+$ 442.9792, found 442.9791.



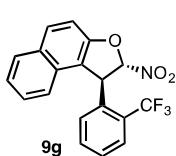
(1*S*,2*S*)-2-Nitro-1-(6-nitrobenzo[*d*][1,3]dioxol-5-yl)-1,2-dihydronaphtho[2,1-*b*]furan (9f).



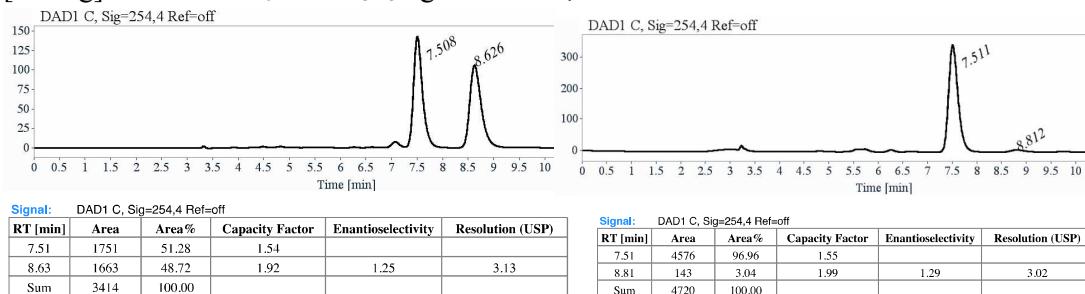
This compound was isolated as a slightly brown solid (556 mg, 83%); **mp** = 201 °C; **R_f** = 0.45 (ethyl acetate/petroleum ether 1:4); **dr** (*trans/cis*) > 20:1; **HPLC** (Chiraldak IE, Heptane/ethanol = 50/50, flow rate = 1.0 mL/min, λ = 254 nm): t_{major} = 8.97 min, t_{minor} = 5.98 min, ee > 99%; $[\alpha]_D^{25}$ (CHCl_3 , c = 1.6) = +234; ^1H NMR (400 MHz, CDCl_3) δ 7.95 (1H, d, J = 9.0 Hz, ArH), 7.92-7.88 (1H, m, ArH), 7.59 (1H, s, ArH), 7.44-7.38 (3H, m, ArH), 7.37-7.31 (1H, m, ArH), 6.16 (1H, s, ArH), 6.13 (2H, s, CH_2), 6.06 (1H, s, CH), 6.02 (1H, s, CH). ^{13}C NMR (100 MHz, CDCl_3) δ 157.1, 152.7, 148.3, 143.0, 132.3, 131.2, 129.5, 129.4, 128.9, 128.3, 125.0, 122.9, 117.7, 112.7, 112.0, 108.7, 106.3, 103.5, 49.3. HRMS (ESI+): $[\text{M}+\text{NH}_4]^+$ calcd for $\text{C}_{19}\text{H}_{16}\text{N}_3\text{O}_7^+$ 398.0983, found 398.0982.



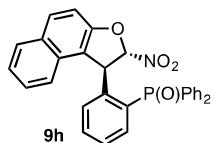
(1*S*,2*S*)-2-nitro-1-(2-(trifluoromethyl)phenyl)-1,2-dihydronaphtho[2,1-*b*]furan (9g). This compound was isolated as a white solid (72 mg, 53%); **mp** = 87-89 °C; **R_f** = 0.30 (ethyl acetate/petroleum ether 1:9); **dr** (*trans/cis*) > 20:1; **HPLC** (Lux-Amylose-2, Heptane/ethanol = 95/5, flow rate = 1.0 mL/min, λ = 254 nm):



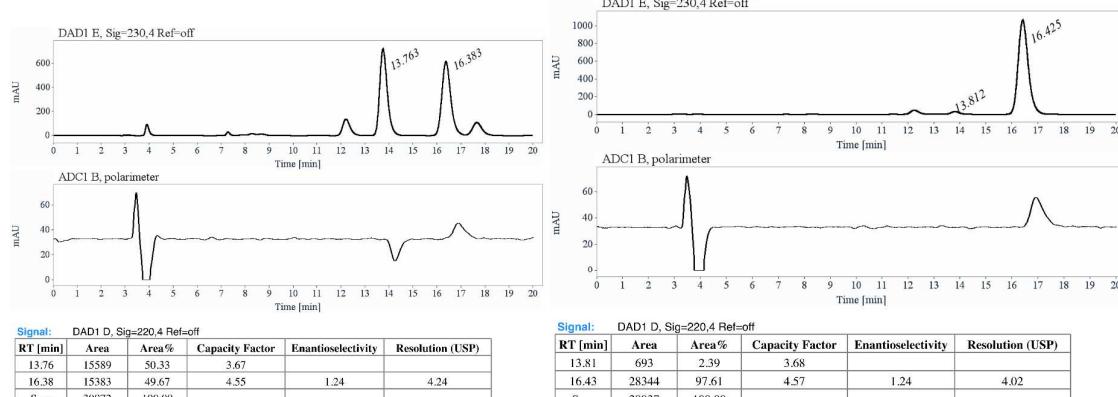
$t_{\text{major}} = 7.51$ min, $t_{\text{minor}} = 8.81$ min, ee = 94%; $[\alpha]_D^{25}$ (CHCl_3 , $c = 1.0$) = +204; **1H NMR** (400 MHz, CDCl_3) δ 7.96 (1H, d, $J = 9.0$ Hz, ArH), 7.92-7.81 (2H, m, ArH), 7.50-7.31 (6H, m, ArH), 6.76 (1H, d, $J = 7.8$ Hz, ArH), 6.07 (1H, s, CH), 5.80 (1H, s, CH). **13C NMR** (100 MHz, CDCl_3) δ 156.8, 136.1, 133.2, 132.1, 131.2, 129.6, 129.4, 129.3, 128.8, 128.4, 128.2, 126.7 (q, $J = 5.8$ Hz), 126.3, 124.9, 122.8, 119.1, 112.3, 111.8, 50.0. **HRMS** (ESI+): $[\text{M}+\text{Ag}]^+$ calcd for $\text{C}_{19}\text{H}_{12}\text{NO}_3\text{F}_3\text{Ag}^+$ 465.9815, found 465.9810.



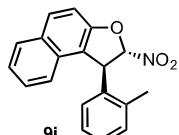
(2-((1*R*,2*S*)-2-nitro-1,2-dihydronaphtho[2,1-*b*]furan-1-yl)phenyl)diphenylphosphine oxide (9h).



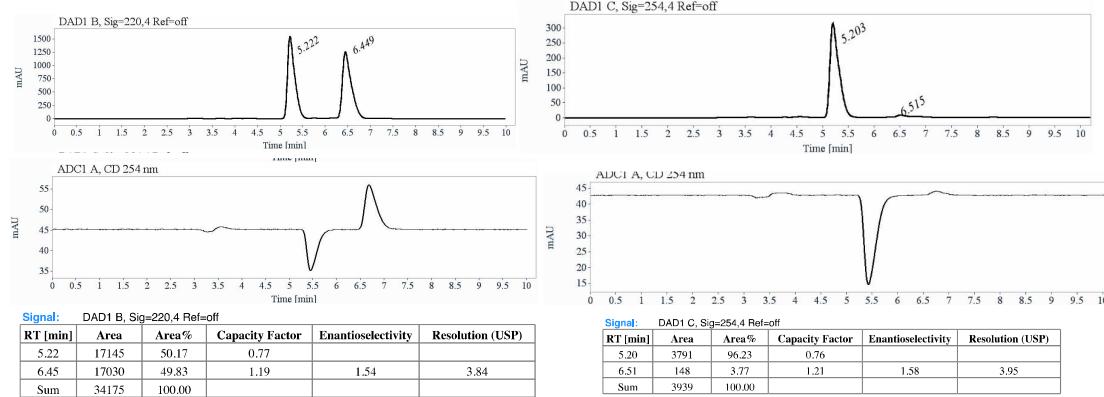
This compound was synthesized on 0.59 mmol and was isolated as a white solid (211 mg, 73%); **mp** = 174-176 °C; **Rf** = 0.47 (ethyl acetate/petroleum ether 1:1); **dr** (*trans/cis*) > 20:1; **HPLC** (Lux-Cellulose-4, Heptane/ethanol = 95/5, flow rate = 1.0 mL/min, $\lambda = 254$ nm): $t_{\text{major}} = 16.43$ min, $t_{\text{minor}} = 13.81$ min, ee = 95%; $[\alpha]_D^{25}$ (CHCl_3 , $c = 1.0$) = +393; **1H NMR** (400 MHz, CDCl_3) δ 7.86-7.74 (6H, m, ArH), 7.66-7.52 (6H, m, ArH), 7.34-7.14 (7H, m, ArH), 6.74 (1H, dd, $J = 7.7, 4.0$ Hz, ArH), 6.44 (1H, s, CH), 6.11 (1H, s, CH). **13C NMR** (100 MHz, CDCl_3) δ 156.5, 142.8 (d, $J = 7.0$ Hz), 133.8 (d, $J = 12.7$ Hz), 133.0, 132.6, 132.5, 132.4 (d, $J = 9.9$ Hz), 132.0, 131.5 (d, $J = 5.4$ Hz), 131.3, 131.0, 130.0 (d, $J = 8.9$ Hz), 129.8, 129.0 (d, $J = 12.1$ Hz), 128.9, 126.7 (d, $J = 12.1$ Hz), 127.6, 124.5, 123.9, 120.2, 113.5, 111.8, 49.9. **31P NMR** (162 MHz, CDCl_3) δ 31.8. **HRMS** (ESI+): $[\text{M}+\text{Ag}]^+$ calcd for $\text{C}_{30}\text{H}_{22}\text{NO}_4\text{F}_3\text{Ag}^+$ 598.0332, found 598.0342.



(1*S*,2*S*)-2-nitro-1-(*o*-tolyl)-1,2-dihydronaphtho[2,1-*b*]furan (9i). This compound was isolated as a white solid (165 mg, 89%); **mp** = 127-128 °C; **Rf** = 0.60 (ethyl acetate/petroleum ether 1:9); **dr** (*trans/cis*) > 20:1; **HPLC** (Chiraldak AD-H, Heptane/ethanol = 80/20, flow rate = 1.0 mL/min, $\lambda = 254$ nm): $t_{\text{major}} =$

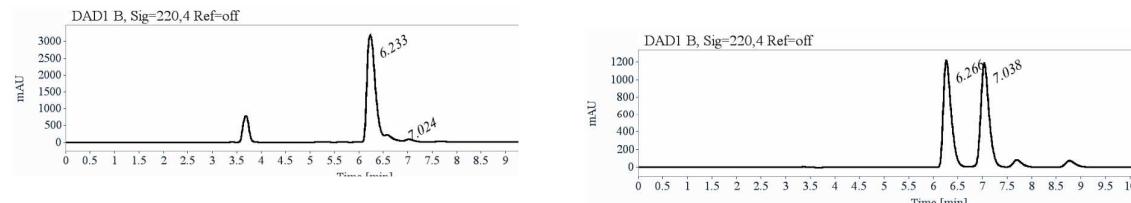


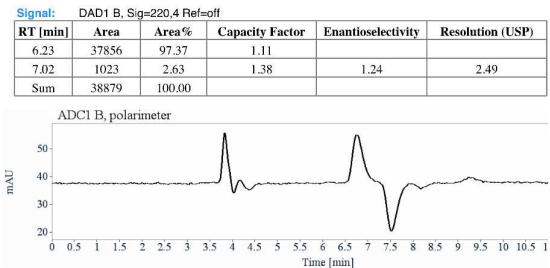
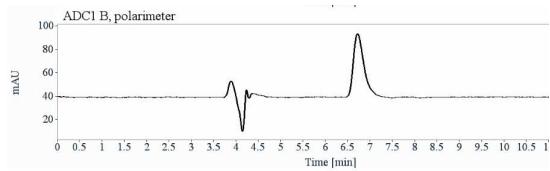
5.20 min, $t_{\text{minor}} = 6.52$ min, ee = 93%; $[\alpha]_D^{25}$ (CHCl_3 , $c = 1.0$) = +131; **$^1\text{H NMR}$** (400 MHz, CDCl_3) δ 7.98-7.86 (2H, m, ArH), 7.48 (1H, d, $J = 8.7$, ArH), 7.41-7.32 (3H, m, ArH), 7.28-7.18 (2H, m, ArH), 7.00 (1H, dd, $J = 7.9, 7.3$ Hz, ArH), 6.52 (1H, d, $J = 7.2$ Hz, ArH), 6.05 (1H, d, $J = 1.5$ Hz, CH), 5.55 (1H, s, CH), 2.76 (3H, s, CH_3). **$^{13}\text{C NMR}$** (100 MHz, CDCl_3) δ 156.7, 136.2, 135.9, 131.5, 131.4, 131.1, 129.6, 129.2, 128.5, 127.9, 127.6, 127.1, 124.7, 123.1, 119.1, 112.4, 112.0, 51.8, 20.3. **HRMS** (ESI+): $[\text{M}+\text{Ag}]^+$ calcd for $\text{C}_{19}\text{H}_{15}\text{NO}_3\text{Ag}^+$ 412.0097, found 412.0096.



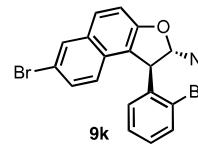
(1*R*,2*S*)-8-bromo-1-(2-bromophenyl)-2-nitro-1,2-dihydronaphtho[2,1-*b*]furan (9j)

This compound was isolated as a white solid (140 mg, 82%); **mp** = 166-167 °C; **R_f** = 0.40 (ethyl acetate/petroleum ether 1:9); **dr** (*trans/cis*) > 20:1; **HPLC** (Chiraldak IB, Heptane/ethanol = 95/5, flow rate = 1.0 mL/min, $\lambda = 254$ nm): $t_{\text{major}} = 6.23$ min, $t_{\text{minor}} = 7.02$ min, ee = 96%; $[\alpha]_D^{25}$ (CHCl_3 , $c = 1.0$) = +128; **$^1\text{H NMR}$** (400 MHz, CDCl_3) δ 7.88 (1H, d, $J = 8.9$ Hz, ArH), 7.76-7.70 (2H, m, ArH), 7.50 (1H, d, $J = 1.6$ Hz, ArH), 7.46-7.41 (2H, m, ArH), 7.21-7.09 (2H, m, ArH), 6.57 (1H, brs, ArH), 6.08 (1H, s, CH), 5.81 (1H, s, CH). **$^{13}\text{C NMR}$** (100 MHz, CDCl_3) δ 157.5, 136.5, 134.0, 131.9, 130.9, 130.8, 130.4, 129.5, 129.2, 128.7, 128.5, 125.5, 124.1, 122.7, 118.2, 112.4, 112.0, 53.6. **HRMS** (ESI+): $[\text{M}+\text{NH}_4]^+$ calcd for $\text{C}_{18}\text{H}_{15}\text{Br}_2\text{N}_2\text{O}_3$ 466.9425, found 466.9425.

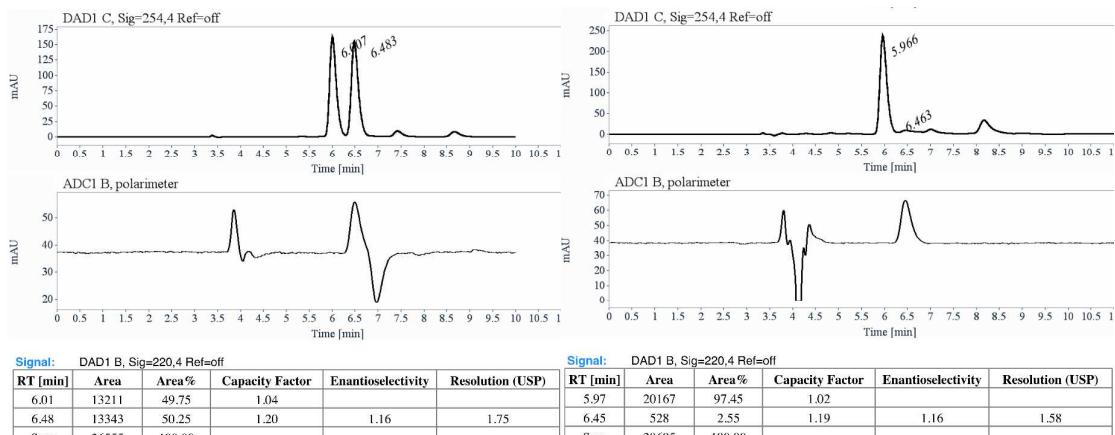




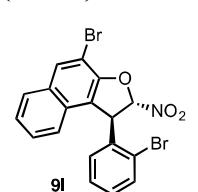
(1*R*,2*S*)-7-bromo-1-(2-bromophenyl)-2-nitro-1,2-dihydronaphtho[2,1-*b*]furan (9j)



This compound was isolated as a white solid (136 mg, 80%); **mp** = 145-147 °C; **R_f** = 0.45 (ethyl acetate/petroleum ether 1:9); **dr** (*trans/cis*) > 20:1; **HPLC** (Chiraldak IB, Heptane/ethanol = 95/5, flow rate = 1.0 mL/min, λ = 254 nm): t_{major} = 5.97 min, t_{minor} = 6.46 min, ee = 95%; $[\alpha]_D^{25}$ (CHCl₃, *c* = 1.0) = +128; **1H NMR** (400 MHz, CDCl₃) δ 8.05 (1H, d, *J* = 1.8 Hz, ArH), 7.84 (1H, d, *J* = 8.9 Hz, ArH), 7.73 (1H, dd, *J* = 8.0, 1.3 Hz, ArH), 7.49-7.44 (2H, m, ArH), 7.27-7.09 (3H, m, ArH), 6.58 (1H, brs, ArH), 6.11 (1H, s, CH), 5.87 (1H, s, CH). **13C NMR** (100 MHz, CDCl₃) δ 156.9, 136.7, 133.8, 132.2, 131.4, 131.2, 131.0, 130.4, 129.3, 128.7, 128.1, 124.9, 124.1, 119.3, 118.6, 113.1, 112.0, 53.7. **HRMS** (ESI+): [M+Na]⁺ calcd for C₁₈H₁₁Br₂NO₃Na⁺ 471.8978, found 471.8980.

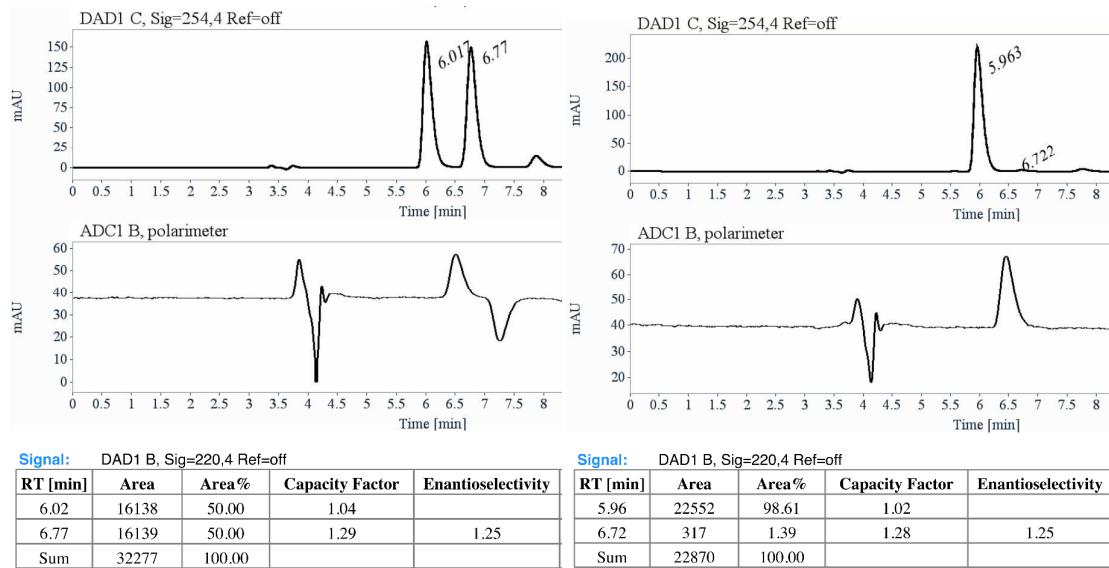


(1*R*,2*S*)-7-bromo-1-(2-bromophenyl)-2-nitro-1,2-dihydronaphtho[2,1-*b*]furan (9j)



This compound was isolated as a white solid (131 mg, 77%); **mp** = 154-156 °C; **R_f** = 0.50 (ethyl acetate/petroleum ether 1:9); **dr** (*trans/cis*) > 20:1; **HPLC** (Chiraldak IB, Heptane/ethanol = 95/5, flow rate = 1.0 mL/min, λ =

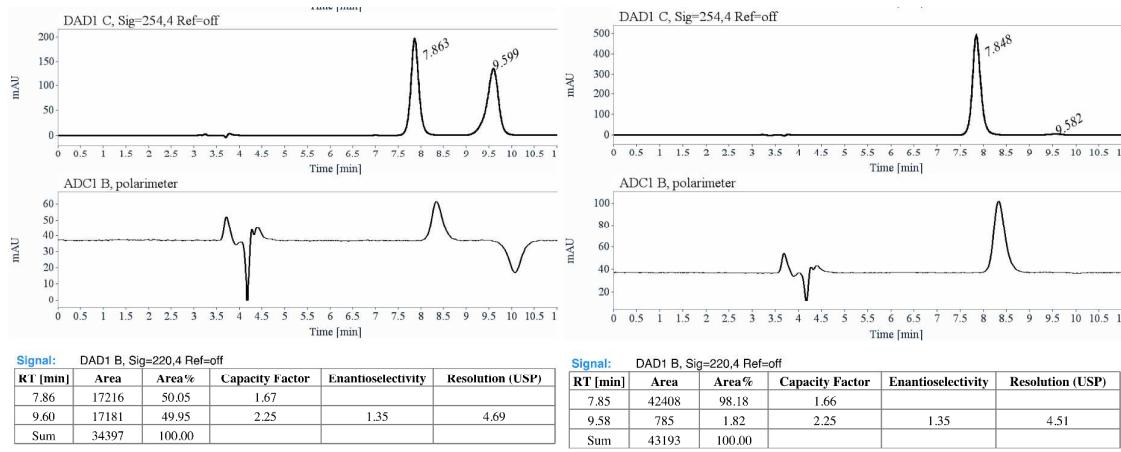
254 nm): $t_{\text{major}} = 5.96$ min, $t_{\text{minor}} = 6.72$ min, ee = 97%; $[\alpha]_D^{25}$ (CHCl_3 , $c = 1.0$) = +152; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 8.14 (1H, s, ArH), 7.86-7.81 (1H, m, ArH), 7.76 (1H, dd, $J = 7.7$, 1.5 Hz, ArH), 7.46-7.39 (2H, m, ArH), 7.37-7.32 (1H, m, ArH), 7.24-7.13 (2H, m, ArH), 6.67 (1H, brs, ArH), 6.18 (1H, s, CH), 6.00 (1H, s, CH). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 153.8, 136.4, 133.9, 133.7, 132.1, 130.4, 129.4, 128.7, 128.5, 128.4, 128.3, 125.8, 124.0, 123.3, 120.4, 111.4, 104.4, 54.8. HRMS (ESI+): $[\text{M}+\text{NH}_4]^+$ calcd for $\text{C}_{18}\text{H}_{15}\text{Br}_2\text{N}_2\text{O}_3^+$ 466.9425, found 466.9427.



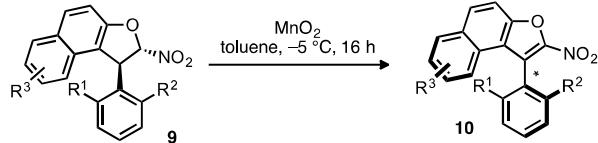
(1*R*,2*S*)-1-(2-bromophenyl)-8-methoxy-2-nitro-1,2-dihydronaphtho[2,1-*b*]furan (9m)



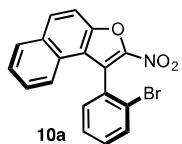
This compound was isolated as a white solid (133 mg, 88%); **mp** = 161-162 °C; **Rf** = 0.42 (ethyl acetate/petroleum ether 1:9); **dr** (*trans/cis*) > 20:1; **HPLC** (Chiralpak IB, Heptane/ethanol = 95/5, flow rate = 1.0 mL/min, $\lambda = 254$ nm): $t_{\text{major}} = 7.85$ min, $t_{\text{minor}} = 9.58$ min, ee = 96%; $[\alpha]_D^{25}$ (CHCl_3 , $c = 1.0$) = +227; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.82 (1H, d, $J = 8.9$ Hz, ArH), 7.75 (1H, d, $J = 8.9$ Hz, ArH), 7.73-7.69 (1H, m, ArH), 7.29 (1H, d, $J = 8.9$ Hz, ArH), 7.19-7.14 (1H, m, ArH), 7.01 (1H, dd, $J = 8.9$, 2.5 Hz, ArH), 6.74 (1H, brs, ArH), 6.68 (1H, d, $J = 2.5$ Hz, ArH), 6.12 (1H, s, CH), 5.86 (1H, s, CH). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 159.3, 156.9, 137.2, 133.5, 131.4, 130.9, 130.7, 130.1, 129.3, 128.8, 126.4, 123.9, 118.6, 117.6, 112.1, 109.2, 101.6, 55.6, 53.9. HRMS (ESI+): $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{19}\text{H}_{14}\text{BrNO}_4\text{Na}^+$ 421.9998, found 421.0000.



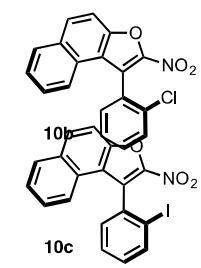
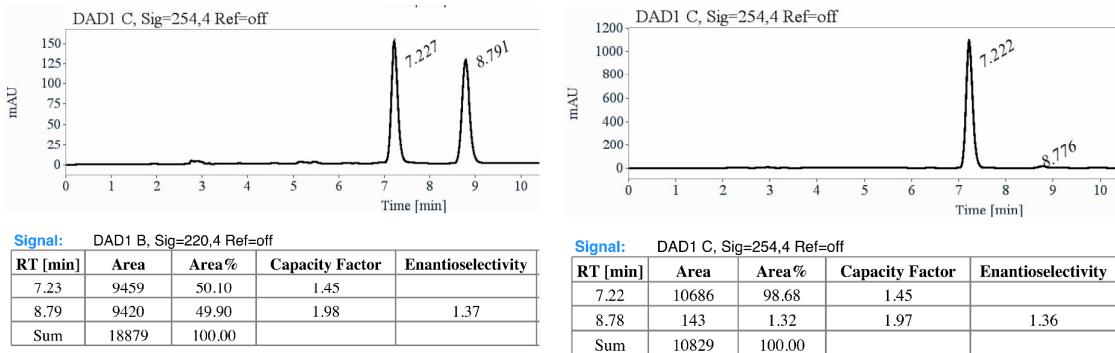
General procedure for the oxidation of dihydrofuran **9a-m** to furan atropisomers **10a-m**



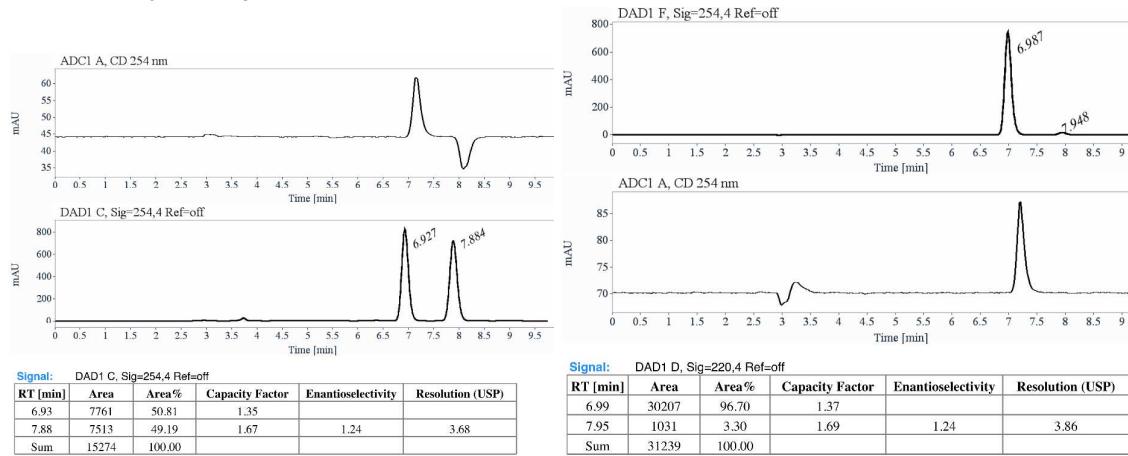
MnO₂ (20 equiv) was added in a clear solution of the corresponding enantioenriched 2,3-dihydrofuran in toluene **9a-m** at -5 °C and reaction mass stirred for 16 h at same temperature. The progress of the reaction was monitored by TLC. After completion of the reaction, the crude product was directly purified by means of flash silica gel column chromatography using pad of celite on the top of silica in column to give the desired furan **10a-m**.



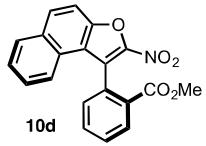
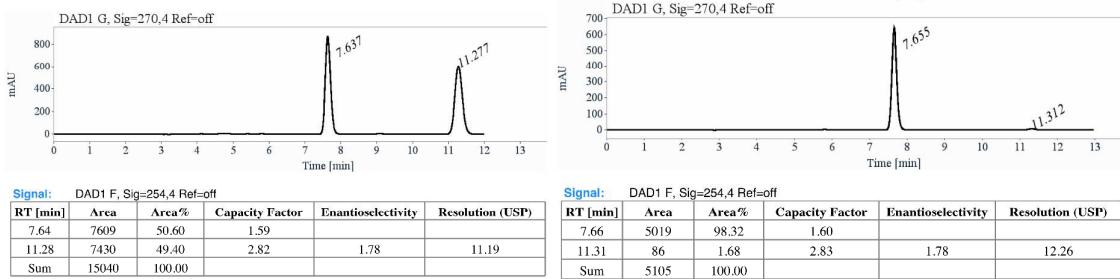
(aS)-1-(2-bromophenyl)-2-nitronaphtho[2,1-b]furan (10a). This compound was isolated as a yellow solid (32 mg, 84%); **mp** = 140-142 °C; **R_f** = 0.36 (ethyl acetate/petroleum ether 1:9); **HPLC** (Lux-Cellulose 2, Heptane/ethanol = 80/20, flow rate = 1.0 mL/min, λ = 254 nm): **t_{major}** = 7.23 min, **t_{minor}** = 8.79 min, ee = 97%; **Chirality conversion percentage** = 100%; $[\alpha]_D^{25}$ (CHCl₃, *c* = 1.0) = -9; **¹H NMR** (400 MHz, CDCl₃) δ 8.06 (1H, d, *J* = 9.0 Hz, ArH), 7.98 (1H, d, *J* = 8.1 Hz, ArH), 7.88-7.84 (1H, m, ArH), 7.59 (1H, s, ArH), 7.76 (1H, d, *J* = 9.0 Hz, ArH), 7.58-7.46 (4H, m, ArH), 7.44-7.35 (2H, m, ArH). **¹³C NMR** (100 MHz, CDCl₃) δ 150.6, 148.2, 133.5, 132.9, 132.1, 131.4, 131.1, 130.7, 129.8, 128.6, 128.4, 128.3, 126.4, 123.5, 123.2, 122.8, 121.5, 112.4. **HRMS** (ESI+): [M+H]⁺ calcd for C₁₈H₁₁NO₃Br⁺ 367.9917, found 367.9917.



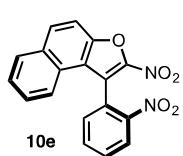
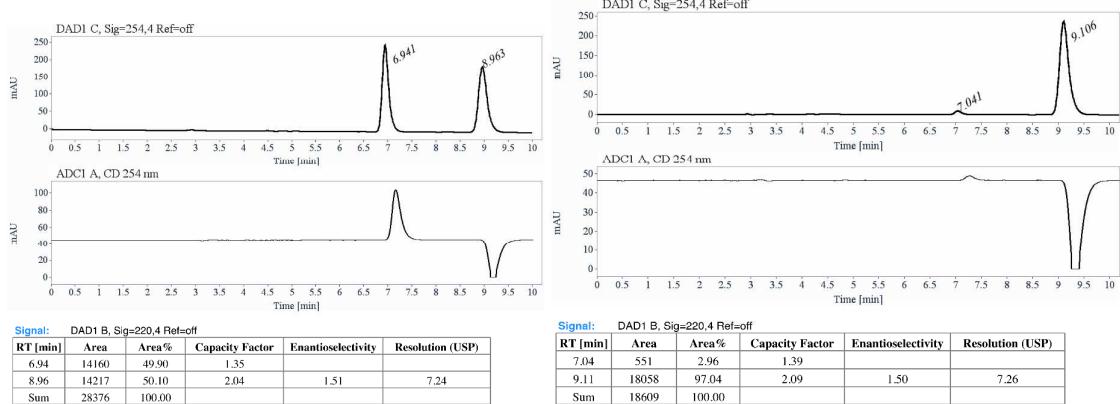
(aS)-1-(2-chlorophenyl)-2-nitronaphtho[2,1-b]furan (10b). This compound was isolated as a yellow solid (106 mg, 92%); **mp** = 182-184 °C; **R_f** = 0.50 (ethyl acetate/petroleum ether 1:9); **HPLC** (Lux-Cellulose 2, Heptane/ethanol = 80/20, flow rate = 1.0 mL/min, λ = 254 nm): t_{major} = 6.99 min, t_{minor} = 7.95 min, ee = 93%; **Chirality conversion percentage** = 96%; $[\alpha]_D^{25}$ (CHCl₃, *c* = 1.0) = -34; **¹H NMR** (400 MHz, CDCl₃) δ 8.06 (1H, d, *J* = 9.0 Hz, ArH), 7.98 (1H, d, *J* = 8.2 Hz, ArH), 7.76 (1H, d, *J* = 9.0 Hz, ArH), 7.68 (1H, ddd, *J* = 8.0, 0.9, 0.9 Hz, ArH), 7.60-7.48 (4H, m, ArH), 7.44-7.39 (2H, m, ArH). **¹³C NMR** (100 MHz, CDCl₃) δ 150.6, 148.3, 133.9, 132.8, 131.4, 131.0, 130.8, 130.3, 129.9, 129.8, 128.6, 128.3, 127.7, 126.4, 122.7, 121.6, 121.4, 112.4. **HRMS** (ESI+): [M+H]⁺ calcd for C₁₈H₁₁NO₃Cl⁺ 324.0422, found 324.0421.



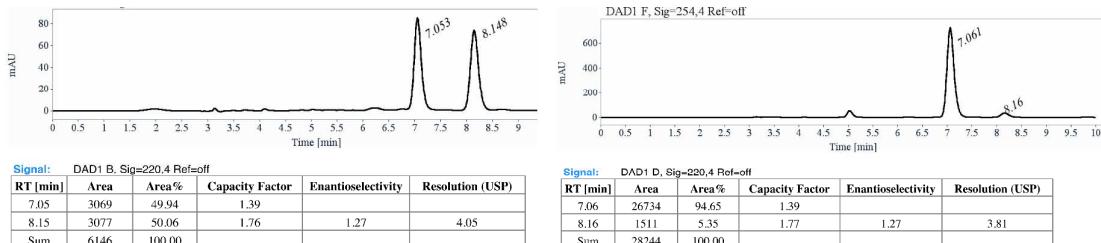
(aS)-1-(2-iodophenyl)-2-nitronaphtho[2,1-b]furan (10c). This compound was isolated as a dark-green solid (447 mg, 90%); **mp** = 180-181 °C; **R_f** = 0.40 (ethyl acetate/petroleum ether 1:9); **HPLC** (Lux-Cellulose 2, Heptane/ethanol = 80/20, flow rate = 1.0 mL/min, λ = 254 nm): t_{major} = 7.66 min, t_{minor} = 11.31 min, ee = 97%; **Chirality conversion percentage** = 100%; $[\alpha]_D^{25}$ (CHCl₃, *c* = 1.0) = -3; **¹H NMR** (400 MHz, CDCl₃) δ 8.12 (1H, dd, *J* = 8.2, 1.1 Hz, ArH), 8.07 (1H, d, *J* = 9.1 Hz, ArH), 7.99 (1H, d, *J* = 8.2 Hz, ArH), 7.76 (1H, d, *J* = 9.1 Hz, ArH), 7.62-7.52 (2H, m, ArH), 7.47-7.38 (2H, m, ArH), 7.35-7.28 (2H, m, ArH). **¹³C NMR** (100 MHz, CDCl₃) δ 150.6, 148.0, 139.8, 136.3, 132.9, 131.4, 131.0, 129.9, 129.8, 129.1, 128.6, 128.4, 126.5, 126.3, 122.9, 121.4, 112.4, 98.5. **HRMS** (ESI+): [M+H]⁺ calcd for C₁₈H₁₁NO₃I⁺ 415.9778, found 415.9780.



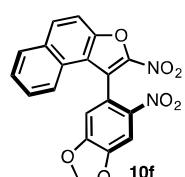
(*aS*)-methyl 2-(2-nitronaphtho[2,1-*b*]furan-1-yl)benzoate (10d). This compound was isolated as a yellow solid (100 mg, 89%); **mp** = 156-159 °C; **R_f** = 0.45 (ethyl acetate/petroleum ether 1:9); **HPLC** (Lux-Cellulose 4, Heptane/ethanol = 60/40, flow rate = 1.0 mL/min, λ = 254 nm): t_{major} = 8.94 min, t_{minor} = 6.95 min, ee = 95%; **Chirality conversion percentage** = 99%; $[\alpha]_D^{25}$ (CHCl_3 , c = 1.0) = +102; **¹H NMR** (400 MHz, CDCl_3) δ 8.34 (1H, dd, J = 7.7, 1.7 Hz, ArH), 8.04 (1H, d, J = 9.1 Hz, ArH), 7.97 (1H, d, J = 8.2 Hz, ArH), 7.79-7.68 (3H, m, ArH), 7.51-7.47 (2H, m, ArH), 7.35 (1H, ddd, J = 8.2, 7.0, 1.4 Hz, ArH), 7.26 (1H, d, J = 8.2 Hz, ArH). **¹³C NMR** (100 MHz, CDCl_3) δ 166.2, 150.6, 147.8, 133.3, 132.6, 132.4, 131.7, 131.4, 130.5, 129.8, 129.7, 129.6, 128.7, 128.1, 126.2, 124.8, 122.8, 122.2, 112.5, 52.4. **HRMS** (ESI+): $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{20}\text{H}_{14}\text{NO}_5^+$ 348.0866, found 348.0866.



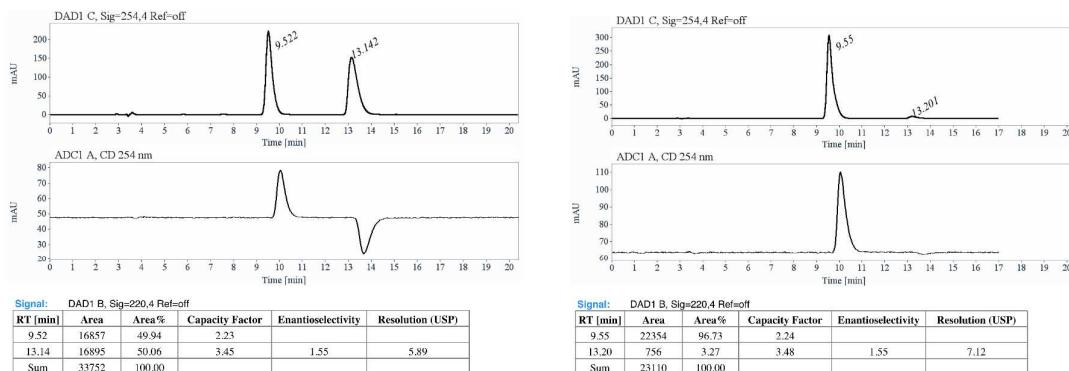
(*aS*)-2-nitro-1-(2-nitrophenyl)naphtho[2,1-*b*]furan (10e). This compound was isolated as a yellow solid (90 mg, 87%); **mp** = 222-224 °C; **R_f** = 0.35 (ethyl acetate/petroleum ether 1:4); **HPLC** (Chiralpak IC, Heptane/ethanol = 80/20, flow rate = 1.0 mL/min, λ = 254 nm): t_{major} = 7.06 min, t_{minor} = 8.16 min, ee = 89%; **Chirality conversion percentage** = 93%; $[\alpha]_D^{25}$ (CHCl_3 , c = 1.0) = +110; **¹H NMR** (400 MHz, CDCl_3) δ 8.45 (1H, dd, J = 8.0, 1.6 Hz, ArH), 8.07 (1H, d, J = 9.0 Hz, ArH), 7.99 (1H, d, J = 8.3 Hz, ArH), 7.87 (1H, ddd, J = 7.3, 7.3, 1.6 Hz, ArH), 7.82 (1H, ddd, J = 7.6, 7.6, 1.6 Hz, ArH), 7.76 (1H, d, J = 9.0 Hz, ArH), 7.58 (1H, dd, J = 7.3, 1.8 Hz, ArH), 7.53 (1H, ddd, J = 7.6, 7.3, 1.3 Hz, ArH), 7.37 (1H, ddd, J = 8.3, 7.0, 1.3 Hz, ArH), 7.25 (1H, d, J = 8.3 Hz, ArH). **¹³C NMR** (100 MHz, CDCl_3) δ 150.9, 148.1, 147.5, 134.5, 133.1, 131.7, 131.5, 130.9, 130.1, 128.4, 128.3, 126.6, 126.5, 125.9, 122.4, 121.6, 121.1, 112.5. **HRMS** (ESI+): $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{18}\text{H}_{11}\text{N}_2\text{O}_5^+$ 335.0662, found 335.0661.



(aS)-2-Nitro-1-(6-nitrobenzo[*d*][1,3]dioxol-5-yl)naphtho[2,1-*b*]furan (10f). This

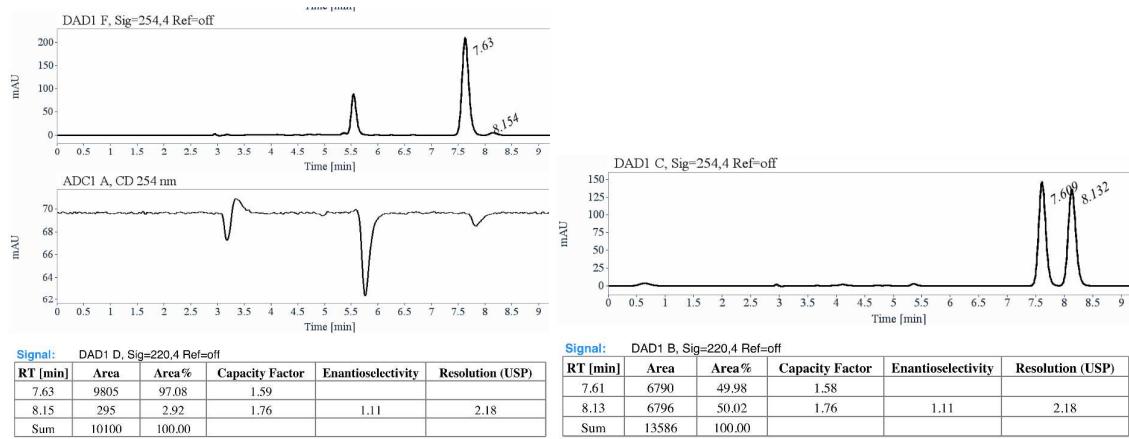


compound was isolated as a yellow-brown solid (200 mg, 85%); **mp** = 203-204 °C; **R_f** = 0.41 (ethyl acetate/petroleum ether 1:4); **HPLC** (Chiralpak IB, Heptane/ethanol = 80/20, flow rate = 1.0 mL/min, λ = 254 nm): $t_{\text{major}} = 9.52 \text{ min}$, $t_{\text{minor}} = 13.14 \text{ min}$, ee = 93%; **Chirality conversion percentage** = 93%; $[\alpha]_D^{25} (\text{CHCl}_3, c = 1.0) = +91.7$; **¹H NMR** (400 MHz, CDCl₃) δ 8.06 (1H, d, J = 9.0 Hz, ArH), 7.99 (1H, d, J = 8.3 Hz, ArH), 7.92 (1H, s, ArH), 7.59 (1H, s, ArH), 7.75 (1H, d, J = 9.0 Hz, ArH), 7.55 (1H, ddd, J = 8.2, 6.6, 1.6 Hz, ArH), 7.47-7.39 (2H, m, ArH), 6.87 (1H, s, ArH), 6.32 (1H, d, J = 1.0 Hz, CHH), 6.28 (1H, d, J = 1.0 Hz, CHH). **¹³C NMR** (100 MHz, CDCl₃) δ 152.8, 150.7, 149.3, 142.2, 132.8, 131.3, 129.8, 128.3, 128.2, 126.3, 122.3, 122.2, 121.3, 121.2, 112.3, 109.8, 106.4, 103.8. **HRMS** (ESI+): [M+H]⁺ calcd for C₁₉H₁₁N₂O₇⁺ 379.0561, found 379.0563.

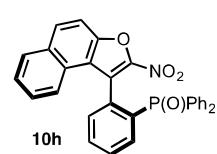


(aS)-2-nitro-1-(2-(trifluoromethyl)phenyl)naphtho[2,1-*b*]furan (10g).

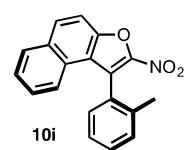
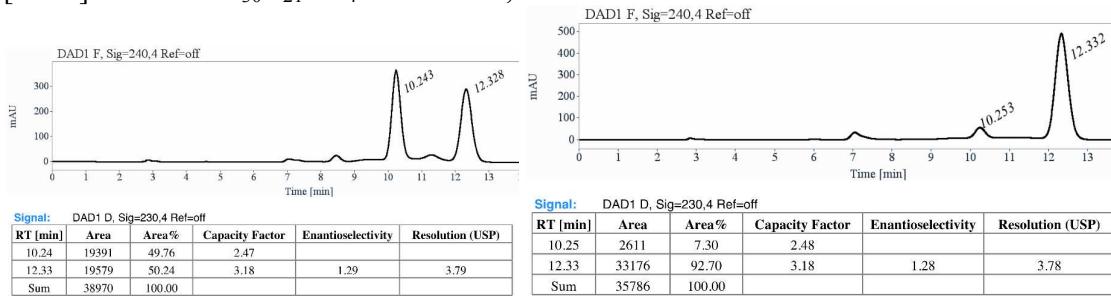
This compound was isolated as a yellow solid (113 mg, 83%); **mp** = 170-172 °C; **R_f** = 0.27 (ethyl acetate/petroleum ether 1:9); **HPLC** (Lux-Cellulose-2, Heptane/ethanol = 90/10, flow rate = 1.0 mL/min, λ = 254 nm): $t_{\text{major}} = 7.63 \text{ min}$, $t_{\text{minor}} = 8.15 \text{ min}$, ee = 97%; **Chirality conversion percentage** = 100%; $[\alpha]_D^{25} (\text{CHCl}_3, c = 1.0) = +152$; **¹H NMR** (400 MHz, CDCl₃) δ 8.06 (1H, d, J = 9.1 Hz, ArH), 8.01-7.95 (2H, m, ArH), 7.82-7.71 (3H, m, ArH), 7.55-7.49 (2H, m, ArH), 7.36 (1H, ddd, J = 8.5, 7.0, 1.3 Hz, ArH), 7.17 (1H, d, J = 8.4 Hz, ArH). **¹³C NMR** (100 MHz, CDCl₃) δ 150.4, 148.4, 132.9, 132.8, 131.5, 130.6, 129.8, 129.7, 129 (q, J = 30.5 Hz), 128.4, 128.2, 127.1 (q, J = 5.1 Hz), 126.4, 125.1, 122.9, 122.4, 122.3, 120.9, 112.3. **HRMS** (ESI+): [M+H]⁺ calcd for C₁₉H₁₁NO₃F₃⁺ 358.0686, found 358.0683.



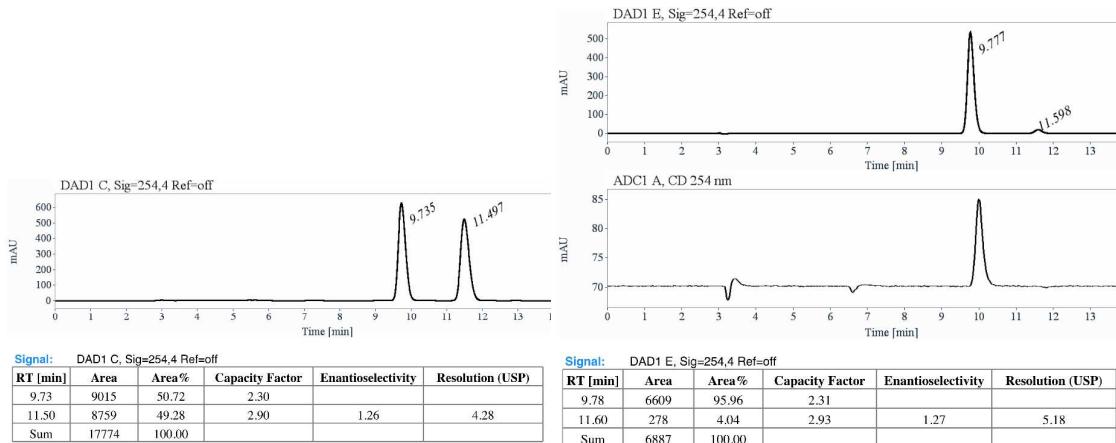
(*aS*)-(2-(2-nitronaphtho[2,1-*b*]furan-1-yl)phenyl)diphenylphosphine oxide (10h).



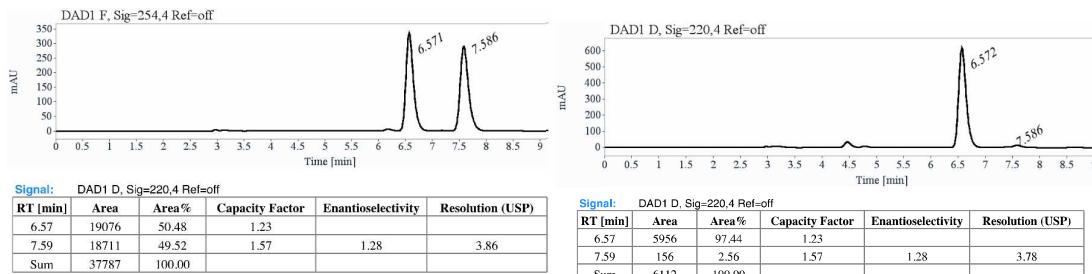
This compound was isolated as a dark green solid (65 mg, 85%); **mp** = 169-170 °C; **R_f** = 0.22 (ethyl acetate/petroleum ether 1:1); **HPLC** (Lux-Cellulose-2, Heptane/ethanol = 50/50, flow rate = 1.0 mL/min, λ = 254 nm): t_{major} = 12.33 min, t_{minor} = 10.25 min, ee = 85%; **Chirality conversion percentage** = 89%; $[\alpha]_D^{25}$ (CHCl_3 , c = 1.0) = +17; **¹H NMR** (400 MHz, CDCl_3) δ 7.90 (1H, d, J = 9.1 Hz, ArH), 7.87 (1H, d, J = 8.2 Hz, ArH), 7.80-7.71 (1H, m, ArH), 7.68-7.60 (2H, m, ArH), 7.59-7.37 (8H, m, ArH), 7.30-7.23 (3H, m, ArH), 7.18 (1H, d, J = 8.3 Hz, ArH), 7.15-7.09 (1H, m, ArH), 7.04-6.97 (2H, m, ArH). **¹³C NMR** (100 MHz, CDCl_3) δ 149.8, 148.8, 135.4 (d, J = 6.7 Hz), 134.5 (d, J = 11.1 Hz), 132.7, 132.0 (d, J = 9.8 Hz), 131.9, 131.6 (d, J = 9.6 Hz), 131.4, 131.2, 130.9 (d, J = 9.2 Hz), 129.4, 129.0 (d, J = 12.0 Hz), 128.7, 128.3 (d, J = 12.2 Hz), 128.1 (d, J = 12.2 Hz), 127.5, 126.0, 123.4, 122.5, 122.1 (d, J = 4.3 Hz), 112.2. **³¹P NMR** (162 MHz, CDCl_3) δ 26.8. **HRMS** (ESI+): $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{30}\text{H}_{21}\text{NO}_4\text{P}^+$ 490.1203, found 490.1207.



(*aS*)-2-nitro-1-(*o*-tolyl)naphtho[2,1-*b*]furan (10i). This compound was isolated as a yellow solid (113 mg, 83%); **mp** = 140-142 °C; **R_f** = 0.57 (ethyl acetate/petroleum ether 1:9); **HPLC** (Lux-Cellulose-2, Heptane/isopropanol = 95/5, flow rate = 1.0 mL/min, λ = 254 nm): t_{major} = 9.78 min, t_{minor} = 11.60 min, ee = 92%; **Chirality conversion percentage** = 100%; $[\alpha]_D^{25}$ (CHCl_3 , c = 1.0) = -36; **¹H NMR** (400 MHz, CDCl_3) δ 8.06 (1H, d, J = 9.1 Hz, ArH), 7.97 (1H, d, J = 8.3 Hz, ArH), 7.76 (1H, d, J = 9.1 Hz, ArH), 7.55-7.45 (3H, m, ArH), 7.43-7.33 (4H, m, ArH), 2.17 (3H, s, CH_3). **¹³C NMR** (100 MHz, CDCl_3) δ 150.6, 148.2, 136.7, 132.8, 131.3, 130.7, 130.0, 129.7, 128.9, 128.3, 126.7, 126.3, 124.1, 122.6, 121.7, 112.4, 20.0. **HRMS** (ESI+): $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{19}\text{H}_{14}\text{NO}_3^+$ 304.0968, found 304.0969.

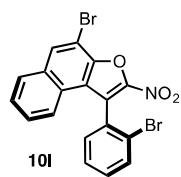
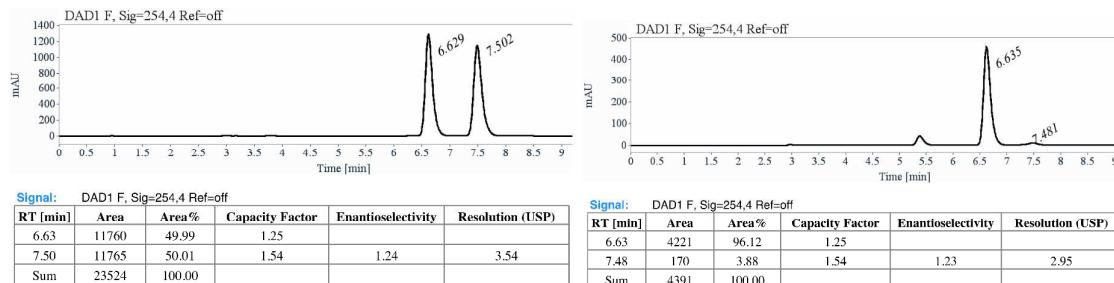


(*aS*)-8-bromo-1-(2-bromophenyl)-2-nitronaphtho[2,1-*b*]furan (10j). This compound was isolated as a dark-green solid (48 mg, 98%); **mp** = 233-234 °C; **R_f** = 0.50 (ethyl acetate/petroleum ether 1:9); **HPLC** (Lux-Cellulose-4, Heptane/isopropanol = 80/20, flow rate = 1.0 mL/min, λ = 254 nm): t_{major} = 6.57 min, t_{minor} = 7.58 min, ee = 95%; **Chirality conversion percentage** = 100%; $[\alpha]_D^{25}$ (CHCl₃, *c* = 1.0) = -17; **1H NMR** (400 MHz, CDCl₃) δ 8.02 (1H, d, *J* = 9.1 Hz, ArH), 7.88 (1H, dd, *J* = 7.9, 1.3 Hz, ArH), 7.83 (1H, d, *J* = 8.7 Hz, ArH), 7.77 (1H, d, *J* = 9.1 Hz, ArH), 7.64-7.59 (1H, m, ArH), 7.57 (1H, dd, *J* = 7.5, 1.3 Hz, ArH), 7.54-7.44 (3H, m, ArH). **13C NMR** (100 MHz, CDCl₃) δ 150.6, 148.1, 133.4, 132.2, 131.1, 131.1, 131.0, 130.5, 129.7, 129.6, 129.5, 128.2, 125.3, 123.3, 122.7, 122.6, 120.5, 112.7. **HRMS** (ESI⁺): [M+H]⁺ calcd for C₁₈H₁₀NO₃Br₂⁺ 447.9003, found 447.9004.



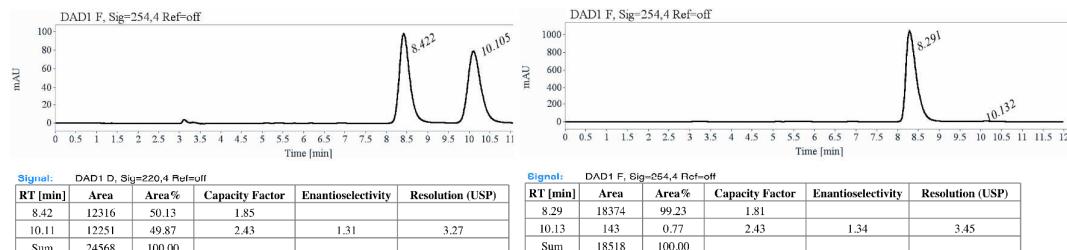
(*aS*)-7-bromo-1-(2-bromophenyl)-2-nitronaphtho[2,1-*b*]furan (10k). This compound was isolated as a dark-green solid (47 mg, 96%); **mp** = 168-169 °C; **R_f** = 0.47 (ethyl acetate/petroleum ether 1:9); **HPLC** (Lux-Cellulose-4, Heptane/isopropanol = 80/20, flow rate = 1.0 mL/min, λ = 254 nm): t_{major} = 6.63 min, t_{minor} = 7.48 min, ee = 92%; **Chirality conversion percentage** = 97%; $[\alpha]_D^{25}$ (CHCl₃, *c* = 1.0) = -52; **1H NMR** (400 MHz, CDCl₃) δ 8.13 (1H, d, *J* = 2.1 Hz, ArH), 7.96 (1H, d, *J* = 9.1 Hz, ArH), 7.85 (1H, dd, *J* = 7.9, 1.5 Hz, ArH), 7.79 (1H, d, *J* = 9.1 Hz, ArH), 7.58-7.44 (4H, m, ArH), 7.23 (1H, d, *J* = 9.1 Hz, ArH). **13C NMR** (100 MHz, CDCl₃) δ 150.4, 148.3, 133.6, 132.7, 131.8, 131.7, 131.6, 131.5, 131.3, 130.7, 128.4, 127.2,

124.3, 123.5, 122.7, 121.6, 120.4, 113.7. **HRMS** (ESI+): $[M+H]^+$ calcd for $C_{18}H_{10}NO_3Br_2^+$ 447.9003, found 447.9000.



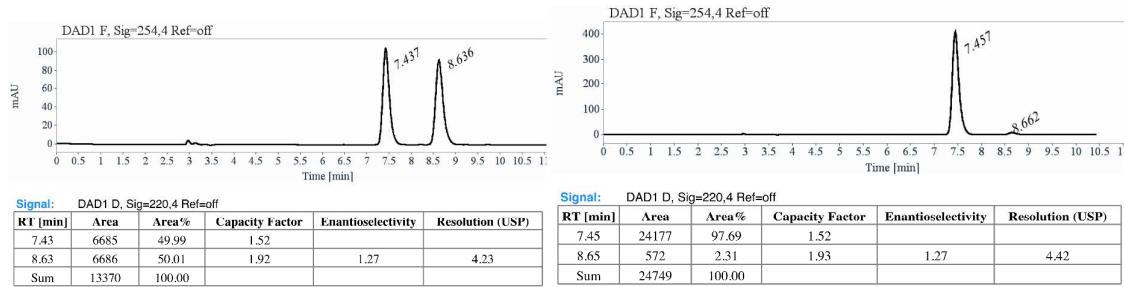
(*aS*)-4-bromo-1-(2-bromophenyl)-2-nitronaphtho[2,1-*b*]furan (**10l**).

This compound was isolated as a dark-green solid (48 mg, 96%); **mp** = 246-247 °C; **R_f** = 0.47 (ethyl acetate/petroleum ether 1:9); **HPLC** (Lux-Cellulose-4, Heptane/isopropanol = 80/20, flow rate = 1.0 mL/min, λ = 254 nm): $t_{\text{major}} = 8.29$ min, $t_{\text{minor}} = 10.13$ min, ee = 98%; **Chirality conversion percentage** = 100%; $[\alpha]_D^{25}$ ($CHCl_3$, $c = 1.0$) = -29; **1H NMR** (400 MHz, $CDCl_3$) δ 8.25 (1H, s, ArH), 7.91 (1H, d, $J = 8.2$ Hz, ArH), 7.86 (1H, dd, $J = 7.9, 1.2$ Hz, ArH), 7.60-7.39 (5H, m, ArH), 7.33 (1H, d, $J = 8.2$ Hz, ArH). **13C NMR** (100 MHz, $CDCl_3$) δ 148.3, 147.5, 134.4, 133.6, 132.4, 131.6, 131.4, 130.7, 128.9, 128.6, 128.4, 127.8, 127.2, 123.5, 123.3, 122.9, 122.5, 104.6. **HRMS** (ESI+): $[M+H]^+$ calcd for $C_{18}H_{10}NO_3Br_2^+$ 447.9003, found 447.8999.



(*aS*)-1-(2-bromophenyl)-8-methoxy-2-nitronaphtho[2,1-*b*]furan (**10m**).

This compound was isolated as a dark-green solid (51 mg, 98%); **mp** = 181-182 °C; **R_f** = 0.40 (ethyl acetate/petroleum ether 1:9); **HPLC** (Lux-Cellulose-4, Heptane/isopropanol = 80/20, flow rate = 1.0 mL/min, λ = 254 nm): $t_{\text{major}} = 7.45$ min, $t_{\text{minor}} = 8.65$ min, ee = 95%; **Chirality conversion percentage** = 99%; $[\alpha]_D^{25}$ ($CHCl_3$, $c = 1.0$) = -6; **1H NMR** (400 MHz, $CDCl_3$) δ 7.97 (1H, d, $J = 9.0$ Hz, ArH), 7.89-7.82 (2H, m, ArH), 7.62-7.42 (4H, m, ArH), 7.14 (1H, dd, $J = 9.0, 2.5$ Hz, ArH). **13C NMR** (100 MHz, $CDCl_3$) δ 159.5, 151.3, 147.9, 133.4, 132.5, 132.4, 131.1, 131.0, 130.9, 130.2, 128.2, 126.2, 123.7, 123.2, 120.8, 118.2, 109.7, 102.4, 55.0. **HRMS** (ESI+): $[M+H]^+$ calcd for $C_{19}H_{13}NO_4Br^+$ 398.0022, found 398.0023.



6. Enantiomerisation barrier determination.

The enantiomerisation barrier, corresponding to barrier to rotation for the following atropisomers, was obtained by kinetic of racemisation of an enantiomer. The slope of the first-order kinetic line gives the racemisation constant ($k_{\text{racemisation}} = 2 \times k_{\text{enantiomerisation}}$). Eyring equation gives the enantiomerisation barrier ($\Delta G^{\ddagger}_{\text{enantiomerisation}}$) from enantiomerisation constant ($K_{\text{enantiomerisation}}$), $R = 8.31451 \text{ J.K}^{-1}.\text{mol}^{-1}$, $h = 6.62608 \text{ } 10^{-34} \text{ J.s}$ and $k_B = 1.38066 \text{ } 10^{-23} \text{ J.K}^{-1}$.

The calculated values of $\Delta G^{\ddagger}_{\text{enantiomerisation}}$ are $\pm 0.5 \text{ kJ/mol}$.

The half-life time $t_{1/2}$, given in the following pages of supplementary material, is $t_{1/2}$ at the temperature used for the kinetic. The half-life time $t_{1/2}$, given in the article, is the half-life time at 25°C , calculated with the hypothesis that the enantiomerisation barrier $\Delta G^{\ddagger}_{\text{enantiomerisation}}$ is independent of temperature.

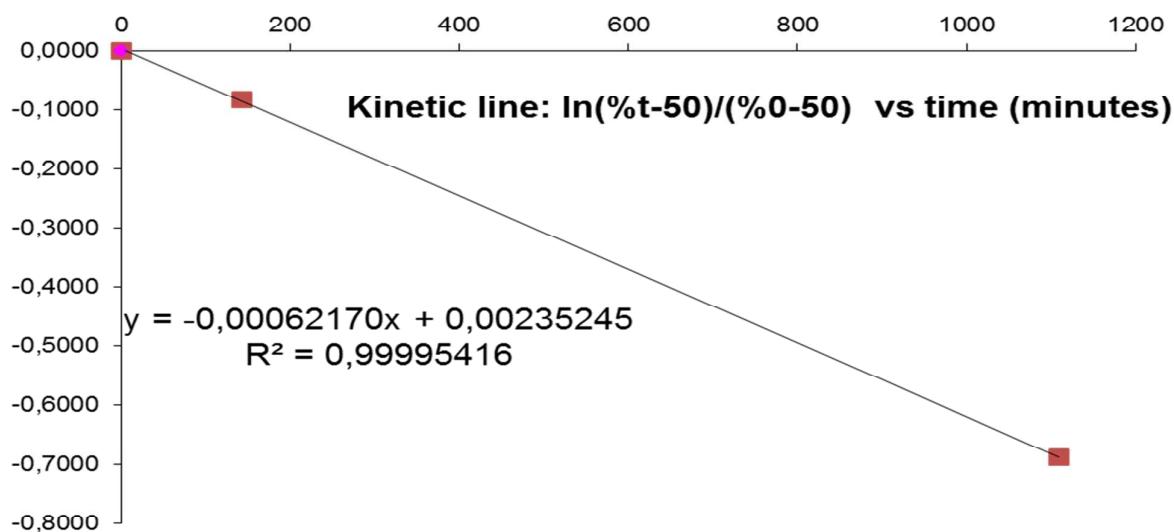
Enantiomerisation barrier for (4a)

About 3 mg of enantio-enriched (**4a**) were refluxed in 15 mL of ethanol. Samples of 10 μ L of this solution were injected on Chiralpak IC (heptane / ethanol 70/30, 1 mL/min, UV detection at 254 nm) to monitor the percentage decrease of the second eluted enantiomer over time.

Solvent : ethanol
 Temperature = 78.29 °C

Time (min)	% second eluted enantiomer (%t)	ln ((%t-50)/(%0-50))
0	99.210	0.0000
142	95.280	-0.0832
1110	74.730	-0.6881

$$\begin{aligned}
 k_{\text{racemisation}} &= 1.03617 \times 10^{-5} \text{ s}^{-1} \\
 k_{\text{enantiomerisation}} &= 5.1808 \times 10^{-6} \text{ s}^{-1} \\
 \Delta G^\ddagger_{\text{enantiomerisation}} &= 122.12 \text{ kJ.mol}^{-1} \\
 &\quad 29.22 \text{ kcal.mol}^{-1} \\
 \text{half-life time } t_{1/2} &= 66895 \text{ seconds} \\
 &\quad 1114.92 \text{ minutes} \\
 &\quad 18.58 \text{ hours} \\
 &\quad 0.77 \text{ days}
 \end{aligned}$$



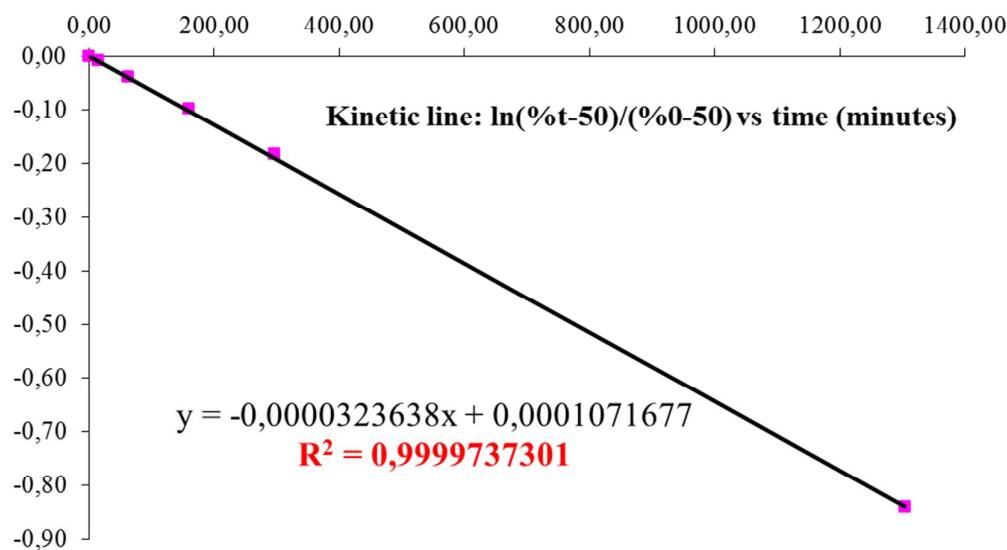
Enantiomerisation barrier for (4b)

About 3 mg of enantio-enriched (**4b**) were refluxed in 15 mL of ethanol. Samples of 10 μ L of this solution were injected on Lux-Cellulose-2 (heptane / ethanol 50/50, 1 mL/min, UV detection at 254 nm) to monitor the percentage decrease of the second eluted enantiomer over time.

Solvent : ethanol
 Temperature = 78.29 °C

Time (min)	% second eluted enantiomer (%t)	ln ((%t-50)/(%0-50))
0.00	91.08	0.0000
15.00	90.73	-0.0086
62.00	89.54	-0.0382
160.00	87.23	-0.0984
297.00	84.23	-0.1824
1304.00	67.73	-0.8403

$$\begin{aligned}
 k_{\text{racemisation}} &= 1.07597 \times 10^{-5} \text{ s}^{-1} \\
 k_{\text{enantiomerisation}} &= 5.3799 \times 10^{-6} \text{ s}^{-1} \\
 \Delta G^\ddagger_{\text{enantiomerisation}} &= 122.01 \text{ kJ.mol}^{-1} \\
 &\quad 29.19 \text{ kcal.mol}^{-1} \\
 \text{half-life time } t_{1/2} &= 64420 \text{ seconds} \\
 &\quad 1073.67 \text{ minutes} \\
 &\quad 17.89 \text{ hours} \\
 &\quad 0.75 \text{ days}
 \end{aligned}$$



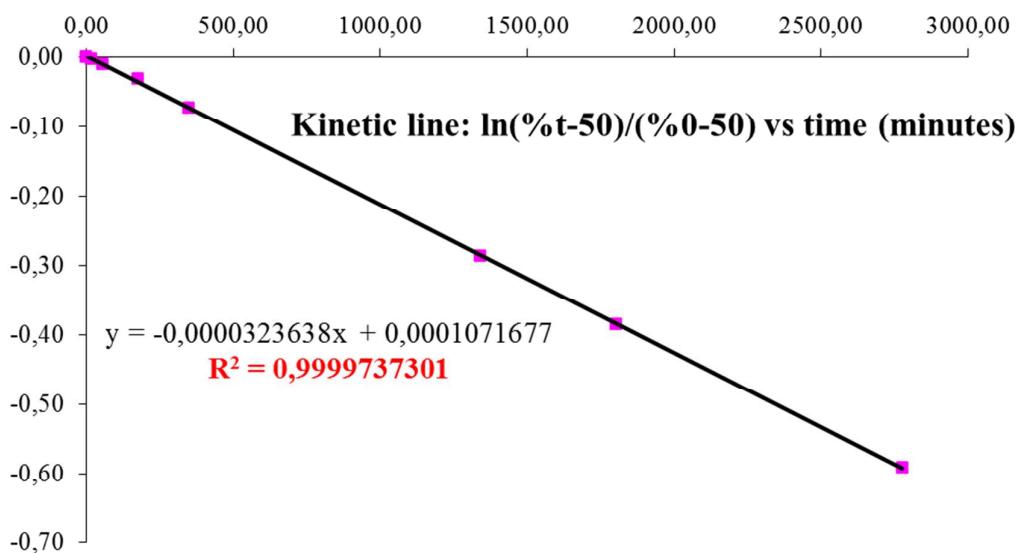
Enantiomerisation barrier for (4c)

About 3 mg of enantio-enriched (**4c**) were refluxed in 15 mL of ethanol. Samples of 10 μ L of this solution were injected on Chiralpak AZ-H (heptane / ethanol 50/50, 1 mL/min, UV detection at 254 nm) to monitor the percentage decrease of the second eluted enantiomer over time.

Solvent : ethanol
 Temperature = 78.29 °C

Time (min)	% second eluted enantiomer (%t)	ln ((%t-50)/(%0-50))
0	94.34	0.0000
16	94.17	-0.0038
57	93.84	-0.0113
175	92.97	-0.0314
350	91.22	-0.0730
1340	83.30	-0.2863
1804	80.14	-0.3860
2777	74.55	-0.5912

$$\begin{aligned} k_{\text{racemisation}} &= 3.56395 \times 10^{-6} \text{ s}^{-1} \\ k_{\text{enantiomerisation}} &= 1.7820 \times 10^{-6} \text{ s}^{-1} \\ \Delta G^\ddagger_{\text{enantiomerisation}} &= \mathbf{125.24} \text{ kJ.mol}^{-1} \\ &\quad 29.96 \text{ kcal.mol}^{-1} \\ \text{half-life time } t_{1/2} &= 194488 \text{ seconds} \\ &\quad 3241.47 \text{ minutes} \\ &\quad 54.02 \text{ hours} \\ &\quad 2.25 \text{ days} \end{aligned}$$



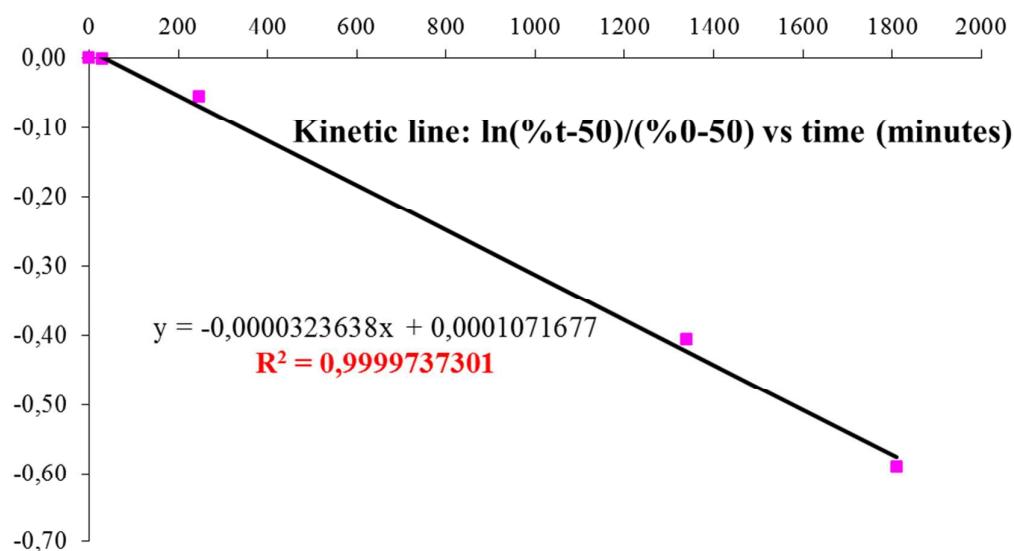
Enantiomerisation barrier for (4d)

About 3 mg of enantio-enriched (**4d**) were refluxed in 15 mL of ethanol. Samples of 10 μ L of this solution were injected on Lux-Cellulose-2 (heptane / ethanol 50/50, 1 mL/min, UV detection at 220 nm) to monitor the percentage decrease of the second eluted enantiomer over time.

Solvent : ethanol
 Temperature = 78.29 °C

Time (min)	% second eluted enantiomer (%t)	ln ((%t-50)/(%0-50))
0	90.03	0.0000
30	89.95	-0.0020
247	87.86	-0.0557
1340	76.66	-0.4065
1811	72.18	-0.5904

$$\begin{aligned}
 k_{\text{racemisation}} &= 5.41033 \times 10^{-6} \text{ s}^{-1} \\
 k_{\text{enantiomerisation}} &= 2.7052 \times 10^{-6} \text{ s}^{-1} \\
 \Delta G^\ddagger_{\text{enantiomerisation}} &= \mathbf{123.91} \text{ kJ.mol}^{-1} \\
 &\quad 29.64 \text{ kcal.mol}^{-1} \\
 \text{half-life time } t_{1/2} &= 128115 \text{ seconds} \\
 &\quad 2135.26 \text{ minutes} \\
 &\quad 35.59 \text{ hours} \\
 &\quad 1.48 \text{ days}
 \end{aligned}$$



Enantiomerisation barrier for (**4e**)

About 3 mg of enantio-enriched (**4e**) were refluxed in 15 mL of chlorobenzene. Samples of 10 μ L of this solution were injected on Chiraldak IB (heptane / ethanol/ chloroform 50/40/10, 1 mL/min, UV detection at 290 nm) to monitor the percentage decrease of the second eluted enantiomer over time.

Solvent : chlorobenzene

Temperature = 131 °C

Time (min)	% second eluted enantiomer (%t)	ln ((%t-50)/(%0-50))
0	89.64	0.0000
12	87.71	-0.0499
24	85.89	-0.0994
36	84.03	-0.1526
50	82.34	-0.2035
73	79.49	-0.2958
110	74.91	-0.4646
141	72.59	-0.5623
168	69.88	-0.6901

$$k \text{ racemisation} = 6.79291 \times 10^{-5} \text{ s}^{-1}$$

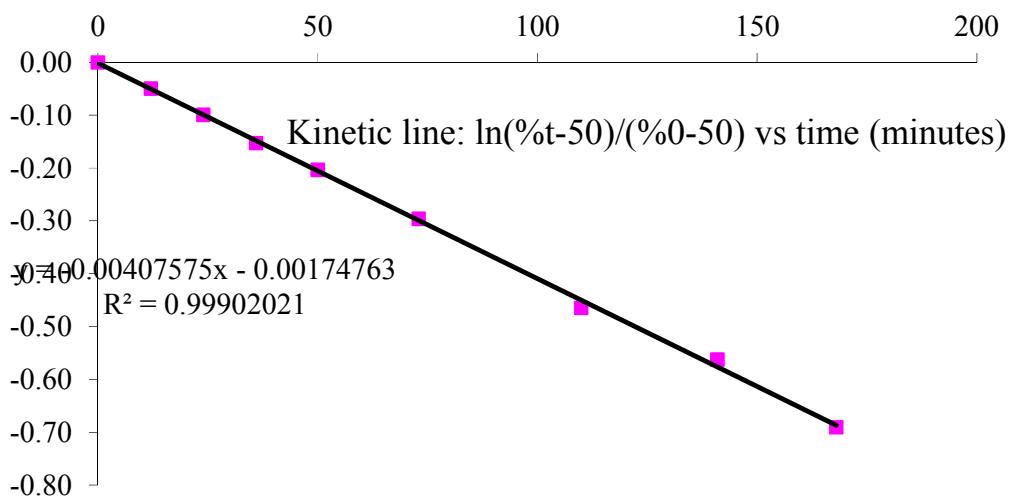
$$k \text{ enantiomerisation} = 3.3965 \times 10^{-5} \text{ s}^{-1}$$

$$\Delta G^\ddagger \text{ enantiomerisation} = 134.59 \text{ kJ.mol}^{-1}$$

$$32.20 \text{ kcal.mol}^{-1}$$

$$\text{half-life time } t_{1/2} =$$

10204 seconds
170.07 minutes
2.83 hours
0.12 days



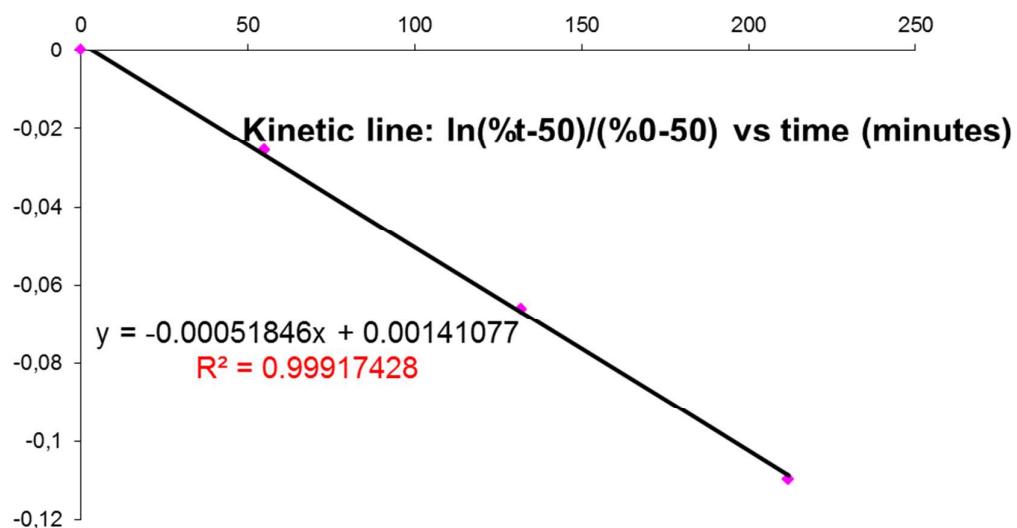
Enantiomerisation barrier for (10a)

About 3 mg of enantio-enriched (**10a**) were refluxed in 15 mL of ethanol. Samples of 10 μ L of this solution were injected on Lux-Cellulose-2 (heptane / ethanol 80/20, 1 mL/min, UV detection at 220 nm) to monitor the percentage decrease of the second eluted enantiomer over time.

Solvent : ethanol
 Temperature = 78.29 °C

Time (min)	% second eluted enantiomer (%t)	ln ((%t-50)/(%0-50))
0	99.02	0
55	97.78	-0.025621236
132	95.88	-0.066199086
212	93.94	-0.109403311

$$\begin{aligned}
 k_{\text{racemisation}} &= 8.64105 \times 10^{-6} \text{ s}^{-1} \\
 k_{\text{enantiomerisation}} &= 4.3205 \times 10^{-6} \text{ s}^{-1} \\
 \Delta G^\ddagger_{\text{enantiomerisation}} &= \mathbf{122.64} \text{ kJ.mol}^{-1} \\
 &\quad 29.34 \text{ kcal.mol}^{-1} \\
 \text{half-life time } t_{1/2} &= 80216 \text{ seconds} \\
 &\quad 1336.93 \text{ minutes} \\
 &\quad 22.28 \text{ hours} \\
 &\quad 0.928421669 \text{ days}
 \end{aligned}$$



Enantiomerisation barrier for (10b)

About 3 mg of enantio-enriched (**10b**) were refluxed in 15 mL of ethanol. Samples of 10 μ L of this solution were injected on Lux-Cellulose-2 (heptane / ethanol 80/20, 1 mL/min, UV detection at 205 nm) to monitor the percentage decrease of the second eluted enantiomer over time.

Solvent : ethanol
 Temperature = 78.29 °C

Time (min)	% second eluted enantiomer (%t)	ln ((%t-50)/(%0-50))
0	93.89	0
13	90.09	-0.090559577
26	86.65	-0.180273075
39	83.49	-0.270439617
52	80.63	-0.359706583
65	78.03	-0.448411139
78	75.63	-0.537922963
91	73.42	-0.628096146

$$k_{\text{racemisation}} = 1.14884 \times 10^{-4} \text{ s}^{-1}$$

$$k_{\text{enantiomerisation}} = 5.7442 \times 10^{-5} \text{ s}^{-1}$$

$$\Delta G^{\ddagger}_{\text{enantiomerisation}} = 115.08 \text{ kJ.mol}^{-1}$$

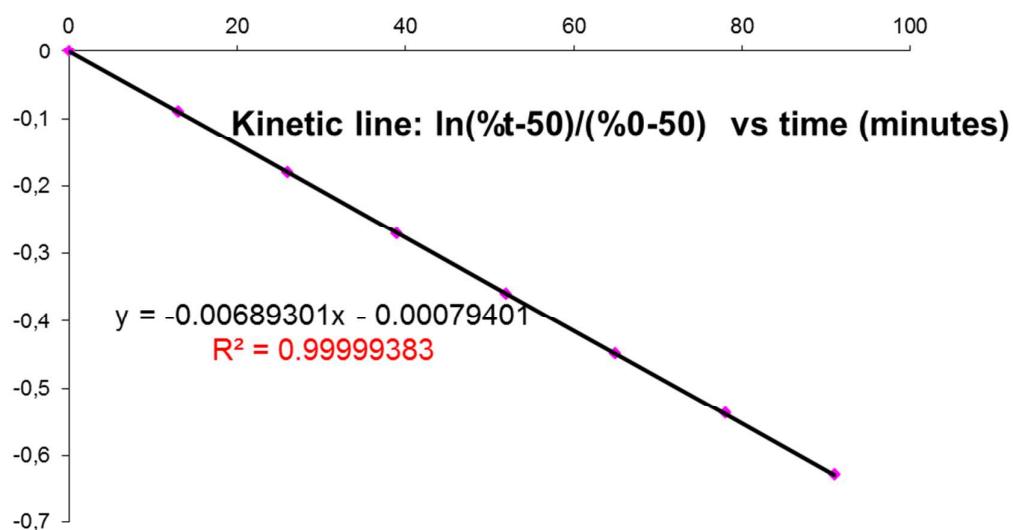
$$27.53 \text{ kcal.mol}^{-1}$$

$$\text{half-life time } t_{1/2} = 6033 \text{ seconds}$$

$$100.56 \text{ minutes}$$

$$1.68 \text{ hours}$$

$$0.069831894 \text{ days}$$



Enantiomerisation barrier for (**10c**)

About 3 mg of enantio-enriched (**10c**) were refluxed in 15 mL of chlorobenzene. Samples of 10 μ L of this solution were injected on Chiralpak IC (heptane / isopropanol 95/5, 1 mL/min, UV detection at 300 nm) to monitor the percentage decrease of the second eluted enantiomer over time.

Solvent : chlorobenzene

Temperature = 131 °C

Time (min)	% second eluted enantiomer (%t)	ln ((%t-50)/(%0-50))
0	89.77	0.0000
19	84.22	-0.1503
37	79.78	-0.2893
55	75.64	-0.4390
74	71.84	-0.5994

$$k_{\text{racemisation}} = 0.000134734 \text{ s}^{-1}$$

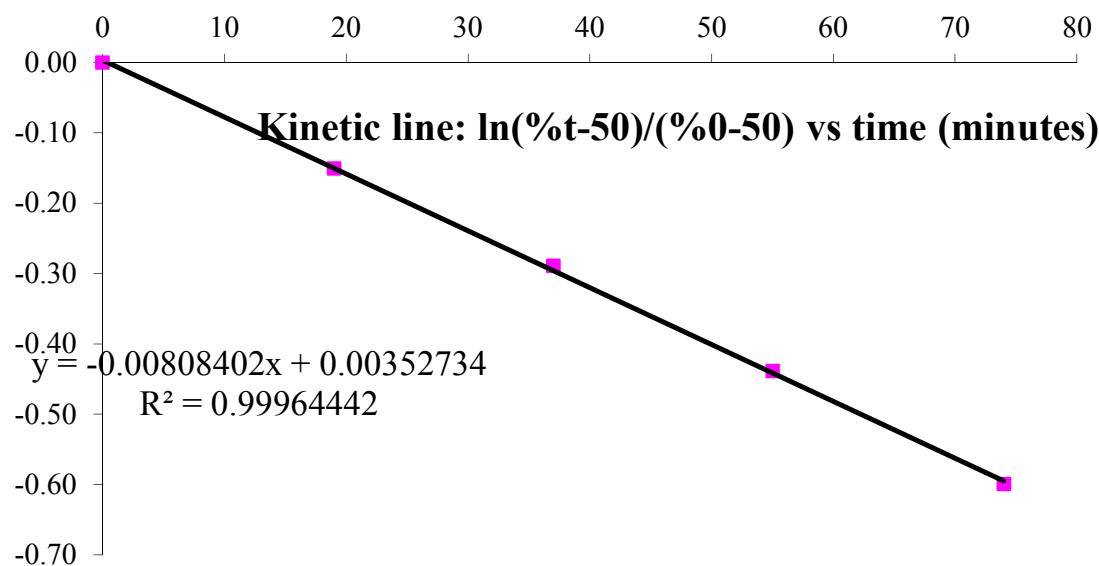
$$k_{\text{enantiomerisation}} = 6.7367 \text{E-}05 \text{ s}^{-1}$$

$$\Delta G^\ddagger_{\text{enantiomerisation}} = 132.29 \text{ kJ.mol}^{-1}$$

$$31.65 \text{ kcal.mol}^{-1}$$

$$\text{half-life time } t_{1/2} =$$

5145	seconds
85.74	minutes
1.43	hours
0.06	days



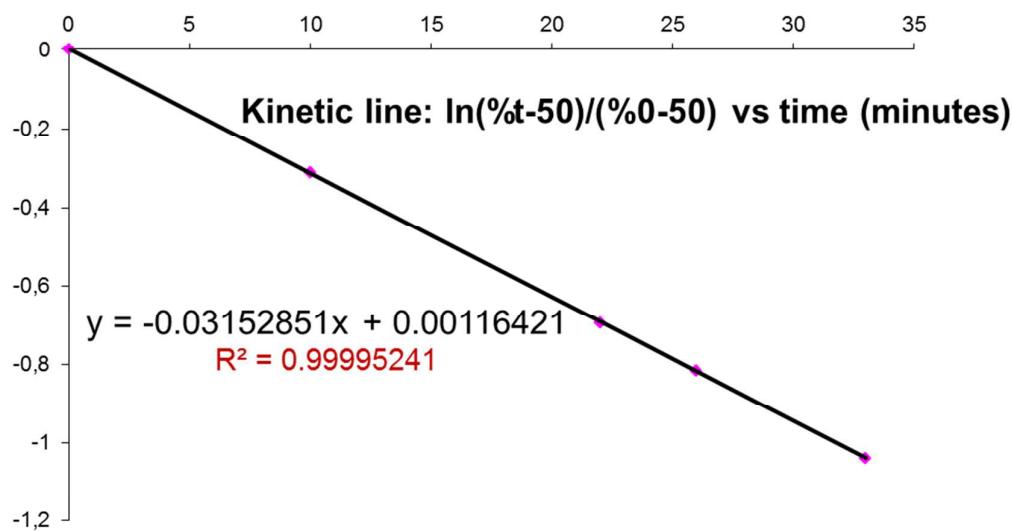
Enantiomerisation barrier for (10d)

About 3 mg of enantio-enriched (**10d**) were refluxed in 15 mL of ethanol. Samples of 10 μ L of this solution were injected on Lux-Cellulose-4 (heptane / ethanol 60/40, 1 mL/min, UV detection at 220 nm) to monitor the percentage decrease of the second eluted enantiomer over time.

Solvent : ethanol
 Temperature = 78.29 °C

Time (min)	% second eluted enantiomer (%t)	ln ((%t-50)/(%0-50))
0	86.4	0
10	76.66	-0.31140446
22	68.14	-0.69644933
26	66.1	-0.815749503
33	62.87	-1.039669753

$$\begin{aligned} k_{\text{racemisation}} &= 5.25475 \times 10^{-4} \text{ s}^{-1} \\ k_{\text{enantiomerisation}} &= 2.6274 \times 10^{-4} \text{ s}^{-1} \\ \Delta G^\ddagger_{\text{enantiomerisation}} &= 110.64 \text{ kJ.mol}^{-1} \\ &\quad 26.47 \text{ kcal.mol}^{-1} \\ \text{half-life time } t_{1/2} &= 1319 \text{ seconds} \\ &\quad 21.98 \text{ minutes} \\ &\quad 0.37 \text{ hours} \\ &\quad 0.015267206 \text{ days} \end{aligned}$$



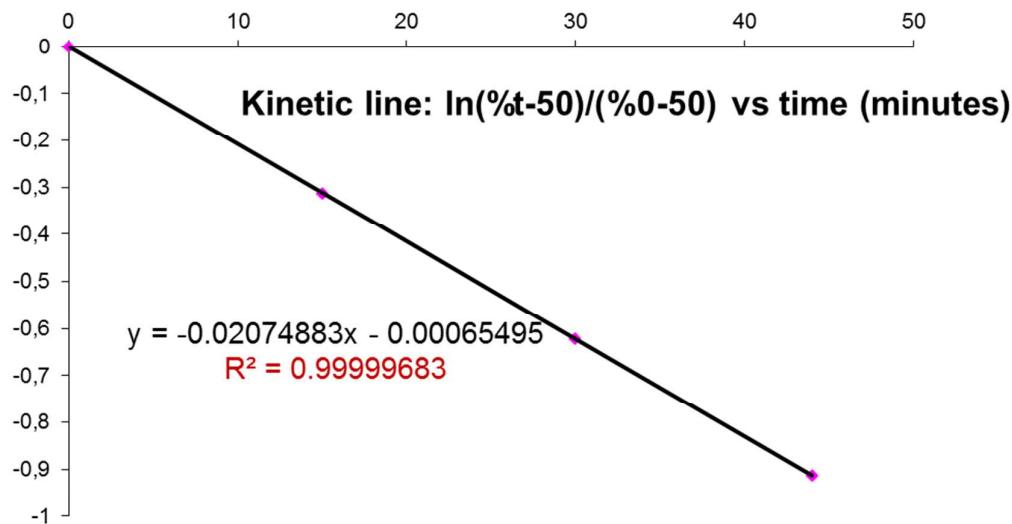
Enantiomerisation barrier for (**10e**)

About 3 mg of enantio-enriched (**10e**) were refluxed in 15 mL of ethanol. Samples of 10 μ L of this solution were injected on Chiralpak IC (heptane / ethanol 80/20, 1 mL/min, UV detection at 254 nm) to monitor the percentage decrease of the second eluted enantiomer over time.

Solvent : ethanol
 Temperature = 78.29 °C

Time (min)	% second eluted enantiomer (%t)	ln ((%t-50)/(%0-50))
0	88.68	0
15	78.29	-0.312814285
30	70.74	-0.623258468
44	65.52	-0.913193156

$$\begin{aligned} k_{\text{racemisation}} &= 3.45814 \times 10^{-4} \text{ s}^{-1} \\ k_{\text{enantiomerisation}} &= 1.7291 \times 10^{-4} \text{ s}^{-1} \\ \Delta G^\ddagger_{\text{enantiomerisation}} &= \mathbf{111.86} \text{ kJ.mol}^{-1} \\ &\quad 26.76 \text{ kcal.mol}^{-1} \\ \text{half-life time } t_{1/2} &= 2004 \text{ seconds} \\ &\quad 33.41 \text{ minutes} \\ &\quad 0.56 \text{ hours} \\ &\quad 0.023199002 \text{ days} \end{aligned}$$



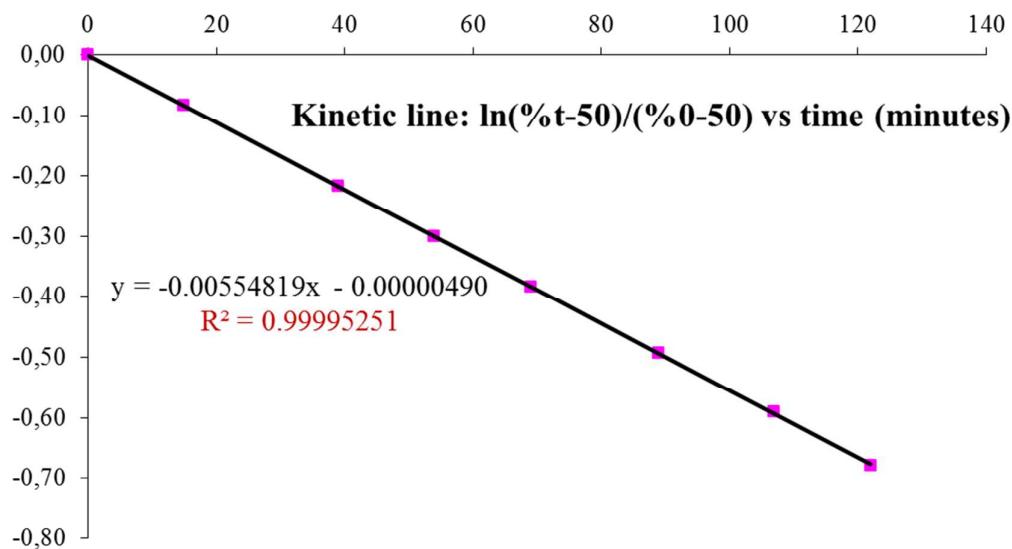
Enantiomerisation barrier for (**10f**)

About 3 mg of enantio-enriched (**10f**) were refluxed in 15 mL of ethanol. Samples of 10 μ L of this solution were injected on Chiraldak IB (heptane / ethanol 80/20, 1 mL/min, UV detection at 220 nm) to monitor the percentage decrease of the second eluted enantiomer over time.

Solvent : ethanol
 Temperature = 78.29 °C

Time (min)	% second eluted enantiomer (%t)	ln ((%t-50)/(%0-50))
0	96.18	0.0000
15	92.51	-0.0828
39	87.18	-0.2168
54	84.21	-0.3000
69	81.46	-0.3838
89	78.20	-0.4932
107	75.59	-0.5903
122	73.41	-0.6794

$$\begin{aligned} k_{\text{racemisation}} &= 9.24699 \times 10^{-5} \text{ s}^{-1} \\ k_{\text{enantiomerisation}} &= 4.6235 \times 10^{-5} \text{ s}^{-1} \\ \Delta G^\ddagger_{\text{enantiomerisation}} &= \textcolor{red}{115.63} \text{ kJ.mol}^{-1} \\ &\quad 27.66 \text{ kcal.mol}^{-1} \\ \text{half-life time } t_{1/2} &= 7496 \text{ seconds} \\ &\quad 124.93 \text{ minutes} \\ &\quad 2.08 \text{ hours} \\ &\quad 0.09 \text{ days} \end{aligned}$$



Enantiomerisation barrier for (10g)

About 3 mg of enantio-enriched (**10g**) were refluxed in 15 mL of chlorobenzene. Samples of 10 μ L of this solution were injected on (S,S)-Whelk-O1 (heptane / isopropanol 95/5, 1 mL/min, UV detection at 220 nm) to monitor the percentage decrease of the second eluted enantiomer over time.

Solvent : chlorobenzene

Temperature = 132 °C

Time (min)	% second eluted enantiomer (%t)	ln ((%t-50)/(%0-50))
0	96.8	0
59	95.28	-0.033017769
1181	75.79	-0.595896383

$$k_{\text{racemisation}} = 8.38783 \times 10^{-6} \text{ s}^{-1}$$

$$k_{\text{enantiomerisation}} = 4.1939 \times 10^{-6} \text{ s}^{-1}$$

$$\Delta G^\ddagger_{\text{enantiomerisation}} = 141.97 \text{ kJ.mol}^{-1}$$

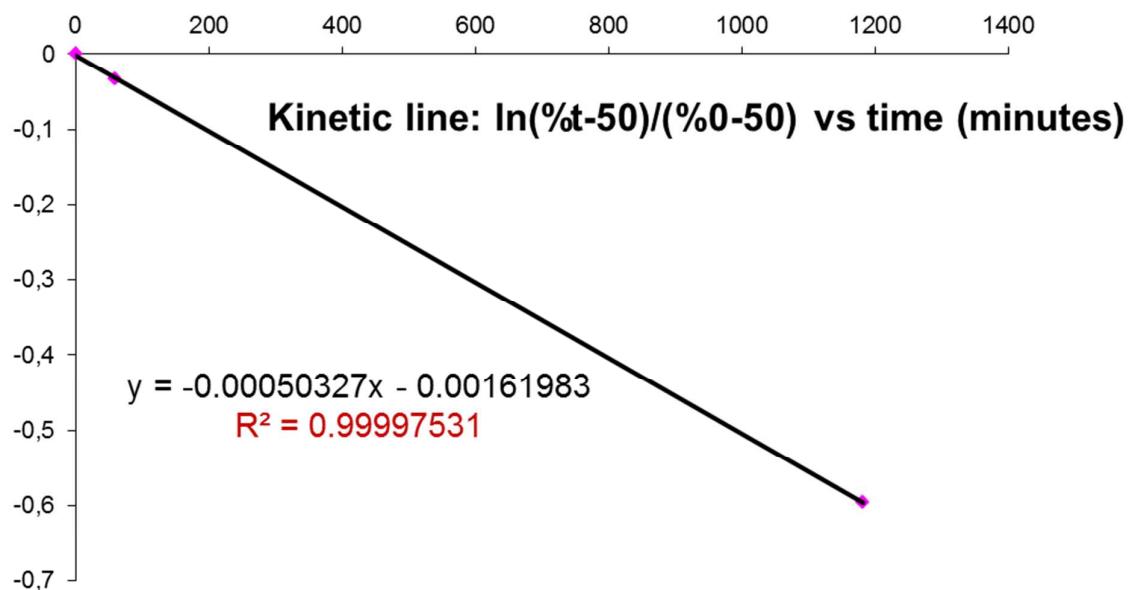
$$33.96 \text{ kcal.mol}^{-1}$$

$$\text{half-life time } t_{1/2} = 82637 \text{ seconds}$$

$$1377.29 \text{ minutes}$$

$$22.95 \text{ hours}$$

$$0.956449469 \text{ days}$$



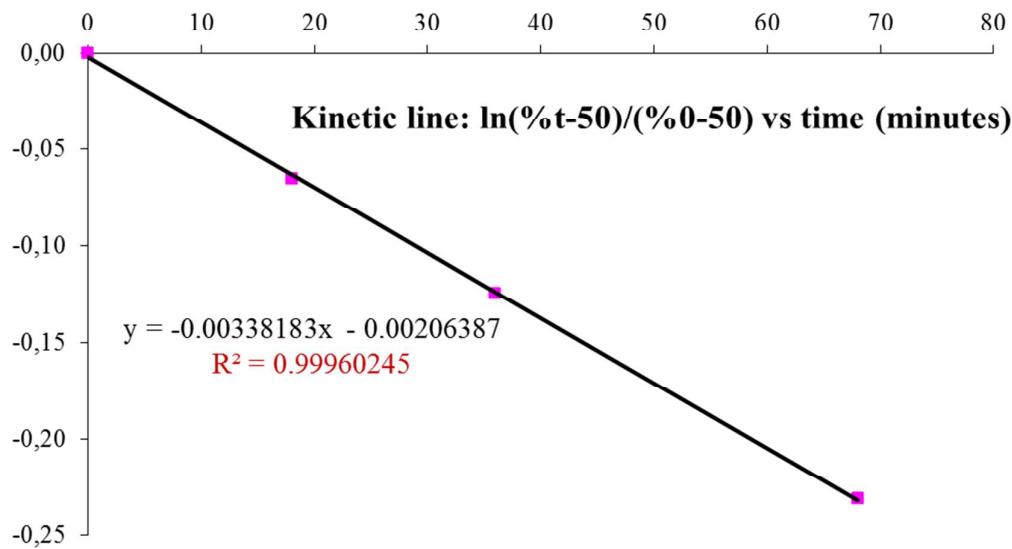
Enantiomerisation barrier for (10h)

About 3 mg of enantio-enriched (**10h**) were refluxed in 15 mL of 1,2-dichlorobenzene. Samples of 10 μ L of this solution were injected on (S,S)-Whelk-O1 (heptane / ethanol 50/50, 1 mL/min, UV detection at 230 nm) to monitor the percentage decrease of the second eluted enantiomer over time.

Solvent : 1,2-dichlorobenzene
 Temperature = 182 °C

Time (min)	% second eluted enantiomer (%t)	ln ((%t-50)/(%0-50))
0	91.39	0.0000
18	88.77	-0.0654
36	86.55	-0.1244
68	82.85	-0.2311

$$\begin{aligned}
 k_{\text{racemisation}} &= 5.63638 \times 10^{-5} \text{ s}^{-1} \\
 k_{\text{enantiomerisation}} &= 2.8182 \times 10^{-5} \text{ s}^{-1} \\
 \Delta G^\ddagger_{\text{enantiomerisation}} &= \mathbf{152.73} \text{ kJ.mol}^{-1} \\
 &\quad 36.54 \text{ kcal.mol}^{-1} \\
 \text{half-life time } t_{1/2} &= 12298 \text{ seconds} \\
 &\quad 204.96 \text{ minutes} \\
 &\quad 3.42 \text{ hours} \\
 &\quad 0.14 \text{ days}
 \end{aligned}$$



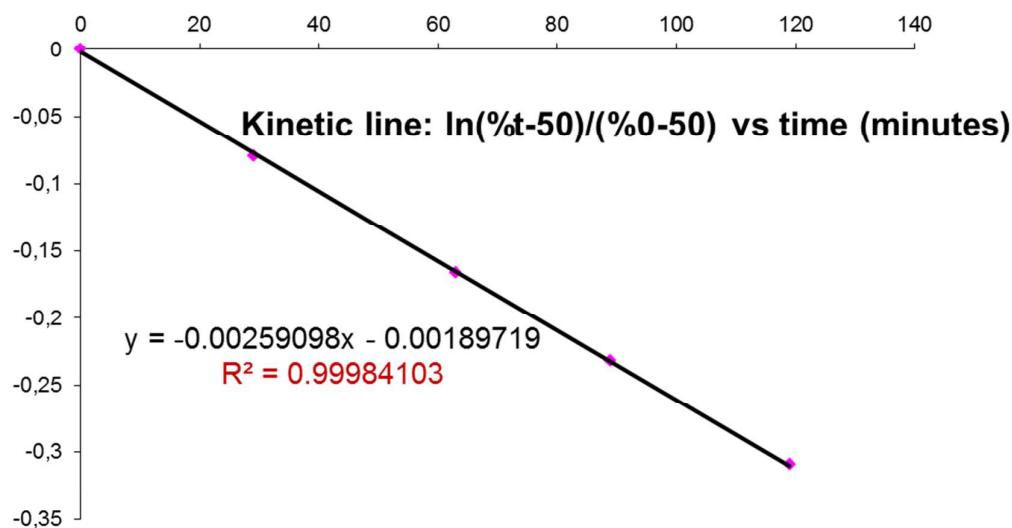
Enantiomerisation barrier for (**10i**)

About 3 mg of enantio-enriched (**10i**) were refluxed in 15 mL of ethanol. Samples of 10 μ L of this solution were injected on Lux-Cellulose-2 (heptane / isopropanol 95/5, 1 mL/min, UV detection at 220 nm) to monitor the percentage decrease of the second eluted enantiomer over time.

Solvent : Ethanol
 Temperature = 78.29 °C

Time (min)	% second eluted enantiomer (%t)	ln ((%t-50)/(%0-50))
0	93.95	0
29	90.61	-0.079038282
63	87.22	-0.166206373
89	84.85	-0.231999487
119	82.25	-0.309534581

$$\begin{aligned} k_{\text{racemisation}} &= 4.31829 \times 10^{-5} \text{ s}^{-1} \\ k_{\text{enantiomerisation}} &= 2.1591 \times 10^{-5} \text{ s}^{-1} \\ \Delta G^\ddagger_{\text{enantiomerisation}} &= \mathbf{117.94} \text{ kJ.mol}^{-1} \\ &\quad 28.22 \text{ kcal.mol}^{-1} \\ \text{half-life time } t_{1/2} &= 16051 \text{ seconds} \\ &\quad 267.52 \text{ minutes} \\ &\quad 4.46 \text{ hours} \\ &\quad 0.185780272 \text{ days} \end{aligned}$$



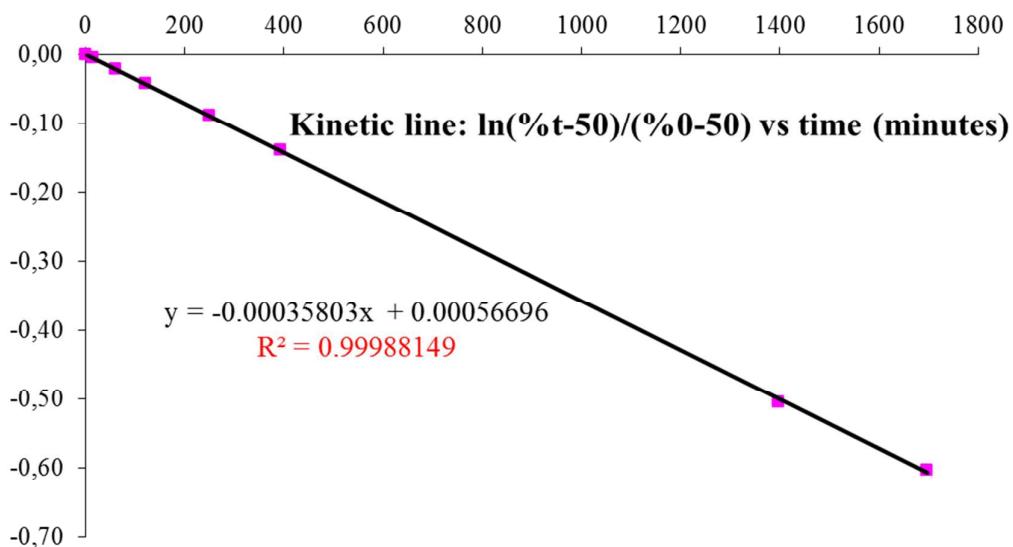
Enantiomerisation barrier for (10j)

About 3 mg of enantio-enriched (**10j**) were refluxed in 15 mL of ethanol. Samples of 10 μ L of this solution were injected on Lux-Cellulose-4 (heptane / ethanol 80/20, 1 mL/min, UV detection at 220 nm) to monitor the percentage decrease of the second eluted enantiomer over time.

Solvent : ethanol
 Temperature = 78.29 °C

Time (min)	% second eluted enantiomer (%t)	ln ((%t-50)/(%0-50))
0	97.77	0.0000
14	97.56	-0.0044
60	96.77	-0.0212
120	95.80	-0.0421
249	93.76	-0.0877
393	91.56	-0.1393
1397	78.83	-0.5050
1697	76.14	-0.6029

$$\begin{aligned} k_{\text{racemisation}} &= 5.96718 \times 10^{-6} \text{ s}^{-1} \\ k_{\text{enantiomerisation}} &= 2.9836 \times 10^{-6} \text{ s}^{-1} \\ \Delta G^\ddagger_{\text{enantiomerisation}} &= \mathbf{123.73} \text{ kJ.mol}^{-1} \\ &\quad 29.60 \text{ kcal.mol}^{-1} \\ \text{half-life time } t_{1/2} &= 116160 \text{ seconds} \\ &\quad 1936.00 \text{ minutes} \\ &\quad 32.27 \text{ hours} \\ &\quad 1.34 \text{ days} \end{aligned}$$



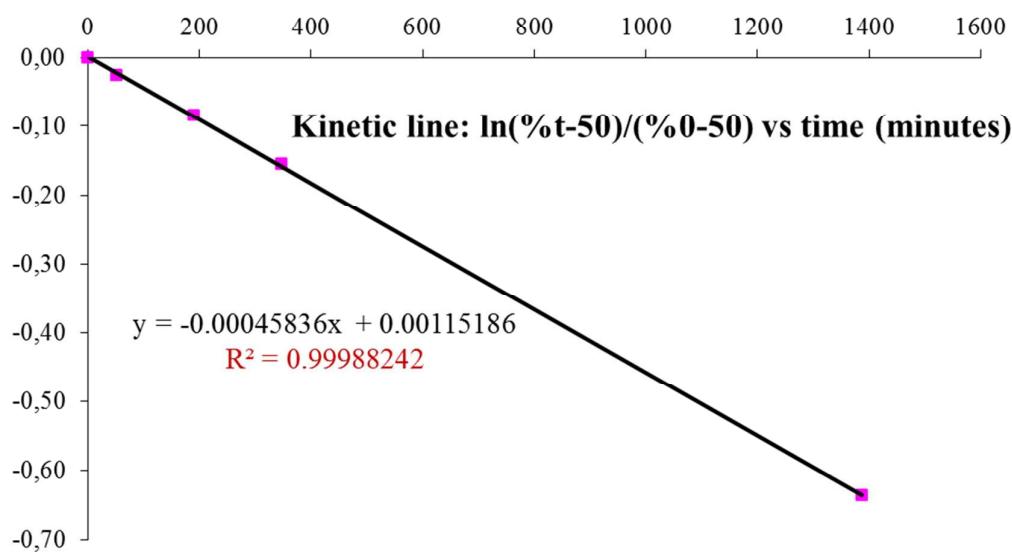
Enantiomerisation barrier for (**10k**)

About 3 mg of enantio-enriched (**10k**) were refluxed in 15 mL of ethanol. Samples of 10 μ L of this solution were injected on Lux-Cellulose-4 (heptane / ethanol 80/20, 1 mL/min, UV detection at 220 nm) to monitor the percentage decrease of the second eluted enantiomer over time.

Solvent : Ethanol
 Temperature = 78.29 °C

Time (min)	% second eluted enantiomer (%t)	ln ((%t-50)/(%0-50))
0	96.15	0.0000
52	94.96	-0.0261
190	92.43	-0.0840
348	89.54	-0.1546
1387	74.44	-0.6357

$$\begin{aligned} k_{\text{racemisation}} &= 7.63939 \times 10^{-6} \text{ s}^{-1} \\ k_{\text{enantiomerisation}} &= 3.8197 \times 10^{-6} \text{ s}^{-1} \\ \Delta G^\ddagger_{\text{enantiomerisation}} &= \mathbf{123.01} \text{ kJ.mol}^{-1} \\ &\quad 29.43 \text{ kcal.mol}^{-1} \\ \text{half-life time } t_{1/2} &= 90733 \text{ seconds} \\ &\quad 1512.22 \text{ minutes} \\ &\quad 25.20 \text{ hours} \\ &\quad 1.05 \text{ days} \end{aligned}$$



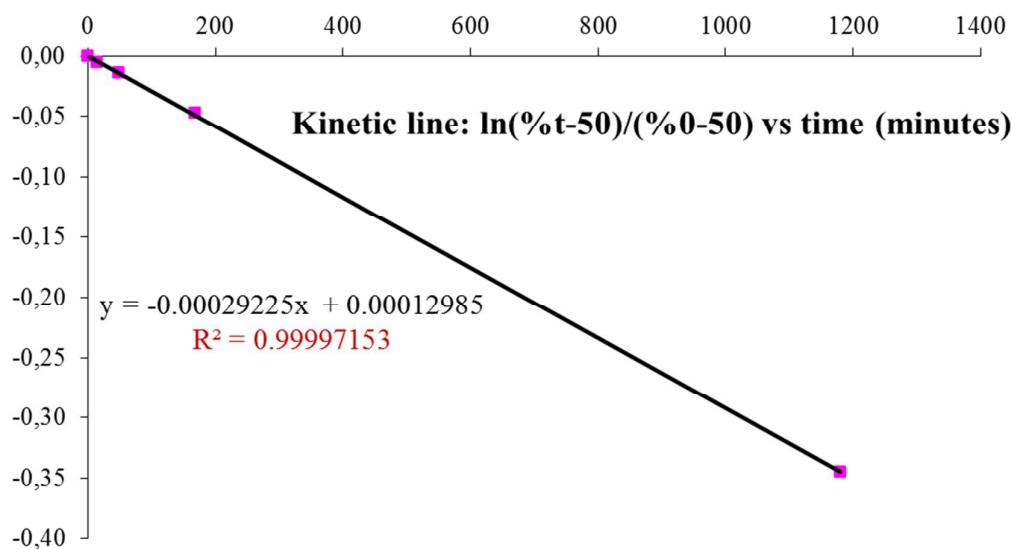
Enantiomerisation barrier for (10I)

About 3 mg of enantio-enriched (**10I**) were refluxed in 15 mL of ethanol. Samples of 10 μ L of this solution were injected on Lux-Cellulose-3 (heptane / ethanol 80/20, 1 mL/min, UV detection at 220 nm) to monitor the percentage decrease of the second eluted enantiomer over time.

Solvent : Ethanol
 Temperature = 78.29 °C

Time (min)	% second eluted enantiomer (%t)	$\ln((\%t-50)/(\%0-50))$
0	97.72	0.0000
15	97.48	-0.0050
48	97.05	-0.0141
168	95.50	-0.0476
1180	83.80	-0.3449

$$\begin{aligned} k_{\text{racemisation}} &= 4.87077 \times 10^{-6} \text{ s}^{-1} \\ k_{\text{enantiomerisation}} &= 2.4354 \times 10^{-6} \text{ s}^{-1} \\ \Delta G^\ddagger_{\text{enantiomerisation}} &= \mathbf{124.33} \text{ kJ.mol}^{-1} \\ &\quad 29.74 \text{ kcal.mol}^{-1} \\ \text{half-life time } t_{1/2} &= 142308 \text{ seconds} \\ &\quad 2371.79 \text{ minutes} \\ &\quad 39.53 \text{ hours} \\ &\quad 1.65 \text{ days} \end{aligned}$$



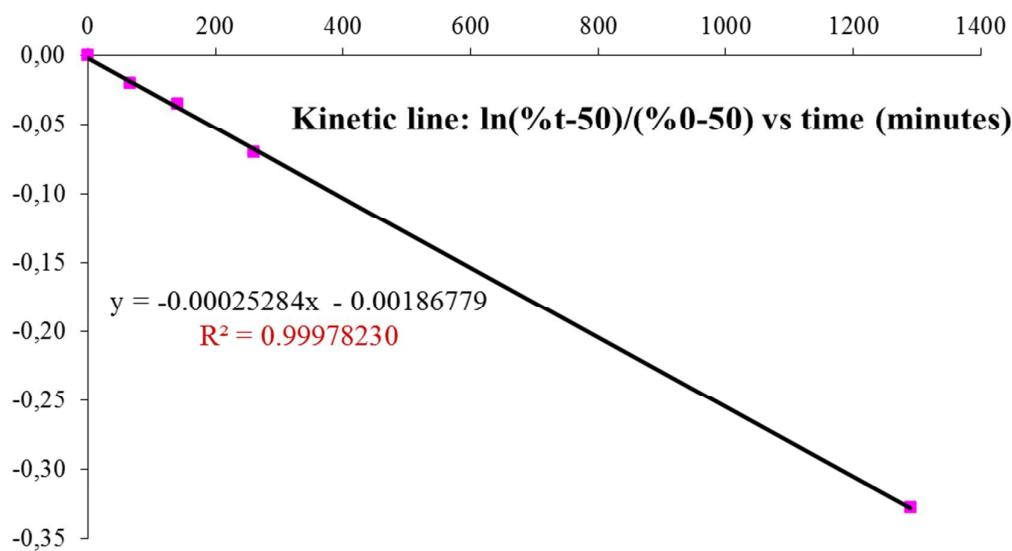
Enantiomerisation barrier for (10m)

About 3 mg of enantio-enriched (**10m**) were refluxed in 15 mL of ethanol. Samples of 10 μ L of this solution were injected on Lux-Cellulose-4 (heptane / ethanol 80/20, 1 mL/min, UV detection at 220 nm) to monitor the percentage decrease of the second eluted enantiomer over time.

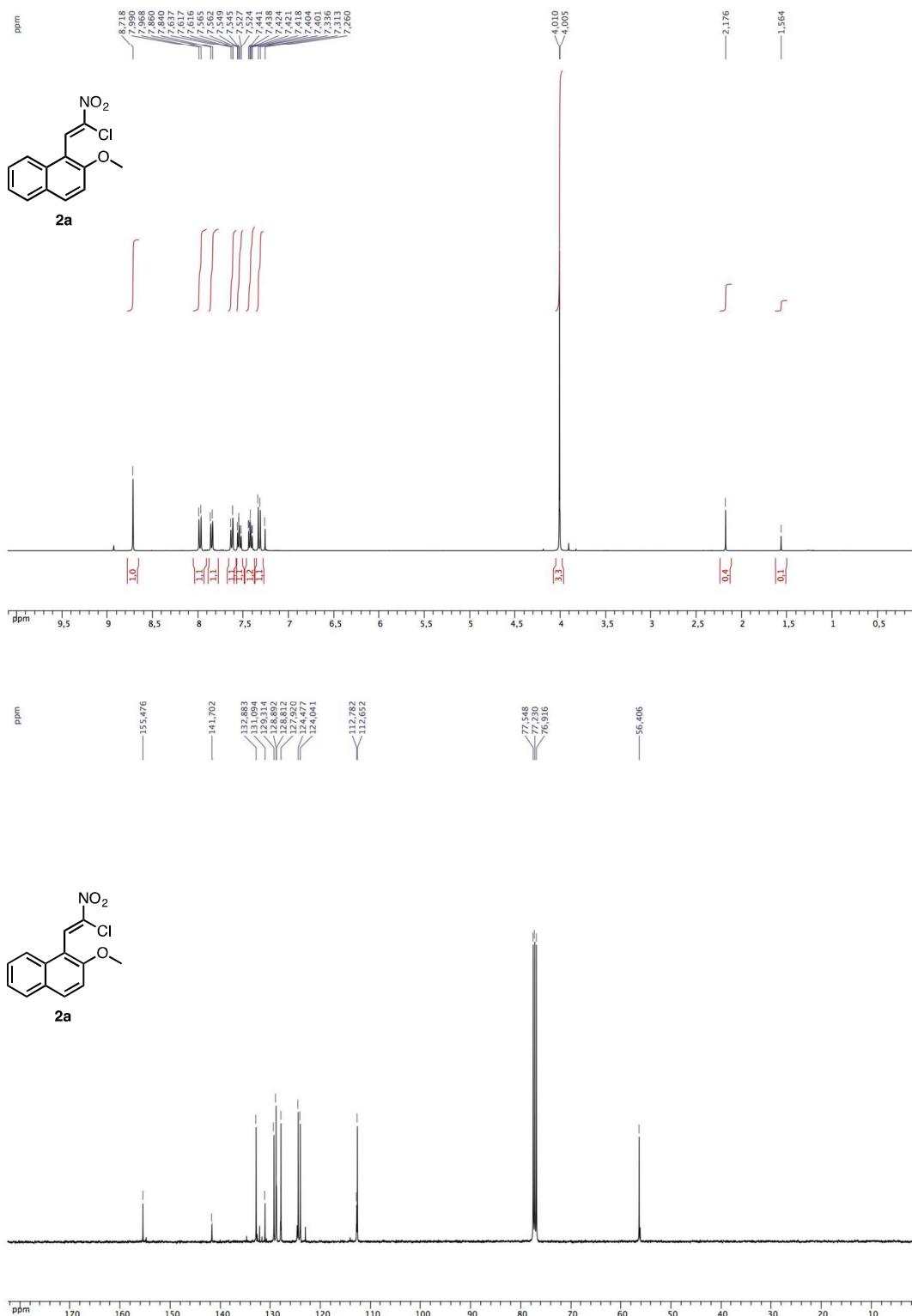
Solvent : Ethanol
 Temperature = 78.29 °C

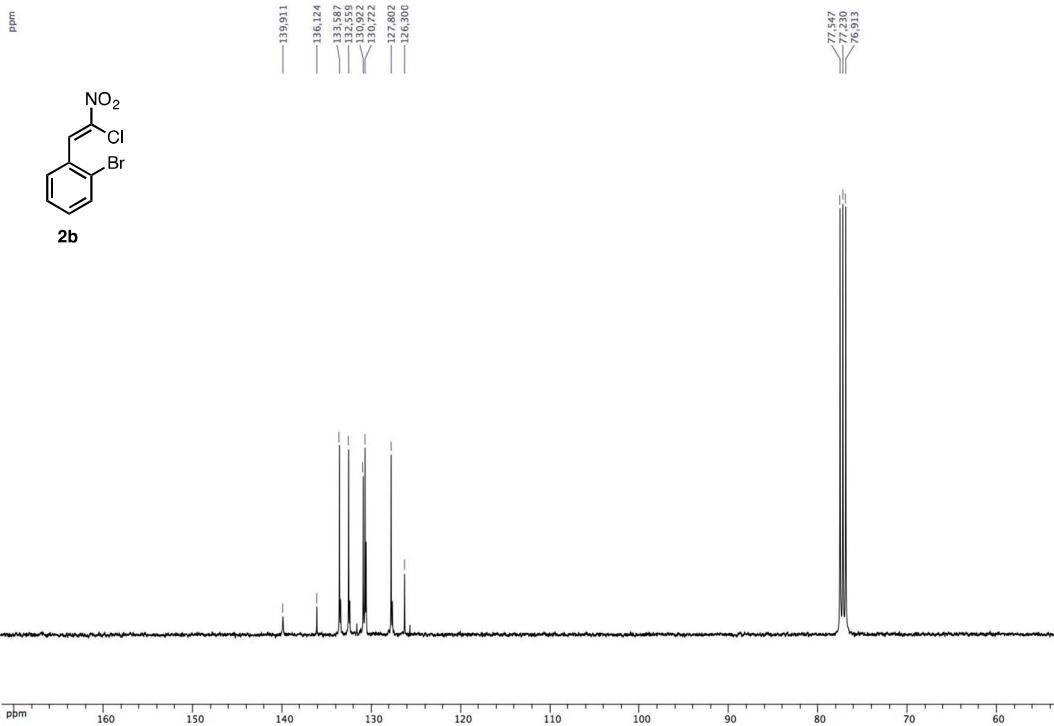
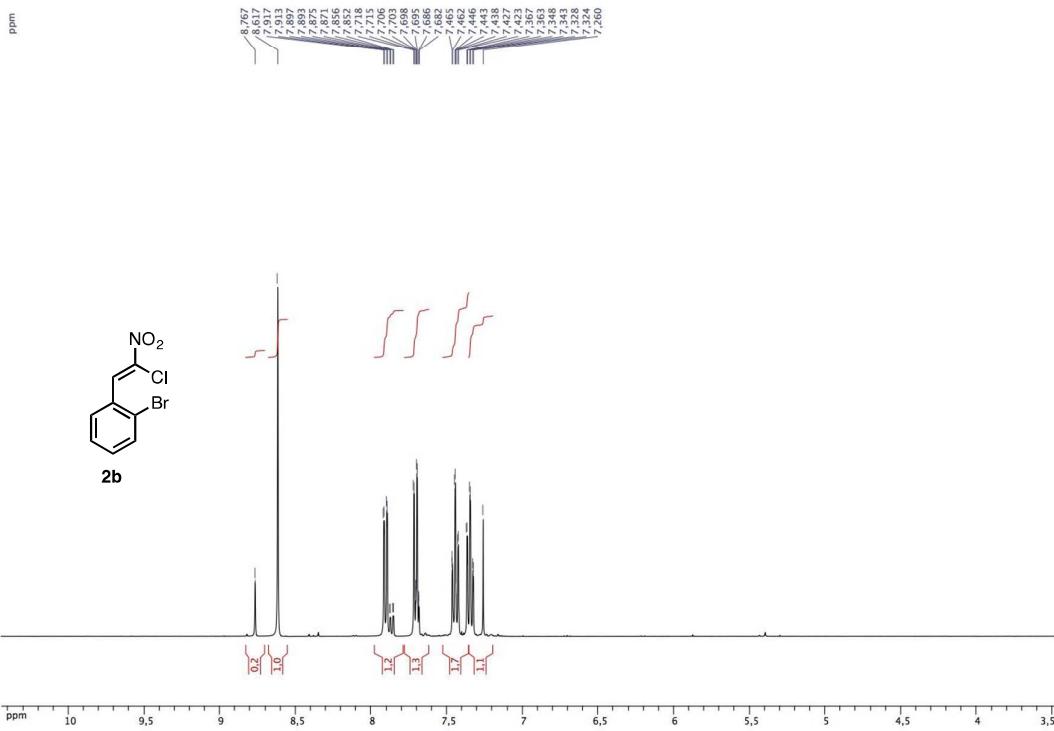
Time (min)	% second eluted enantiomer (%t)	ln ((%t-50)/(%0-50))
0	97.93	0.0000
66	96.98	-0.0200
140	96.26	-0.0355
260	94.68	-0.0702
1290	84.54	-0.3276

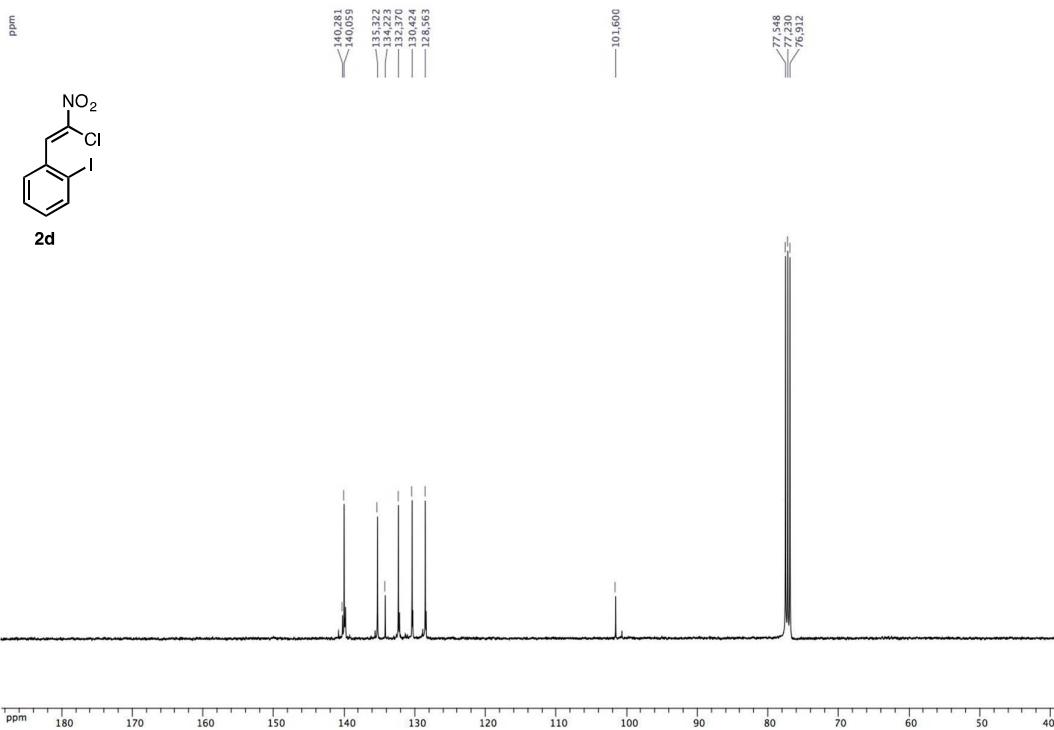
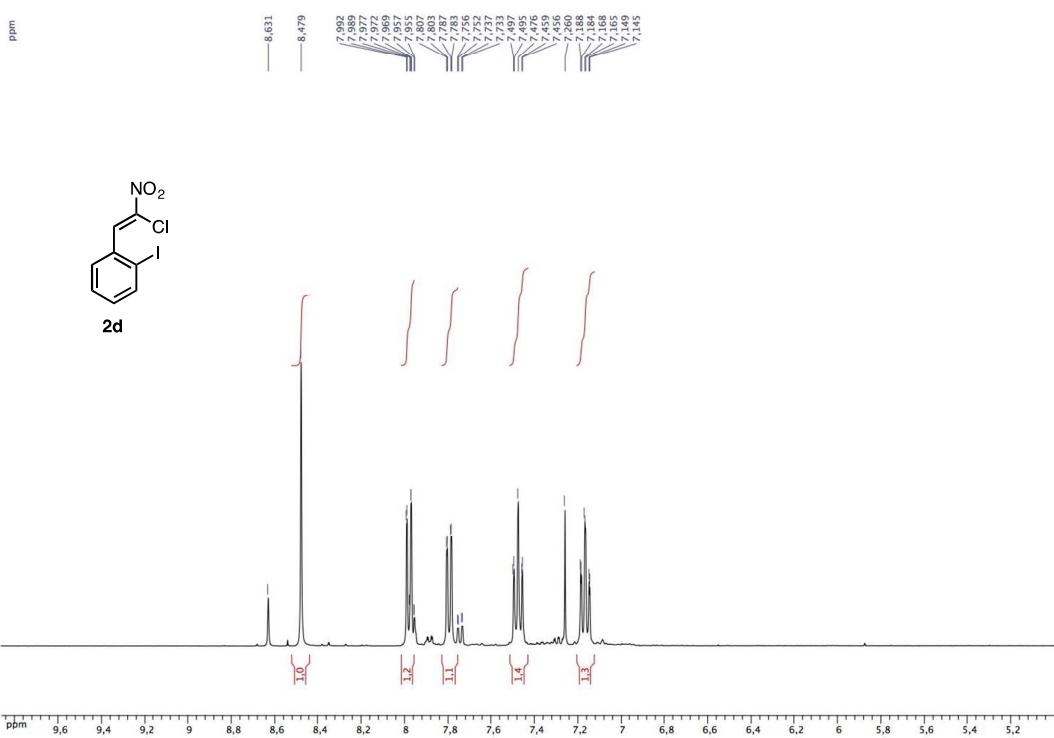
$$\begin{aligned}
 k_{\text{racemisation}} &= 4.21397 \times 10^{-6} \text{ s}^{-1} \\
 k_{\text{enantiomerisation}} &= 2.1070 \times 10^{-6} \text{ s}^{-1} \\
 \Delta G^\ddagger_{\text{enantiomerisation}} &= \mathbf{124.75} \text{ kJ.mol}^{-1} \\
 &\quad 29.84 \text{ kcal.mol}^{-1} \\
 \text{half-life time } t_{1/2} &= 164488 \text{ seconds} \\
 &\quad 2741.47 \text{ minutes} \\
 &\quad 45.69 \text{ hours} \\
 &\quad 1.90 \text{ days}
 \end{aligned}$$

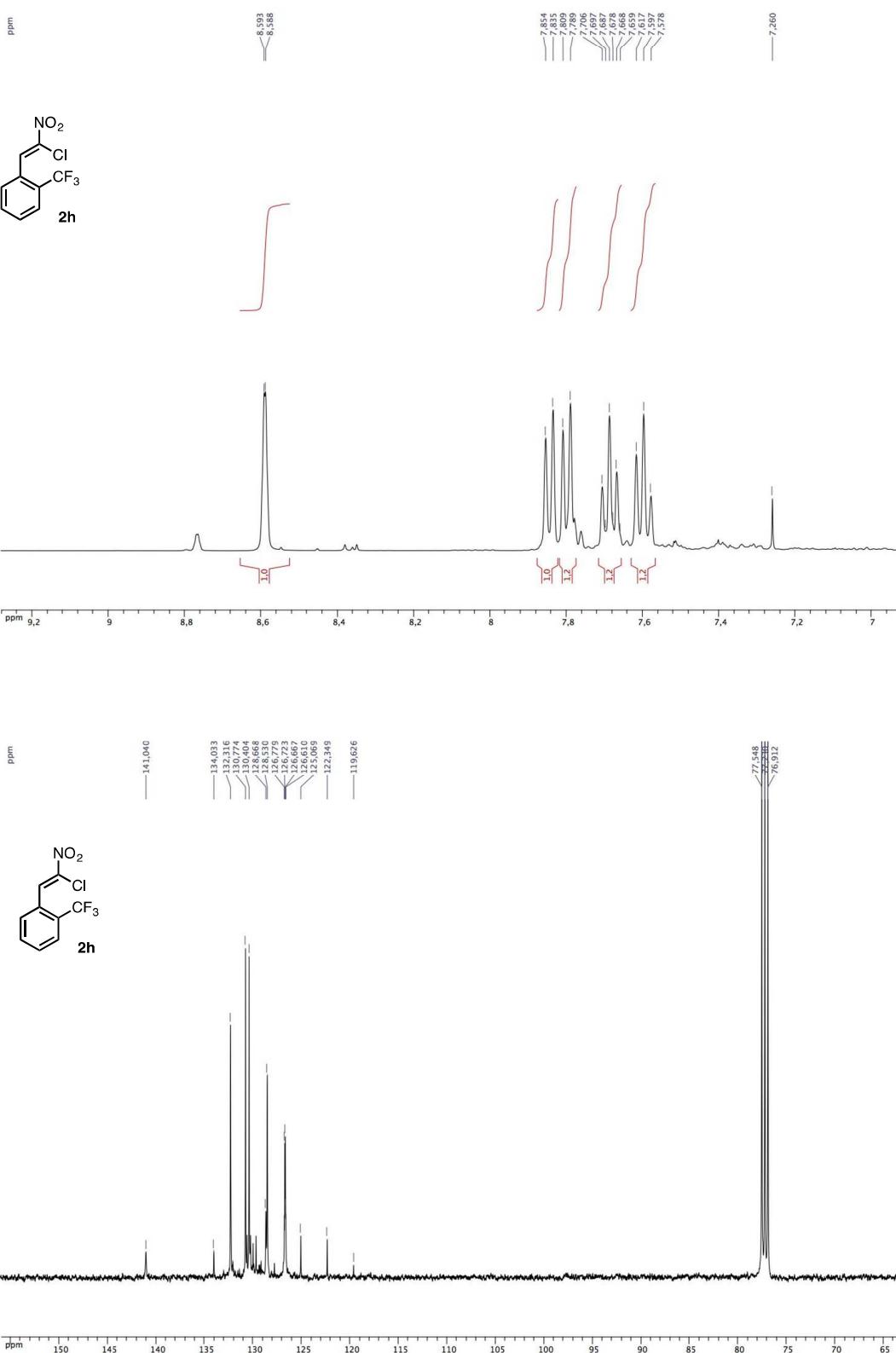


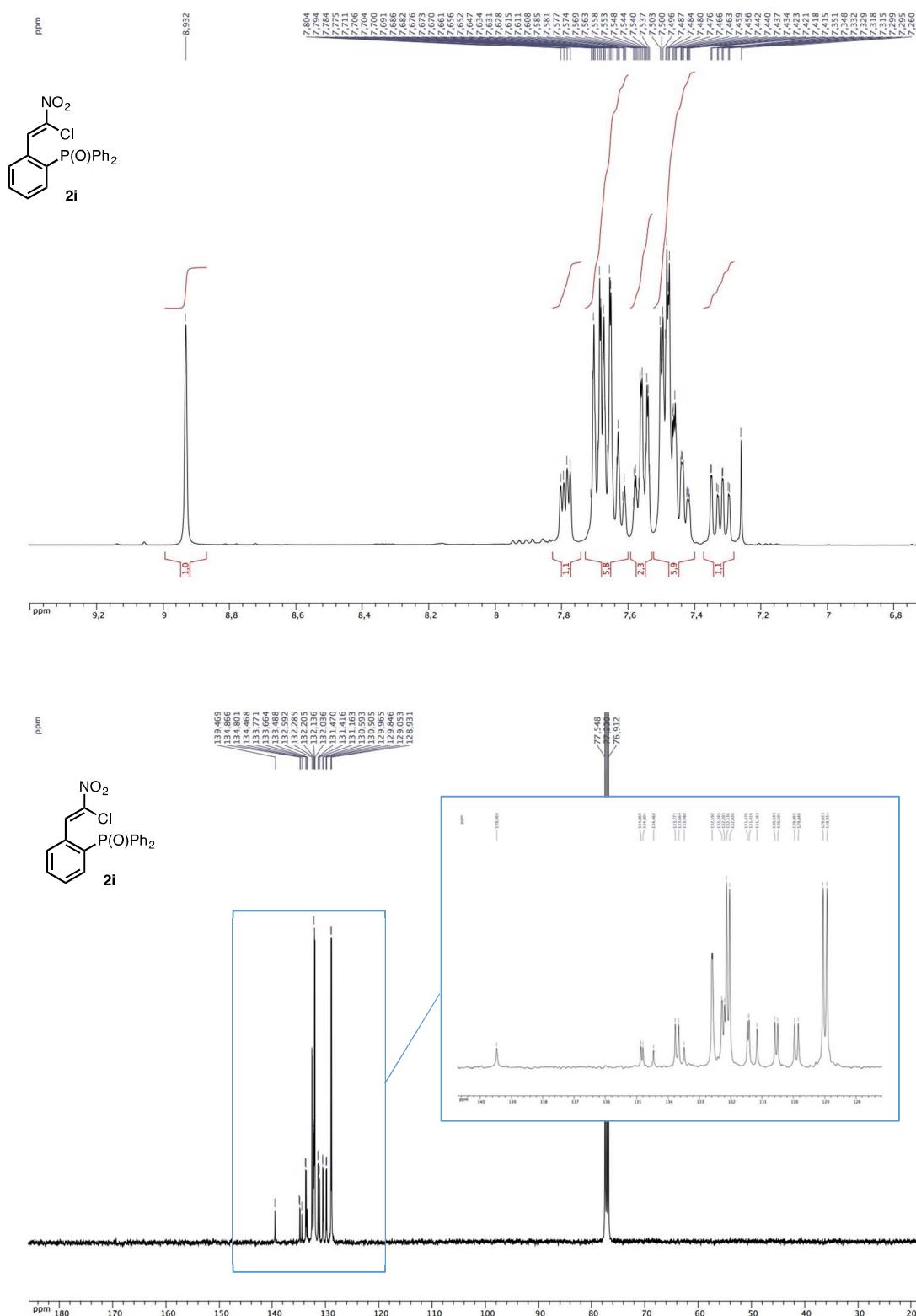
7. ^1H and ^{13}C NMR spectra

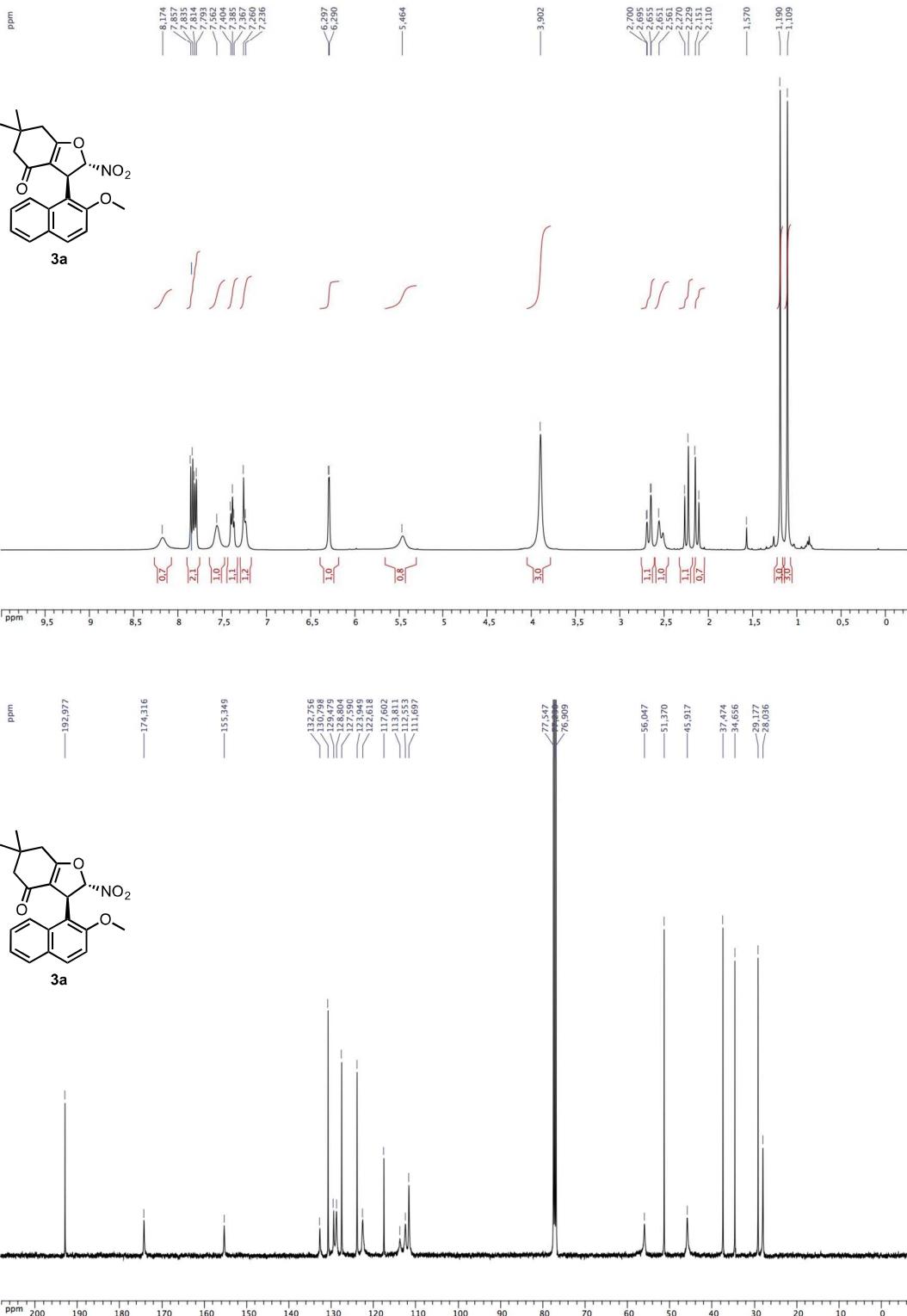


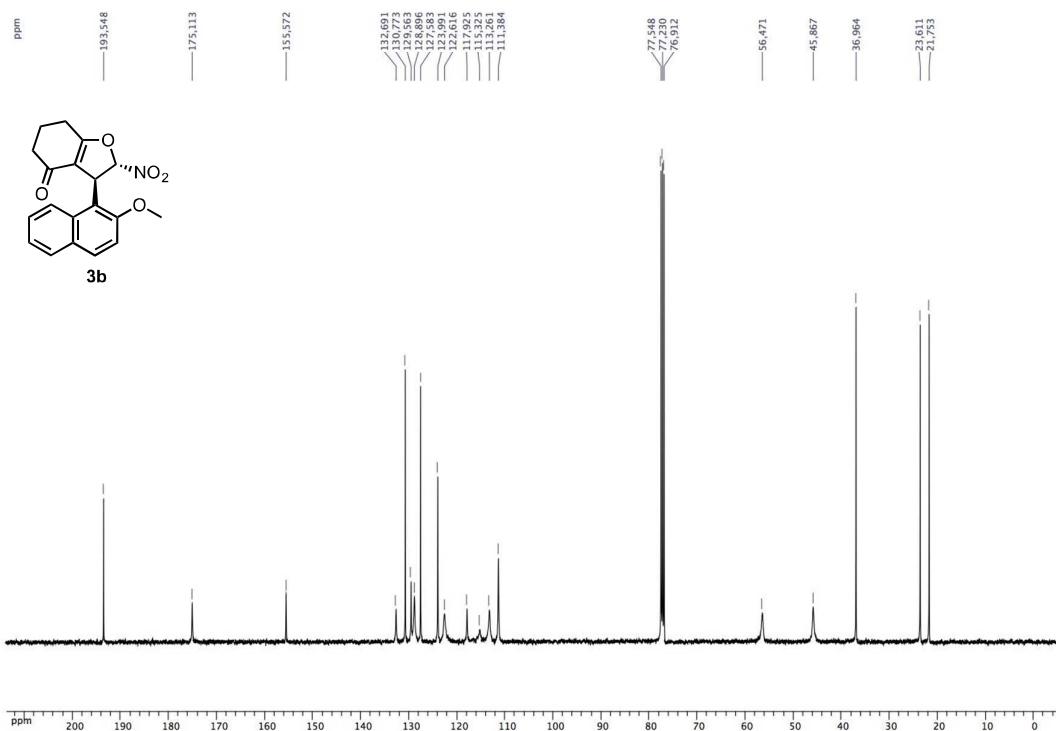
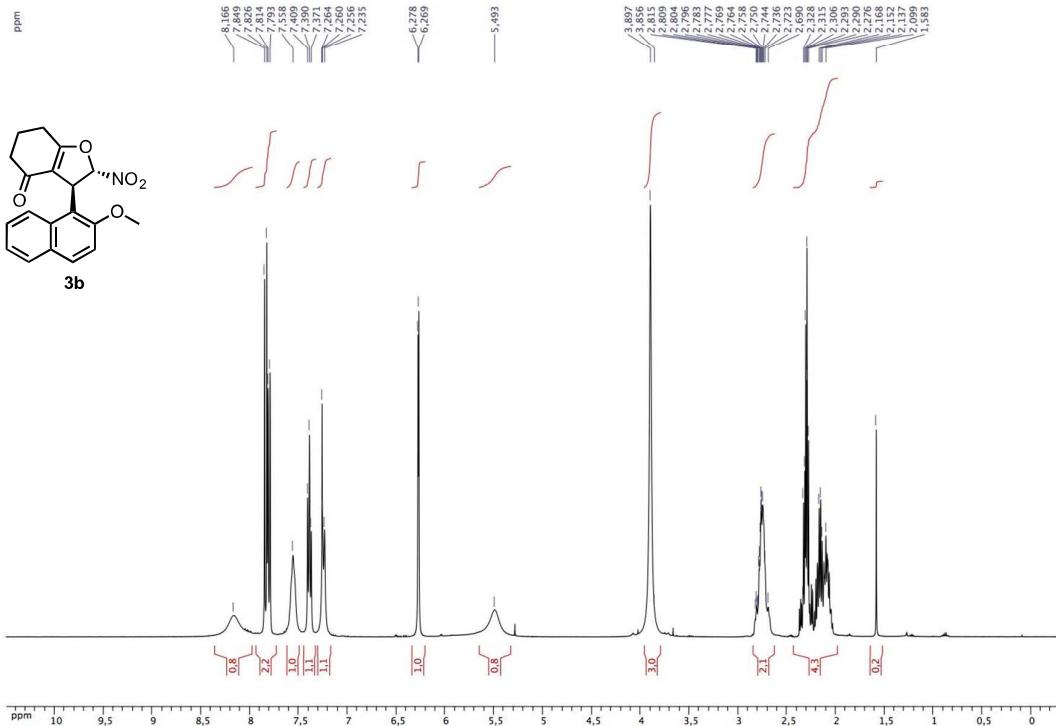


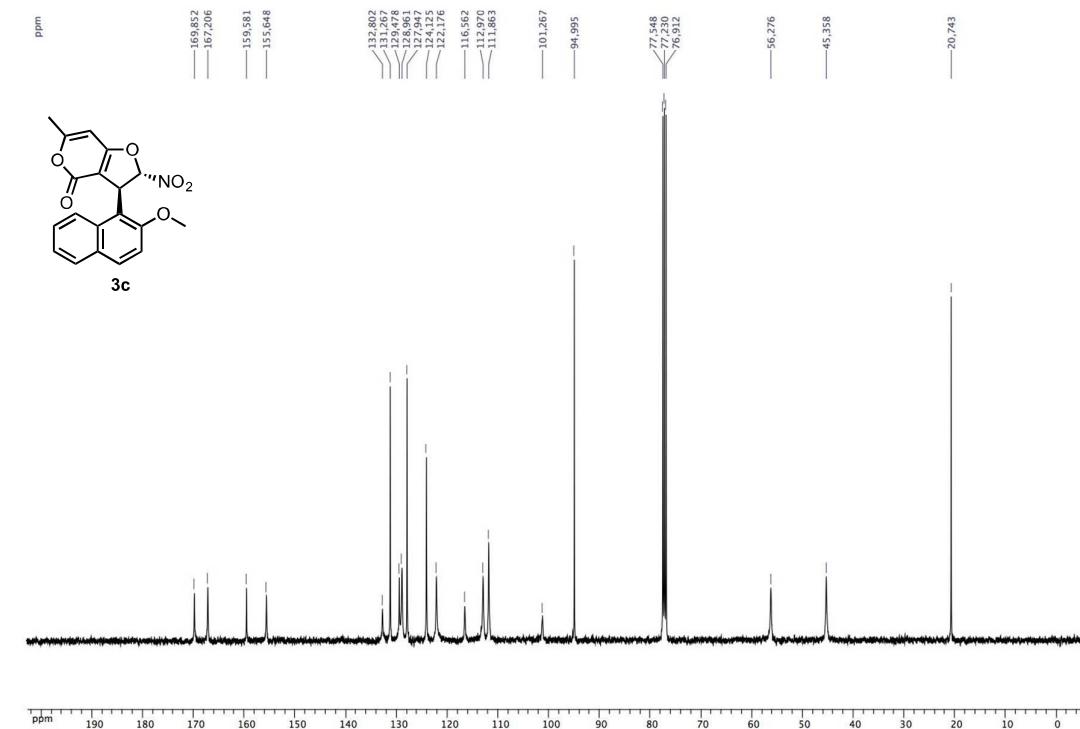
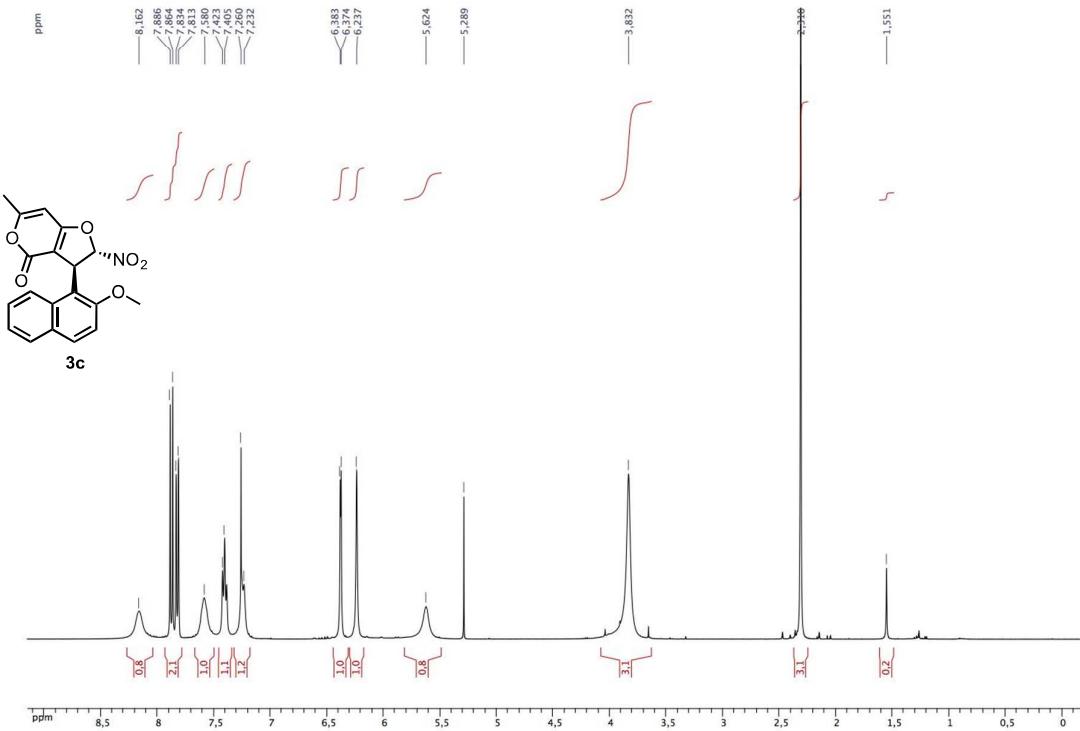


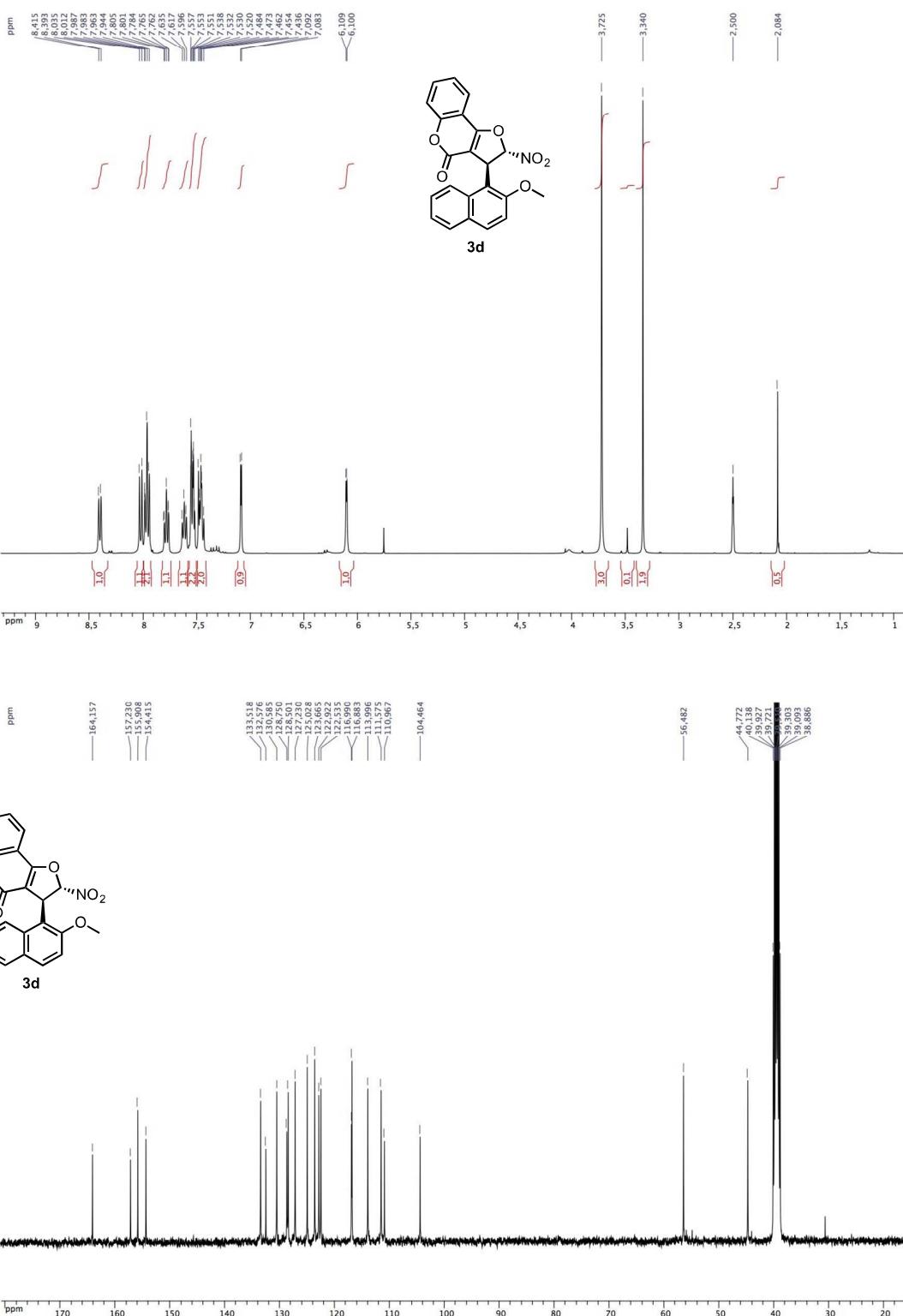


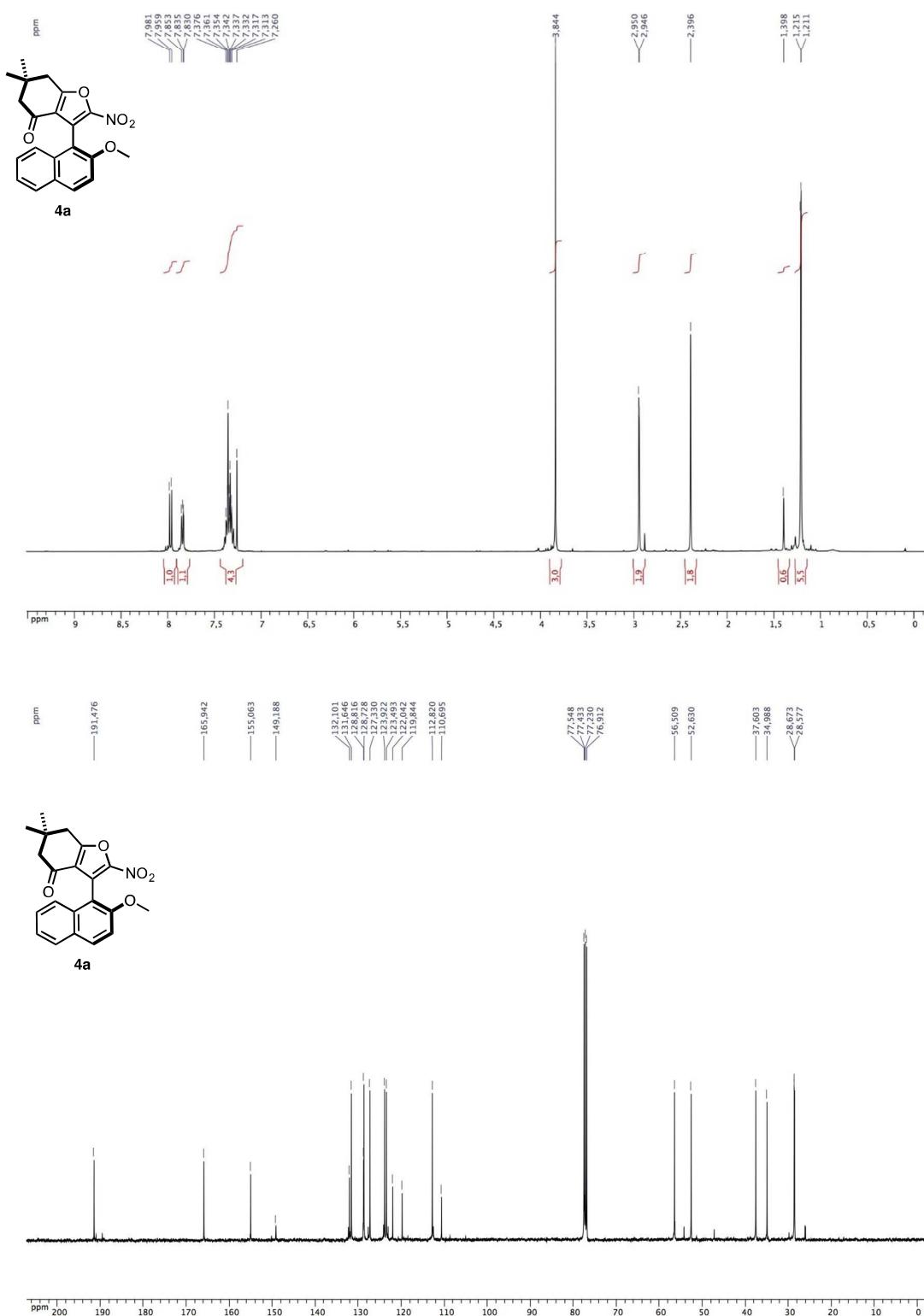


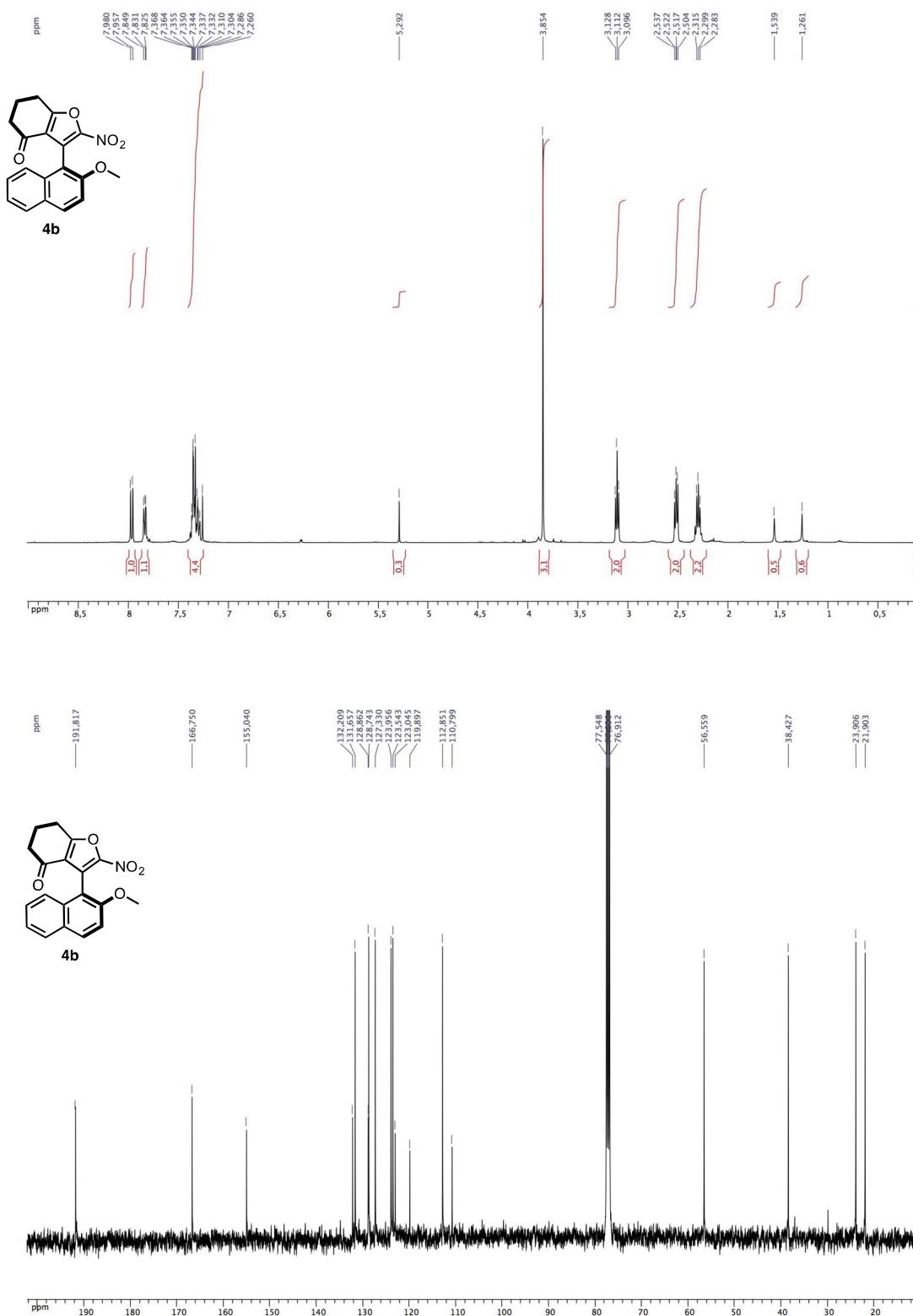


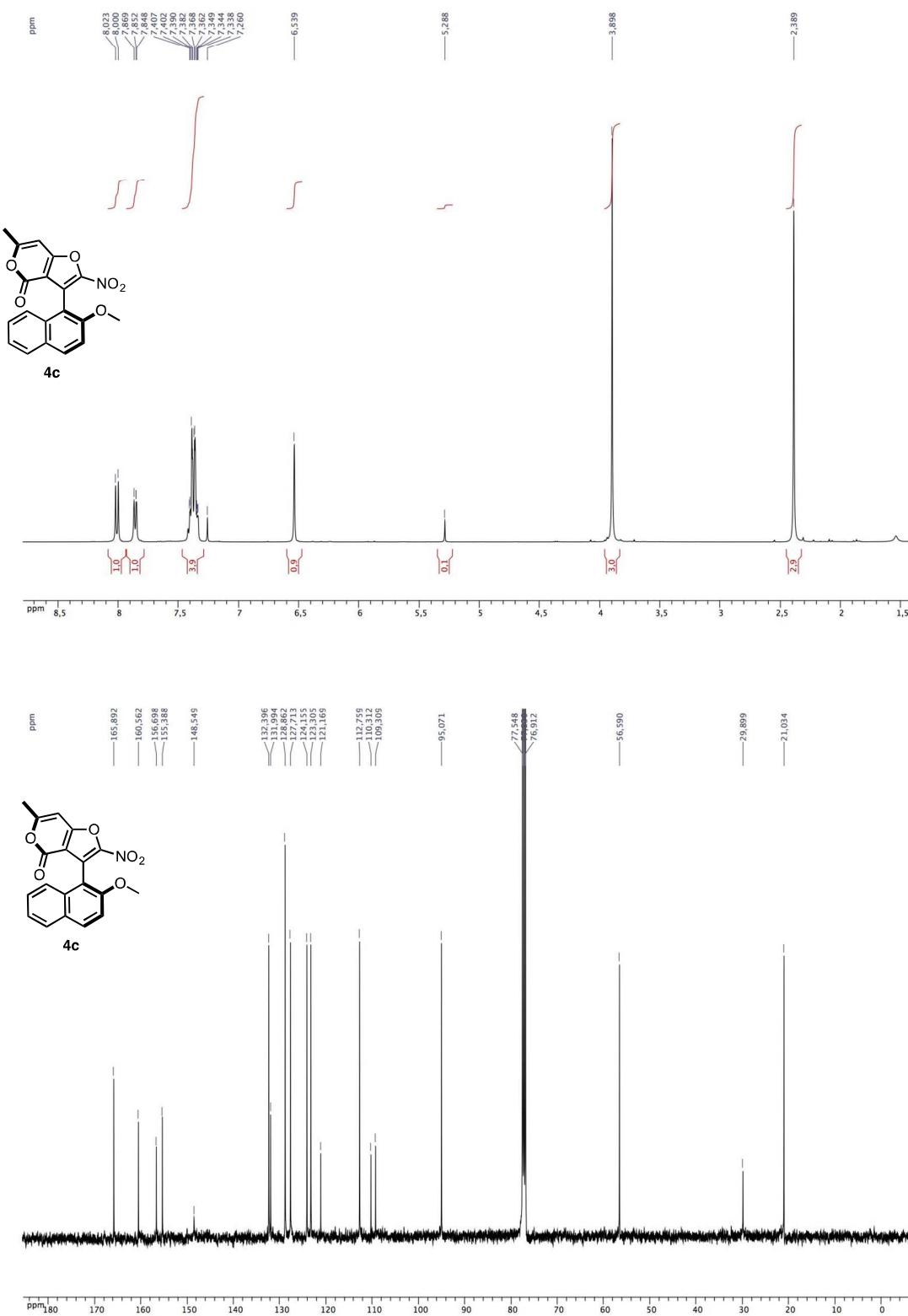


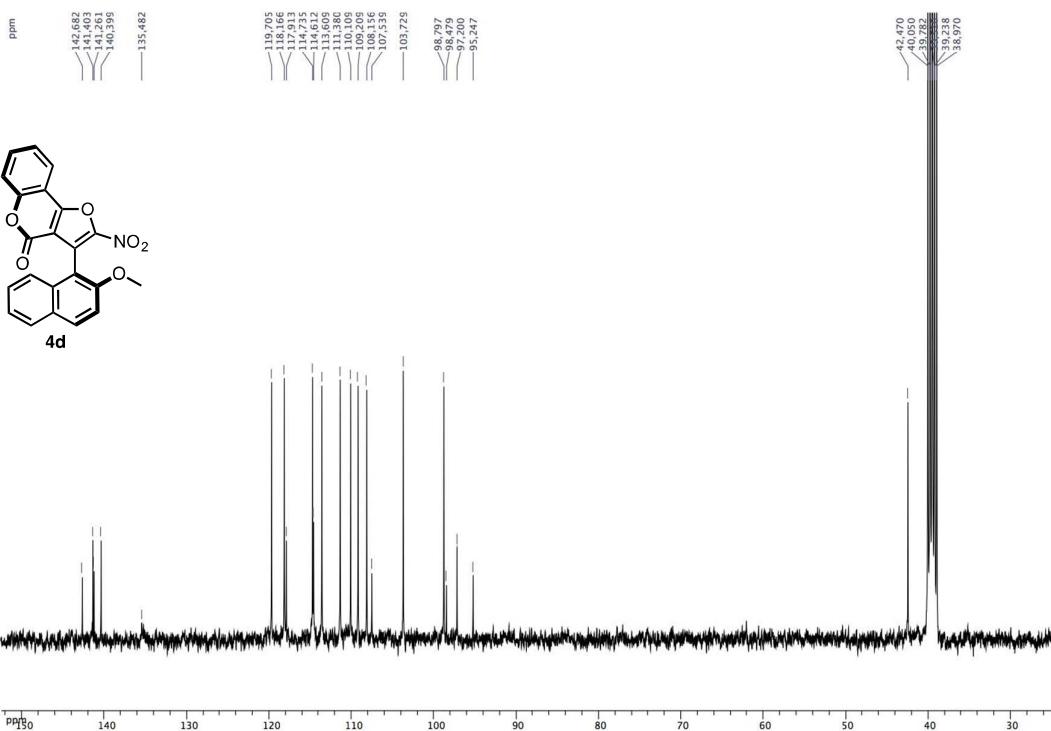
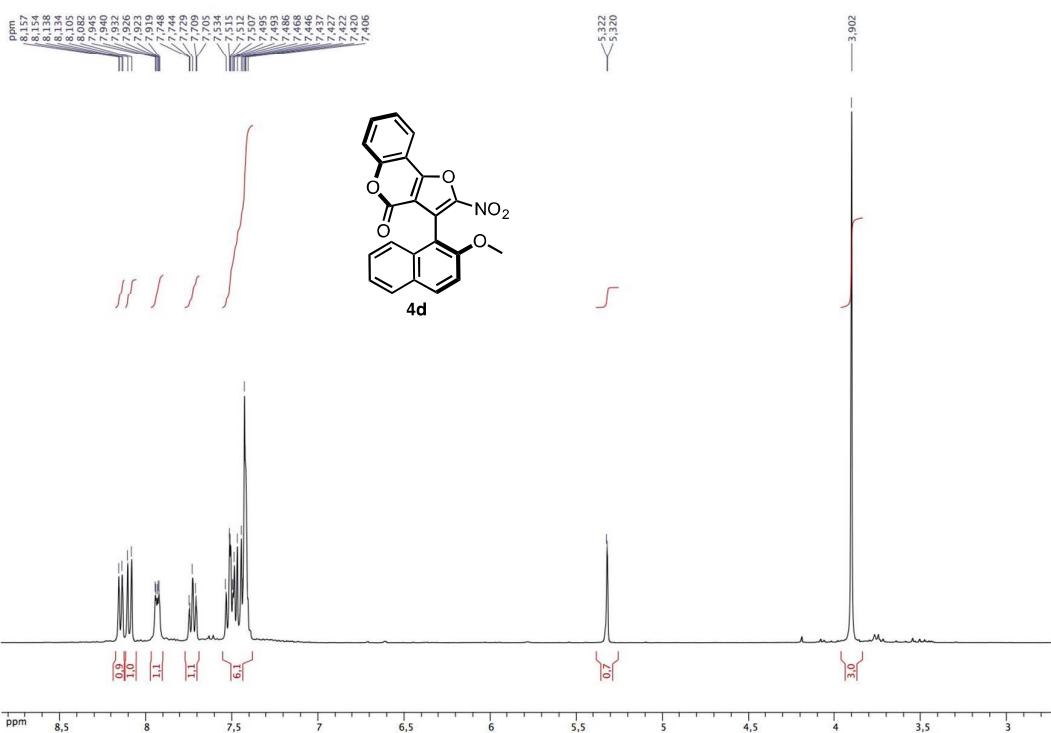


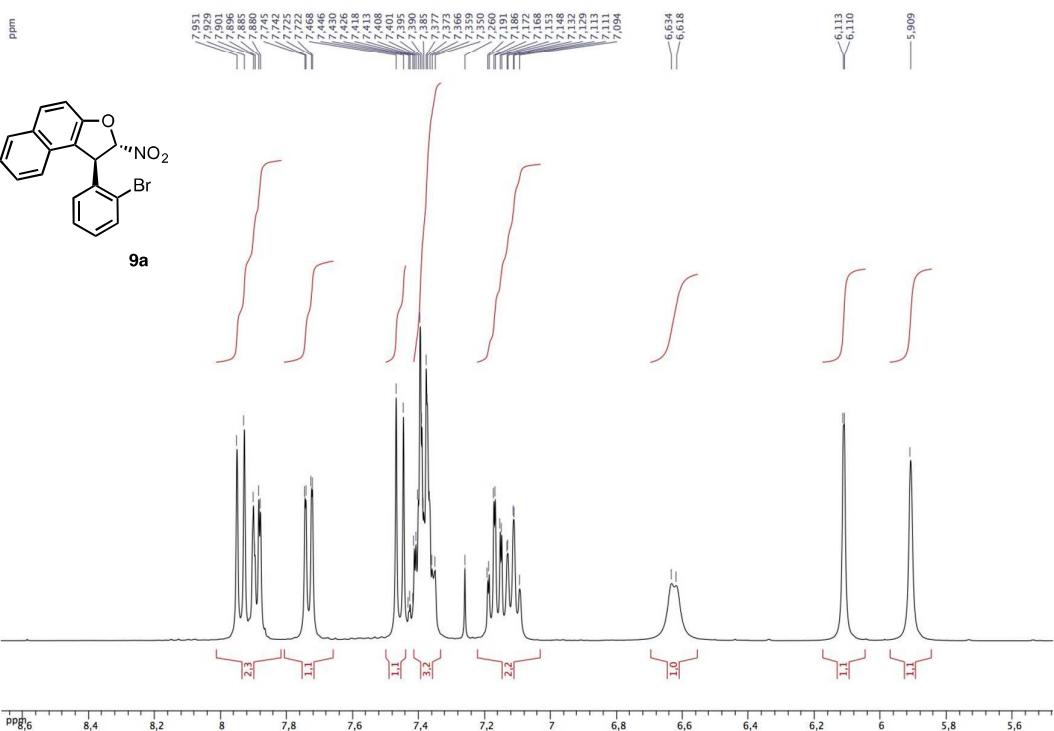
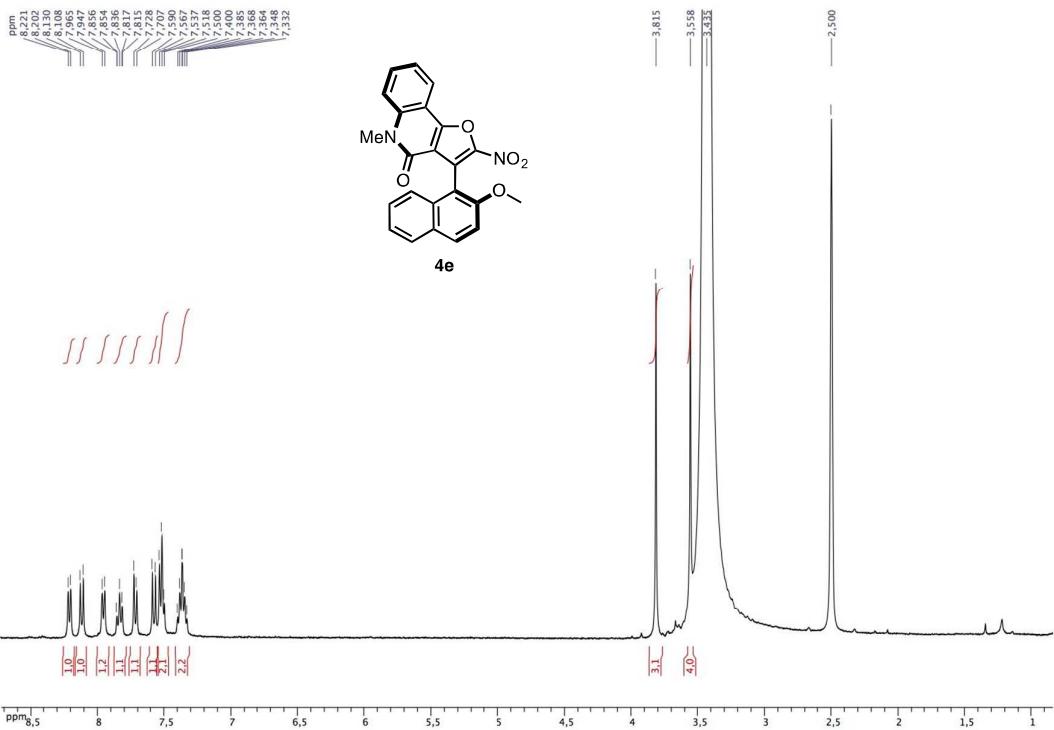


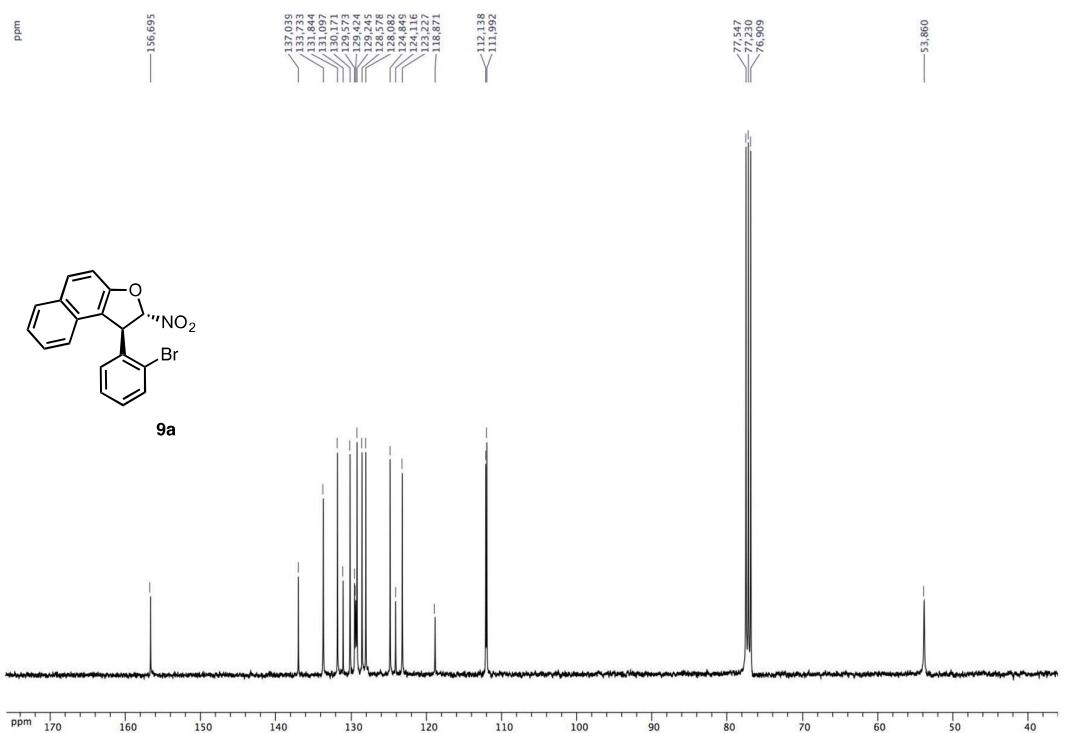


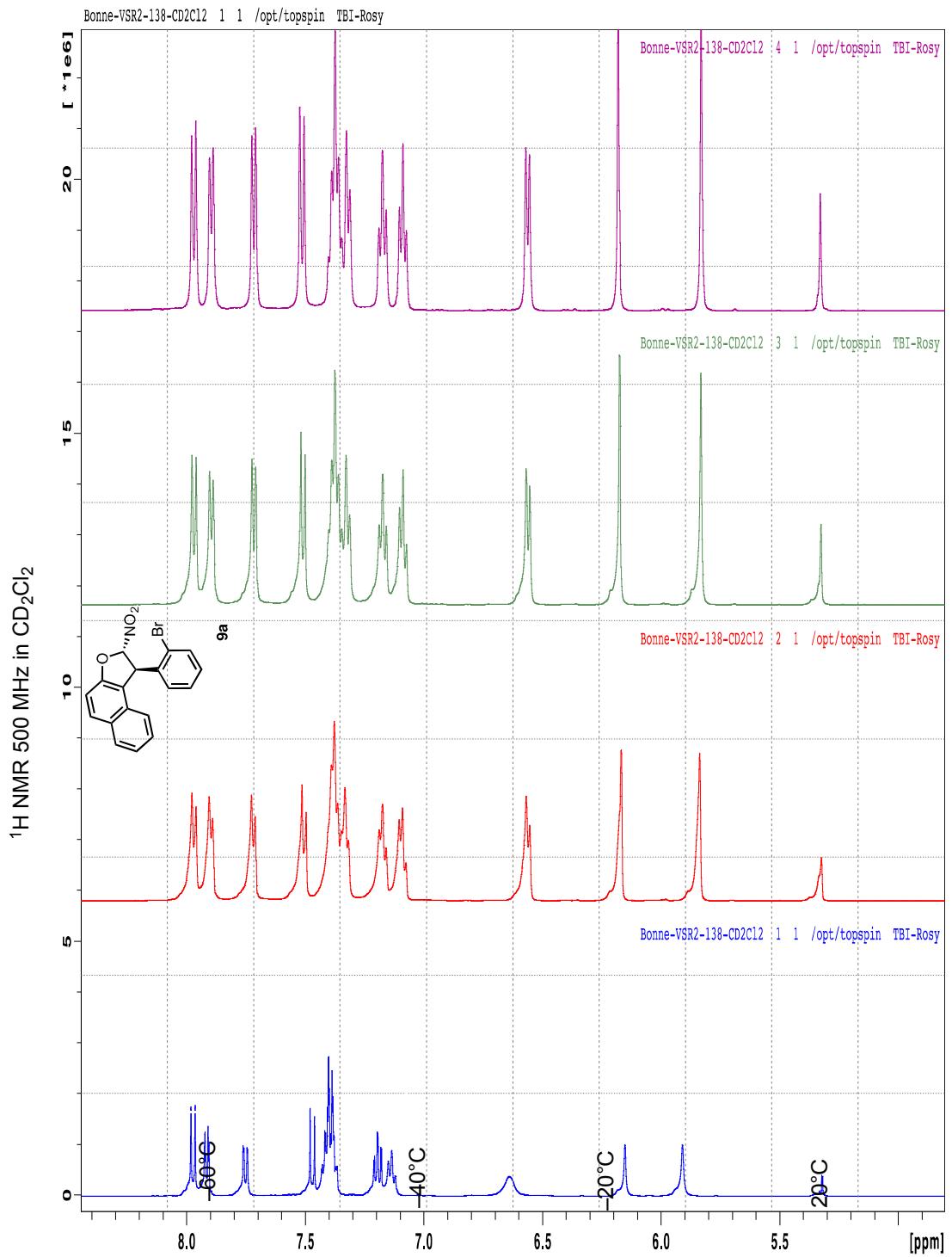


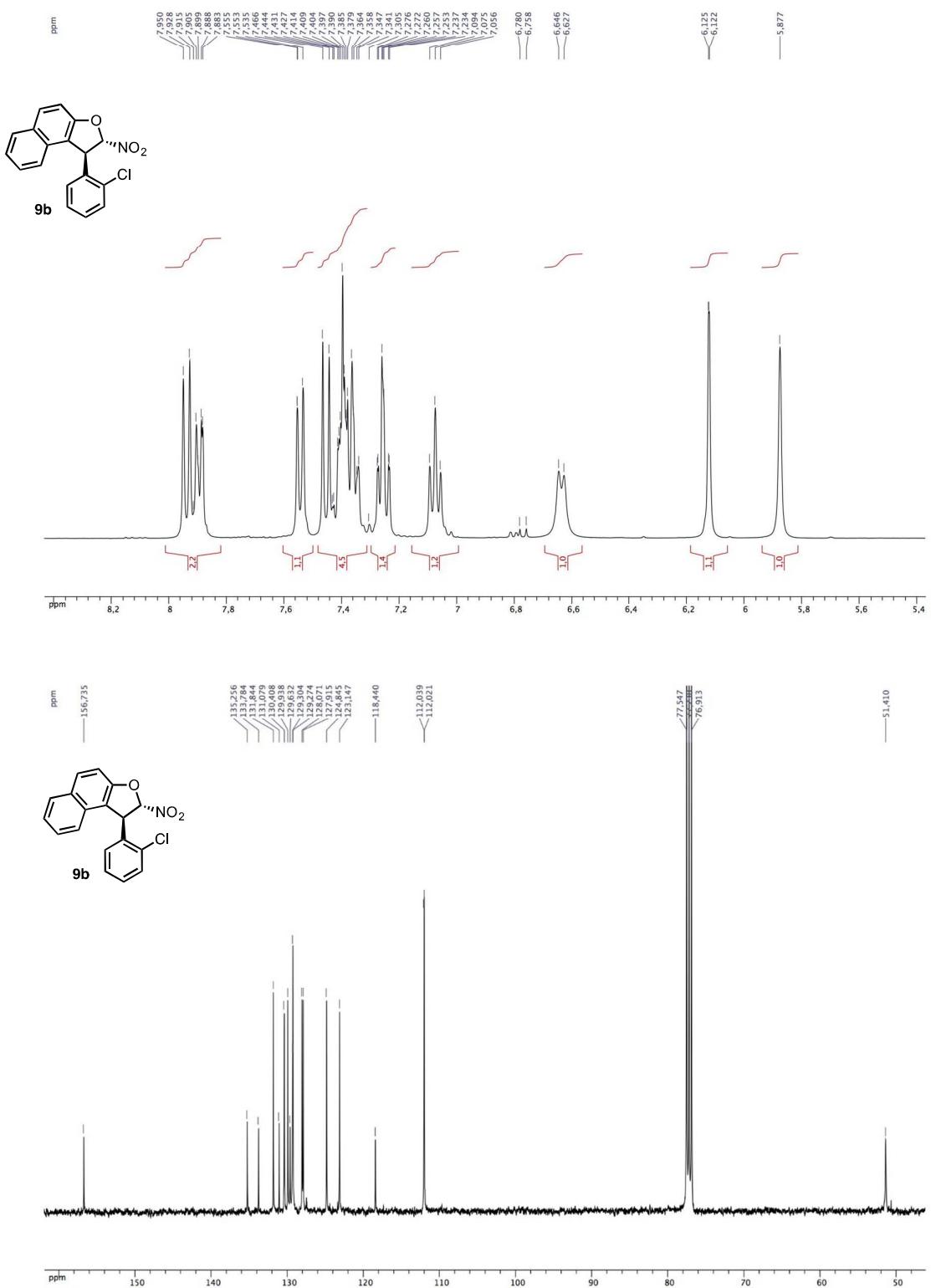


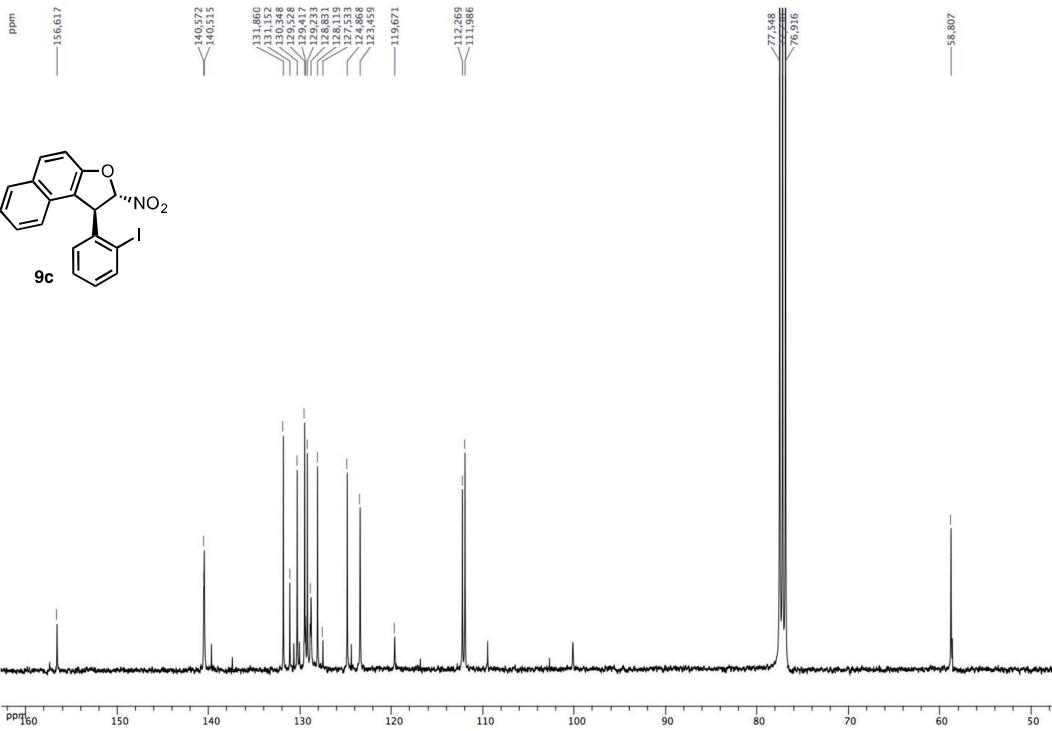
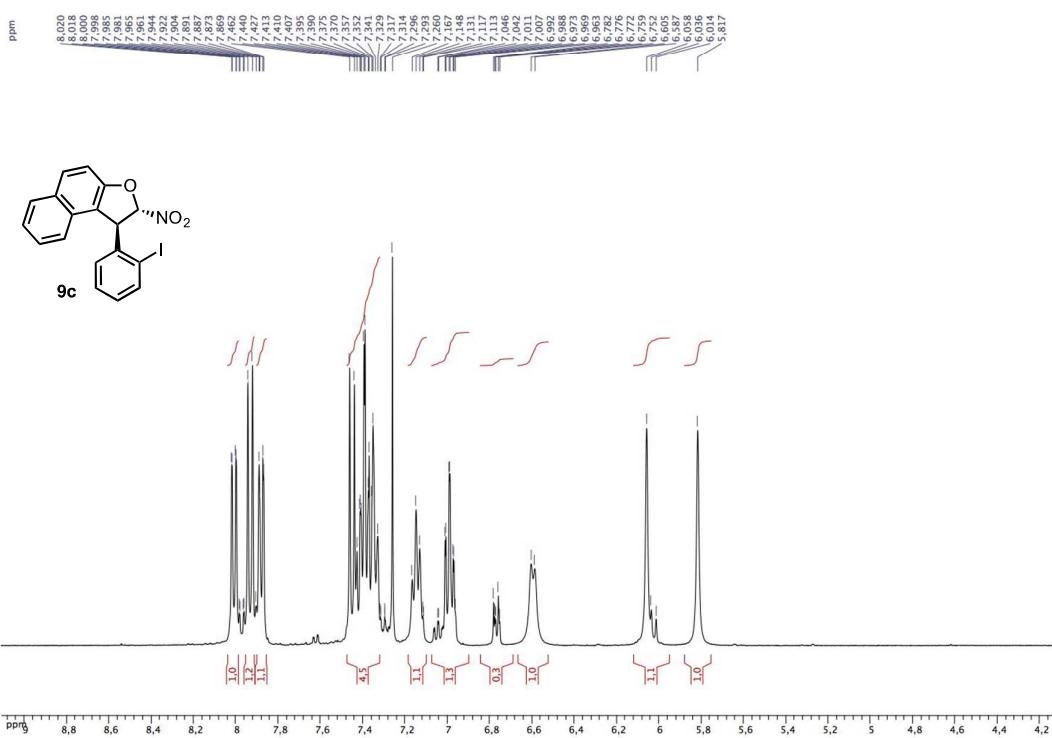


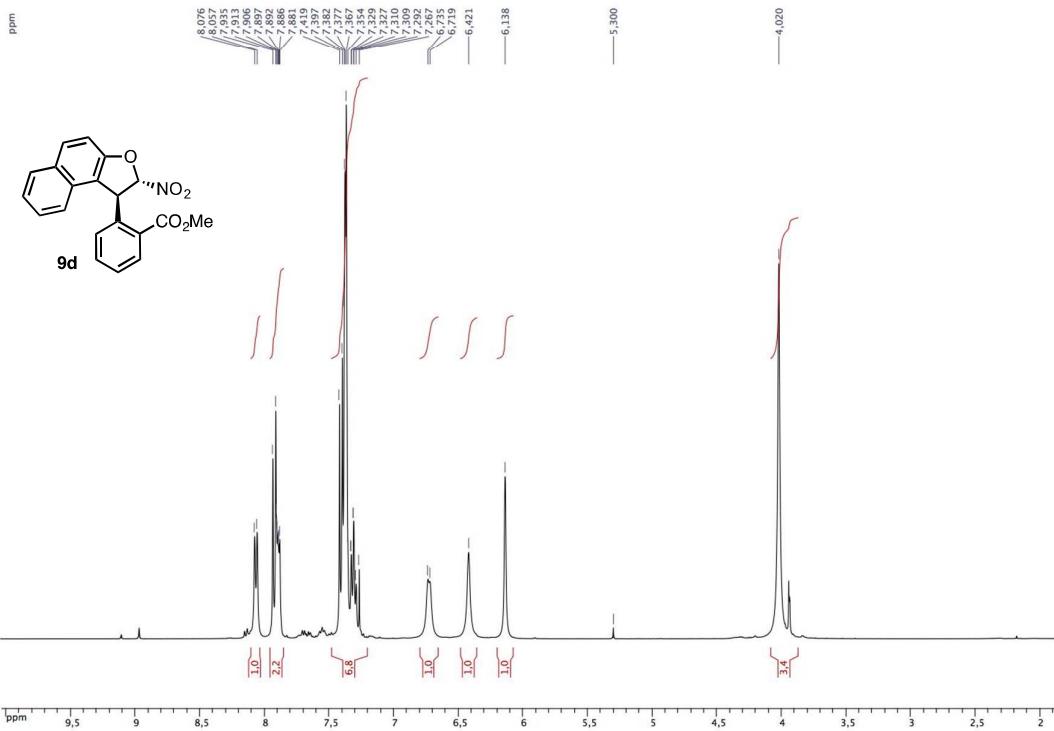


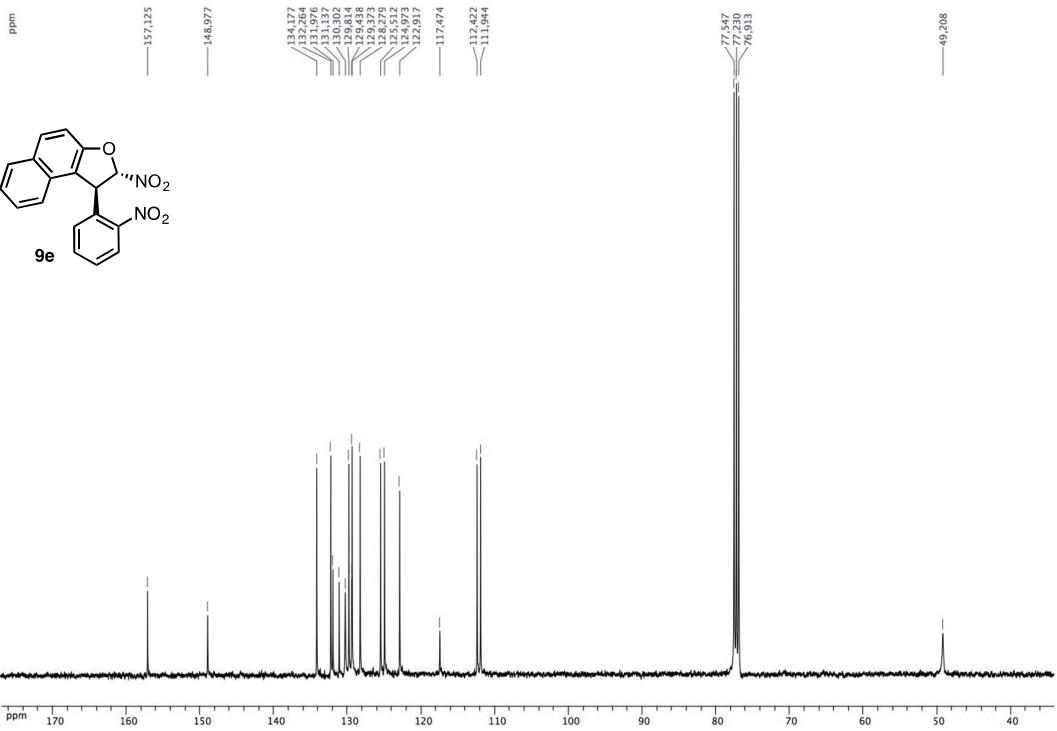
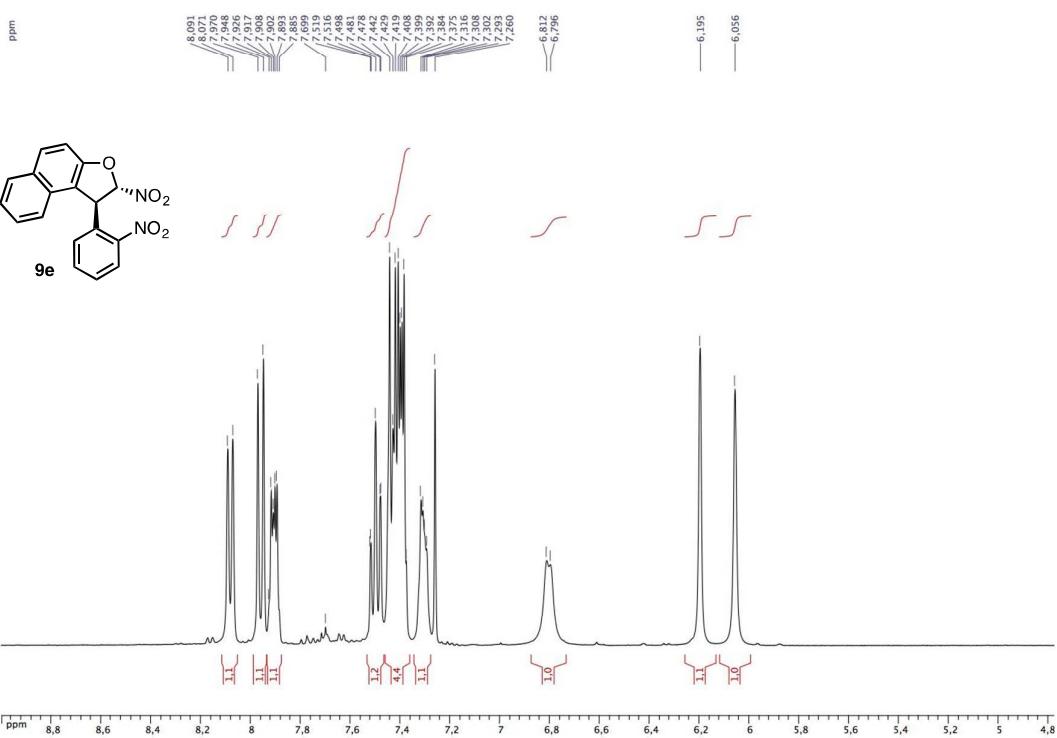


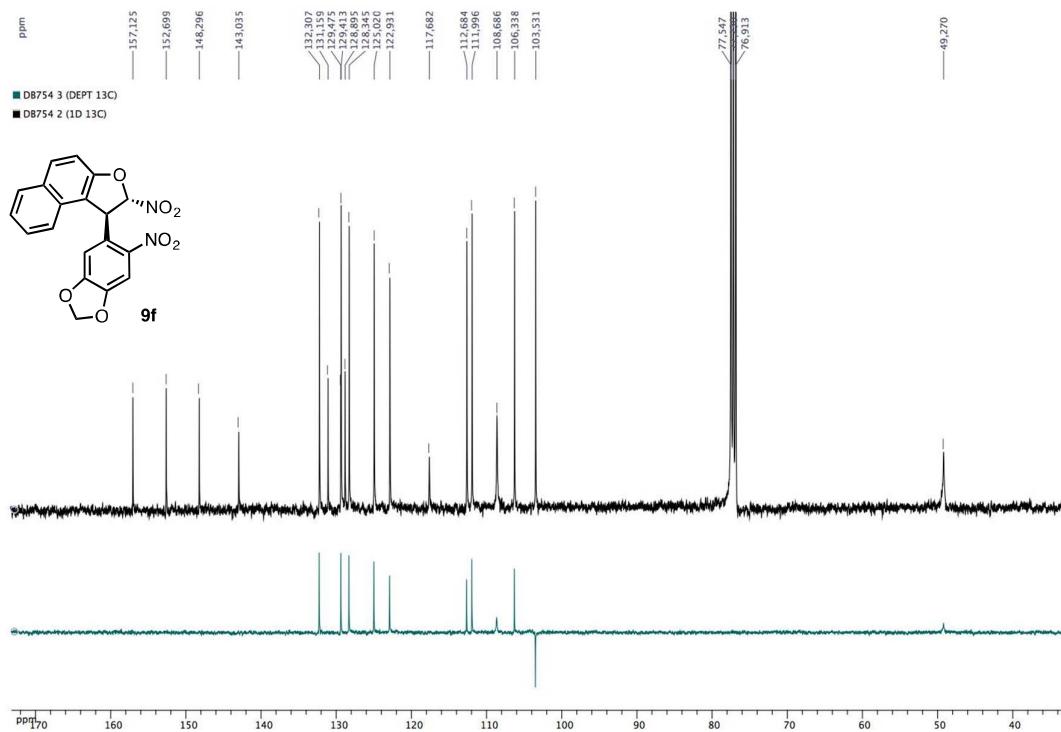
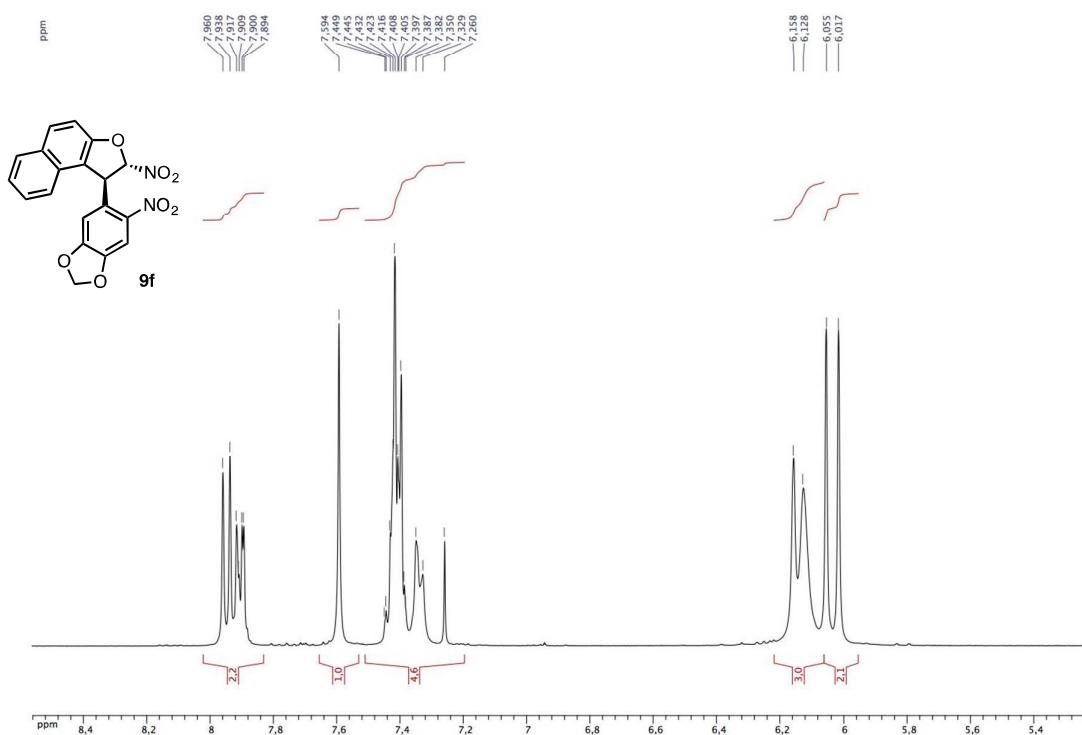


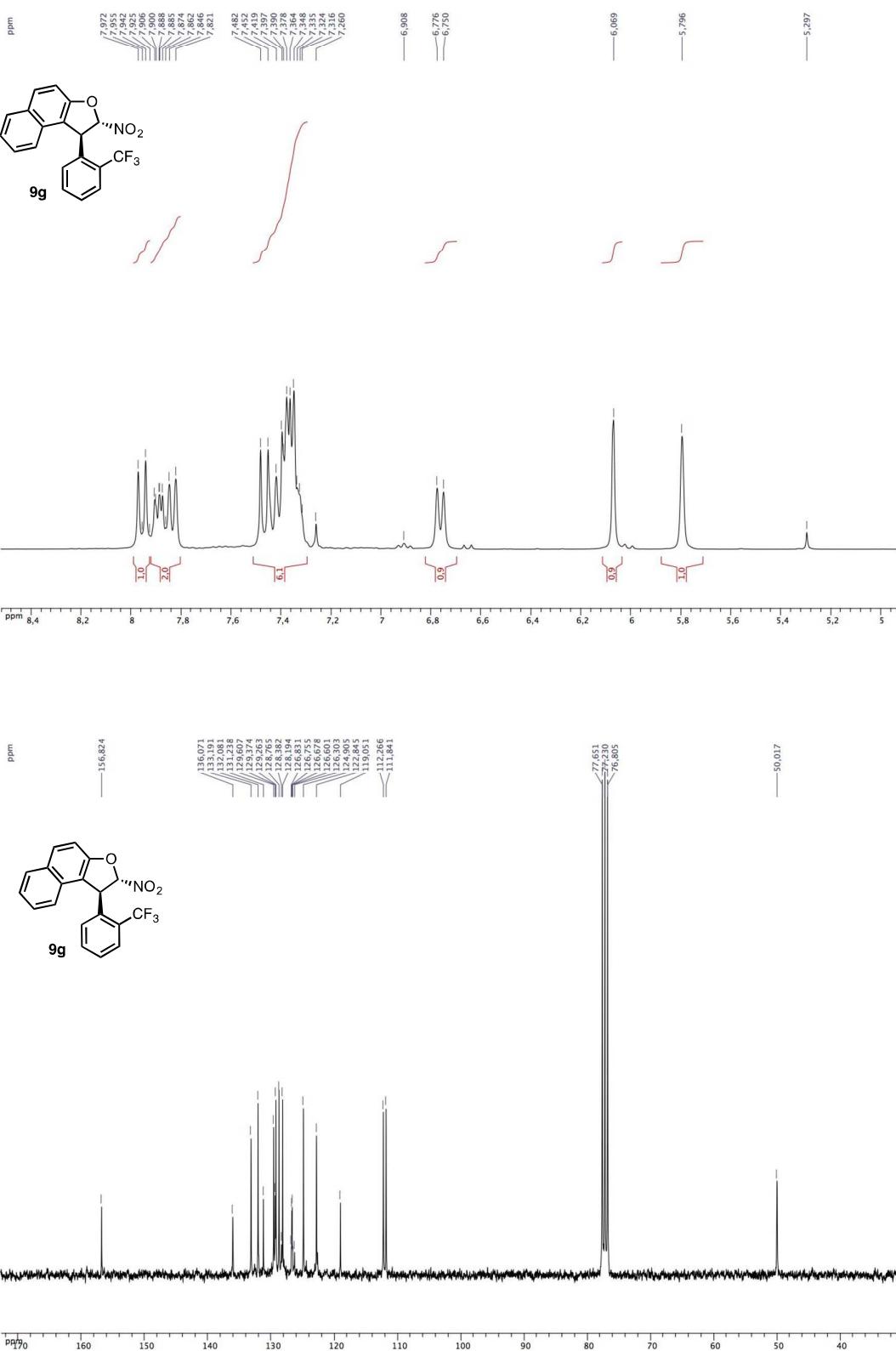


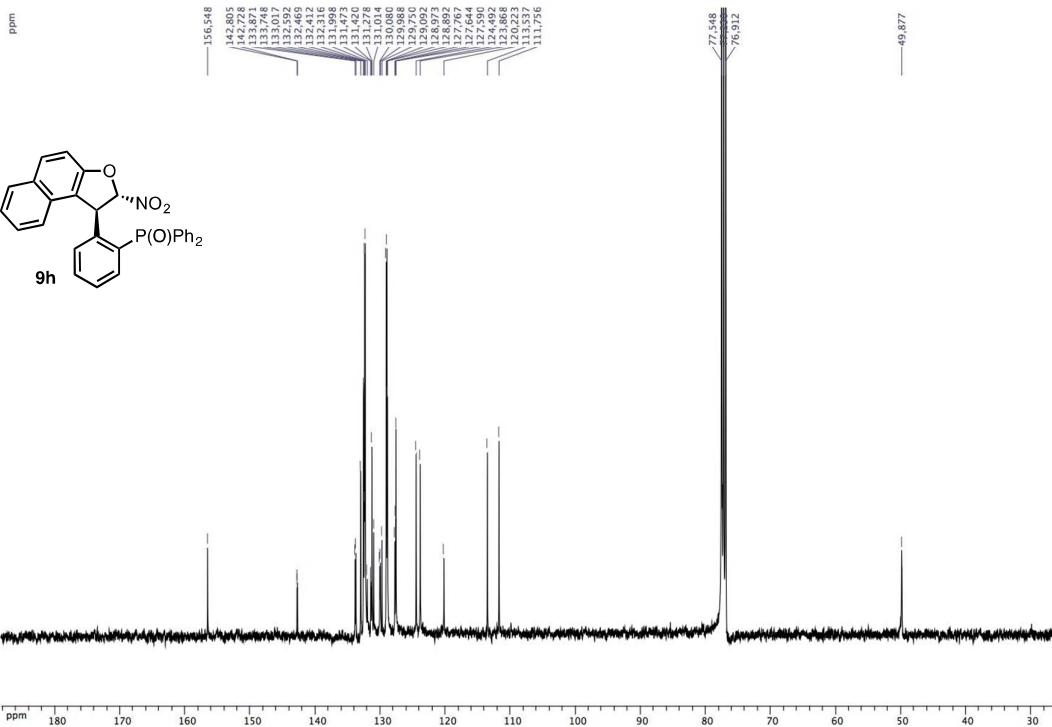
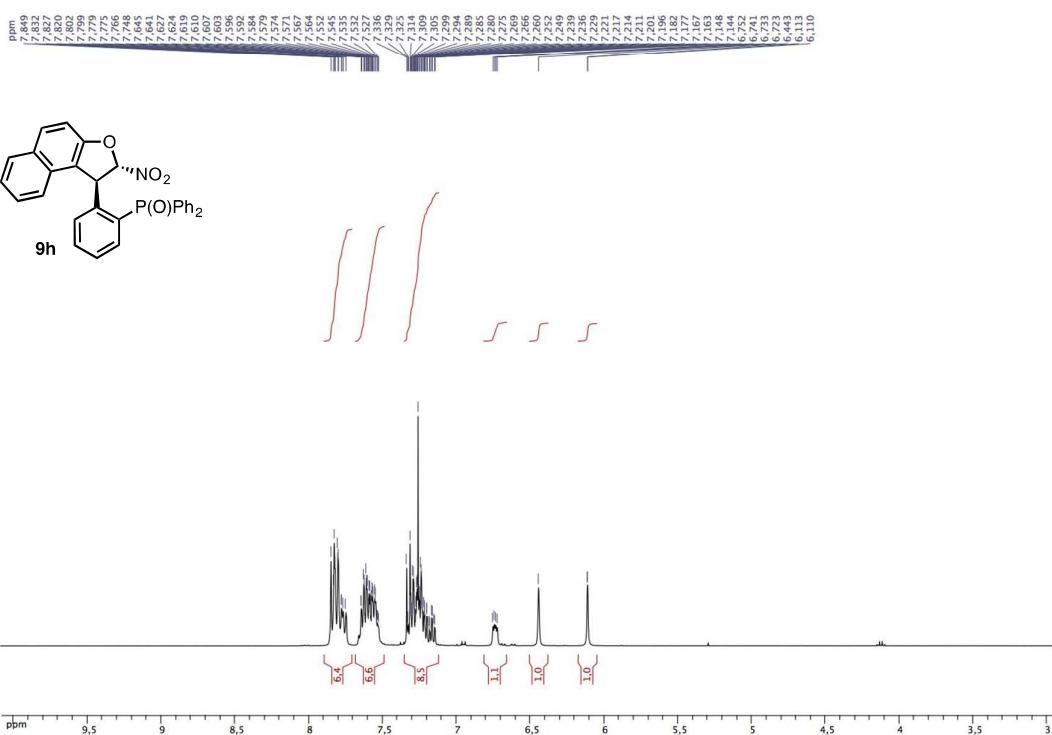


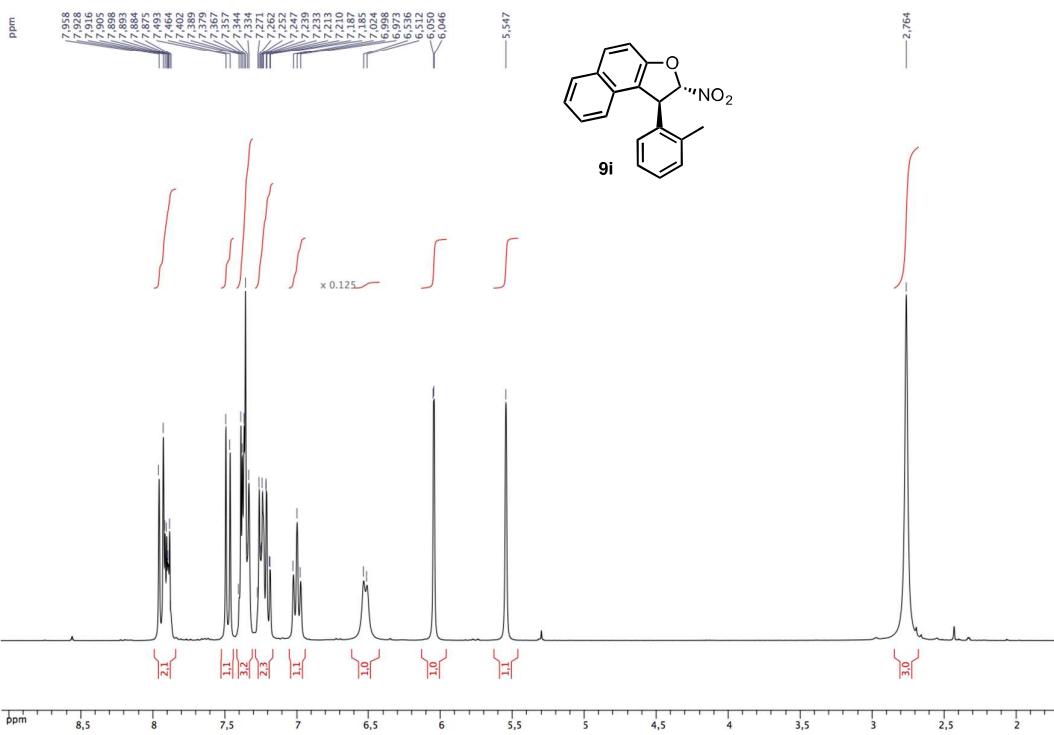
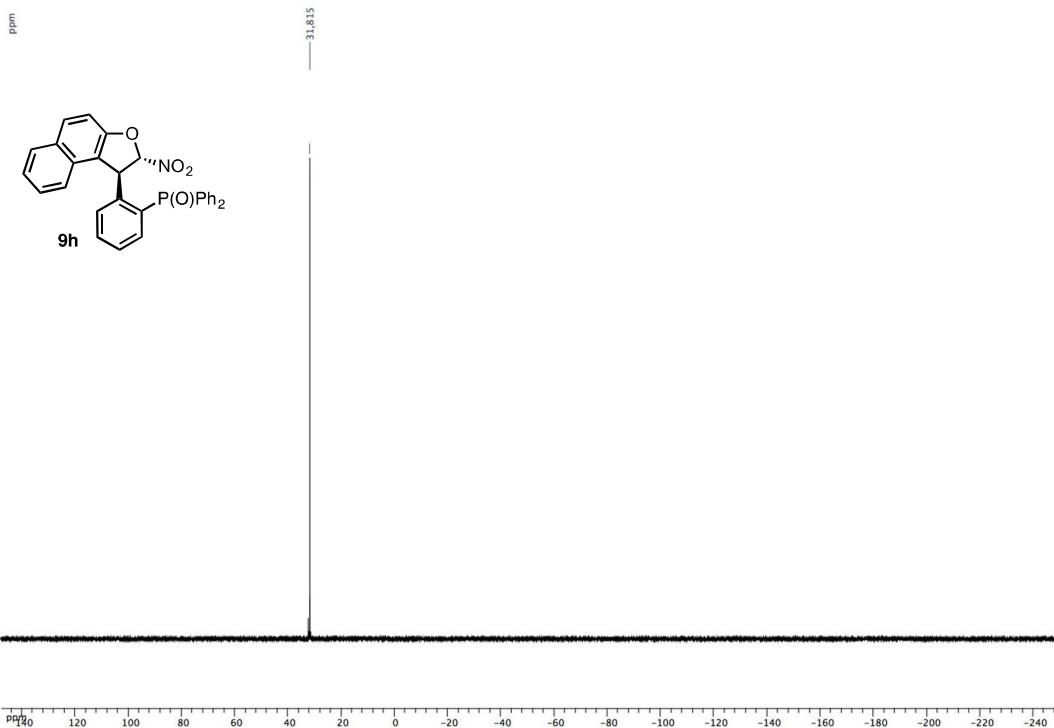


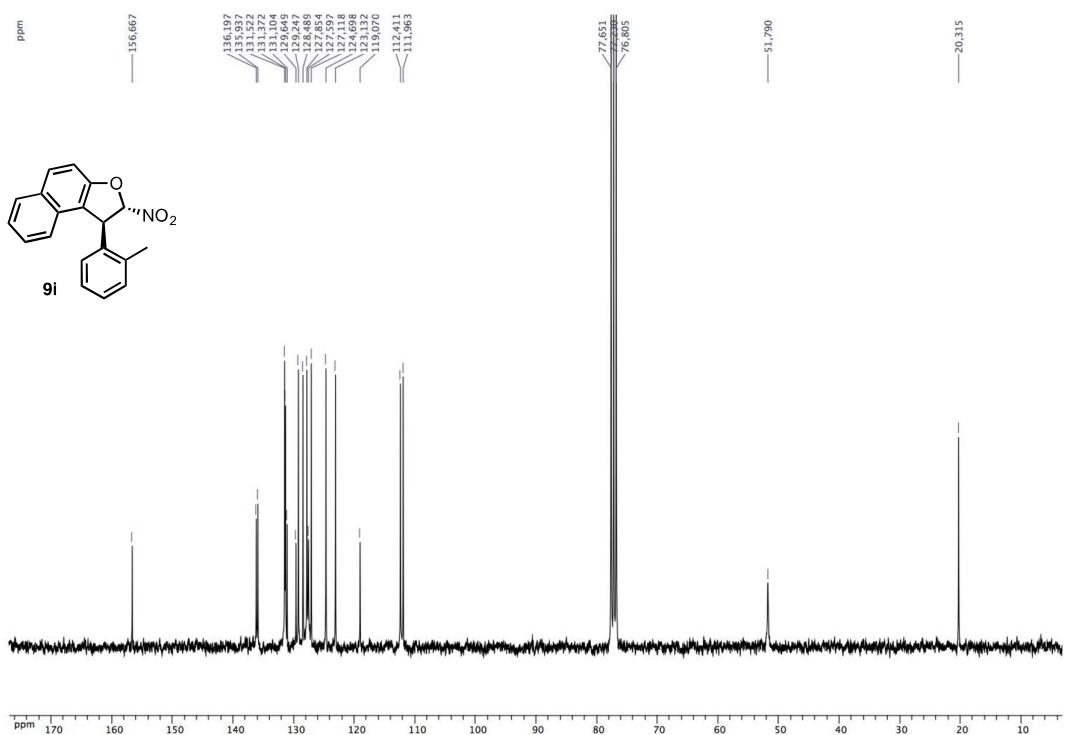


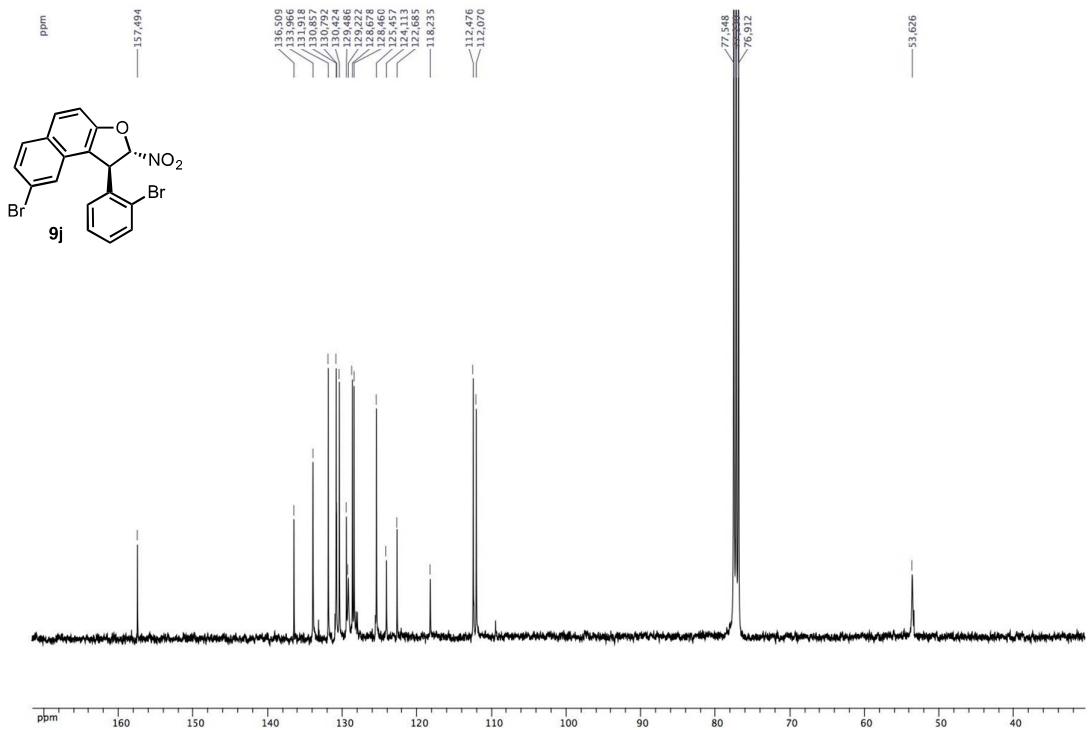
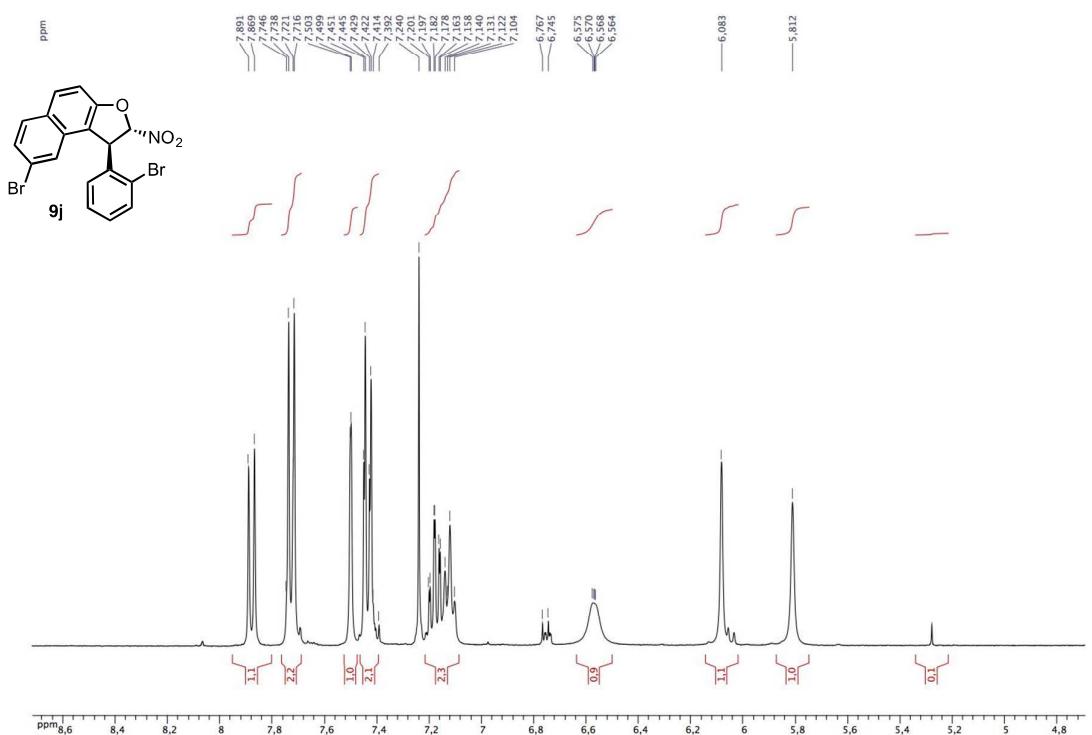


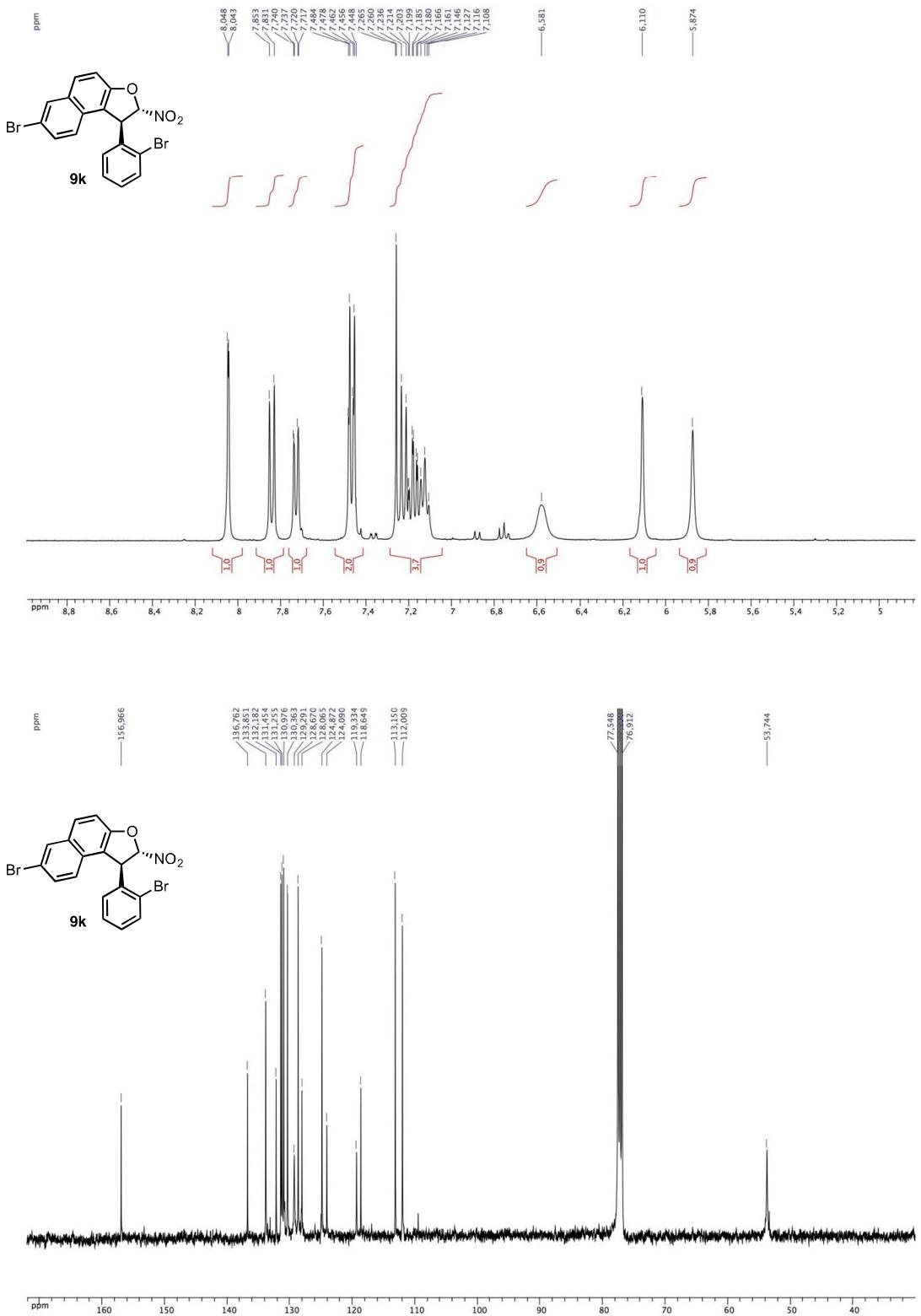


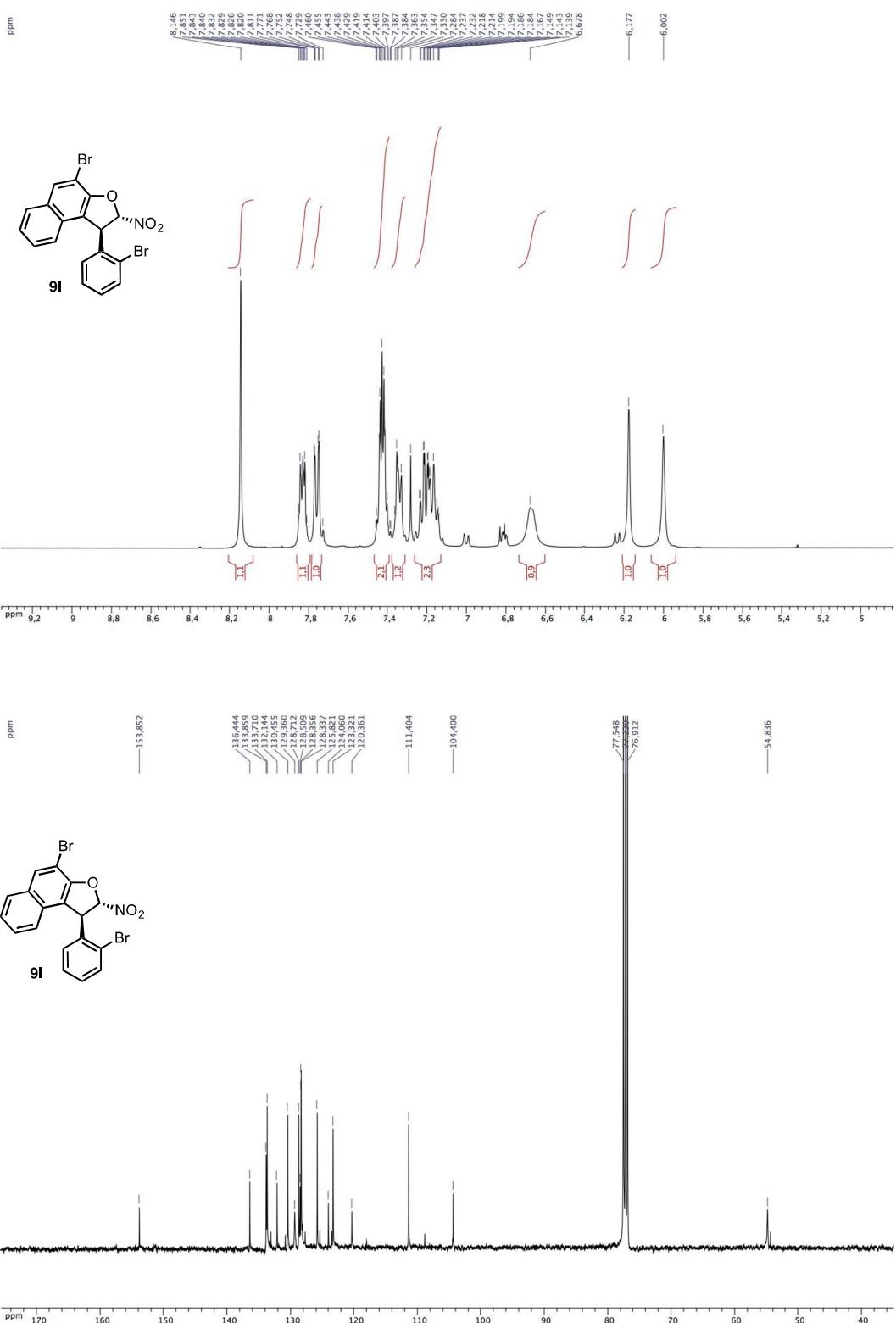


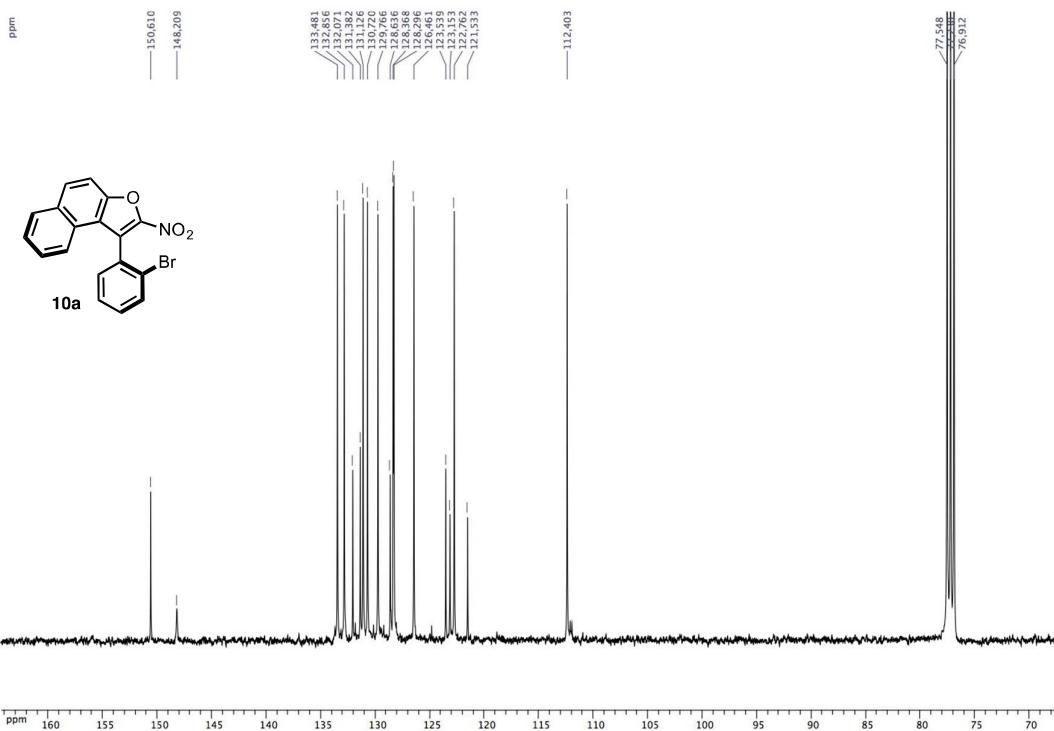
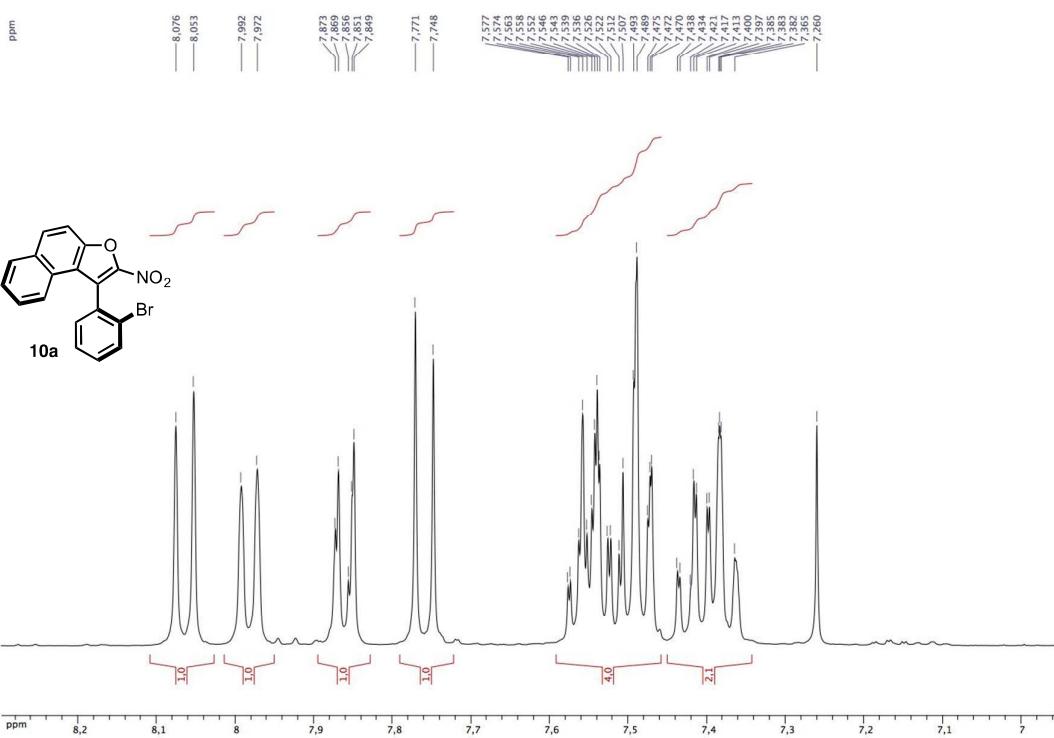


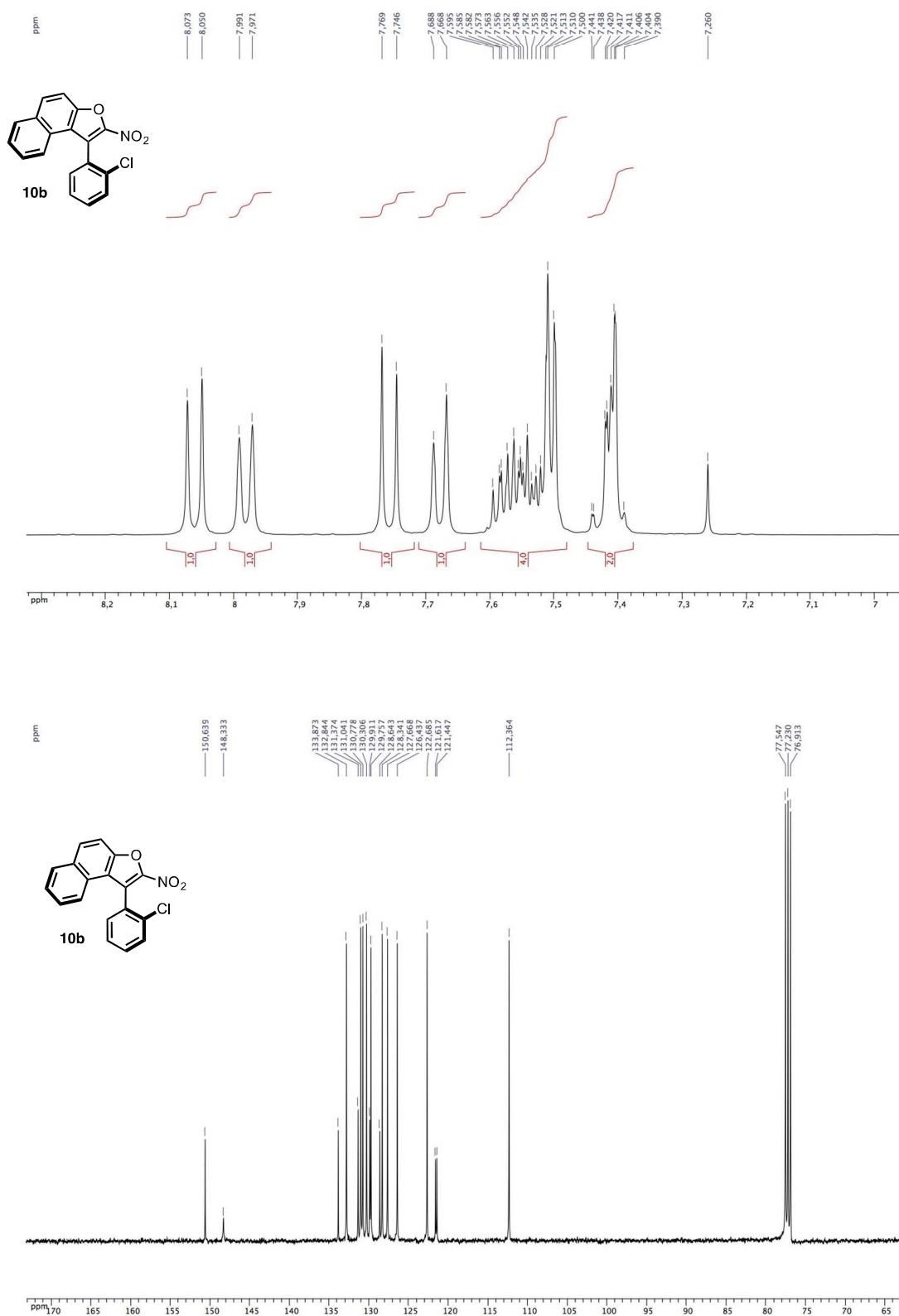


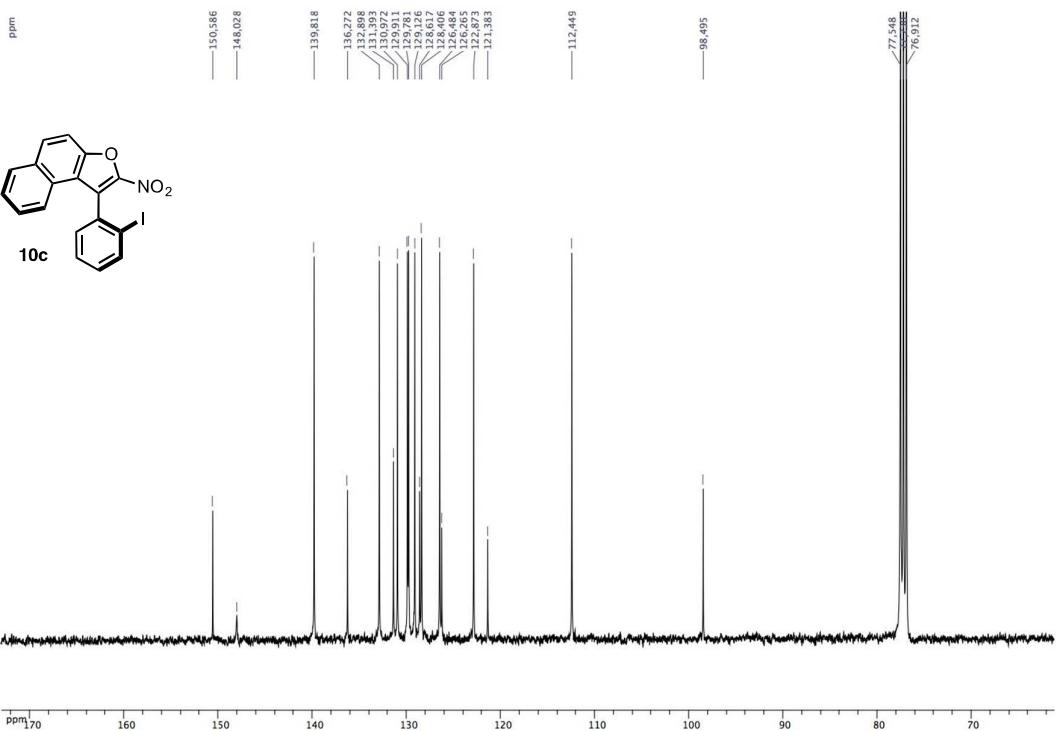
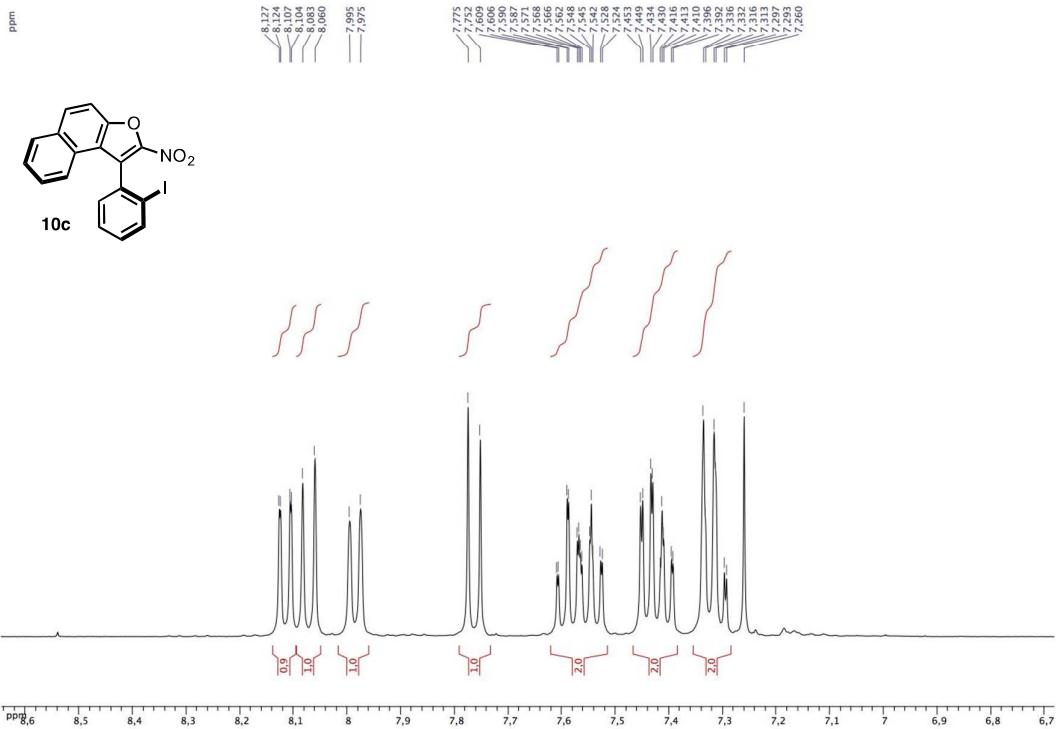


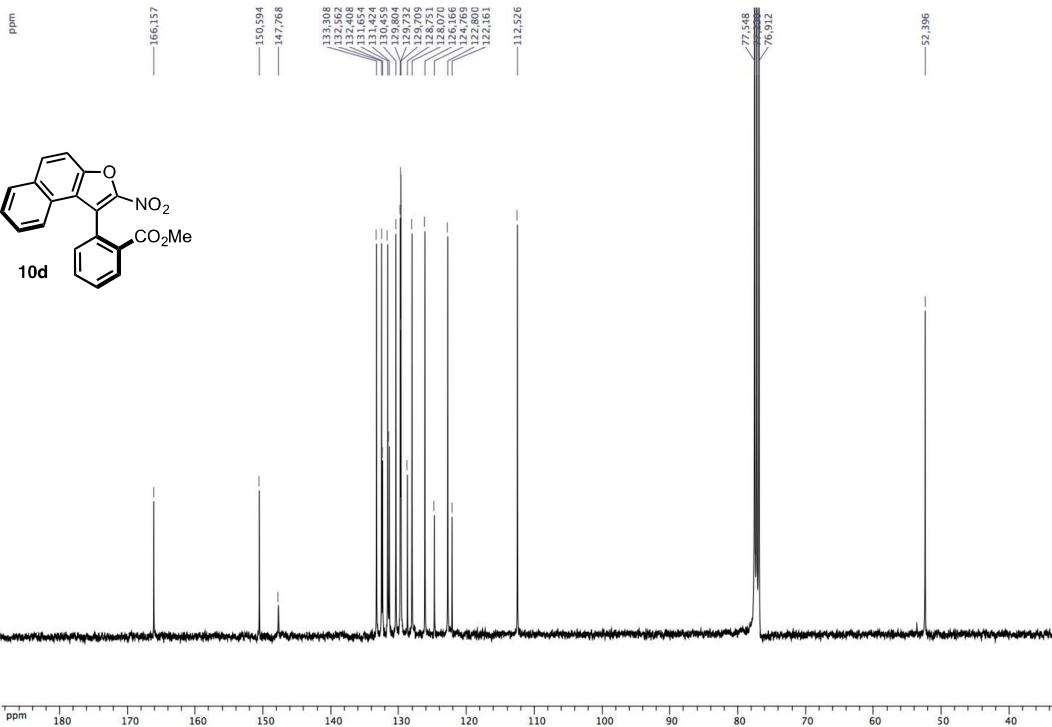
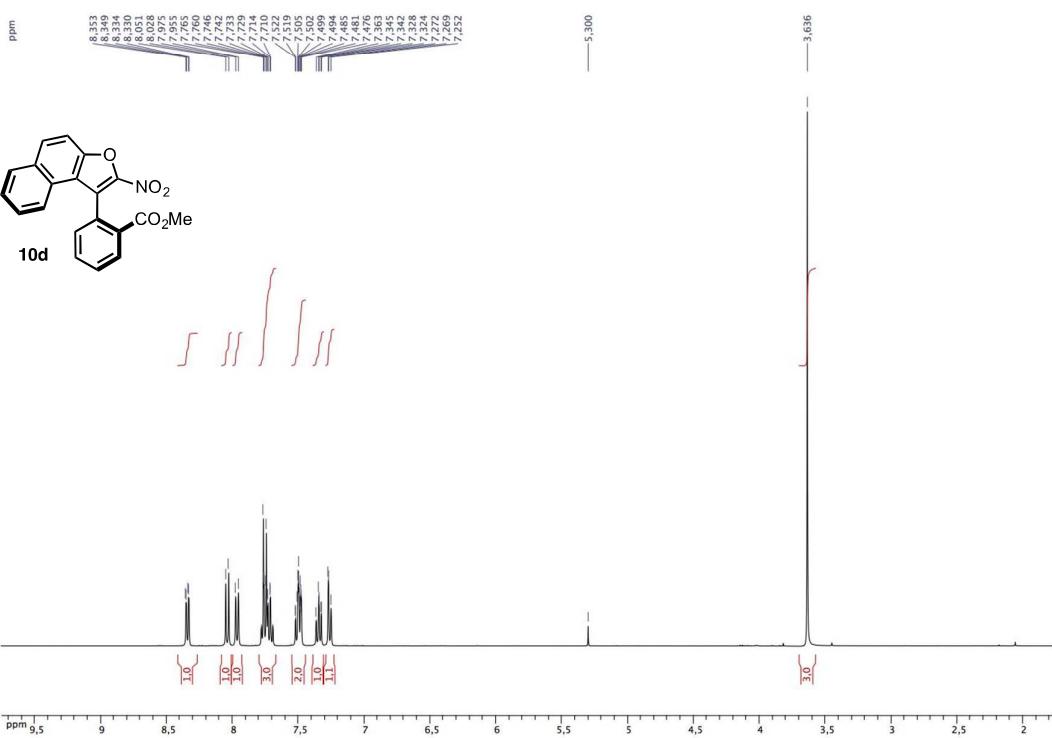


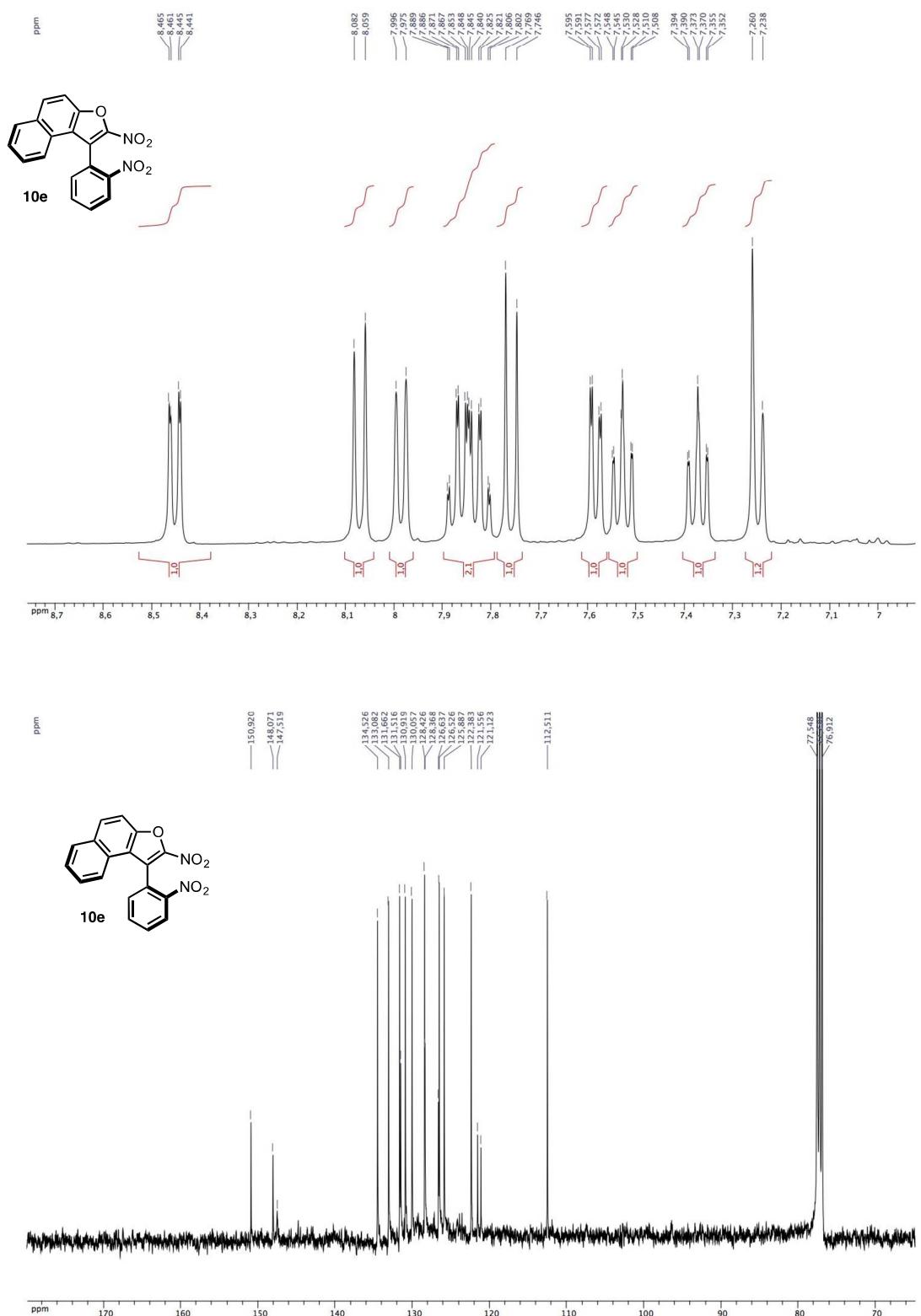


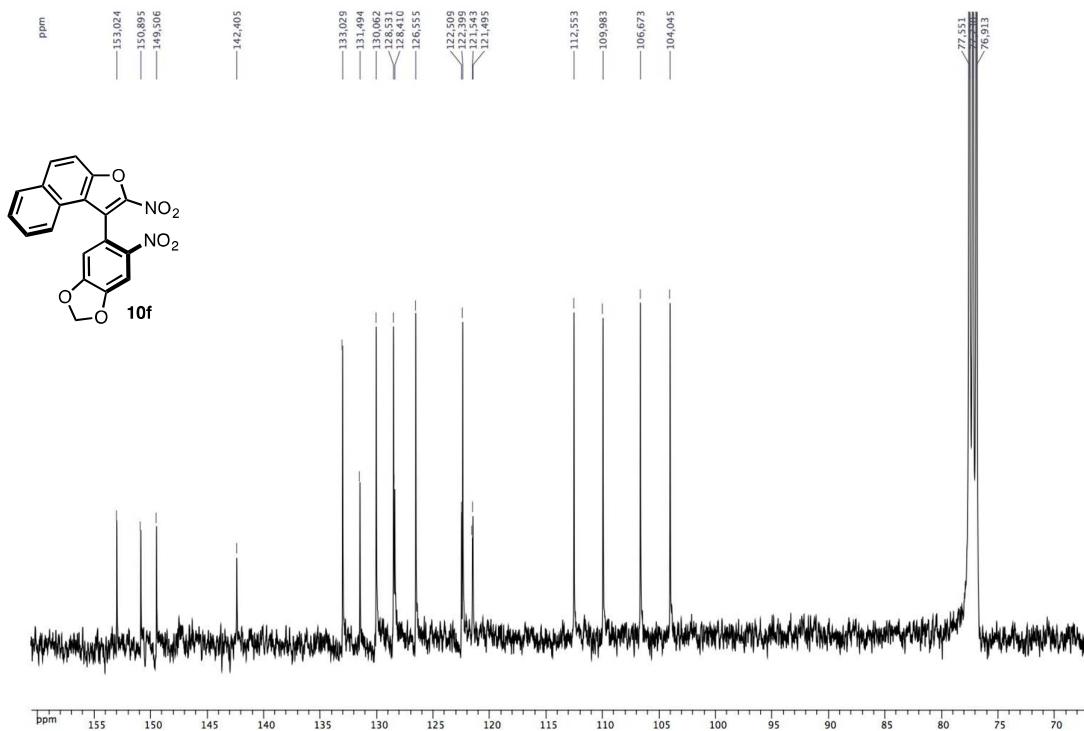
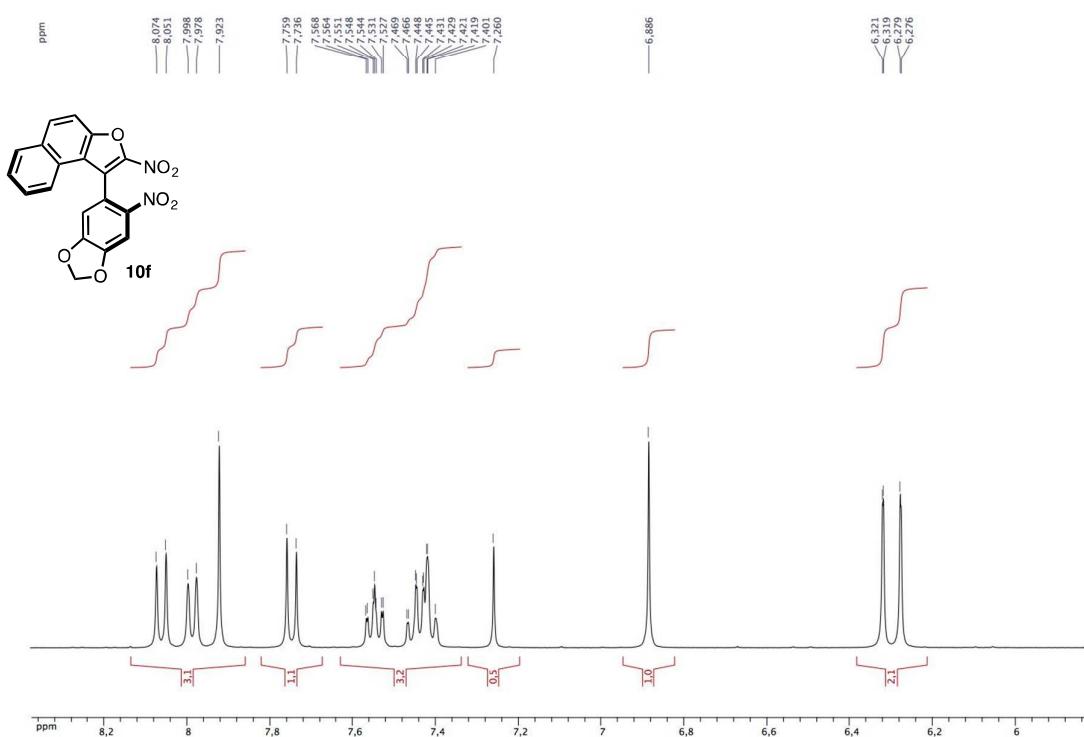


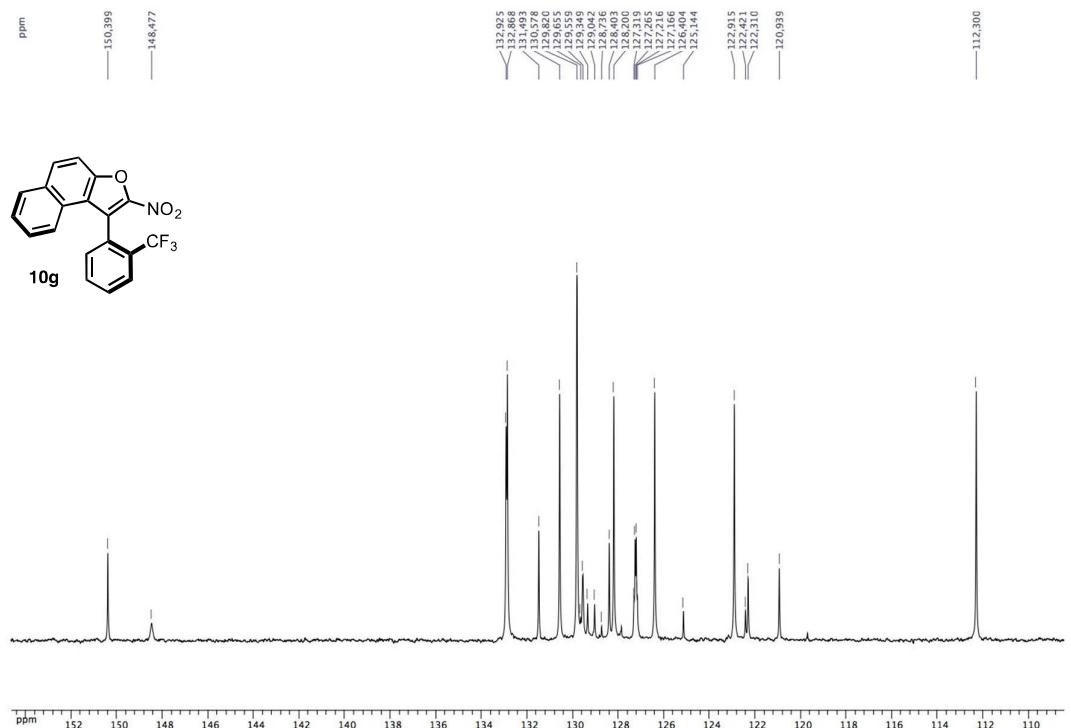
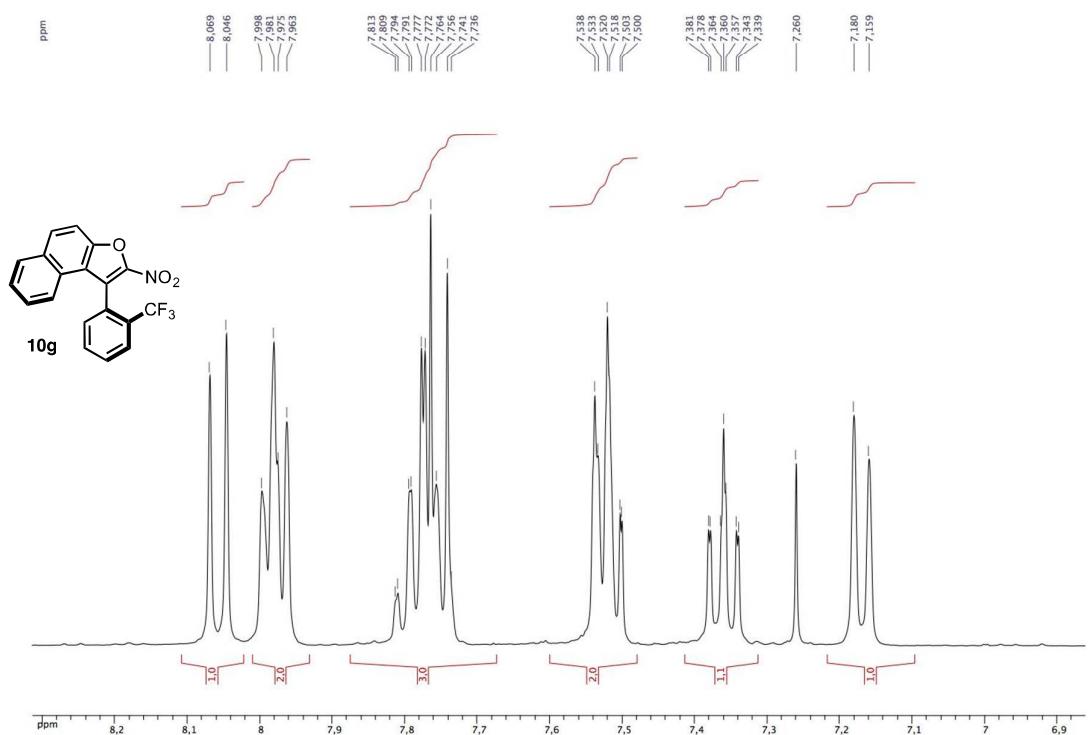


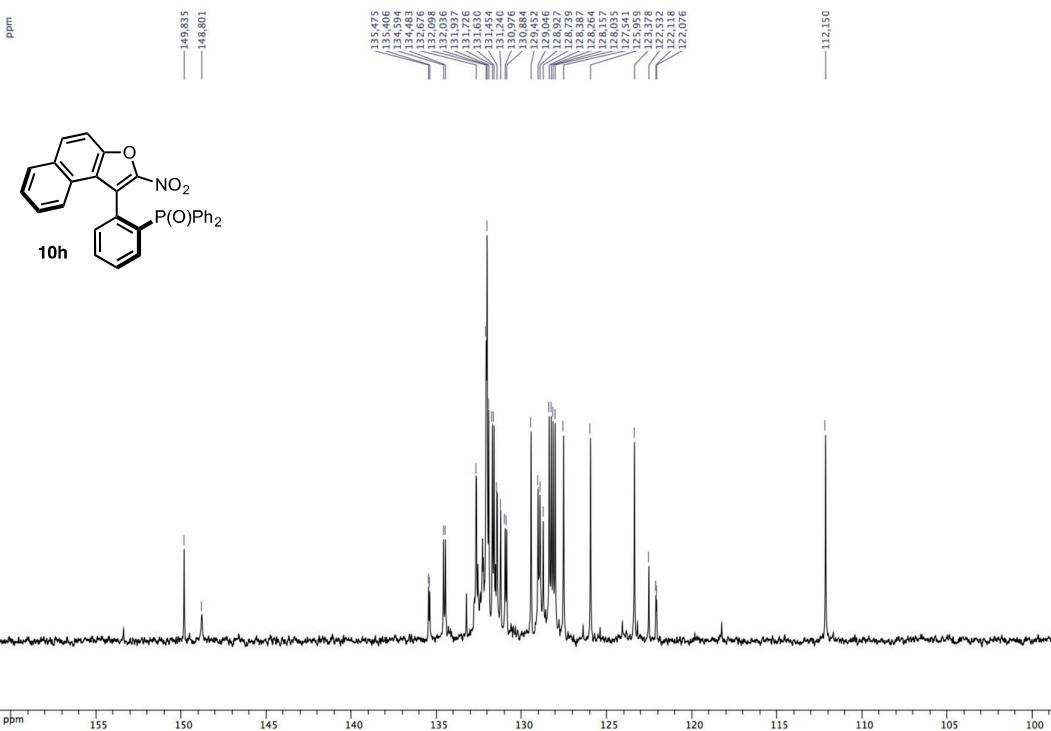
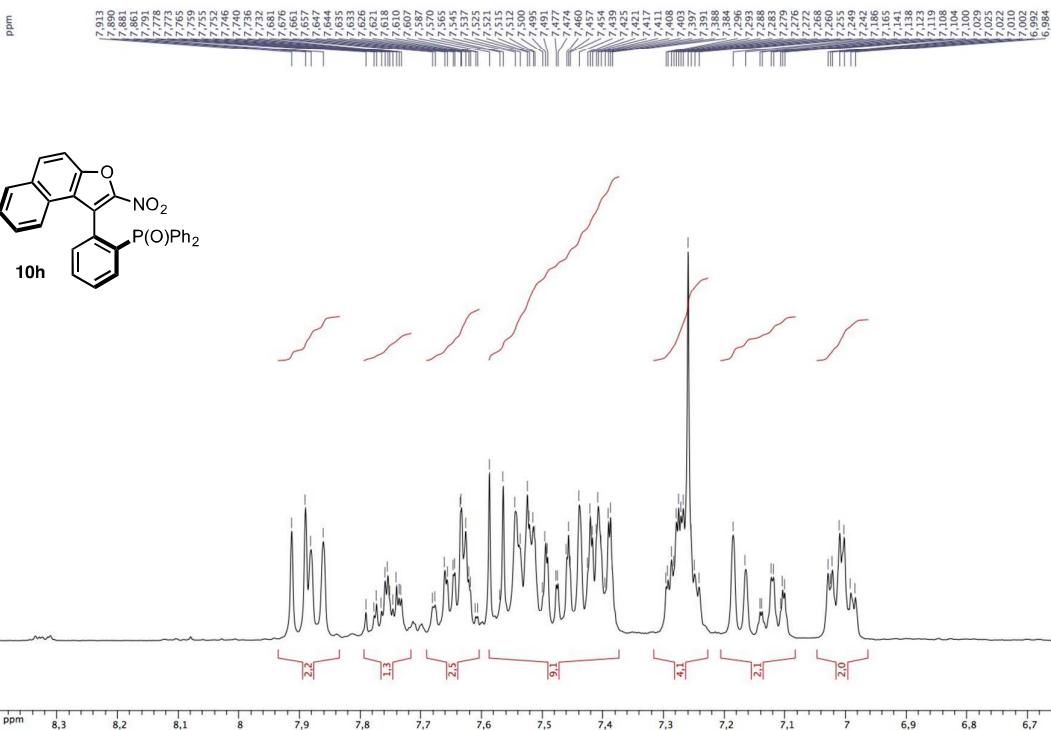


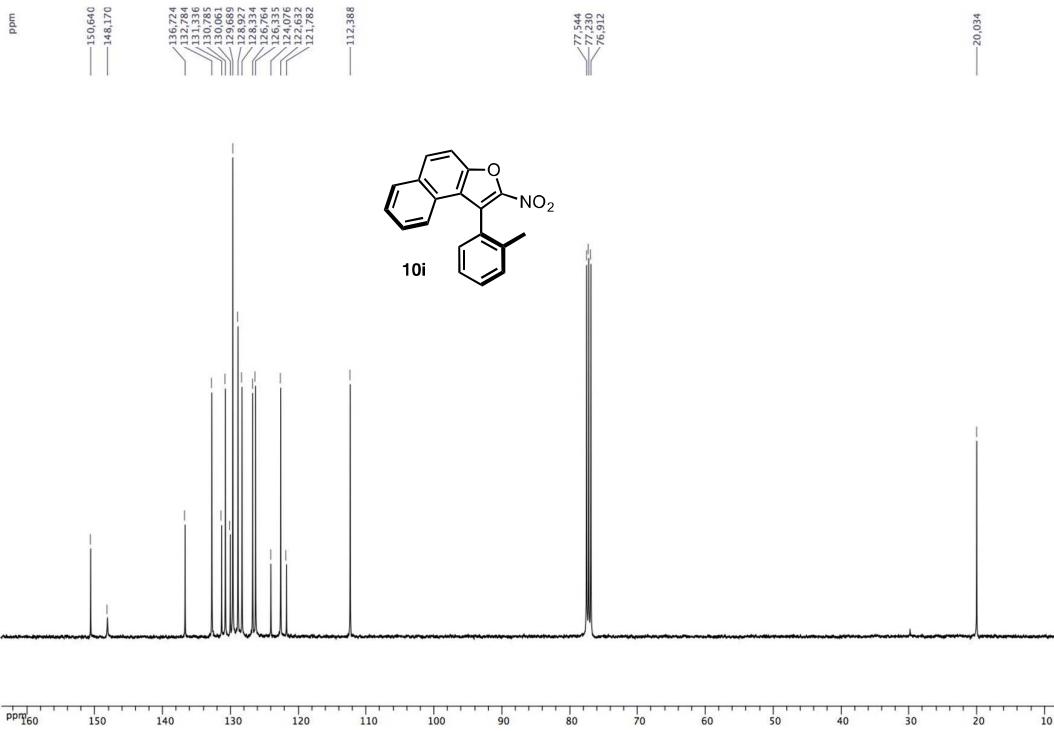
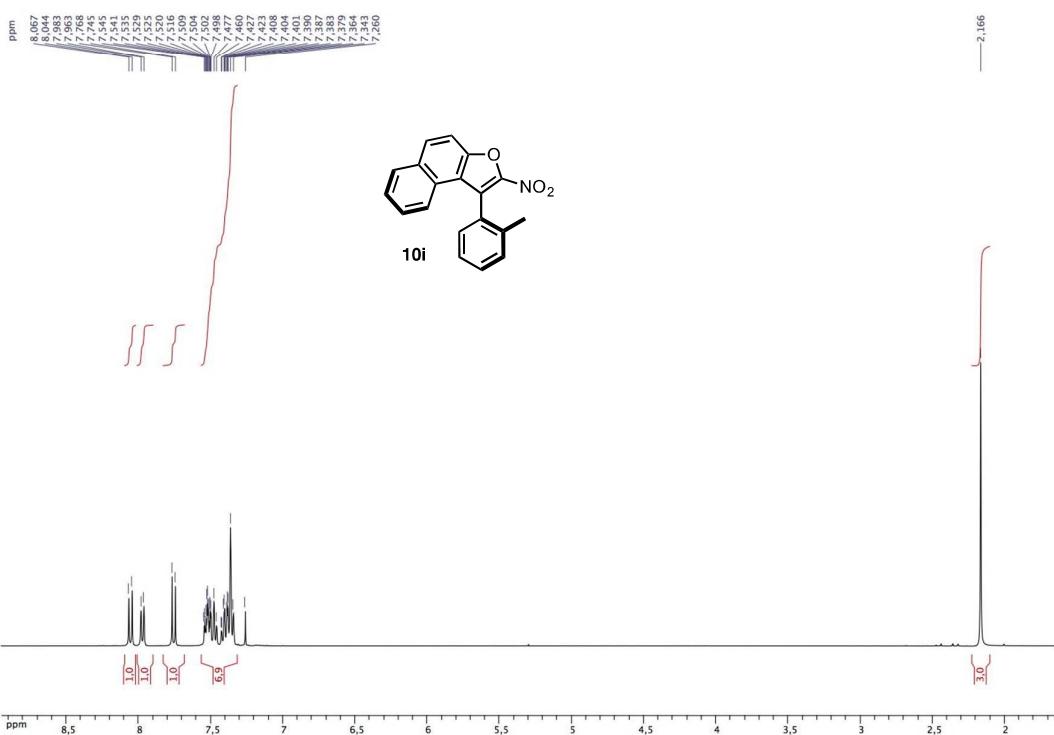


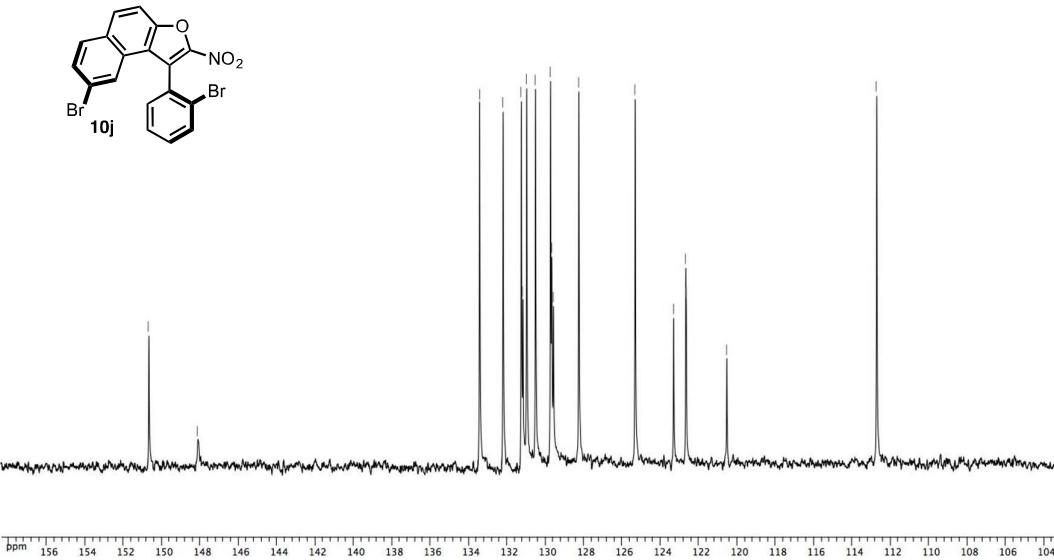
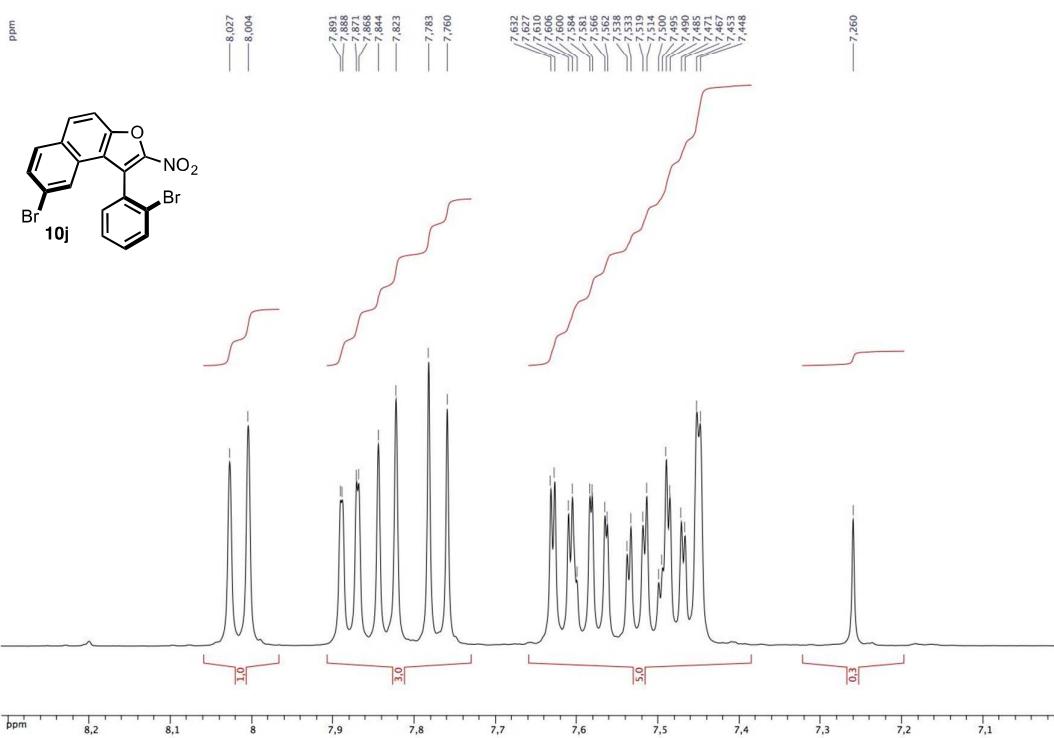


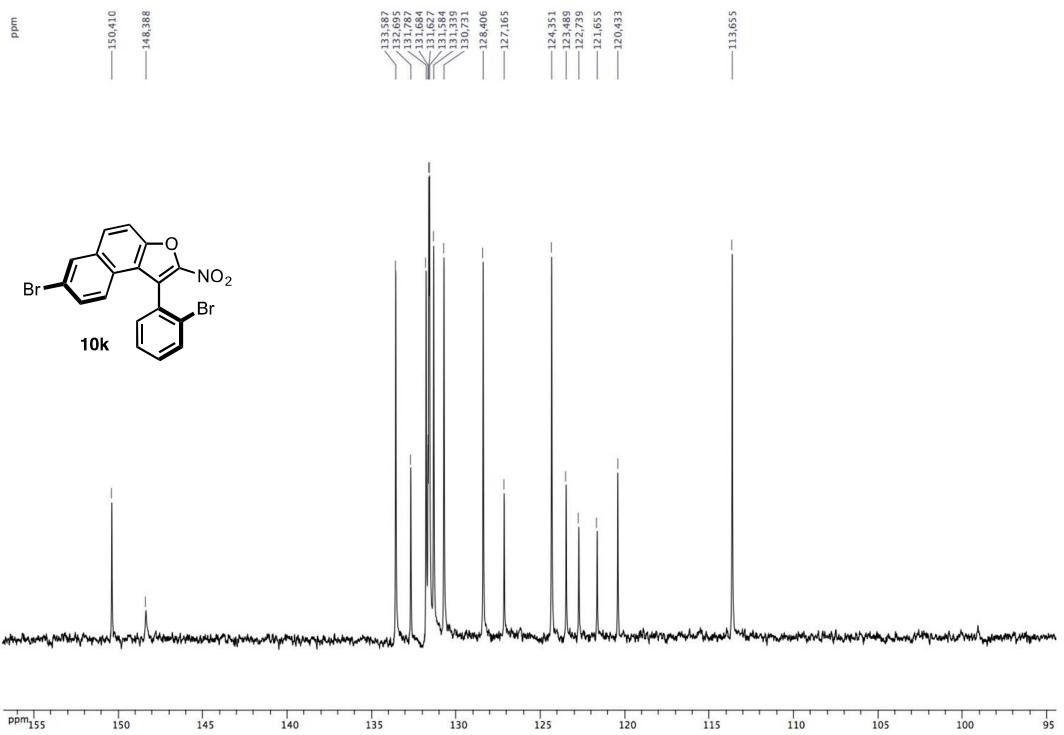
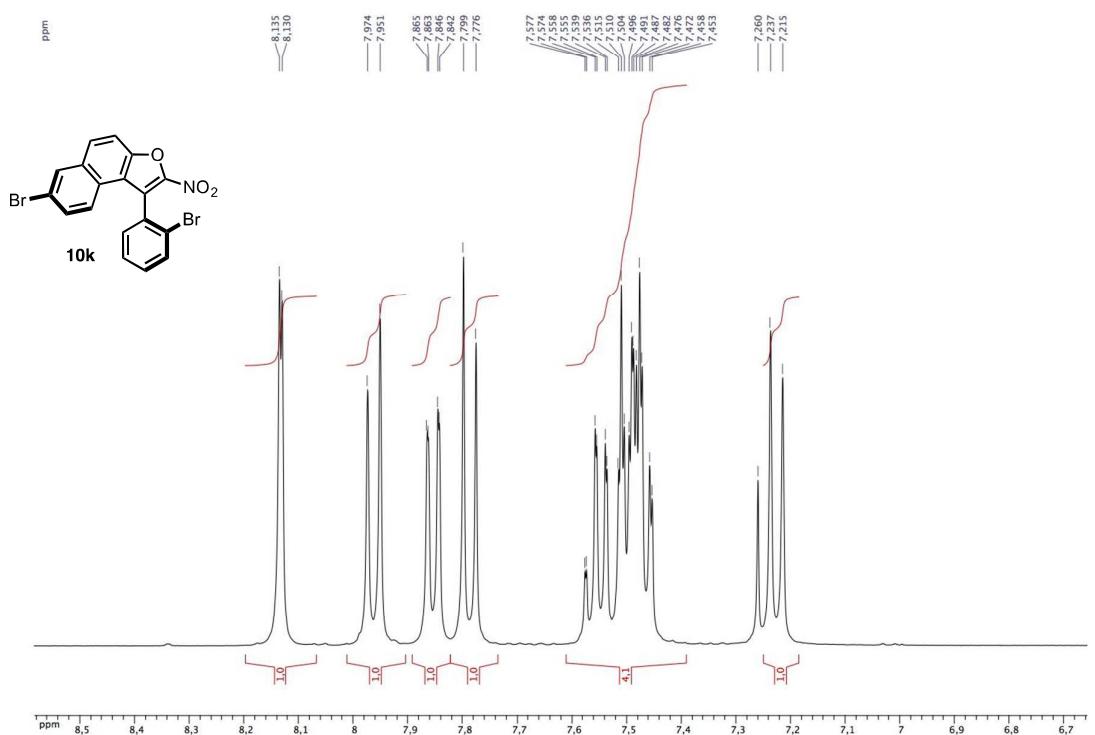


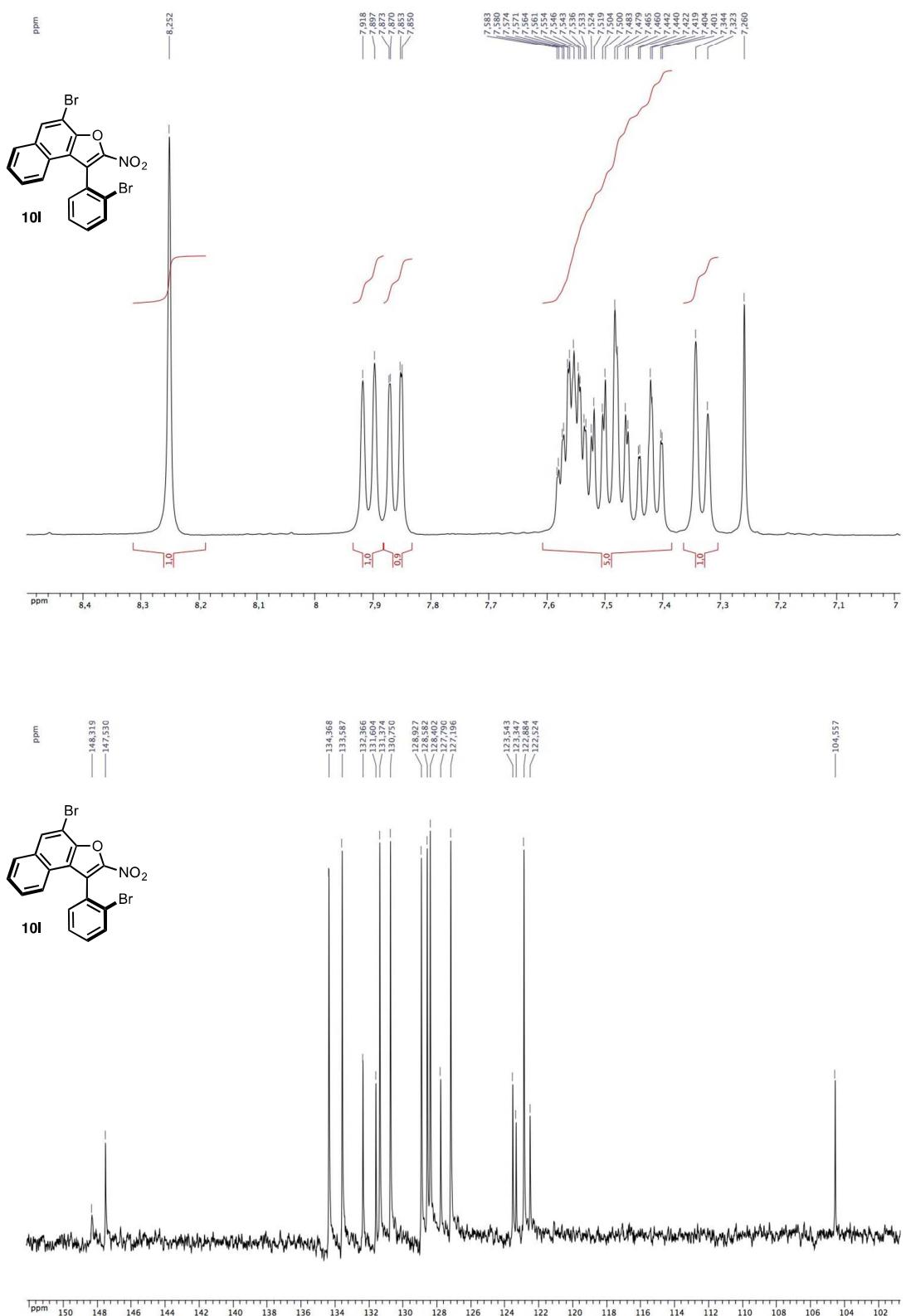


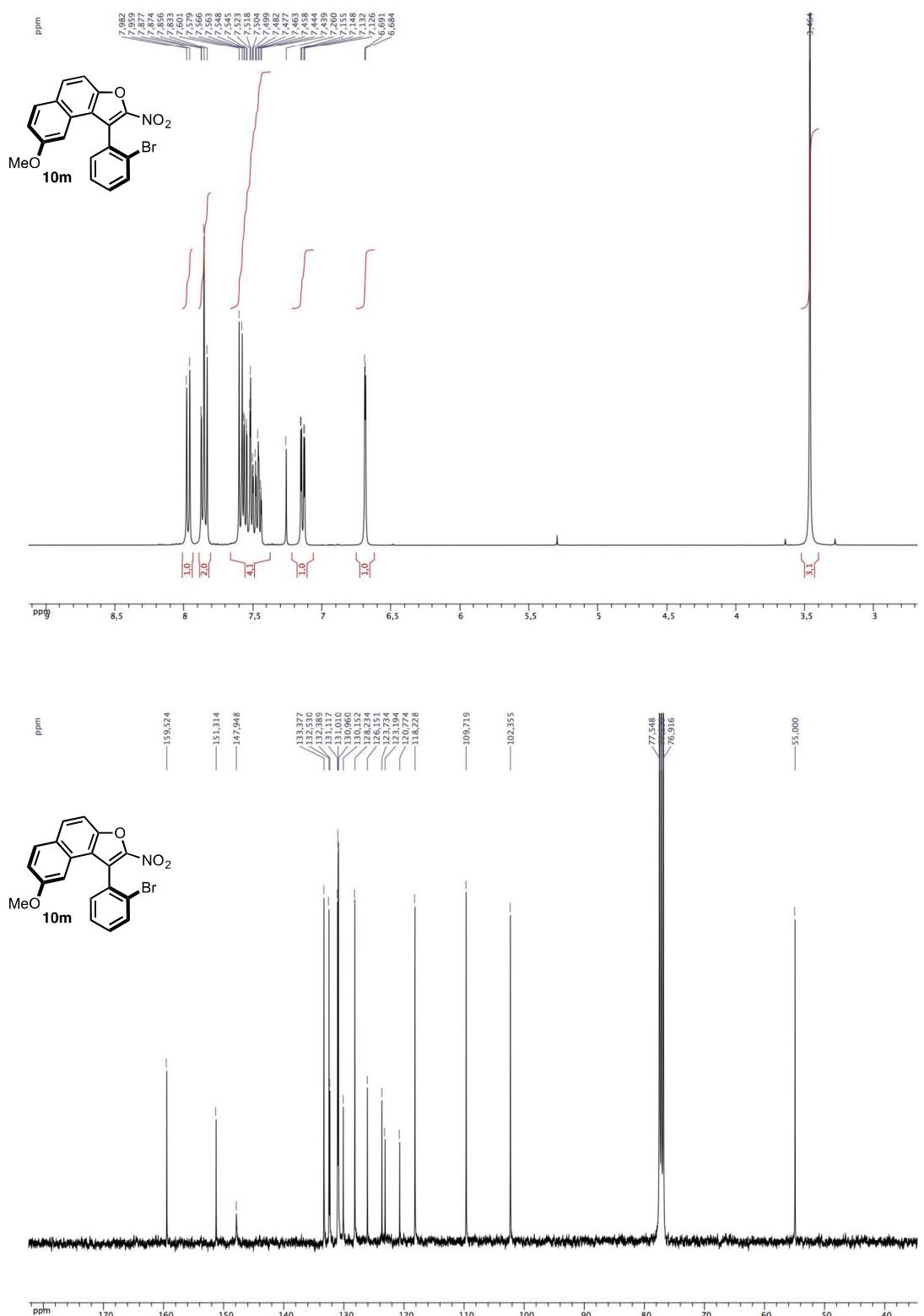












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